Featured Article

Molecular Profiling of Inflammatory Breast Cancer: Identification of a Poor-Prognosis Gene Expression Signature

Ivan Bièche,1,2 Florence Lerebours,1 Sengül Tozlu,1 Marc Espie,3 Michel Marty,4 and Rosette Lidereau1

1Laboratoire d’Oncogénétique - INSERM E0017, Centre René Huguenin, St-Cloud; 2Laboratoire de Génétique Moléculaire - UPRES EA 3618, Faculté des Sciences Pharmaceutiques et Biologiques, Université Paris V, Paris; 3Service d’Oncologie Médicale, Hôpital Saint-Louis, Paris; 4Direction de la Recherche Thérapeutique, Institut Gustave Roussy, Villejuif, France

ABSTRACT

Purpose: Inflammatory breast cancer (IBC) is a rare but particularly aggressive form of primary breast cancer. The molecular mechanisms responsible for IBC are largely unknown.

Experimental Design: To obtain further insight into the molecular pathogenesis of IBC, we used real-time quantitative reverse transcription (RT)-PCR to quantify the mRNA expression of 538 selected genes in IBC relative to non-IBC.

Results: Twenty-seven (5.0%) of the 538 genes were significantly up-regulated in IBC compared with non-IBC. None were down-regulated. The 27 up-regulated genes mainly encoded transcription factors (JUN, EGR1, JUNB, FOS, FOSB, MYCN, and SNAIL1), growth factors (VEGF, DTR/HB-EGF, IGFBP7, IL6, ANGPT2, EREG, CCL3/MIP1α, and CCL5/RANTES) and growth factor receptors (TBX2A/R, TNFRSF10A/TRAILR1, and ROBO2). We also identified a gene expression profile, based on MYCN, EREG, and SHH, which discriminated subgroups of IBC patients with good, intermediate, and poor outcome.

Conclusion: Our study has identified a limited number of signaling pathways that require inappropriate activation for IBC development. Some of the up-regulated genes identified here could offer useful diagnostic or prognostic markers and could form the basis of novel therapeutic strategies.

Received 2/17/04; revised 6/17/04; accepted 6/28/04.

Grant support: Comité Régional des Hauts-de-Seine de la Ligue Nationale Contre le Cancer. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Note: Supplementary data for this article can be found at Clinical Cancer Research Online (http://clincancerres.aacrjournals.org).

Requests for reprints: Ivan Bièche, Laboratoire d’Oncogénétique, Centre René Huguenin, St-Cloud, France, 92210. Phone: 33-1-47-11-15-66; Fax: 33-1-47-11-16-96; E-mail: i.biache@stcloud-huguenin.org.

©2004 American Association for Cancer Research.

INTRODUCTION

Inflammatory breast cancer (IBC) is the most lethal form of primary breast cancer (1). It is characterized clinically by erythema, dimpling of the skin (“peau d’orange”), and rapid onset (2). The 3-year survival rate is about 40% among patients with IBC, compared with 85% among patients with non-IBC (1). Little is known of the molecular mechanisms underlying the genesis and progression of IBC (3). TP53 and ERBB2, genes involved in breast carcinogenesis, are more frequently altered in IBC than in non-IBC (4). A few genetic alterations, such as WISP3/LIBC, ARHC/RhoC, and CDH1/E-cadherin, have been specifically linked to the inflammatory phenotype in this setting (5, 6). Several authors have reported that IBC tends to be highly vascularized (7, 8).

The recent advent of efficient tools for large-scale assessment of gene expression has already provided new insights into the involvement of gene networks and regulatory pathways in various tumoral processes (9). cDNA microarrays can be used to test the expression of thousands of genes at a time, whereas real-time reverse transcription (RT)-PCR is a more accurate and quantitative assay method applicable to smaller numbers of selected candidate genes (10, 11).

To obtain further insight into the molecular pathogenesis of IBC, we used real-time quantitative RT-PCR to quantify the mRNA expression of many selected genes in pooled IBC samples, in comparison with pooled non-IBC samples. We assessed the expression level of 538 genes known to be involved in various cellular and molecular mechanisms associated with tumorigenesis, focusing on genes related to angiogenesis and inflammation. Genes of interest were further investigated in 36 individual IBC samples in comparison with 22 non-IBC samples.

MATERIALS AND METHODS

Patients and Samples

IBC samples were surgical biopsy specimens obtained from 36 women with clinical IBC treated at Saint-Louis Hospital (Paris, France). All tumors were diagnosed on the basis of rapidly progressive signs such as localized or generalized induration, redness, and edema of the breast, and thus classified T4d (International Union Against Cancer classification, 1977). All biopsies were done before treatment, and all confirmed the diagnosis of infiltrating carcinoma. All patients underwent a first-line anthracycline-based high-dose chemotherapy followed by the local treatment. At the time of this analysis, 27 patients had relapsed and nine remained disease-free.

As “non-IBC” controls, we used 22 specimens of non-inflammatory locally advanced breast cancers including 6 stage Iib and 16 stage III. These 22 non-IBC controls were all high-grade invasive ductal carcinomas, i.e., Scarf-Bloom-Richardson histopathological grade III. The expression levels of...
the 538 genes in IBCs were expressed relative to the expression levels in non-IBCs.

The tumor samples were flash-frozen in liquid nitrogen and stored at −80°C until RNA extraction. Tumor samples containing >70% of tumor cells were considered suitable for the study.

Real-Time RT-PCR

Theoretical Basis. Reactions are characterized by the point during cycling when amplification of the PCR product is first detected, rather than the amount of PCR product accumulated after a fixed number of cycles. The larger the starting quantity of the target molecule, the earlier a substantial increase in fluorescence is observed. The parameter threshold cycle (Ct) is defined as the fractional cycle number at which the fluorescence generated by cleavage of a TaqMan probe (or by SYBR green dye-amplicon complex formation) passes a fixed threshold above baseline. The increase in fluorescent signal associated with exponential growth of PCR products is detected by the laser detector of the ABI Prism 7700 Sequence Detection System (Perkin-Elmer Applied Biosystems, Foster City, CA), using PE Biosystems analysis software according to the manufacturer’s manuals.

The precise amount of total RNA added to each reaction mix (based on absorbance) and its quality (i.e., lack of extensive degradation) are both difficult to assess. We therefore also quantified transcripts of two endogenous RNA control genes involved in two cellular metabolic pathways, namely TBP (GenBank accession NM_003194), which encodes the TATA box-binding protein (a component of the DNA-binding protein complex TFIIID), and RPLP0 (NM_001002), which encodes human acidic ribosomal phosphoprotein P0. Each sample was normalized on the basis of its TBP (or RPLP0) content.

We selected TBP as an endogenous control because the prevalence of its transcripts is moderate and because there are no known TBP retropseudogenes (retropseudogenes lead to co-amplification of contaminating genomic DNA and thus interfere with RT-PCR, despite the use of primers in separate exons). We also selected RPLP0 because the prevalence of its transcripts is high as compared with TBP and because this gene is used widely as an endogenous control for Northern blot analysis (known better as 36B4).

Results, expressed as N-fold differences in target gene expression relative to the TBP (or RPLP0) gene, and termed "Ntarget," were determined as Ntarget = 2ΔCt sample, where the ΔCt value of the sample was determined by subtracting the average Ct value of the target gene from the average Ct value of the TBP (or RPLP0) gene (12, 13).

The Ntarget values of the samples were subsequently normalized such that the median of the non-IBC Ntarget values was 1.

Primers and Controls. Primers for TBP, RPLP0, and the 538 target genes (list in Supplemental data) were chosen with the assistance of the Oligo 5.0 computer program (National Biosciences, Plymouth, MN).

We conducted searches in database of Expressed Sequence Tags (dbEST), high-throughput genomic sequence (htgs) database, and protein (nr) database to confirm the total gene specificity of the nucleotide sequences chosen as primers and the absence of single nucleotide polymorphisms. In particular, the primer pairs were selected to be unique relative to the sequences of closely related family member genes or of the corresponding retropseudogenes. To avoid amplification of contaminating genomic DNA, one of the two primers was placed at the junction between two exons, if possible. In general, amplicons were between 70 and 120 nucleotides long. Gel electrophoresis was used to verify the specificity of PCR amplicons.

For each primer pair, we performed no-template control and no-reverse-transcriptase control assays, which produced negligible signals (usually >40 in Ct value), suggesting that primer-dimer formation and genomic DNA contamination effects were negligible.

RNA Extraction. Total RNA was extracted from frozen tumor samples with the acid-phenol guanidinium method. The quality of the RNA samples was determined by electrophoresis through agarose gels and staining with ethidium bromide, the 18S and 28S RNA bands being visualized under UV light.

cDNA Synthesis and PCR Reaction Conditions cDNA Synthesis and PCR reaction conditions have been described previously (13).

Statistical Analysis

Because the mRNA levels did not fit a Gaussian distribution, (a) the mRNA levels in each subgroup of samples were characterized by their median values and ranges, rather than their mean values and coefficients of variation, and (b) relationships between the molecular markers and clinical and histologic parameters were tested using the nonparametric Kruskal-Wallis (14) or Mann-Whitney U test (15).

To visualize the capacity of a given molecular marker to discriminate between two populations (in the absence of an arbitrary cutoff value), we summarized the data in a receiver operating characteristic (ROC) curve (16). The area under curve (AUC) was calculated as a single measure for the discriminatory capacity of each molecular marker. When a molecular marker has no discriminatory value, the ROC curve lies close to the diagonal and the AUC is close to 0.5. In contrast, when a molecular marker has strong discriminatory value, the ROC curve moves to the upper left-hand corner, and the AUC is close to 1.

Hierarchical clustering was done with the GenANOVA software (17).

RESULTS

We first determined the mRNA expression level of the 538 genes in an IBC pool and a non-IBC pool. Genes with an expression in the IBC pool differing (>2-fold) from that in the non-IBC pool were then examined for their mRNA expression in 36 individual IBCs relative to 22 non-IBCs. Gene expression levels in the IBCs were expressed relative to the corresponding levels in the non-IBCs.

Expression of the 538 Genes in the IBC and Non-IBC Pools. The IBC and non-IBC pools were each prepared by mixing identical amounts of tumor RNA from eight patients. The mean TBP gene Ct values for the eight tumor samples were 25.81 ± 0.33 (IBC pool) and 25.64 ± 0.37 (non-IBC pool).

Very low levels of target gene mRNA that were only detectable but not reliably quantifiable by real-time quantitative
RT-PCR assays, mainly based on fluorescence SYBR green methodology (C_r > 32), were observed for 47 (8.7%) of the 538 genes in both the IBC and non-IBC pools.

Forty-eight (9.6%) of the 491 remaining genes were expressed at a different level (>2-fold) in the IBC pool compared with the non-IBC pool; 40 (83.3%) were up-regulated, and 8 (16.7%) were down-regulated.

mRNA Expression of ESR1/ERα, MKI67, and 48 Candidate Genes in 36 IBCs and 22 Non-IBCs. The expression level of the 48 dysregulated genes identified by pooled sample analysis was then determined individually in 36 IBCs and 22 non-IBCs. Twenty-seven (67.5%) of the 40 up-regulated genes identified by the pooled sample analysis were significantly up-regulated in the 36 individual IBCs relative to the 22 non-IBCs (P < 0.05; Table 1). None of the eight down-regulated genes identified by the pooled sample analysis was significantly down-regulated in the individual IBCs.

The 27 up-regulated genes mainly encoded transcription factors (JUN, EGR1, JUNB, FOS, FOSB, MYCN, and SNAIL1), growth factors (VEGF, DTR/HB-EGF, IGFBP7, IL6, ANGPT2, EREG, CCL3/MIP1A, and CCL5/RANTES), and growth factor receptors (TBX2R, TNFRSF10A/TRAILR1, and ROBO2).

ROC curve analysis was then used to test the capacity of each of these 27 genes to discriminate between IBC and non-IBC. The overall diagnostic values of the 27 molecular markers was assessed in terms of their AUC values (Table 1). Fig. 1 shows the mRNA levels of the three most discriminatory genes, namely JUN (AUC-ROC, 0.908), EGR1 (AUC-ROC, 0.896), and DUSP1 (AUC-ROC, 0.896), in each non-IBC and IBC sample.

In the same set of 58 tumors, we also examined the expression of the ESR1/ERα and ERBB2 genes and that of the proliferation-associated gene MKI67 gene that encodes the proliferation-related antigen Ki-67. MKI67 shows a trend toward

### Table 1. List of the significantly dysregulated genes in IBCs relative to non-IBCs

<table>
<thead>
<tr>
<th>Genes</th>
<th>Gene definition</th>
<th>Gene characterisation</th>
<th>Non-IBC (n = 22)</th>
<th>IBC (n = 36)</th>
<th>P*</th>
<th>ROC-AUC</th>
</tr>
</thead>
<tbody>
<tr>
<td>JUN</td>
<td>Jun oncogene</td>
<td>Transcription factor</td>
<td>1.0 (0.2–10.4)†</td>
<td>8.3 (0.8–230.7)</td>
<td>&lt;10^-6</td>
<td>0.908</td>
</tr>
<tr>
<td>EGR1</td>
<td>Early growth response 1</td>
<td>Transcription factor</td>
<td>1.0 (0.1–5.2)</td>
<td>13.8 (0.3–625.1)</td>
<td>&lt;10^-6</td>
<td>0.896</td>
</tr>
<tr>
<td>DUSP1</td>
<td>Dual specificity phosphatase 1 (CL100, MKP-1)</td>
<td>Signaling transduction</td>
<td>1.0 (0.2–4.8)</td>
<td>13.7 (0.6–303.0)</td>
<td>&lt;10^-6</td>
<td>0.896</td>
</tr>
<tr>
<td>JUNB</td>
<td>Jun B oncogene</td>
<td>Transcription factor</td>
<td>1.0 (0.1–4.2)</td>
<td>6.7 (0.6–47.9)</td>
<td>&lt;10^-6</td>
<td>0.893</td>
</tr>
<tr>
<td>FOS</td>
<td>Fos oncogene</td>
<td>Transcription factor</td>
<td>1.0 (0.3–15.9)</td>
<td>30.1 (0.4–1051.5)</td>
<td>&lt;10^-6</td>
<td>0.886</td>
</tr>
<tr>
<td>FOSB</td>
<td>Fos B oncogene</td>
<td>Transcription factor</td>
<td>1.0 (0.1–19.7)</td>
<td>40.5 (0.2–1580.4)</td>
<td>0.000002</td>
<td>0.876</td>
</tr>
<tr>
<td>VEGF</td>
<td>Vascular endothelial growth factor</td>
<td>Angiogenesis</td>
<td>1.0 (0.1–2.4)</td>
<td>2.9 (0.5–13.8)</td>
<td>0.000008</td>
<td>0.852</td>
</tr>
<tr>
<td>DTR/HB-EGF</td>
<td>Dipherthia toxin receptor (heparin-binding EGF-like growth factor)</td>
<td>Growth factor</td>
<td>1.0 (0.3–4.9)</td>
<td>4.7 (0.4–56.4)</td>
<td>0.00004</td>
<td>0.824</td>
</tr>
<tr>
<td>TBX2R</td>
<td>Thromboxane A2 receptor</td>
<td>Angiogenesis</td>
<td>1.0 (0.4–4.0)</td>
<td>4.4 (1.2–22.2)</td>
<td>0.00004</td>
<td>0.824</td>
</tr>
<tr>
<td>PTGS2/COX2</td>
<td>Prostaglandin-endoperoxide synthase 2 (cyclooxygenase-2)</td>
<td>Angiogenesis</td>
<td>1.0 (0.2–22.9)</td>
<td>10.7 (0.2–256.3)</td>
<td>0.00004</td>
<td>0.824</td>
</tr>
<tr>
<td>IGFBP7</td>
<td>Insulin-like growth factor binding protein 7 (IGF-β7)</td>
<td>Growth factor</td>
<td>1.0 (0.1–2.8)</td>
<td>3.1 (0.3–23.1)</td>
<td>0.00012</td>
<td>0.803</td>
</tr>
<tr>
<td>IL6</td>
<td>Interleukin 6</td>
<td>Growth factor</td>
<td>1.0 (0.2–20.6)</td>
<td>9.3 (0.4–93.1)</td>
<td>0.00020</td>
<td>0.793</td>
</tr>
<tr>
<td>NOS3</td>
<td>Nitric oxide synthase 3, endothelial (ENOS)</td>
<td>Oxidative stress</td>
<td>1.0 (0.2–3.6)</td>
<td>3.5 (0.6–9.6)</td>
<td>0.00029</td>
<td>0.785</td>
</tr>
<tr>
<td>MAP3K5/COT</td>
<td>Mitogen-activated protein kinase kinase 8</td>
<td>Signaling transduction</td>
<td>1.0 (0.3–3.0)</td>
<td>2.7 (0.1–11.6)</td>
<td>0.00068</td>
<td>0.768</td>
</tr>
<tr>
<td>RASGRF1</td>
<td>Ras protein-specific guanine nucleotide-releasing factor 1</td>
<td>Signaling transduction</td>
<td>1.0 (0.1–108.0)</td>
<td>6.6 (0.7–183.5)</td>
<td>0.0012</td>
<td>0.755</td>
</tr>
<tr>
<td>KAI1</td>
<td>Kangai 1 (CD82 antigen)</td>
<td>Cell motility</td>
<td>1.0 (0.2–4.1)</td>
<td>3.4 (1.0–10.5)</td>
<td>0.0014</td>
<td>0.752</td>
</tr>
<tr>
<td>THBD</td>
<td>Thrombomodulin</td>
<td>Angiogenesis</td>
<td>1.0 (0.2–4.6)</td>
<td>3.6 (0.5–23.6)</td>
<td>0.0014</td>
<td>0.751</td>
</tr>
<tr>
<td>PPARG1</td>
<td>Peroxisome proliferative activated receptor, gamma, coactivator 1 (PGC1)</td>
<td>Transcription factor coactivator</td>
<td>1.0 (0.0–14.0)</td>
<td>3.0 (0.4–156.6)</td>
<td>0.0024</td>
<td>0.739</td>
</tr>
<tr>
<td>ANGPT2</td>
<td>Angiopoietin 2</td>
<td>Angiogenesis</td>
<td>1.0 (0.2–4.7)</td>
<td>3.0 (0.3–10.7)</td>
<td>0.0024</td>
<td>0.739</td>
</tr>
<tr>
<td>EREG</td>
<td>Epieregulin</td>
<td>Growth factor</td>
<td>1.0 (0.0–173.2)</td>
<td>16.1 (0.2–10744.5)</td>
<td>0.0037</td>
<td>0.729</td>
</tr>
<tr>
<td>TNFRSF10A</td>
<td>Tumor necrosis factor receptor superfamily, member 10a (TRAILR1, DR4)</td>
<td>Apoptosis</td>
<td>1.0 (0.4–2.3)</td>
<td>2.7 (0.2–17.9)</td>
<td>0.0043</td>
<td>0.725</td>
</tr>
<tr>
<td>ROBO2</td>
<td>Roundabout homolog 2</td>
<td>Growth factor receptor</td>
<td>1.0 (0.0–771.6)</td>
<td>14.8 (0.0–1521.1)</td>
<td>0.0073</td>
<td>0.711</td>
</tr>
<tr>
<td>CCL5/MIP1A</td>
<td>Chemokine (C-C motif) ligand 3</td>
<td>Growth factor</td>
<td>1.0 (0.1–5.6)</td>
<td>2.0 (0.2–31.2)</td>
<td>0.019</td>
<td>0.684</td>
</tr>
<tr>
<td>MYCN</td>
<td>N-myc oncogene</td>
<td>Transcription factor</td>
<td>1.0 (0.1–4.8)</td>
<td>3.1 (0.2–144.4)</td>
<td>0.021</td>
<td>0.682</td>
</tr>
<tr>
<td>CCL5/RANTES</td>
<td>Chemokine (C-C motif) ligand 5</td>
<td>Growth factor</td>
<td>1.0 (0.1–3.5)</td>
<td>3.2 (0.1–33.1)</td>
<td>0.029</td>
<td>0.672</td>
</tr>
<tr>
<td>SNAIL1</td>
<td>Snail homolog 1</td>
<td>Transcription factor</td>
<td>1.0 (0.5–3.5)</td>
<td>2.2 (0.5–7.3)</td>
<td>0.043</td>
<td>0.660</td>
</tr>
<tr>
<td>H19</td>
<td>H19, imprinted maternally expressed untranslated mRNA</td>
<td>Development</td>
<td>1.0 (0.1–13.4)</td>
<td>6.4 (0.1–58.6)</td>
<td>0.046</td>
<td>0.657</td>
</tr>
<tr>
<td>MKI67</td>
<td>Proliferation-related Ki-67 antigen</td>
<td>Cell cycle regulation</td>
<td>1.0 (0.1–3.7)</td>
<td>1.6 (0.2–21.2)</td>
<td>NS (0.07)</td>
<td>0.644</td>
</tr>
<tr>
<td>ESR1/ERα</td>
<td>Estrogen receptor 1 (α)</td>
<td>Nuclear receptor</td>
<td>1.0 (0.0–21.1)</td>
<td>0.4 (0.0–21.3)</td>
<td>NS (0.18)</td>
<td>0.606</td>
</tr>
</tbody>
</table>

* Kruskal-Wallis test.
† Median (range) of gene mRNA levels.
overexpression in IBC compared with non-IBC ($P = 0.07$), whereas $ESR1/ER\alpha$ and $ERBB2$ expressions were similar in the two tumor types.

The mRNA levels indicated in Table 1 (calculated as described in Materials and Methods) show the abundance of the target relative to the endogenous control (TBP), to normalize the starting amount and quality of total RNA. Similar results were obtained with a second endogenous control, $RPLP0$ (also known as 36B4). Indeed, the 27 up-regulated genes were also significantly up-regulated in the IBCs relative to the non-IBCs.

**mRNA Expression of the 48 Candidate Genes in IBC, According to Relapse.** Twenty-seven (75%) of the 36 patients with IBC relapsed, a proportion in keeping with published data (18). Comparison of median mRNA levels of the 48 genes between patients who relapsed ($n = 27$) and those who did not relapse ($n = 9$) identified three genes with significantly different

---

**Fig. 1** mRNA levels of $JUN$, $EGR1$, and $DUSP1$ in 22 individual non-IBCs (gray bars) and 36 IBCs (black bars). Median values (and ranges) are indicated for each tumor subgroup.
expression ($P < 0.05$), namely MYCN, EREG, and SHH (Table 2). MYCN was the most discriminatory gene (AUC-ROC 0.765). It is noteworthy that SHH mRNA levels were higher in patients who relapsed, whereas MYCN and EREG mRNA levels were lower in patients who relapsed.

Finally, hierarchical clustering of the samples, based on the expression of these three genes, subdivided the patient population into three groups with significantly different outcomes ($P = 0.003$; Fig. 2): 16 patients with poor outcome (all but 1 relapsed), 11 with intermediate outcome (9 relapsed) and 9 patients with good outcome (only 3 relapsed).

**DISCUSSION**

We first used real-time quantitative RT-PCR to quantify the mRNA expression of 538 selected genes in pooled IBC samples relative to pooled non-IBC samples. The 48 genes of interest thus identified were then investigated in 36 individual IBCs and 22 non-IBCs. Using the same approach, we have shown the involvement of several altered molecular pathways in the genesis of prostate and liver cancer (10, 11).

Real-time quantitative RT-PCR is a promising alternative to cDNA microarrays for molecular tumor profiling. In particular, real-time RT-PCR is far more precise, reproducible, and quantitative than cDNA microarrays. Real-time RT-PCR is also more useful for analyzing weakly expressed genes (such as EREG, ROBO2, and SHH in the present study). Finally, real-time RT-PCR requires smaller amounts of total RNA (about 2 ng/target gene) and is therefore suitable for analyzing small tumor samples or microdissected tumor samples.

We studied a number of genes involved in various cellular and molecular mechanisms associated with tumorigenesis and known to be altered (mainly at the transcriptional level) in various cancers. These genes encode proteins involved in cell cycle control, cell-cell interactions, signal transduction pathways, apoptosis, angiogenesis, etc. (list in Supplemental data). After scrutinizing the literature, we also included many genes involved in angiogenesis and inflammation.

Among the 538 genes analyzed, 27 (5.0%) showed substantial up-regulation in IBC compared with non-IBC, suggest-
ing that several signaling pathways are specifically involved in IBC (Table 1).

Some results of this study are in partial agreement with those reported in the IBC literature. First, MKI67 was overexpressed in IBC (but not significantly), supporting higher cell proliferation rates than in non-IBC. MKI67 encodes Ki-67 antigen, a large protein of unknown function that is a classical histopathological marker associated with cell proliferation. Paradiso et al. (19) have reported higher proliferative activity (measured by [H]thyidine autoradiographic labeling index) in IBC than in non-IBC.

Second, genes involved in angiogenesis (VEGF, TBXA2R, PTGS2/COX2, THBD/thrombomodulin, and ANGPT2/angiopoietin 2) were also up-regulated. Several authors have reported that IBC has strong angiogenic potential, using different approaches and experimental systems including the following: analysis of microvessel density and endothelial cell proliferation in IBC (6) and analysis of histologic characteristics and molecular basis of human IBC xenograft in nude mice (7, 8). However, we found that other best-known major angiogenic genes (VEGF2, VEGF3, VEGF4, VEGFR1, VEGFR2, and VEGFR3) had similar expression levels in IBC and non-IBC.

Likewise, IBC and non-IBC showed similar expression levels of the best-known inflammatory cytokines (see list in Supplemental data), including IFNG, TNF, IL1A, IL1B, IL8, and IL10, tending to confirm the hypothesis that the inflammatory phenotype of IBC is attributable to blockade of the dermal lymphatics by the tumor infiltrate and not to infiltration by inflammatory cells (3). Interestingly, the gene CCR5 encoding the chemokines CCL3/MIP1A and CCL5/RANTES was up-regulated (Table 1). Initially, the gene CCR5 encoding the specific receptor of both CCL3/MIP1A and CCL5/RANTES was also up-regulated in IBC, although not significantly (P = 0.11), and had a very similar expression pattern to CCL3 and CCL5 (r = +0.539, P = 0.00078 and r = +0.891, P < 10^-6, respectively; Spearman rank correlation test). Moreover, the two other CCR5 ligands (CCL4/MIP1B and CCL8/MCP-2), although not significantly up-regulated in IBC, also had a very similar expression pattern to CCR5 (r = +0.889, P < 10^-6 and r = +0.738, P < 10^-6, respectively; Spearman rank correlation test) in IBC samples.

Finally, IBC and non-IBC showed similar expression levels of the genes WISP3/LIBC, ARHC/RhoC, and CDH1/E-cadherin, which has previously been described as specifically altered in IBC (5, 6).

Our results suggest that specific molecular signaling pathways are altered in IBC compared with non-IBC. We found that most components of the AP-1 transcription factor family, including JUN, JUNB, FOS, and FOSB, were up-regulated (Table 1). AP-1 has been implicated in a variety of tumoral processes, including inflammation, cell transformation, invasive growth, angiogenesis, and metastasis (20). Interestingly, we also observed the up-regulation of major downstream target genes of the altered AP-1 pathway, i.e., DTR/HH-EGF (21), PTGS2/COX2 (22), and MAP3K8/COT (23). AP-1 transcription factor activation may be attributable to many different signaling pathways, including the binding of IL6 (the IL6 gene was up-regulated in our study) to its receptor (24).

Other early growth-response genes that code for transcription factors (MYCN and EGR1) or nonreceptor-type protein-tyrosine phosphatase (DUSP1) were up-regulated in IBC. Like AP-1, EGR1 (also known as KROX24) and DUSP1 (also known as CL100 and MKP-1) are mainly activated by hypoxia, environmental stress, and proinflammatory cytokines (25, 26). It is noteworthy that HIF1A (Hypoxia-inducible factor 1α) gene, encoding a key transcription factor induced by hypoxia (27), was also overexpressed in IBC but not significantly (P = 0.06). Taken together, our results suggest that intratumoral hypoxia, possibly attributable to avid embolus formation, may be a prominent feature of IBC. Hypoxia is thought to drive angiogenesis and to be an important contributor to radiation and drug resistance (27).

We attempted to identify markers of IBC aggressiveness by comparing the gene transcription profiles according to relapse status, and we identified a three-gene expression signature predictive of relapse (MYCN, EREG, and SHH).

EREG (epiregulin) belongs to the EGFR growth factor family. This ERBB ligand, like DTR/HH-EGF (also up-regulated in our study), binds both ERBB1 and ERBB4 (28). SHH codes for the most important molecule of the Hedgehog-Gli signaling pathway. Inappropriate activation of the Hedgehog-Gli signaling pathway occurs in several malignancies, including pancreas, brain, and skin tumors (29, 30). The third gene of our gene expression signature, MYCN, is an unexpected gene involved in breast tumorigenesis. Indeed, MYCN alteration is classically described in neuroblastoma and retinoblastoma, in opposite to MYC (not up-regulated in our IBC study) that is altered in numerous malignancies including breast cancer. Interestingly, MYCN was recently identified as a major direct target of SHH pathways in medulloblastoma, a childhood-onset brain tumor (31). However, we observed a negative correlation between SHH and MYCN expression in IBC (r = -0.411, P = 0.012; Spearman rank correlation test). Taken together, these results point to complex regulation of MYCN expression by SHH, through various Gli transcription factors (GLI1, GLI2, GLI3, or GLI4), specific to each cancer type.

In conclusion, this study points to the involvement of several altered molecular pathways in IBC tumorigenesis. Additional studies are necessary to identify the genetic or epigenetic mechanisms responsible for the altered gene expression and to determine the cells types (tumoral epithelial cells or various stromal including fibroblasts, macrophages, lymphocytes, or other cells) responsible for the altered expression of each gene. We identified a gene expression signature of poor-prognosis IBC that warrants validation in larger series. Finally, some of the genes identified here could form the basis for novel IBC treatment strategies. In this regard, it is noteworthy that EREG and SHH encode secreted proteins that could be studied in human xenograft models of IBC.

ACKNOWLEDGMENTS

We thank the staff of Centre René Huguenin and Hôpital Saint-Louis for assistance in specimen collection and patient care.

REFERENCES

1. Chang S, Parker SL, Pham T, Buzdar AU, Hursting SD. Inflammatory breast carcinoma incidence and survival: the surveillance, epide-
Molecular Profiling of Inflammatory Breast Cancer: Identification of a Poor-Prognosis Gene Expression Signature

Ivan Bièche, Florence Lerebours, Sengül Tozlu, et al.


Updated version
Access the most recent version of this article at:
http://clincancerres.aacrjournals.org/content/10/20/6789

Supplementary Material
Access the most recent supplemental material at:
http://clincancerres.aacrjournals.org/content/suppl/2004/12/29/10.20.6789.DC1
http://clincancerres.aacrjournals.org/content/suppl/2005/06/27/10.20.6789.DC2

Cited articles
This article cites 29 articles, 14 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/10/20/6789.full.html#ref-list-1

Citing articles
This article has been cited by 34 HighWire-hosted articles. Access the articles at:
/content/10/20/6789.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.