Dual Inhibition of mTOR and Estrogen Receptor Signaling In vitro Induces Cell Death in Models of Breast Cancer

Anne Boulay, Joelle Rudloff, Jingjing Ye, Sabine Zumstein-Mecker, Terence O'Reilly, Dean B. Evans, Shiuan Chen, and Heidi A. Lane

Abstract

Purpose: RAD001 (everolimus), a mammalian target of rapamycin (mTOR) pathway inhibitor in phase II clinical trials in oncology, exerts potent antiproliferative/antitumor activities. Many breast cancers are dependent for proliferation on estrogens synthesized from androgens (i.e., androstenedione) by aromatase. Letrozole (Femara) is an aromatase inhibitor used for treatment of postmenopausal women with hormone-dependent breast cancers. The role of the mTOR pathway in estrogen-driven proliferation and effects of combining RAD001 and letrozole were examined in vitro in two breast cancer models.

Experimental Design: The role of the mTOR pathway in estrogen response was evaluated in aromatase-expressing MCF7/Aro breast cancer cells by immunoblotting. Effects of RAD001 and letrozole (alone and in combination) on the proliferation and survival of MCF7/Aro and T47D/Aro cells were evaluated using proliferation assays, flow cytometry, immunoblotting, and apoptosis analyses.

Results: Treatment of MCF7/Aro cells with estradiol or androstenedione caused modulation of the mTOR pathway, a phenomenon reversed by letrozole or RAD001. In MCF7/Aro and T47D/Aro cells, both agents inhibited androstenedione-induced proliferation; however, in combination, this was significantly augmented (P < 0.001, two-way ANOVA, synergy by isobologram analysis). Increased activity of the combination correlated with more profound effects on G1 progression and a significant decrease in cell viability (P < 0.01, two-way ANOVA) defined as apoptosis (P < 0.05, Friedman test). Increased cell death was particularly evident with optimal drug concentrations.

Conclusion: mTOR signaling is required for estrogen-induced breast tumor cell proliferation. Moreover, RAD001-letrazole combinations can act in a synergistic manner to inhibit proliferation and trigger apoptotic cell death. This combination holds promise for the treatment of hormone-dependent breast cancers.
and the mitogen-activated protein kinase pathways, have been documented (10, 11). Moreover, the observation that long-term estrogen-deprived MCF7 cells (with increased estrogen sensitivity) exhibit up-regulation of ErbB2 and Erk1/2 (12), and increased Akt phosphorylation and mammalian target of rapamycin (mTOR) effector activation (13) also indicates that up-regulation of growth factor signaling is fundamental to the adaptation of breast cancer cells to low estrogen levels in cultured cells. In support of this hypothesis, tamoxifen treatment in the clinical setting resulted in increased tumor ErbB2 and p38 mitogen-activated protein kinase activation at relapse (14). It is feasible, therefore, that interactions between estrogen/ER and growth factor signal transduction pathways may contribute to both the failure of endocrine therapy as well as the development of resistance.

One strategy to improve the efficacy of aromatase inhibitors and potentially circumvent or delay the development of resistance is to concomitantly target the aromatase/estrogen/ER and growth factor pathways in breast cancer patients. In this respect, the PI3K/Akt pathway plays a major role in breast cancer, with up-regulation associated with a more aggressive clinical phenotype (15) and a worse clinical outcome for endocrine-treated patients (16). Furthermore, this pathway has been heavily implicated in resistance to antiestrogen therapeutics (10, 11, 17). An emerging mediator of PI3K/Akt activities relating to tumor cell growth and proliferation is the mTOR kinase (18, 19). The mTOR pathway is a central sensor for nutrient/energy availability, being further modulated by PI3K/Akt-dependent mechanisms (19). In the presence of mitogenic stimuli and sufficient nutrients and energy, mTOR relays a positive signal to the translational machinery by activating the 40S ribosomal protein S6 kinases (S6K1-2) and inhibiting the eukaryotic initiation factor 4E binding proteins (4E-BP1-3). The S6Ks have been implicated in the translational regulation of mRNAs that typically encode ribosomal proteins as well as components of the translational machinery. The mTOR-dependent phosphorylation of 4E-BP1 mediates its dissociation from the RNA cap-binding protein eIF-4E, thereby allowing reconstitution of a translationally competent initiation factor complex (eIF-4F). The eIF-4F complex also comprises eIF-4GI or eIF-4GII scaffold proteins and the eIF-4A RNA helicase, and activation results in the translation of proteins involved in G1/S phase progression (19). The importance of mTOR signaling in tumor biology is now widely accepted (19, 20). Consequently, a number of agents that selectively target mTOR are being developed in the oncology indication (19).

RAD001 (everolimus), an orally bioavailable, mTOR inhibitor currently in phase II clinical trials in cancer patients (19, 21), RAD001 potently inhibits tumor cell proliferation in vitro and exhibits antitumor activity in a range of animal models (18, 22–25). Irrespective of ER status, breast cancer cell lines seem particularly sensitive to RAD001 (and other rapamycins), with IC50 values for in vitro antiproliferative activity in the sub- to low nanomolar range (24, 26, 27). As the PI3K/Akt pathway is heavily deregulated in breast cancer and up-regulation of this pathway is associated with increased sensitivity to mTOR inhibition (24, 27, 28), the application of RAD001 in this patient population is warranted.

We evaluated the potential for combining letrozole with RAD001 in two in vitro models of breast carcinoma (MCF7 and T47D). We show that estrogen-induced proliferation is largely dependent on mTOR signaling. Furthermore, RAD001 in combination with letrozole has more profound effects on aromatase-mediated estrogen-induced proliferation in aromatase-expressing lines than either agent alone. In MCF7 cells, this translated at the molecular level to a greater modulation of key G1 regulators. Strikingly, combinations of both agents triggered a more profound induction of programmed cell death in both models. These data are highly supportive of the combination of these agents for the therapy of endocrine-dependent breast cancers.

Materials and Methods

Cell culture

MCF7 and T47D human breast carcinoma lines (29) were cultured in MEM EBS (Amimed) or RPMI 1640 (HyClone, Logan, UT), respectively. Supplements included 10% FCS, 2 mmol/L l-glutamine, 1 mmol/L sodium pyruvate, 1% nonessential amino acids (MCDF7), 100 IU/mL penicillin, 100 μg/mL streptomycin, 0.5 μg/mL insulin (T47D), and 0.5 (MCF7) or 0.3 (T47D) mg/mL G418. Cells were steroid deprived using phenol red–free medium supplemented with 10% charcoal-stripped FCS (HyClone/Omega) for 3 days (T47D) or 5 days (MCF7), before E2 or androstenedione (Δ4A) treatment. Treatments were initiated either 2 days post-seeding (MCF7) or immediately (T47D), and cells were treated with ligands and inhibitors every other day for 6 days (except where otherwise mentioned).

Compounds and ligands

Both RAD001 (everolimus), a derivative of rapamycin [40-O-2-hydroxyethyl]-rapamycin] and letrozole (Femara), a nonsteroidal aromatase inhibitor, were synthesized in the laboratories of Novartis Institutes for BioMedical Research (Basel, Switzerland) and were prepared in DMSO (20 mmol/L) and in ethanol (1 mmol/L), respectively. E2 and Δ4A (Sigma-Aldrich, St. Louis, MO) were prepared in ethanol at 1 and 10 mmol/L, respectively. Aliquots were stored at −20°C.

Cell proliferation assays

For E2 and Δ4A titrations, 5 × 104 MCF7/Aro and 6 × 104 MCF7 3 (1) cells were seeded (100 μL per well) into 96-well plates and steroid-deprived cells were treated every other day for 6 days. Effects on proliferation were analyzed using the YO-PRO DNA-binding fluorescent dye technique as previously described (22). To evaluate the antiproliferative effect of RAD001 and letrozole on MCF7/Aro, 105 cells were seeded into 6-well plates. Steroid-deprived cells were treated with 10 mmol/L Δ4A or 1 mmol/L E2 with RAD001 or letrozole (alone or in combination) every second day for 6 days. Cells were harvested by trypsinization, resuspended in PBS and counted using a CASY cell counter (Schärfe System, Reutlingen, Germany). For T47D/Aro, 6 × 104 cells were seeded into 6-well plates. Steroid-deprived cells were treated with 10 nmol/L Δ4A with RAD001 or letrozole (alone or in combination) every second day for 6 days. Cells were harvested by trypsinization, resuspended in PBS and counted using a CASY cell counter (Schärfe System, Reutlingen, Germany). For T47D/Aro, 6 × 104 cells were seeded into 6-well plates. Steroid-deprived cells were treated with 10 nmol/L Δ4A with RAD001 or letrozole (alone or in combination) every second day for 6 days. Cells were dissolved in 0.5 N NaOH and the protein concentration was determined.

Protein extraction and immunoblotting

To evaluate signaling pathways, 1.2 × 106 MCF7/Aro cells were seeded into 10-cm plates. Steroid-deprived cells were treated with vehicle or 1 mmol/L E2 with or without 30 minutes pretreatment with 20 nmol/L RAD001. Alternatively, cells were treated with 10 nmol/L Δ4A and concomitantly treated with 500 nmol/L letrozole or vehicle. Whole cell protein extracts were prepared as previously described (30), and supernatants were stored at −80°C.

To assess effects on cell cycle regulators, 8 × 105 and 106 MCF7/Aro cells were seeded into 10-cm plates, respectively. Steroid-deprived cells were treated with 10 nmol/L Δ4A and concomitantly treated with...
vehicle, 100 nmol/L letrozole, and 2 nmol/L RAD001 (alone or in combination) for 4 hours. Floating cells were collected and adherent cells were harvested by scraping into PBS containing 1 mmol/L phenylmethylsulfonyl fluoride. Pooled cells were extracted and frozen as described.

Immunoblotting was done as previously described (23), with the following antibodies: anti-S6K1, anti-phospho-S6K1 (Thr 389), anti-phospho-S6 (Ser240/Ser244), antihistone H3 (Ser10), anti-4E-BP1, anti-phospho-4E-BP1 (Ser70), anti-akt, anti-phospho-Akt (Ser473), anti-Phospho-Erk1/2 (Thr 202/Tyr204), and anti-phospho-ERK1/2 (Ser185) (Biosource, Camarillo, CA); anti-actin (Chemicon, Temecula, CA); anti-a-tubulin (Tub21, Sigma, St. Louis, MO), and anti-S6 (provided by Dr. J. Mestan, Novartis Institutes for BioMedical Research).

Cell cycle analysis
To assess effects on the cell cycle, 5 × 10⁵ MCF7/Aro cells were seeded into 6- or 10-cm plates. Steroid-deprived cells were treated with 10 nmol/L Δ4A and simultaneously treated with vehicle, 100 nmol/L letrozole, and 0.2 or 2 nmol/L RAD001 (alone or in combination) for 24 hours. Floating cells were collected and adherent cells harvested by trypsinization. Cells were washed once with PBS and then resuspended in propidium iodide buffer (1 mmol/L sodium citrate (pH 4), 1.5 mmol/L NaCl, 5 mmol/L EDTA, 5 mmol/L EGTA, 0.1% NP40, 4 μg of propidium iodide/mL, and 175 μg of RNase A/mL in PBS). After 30 minutes of incubation in the dark on ice, cell cycle distribution was analyzed with a Becton Dickinson FACSCalibur flow cytometer.

Cell viability assays
YO-PRO. Cells (10⁵) in 2 mL medium were seeded into 6-well plates. Steroid-deprived cells were treated with 10 nmol/L Δ4A and concomitantly treated with vehicle, letrozole (alone or in combination with RAD001) every other day for 6 days. Effects on cell viability were analyzed using the YO-PRO DNA-binding fluorescent dye technique as previously described (22).

Terminal deoxynucleotidyl transferase–mediated nick-end labeling. For flow cytometry, 4 × 10⁵ cells were seeded into 10-cm plates. Steroid-deprived cells were treated with 10 nmol/L Δ4A and concomitantly treated with vehicle, letrozole (alone or in combination with RAD001) every second day for 6 days. Cells were stained according to the manufacturers (APO-DIRECT; BD PharMingen, San Diego, CA). FITC-labeled cells were analyzed by flow cytometry using Cell Quest software. For microscopic analyses, 10⁵ MCF7/Aro cells were seeded on gelatin-covered cover slips in 6-well plates. Steroid-deprived cells were treated as above. Terminal deoxynucleotidyl transferase–mediated nick-end labeling (TUNEL) staining was done directly on the adherent cells in a humidified chamber as above. Cell nuclei were counterstained with Hoechst 33258 (Molecular Probes, Eugene, OR) in the dark. Cells were washed thrice with PBS and coverslips were mounted using Vectashield (Vector Laboratories, Burlingame, CA). Apoptotic cells were analyzed by fluorescence microscopy (Leica DM IRB, 20× objective; Kodak DC290 Zoom digital camera).

Statistical analyses
Proliferation and cell viability data were statistically analyzed using two-way ANOVA (with Tukey key test for pairwise comparisons) to test for the effect of RAD001 and letrozole as single agents and for interactions between the compounds. Calculations were made using SigmaStat 3.1 (Systat Software, Systat Software GmbH, Erkrath, Germany), and P < 0.05 was considered statistically significant. To further determine the nature of the letrozole-RAD001 interactions, partial isobolograms were constructed to permit estimating the coefficient g, which represents the value of the equation Ac / (Am + Bc) / Bm, where Ac (or Bc) is the dose of compound A (or B) in combination, Am (or Bm) is the dose of compound A (or B) in monotherapy, that give equivalent activity (31, 32). When g = 1, the combination is additive in nature; when >1, the combination is considered antagonistic; and when <1, the combination may be considered synergistic. The activity of the known doses of letrozole or RAD001 in combination was taken as a reference point for determining, by interpolation of the concentration-response curves, the concentrations of letrozole and RAD001 in monotherapy that would produce the same activity. As some of the antiproliferative activities of the combinations fell outside of the single agent concentration-response curves, these combinations were not available for use in determining interactions. Of the data that satisfied these restrictions, all combinations showed synergy by this calculation procedure as described in Materials and Methods (31, 32). When g = 1, the combination is additive in nature; when >1, the combination is considered antagonistic; and when <1, the combination may be considered synergistic. The activity of the known doses of letrozole or RAD001 in combination was taken as a reference point for determining, by interpolation of the concentration-response curves, the concentrations of letrozole and RAD001 in monotherapy that would produce the same activity. As some of the antiproliferative activities of the combinations fell outside of the single agent concentration-response curves, these combinations were not available for use in determining interactions. Of the data that satisfied these restrictions, all combinations showed synergy by this calculation.
Furthermore, two-way ANOVA was used to determine interactions between RAD001 and letrozole, which when positively interacting, according to Slinker (33) shows drug synergy. For flow cytometry TUNEL analyses, the statistical significance was determined by the Friedman test for multiple comparison of ratios (34), using Systat V11.0 (Systat Software).

**Results**

**Estrogen-driven proliferation of MCF7/Aro cells exhibits mammalian target of rapamycin dependency.** To assess the estrogen sensitivity of MCF7 cells, we evaluated the relative proliferation of MCF7/Aro (stably expressing aromatase) and MCF7 3(1) vector control cells in the presence of E2 or the precursor Δ4A, using the YO-PRO proliferation assay (Fig. 1A and B). Steroid-deprived cells were treated every other day for 6 days. Steroid deprivation completely abolished MCF7/Aro and MCF7 3(1) cell proliferation (data not shown) thus indicating that proliferation is estrogen dependent. Consistent with this observation, both E2 and Δ4A stimulated the proliferation of MCF7/Aro cells in a concentration-dependent manner, while MCF73(1) cells responded only to E2 (Fig. 1A and B). Hence, MCF7/Aro cells can convert Δ4A into estrogens. In agreement with previous reports (35–37), these experiments defined 1 nmol/L E2 and 10 nmol/L Δ4A as effective concentrations for further experiments.

MCF7 parental cells cultured in complete medium are sensitive to mTOR inhibition (26, 27). In complete medium, we also observed that RAD001 potently inhibited MCF7 proliferation (IC50 = 0.6 ± 0.1 nmol/L), and MCF7 3(1) and MCF7/Aro cells exhibited a similar response to optimal RAD001 concentrations (20 nmol/L RAD001 induces a ~20% increase in G1 population after 24 hours of incubation; data not shown). To examine whether estrogen-driven proliferation of MCF7/Aro cells was dependent on mTOR signaling, steroid-deprived cells were stimulated with E2 or Δ4A in the presence of increasing concentrations of RAD001 or vehicle (Fig. 1C). Interestingly, RAD001 dramatically impaired E2- and Δ4A-induced proliferation of MCF7/Aro cells in a concentration-dependent manner, with maximal effects elicited at 2 to 20 nmol/L RAD001 (2 nmol/L, 68% and 67%; 20 nmol/L, 79% and 74% inhibition of E2- and Δ4A-dependent proliferation, respectively), and partial inhibition at 0.2 nmol/L RAD001 (Fig. 1C). From these data, 0.2 and 2 nmol/L were defined as suboptimal and optimal RAD001 concentrations, respectively.

The effect of E2 on mTOR signaling was also analyzed. Steroid-deprived MCF7/Aro cells were either vehicle-treated or treated for up to 24 hours with E2, with or without a 30 minutes pretreatment with 20 nmol/L RAD001 (Fig. 2). Vehicle had little effect on mTOR pathway components. In contrast, E2 treatment induced prolonged phosphorylation of S6K1 and its substrate...
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Figure 3. Combinations of RAD001 and letrozole result in enhanced antiproliferative activity in androstenedione-driven MCF7/Aro cells. Steroid-deprived MCF7/Aro cells were treated with 10 nmol/L Δ4A and increasing concentrations of letrozole (Lct) alone or in combination with 0.2 or 2 nmol/L RAD001 (C) for 6 days. Relative proliferation was assessed by direct cell counting. Columns, means of triplicate values; bars, ±SD. Stars, \( P < 0.001 \), two-way ANOVA using Tukey’s test for pairwise comparisons (synergistic drug interaction). B, cells were left untreated or treated with 10 nmol/L Δ4A with concomitant treatment with 500 nmol/L letrozole or vehicle for 4 or 24 hours. Whole cell protein extracts were resolved by SDS-PAGE, transferred onto a polyvinylidene difluoride membrane, and probed with the indicated antibodies. β-Tubulin was used as a loading control.

The 40S ribosomal protein S6, which was evident after 2 hours and maintained for 24 hours (Fig. 2, left). Consistent with these changes being mTOR regulated, both responses were inhibited by RAD001 pretreatment (Fig. 2, right). Similarly, E2 led to decreased electrophoretic mobility of the translational repressor 4E-BP1 for 2 to 24 hours, a phenomenon indicative of increased phosphorylation and prolonged functional inactivation and also counteracted by RAD001 treatment (Fig. 2, compare left and right). Of particular interest, E2 treatment caused decreased elf-4E and increased elf-4GI phosphorylation, events that paralleled effects on S6K1 and 4E-BP1 and antagonized by RAD001 (Fig. 2). Moreover, short-term treatment with RAD001 alone resulted in increased phosphorylation of elf-4E, an observation not previously reported (Fig. 2, compare lane C SD with lane C RAD001). In the presence of RAD001, prolonged increased Akt phosphorylation was also observed, consistent with previous observations (38). However, although it has been previously reported that E2 induces transient induction (5-10 minutes) of Akt phosphorylation in MCF7 cells (39), analysis of Akt and mitogen-activated protein kinase phosphorylation suggested no E2-specific modulation of these signal transducers in our experimental setting, where the first measurement was after 10 minutes.

Altogether, these data provide a molecular basis explaining the observation that RAD001 inhibits E2- and Δ4A-induced proliferation in MCF7/Aro cells, with specific modulation of elements downstream of mTOR highlighting a contribution of this pathway to estrogen response.

Combinations of RAD001 and letrozole result in enhanced antiproliferative activity androstenedione-driven MCF7/Aro cells. Letrozole inhibits the conversion of Δ4A into estrogens by inhibiting aromatase activity (1). Hence, treatment of MCF7/Aro cells with letrozole resulted in a concentration-dependent inhibition of Δ4A-driven proliferation (Fig. 3A). The IC50 for letrozole was defined as ~100 nmol/L, with maximal inhibition observed at 500 to 1,000 nmol/L (Fig. 3C and A). Moreover, Δ4A treatment also induced modulation of the mTOR pathway, an effect reversed by the presence of 500 nmol/L letrozole (Fig. 3B). Hence, 100 and 500 nmol/L were defined as suboptimal and optimal concentrations of letrozole, respectively.

The effect of combining suboptimal and optimal concentrations of RAD001 and letrozole on Δ4A-driven proliferation of MCF7/Aro cells was evaluated. As expected (see Fig. 1C and 3A), treatment with RAD001 or letrozole alone inhibited MCF7/Aro proliferation in a concentration-dependent manner (Fig. 3C). Strikingly, combining suboptimal concentrations of RAD001 and letrozole significantly enhanced the antiproliferative activity as compared with either agent alone (0.2 nmol/L RAD001, 45%; 100 nmol/L letrozole, 41%; Combination, 76% inhibition of proliferation). A similar phenomenon was observed with optimal concentrations of the two agents (2 nmol/L RAD001, 80%; 500 nmol/L letrozole, 70%; Combination, 96% inhibition of proliferation). Statistical analysis indicated that highly significant interactions exist between RAD001 and letrozole (\( P < 0.001 \), two-way ANOVA), suggesting strong combination effects and, according to Slinker (33), synergistic drug interaction. Furthermore, using methods that facilitate determination of combination effects based upon limited isobologram data (31, 32), synergistic drug interactions were shown (see Materials and Methods).

Increased antiproliferative potential of RAD001/letrozole combinations correlates with more pronounced G1 accumulation. Both mTOR and estrogen signaling pathways are known to regulate G1-phase progression (19, 40). To investigate the effect of letrozole and RAD001 treatment on cell cycle distribution,
steroid-deprived MCF7/Aro cells were treated with Δ4A in the absence or presence of 100 nmol/L (suboptimal) letrozole and 0.2 nmol/L (suboptimal) or 2 nmol/L (optimal) RAD001, alone or in combination. To observe immediate effects on cell cycle distribution, flow cytometry was done after 24 hours incubation (Fig. 4A). As expected (24), 2 nmol/L RAD001 induced an increase in the G1 population, whereas 0.2 nmol/L RAD001 had no-to-minimal effects (Fig. 4A, top row and see legend). Consistent with a previous report (36), treatment with 100 nmol/L letrozole also affected G1 progression; however, combinations of both agents triggered a more pronounced G1 accumulation (Fig. 4A, bottom row and see legend). Strikingly, this occurred even with the suboptimal 0.2 nmol/L RAD001 concentration, which alone had little effect on the cell cycle.

To further analyze these cell cycle effects, analysis of proteins central to G1-S phase progression was done. Steroid-deprived MCF7/Aro cells were treated for 4 hours with Δ4A in the absence or presence of 100 nmol/L letrozole or 2 nmol/L RAD001, alone or in combination. RAD001 caused S6 dephosphorylation and decreased 4E-BP1 protein mobility (Fig. 4B, left). A minor effect of 100 nmol/L letrozole on these proteins was also observed, consistent with this suboptimal concentration (Fig. 4B and see Fig. 3A). Analysis of the expression of the D-type cyclins, essential subunits of G1 cyclin-dependent kinases (Cdk4/6) suggested to be regulated through both mTOR and ER signaling (19, 40), showed that both RAD001 and letrozole similarly reduced cyclin D1 and D2 protein expression, with cyclin D3 levels unaffected. In combination, however, a further decrease in cyclin D1 and D2 protein was observed (Fig. 4B, middle). Cyclin D complexed with Cdk4 is essential for the phosphorylation of the retinoblastoma tumor suppressor protein, in particular on residue Ser795 (41). Indeed, RAD001 and letrozole caused a slight increase in retinoblastoma mobility, indicative of dephosphorylation and activation as a suppressor of proliferation, but this was accentuated with the combination; in which case dephosphorylation of Ser795 was observed (Fig. 4B, right). Taken together, these data indicate that dual inhibition of E2 and mTOR signaling can result in more profound effects on G1 regulators, culminating in a more pronounced G1 accumulation.

Dual inhibition of mammalian target of rapamycin and estradiol signaling induces apoptosis of MCF7/Aro cells. Steroid-deprived MCF7/Aro cells were treated with 10 nmol/L Δ4A in the absence or presence of 100 or 500 nmol/L letrozole (Let), alone or in combination with 2 nmol/L RAD001 for 6 days. The numbers of nonviable and apoptotic cells were evaluated using the YO-PRO assay (A) or a cytometry-based TUNEL analysis (B), respectively, as described in Materials and Methods. Columns, means of triplicate values (A) or a single value (B); bars, ± SD. A, stars, P < 0.01, two-way ANOVA using Tukey’s test for pairwise comparisons. B, stars, P < 0.05, Friedman test.
was done. MCF7/Aro cells were treated with Δ4A in the absence or presence of 2 nmol/L RAD001 or 100 or 500 nmol/L letrozole alone had minimal effects on apoptosis, a significant potentiation of apoptotic cell death was observed with the combination (P < 0.05, Friedman test; Fig. 5B; ref. 34), again particularly evident with 500 nmol/L letrozole (apoptotic index: 5.6%, 2 nmol/L RAD001; 2.6%, 500 nmol/L letrozole; 20%, Combination). Induction of apoptosis was also confirmed using a fluorescence microscopy-based TUNEL assay. Treatment of MCF7/Aro cells with optimal concentrations of RAD001 or letrozole clearly reduced the number of cells per field, consistent with their antiproliferative effects (Figs. 1C, Fig. 3A, and Fig. 4A), but only 0.6% and 1.8% of the cells stained positively for TUNEL, respectively (Fig. 5C). In comparison, drug combination further reduced the number of cells per field and 16.6% of the cells stained positive for TUNEL. Taken together, these data show that combinations of RAD001 and letrozole trigger apoptotic cell death in MCF7/Aro cells.

**Discussion**

With the development of targeted therapeutics, such as letrozole and RAD001, emerges a potential for combining these agents in rational, mechanism-based approaches to achieve a more potent antitumor effect in the patient. Clearly, in the case of the ER and mTOR signaling pathways, there is a large body of evidence suggesting that these pathways have distinct as well as overlapping signaling cascades and outputs (10, 16, 17, 26, 35, 39). With this in mind, it is possible to design rational strategies to exploit these interactions to enhance the anti-tumor activity of these agents.
in vivo major inhibition, characterized by increased phosphorylation of a
also defined a novel response of tumor cells to mTOR
enhanced phosphorylation of S6K1 and 4E-BP1 (13). We have
mitogenic affect of E2, a phenomenon associated with
depprivation enhances the sensitivity of MCF7 cells to the
modulation of downstream elements of mTOR signaling in
in context of estrogen response. In agreement with
a dependency on mTOR, we further show that E2 treatment
of MCF7/Aro cells resulted in rapid activation of the S6K1/S6
pathway and modulation of 4E-BP1/eIF-4E/eIF-4GI phosphor-
ylation. This occurred within 4 hours of E2 addition, was also
associated with Δ4A-induced proliferation, and was prevented
by concomitant treatment with either RAD001 or letrozole.
These data, therefore, provide the first report defining clear
modulation of downstream elements of mTOR signaling in
response to estrogen signaling. In addition, these data are
consistent with a recent report that long-term estrogen
deprivation enhances the sensitivity of MCF7 cells to the
mitogenic affect of E2, a phenomenon associated with
enhanced phosphorylation of S6K1 and 4E-BP1 (13). We have
also defined a novel response of tumor cells to mTOR
inhibition, characterized by increased phosphorylation of a
major in vivo eIF-4E site (Ser209). We have observed a similar
phenomenon with a number of tumor lines derived from
prostate carcinoma and glioblastoma. The exact role of eIF-4E
Ser209 phosphorylation is controversial (42); however, our data
suggest a negative effect on translational events required for the
proliferative response.

The major aim of our work was to evaluate the potential
for combining RAD001 and letrozole in aromatase-expressing
breast cancer cell lines. Strikingly, drug combinations
significantly enhanced the antiproliferative activity compared
with either agent alone, with statistical analysis indicating a
synergistic interaction. Consistent with previous reports
(24, 36), treatment of MCF7/Aro cells with letrozole or
RAD001 induced G1 accumulation; however, this was clearly
increased with the combination. Both agents caused minor
decreases in cyclin D1 and D2 expression after 4 hours of
treatment, suggesting that these events are causative rather
than a consequence of decreased proliferation. Effects on
cyclin D were transient, as after 24 hours no effect was
observed (data not shown), consistent with previous work
(39, 43). Decreased cyclin D1 expression has also been
reported after E2 ablation or letrozole treatment of MCF7
and aromatase-expressing MCF7 xenografts, respectively
(36, 44, 45). Cyclin D–dependent kinases are essential
regulators of retinoblastoma phosphorylation in early G1 phase
(41); indeed, retinoblastoma phosphorylation and inactivation
as a cell cycle suppressor correlates with cyclin D1 induction
in E2-stimulated steroid-deprived MCF7 cells (39, 43). Here, both
letrozole and RAD001 treatment moderately increased retino-
blasto ma mobility indicative of decreased phosphorylation.
Moreover, combination of both agents caused a more profound
decrease in both cyclin D1 and D2 levels, accompanied by
greater effects on retinoblastoma protein mobility/phosphory-
lation; fully supportive of the increased antiproliferative effect
and G1 accumulation observed. Taken together, our data show
for the first time that concomitant inhibition of the mTOR
pathway and estrogen signaling causes more profound effects on
G1 progression and key G1 regulators.

The aim of cancer therapy is to eradicate tumors rather
than purely delay or halt development. In this context, we
further show potentiation of apoptotic cell death with
combinations of both agents, which alone have minimal
effects on cell viability. This potentiation occurred after
prolonged treatment with the agents (i.e., 6 days). Estrogen
withdrawal or letrozole treatment has been previously
reported to induce apoptotic cell death in MCF7 xenografts
(36, 44, 45). This observation was substantiated in vitro,
where estrogen withdrawal, antiestrogens, and aromatase
inhibitors were shown to induce apoptosis to varying degrees
in aromatase-expressing MCF7 cells (36). In our study,
letrozole alone had minimal effects on cell viability when
used at optimal concentrations, contrary to the more
profound effects reported by others following TUNEL assay
(36). We have confirmed our results using two TUNEL assay
approaches (fluorescence-activated cell sorting and fluores-
cence microscopy) and the YO-PRO survival assay and
suggest that the differences observed between our data and
that of Thianthanawat et al. (36) may be based on the
supraoptimal letrozole concentrations and longer incubation
periods (i.e., 8 days) used in the latter case. In the presence
of mitogens and sufficient nutrients, rapamycins generally act
cytostatically, as observed here for RAD001. However, under
stress conditions, potentiation of cell death has been reported
in certain cellular backgrounds (19, 22, 46, 47). Here we
show that in a “stress” situation that recapitulates E2
deprivation (i.e., in the presence of letrozole), concomitant
RAD001 treatment potentiates a significant induction of
breast tumor cell death. Previous studies have indicated that
E2 protects against cell death by increasing the levels of the
antiapoptotic protein Bcl-2 (48). Similarly, it was reported
that antiestrogens and aromatase inhibitors induce cell death
in vitro by increasing the expression of the proapoptotic
protein Bax and decreasing Bcl-2 expression, a phenomenon
correlating with increased caspase activity (36). Although we
also looked for specific effects on apoptotic regulators in the
combination-treated cells, the analysis was complicated by a
general reduction in protein expression coinciding with the
induction of cell death/apoptosis (data not shown). Hence,
although reduced protein expression is not surprising
considering the role of the mTOR pathway in the regulation
of global protein translation (22), this hindered a concrete
analysis of the molecular basis of the increased cell death
observed with the combination.

Taking all these data together, the more profound effects
of RAD001/letrozole combinations on both cell cycle progression
and survival, in breast lines sensitive to both agents alone, point
to a clear potential for combining these agents for the treatment
of ER-positive breast cancers. However, although endocrine
therapy is one of the most effective systemic therapies for
hormone receptor–positive breast cancer patients, efficacy is

3 Unpublished data.
often limited by the presence/onset of resistance (49). Aromatase inhibitors have a different mode of action than selective estrogen receptor modulators (1, 5), and indeed, agent-selective resistance seems to exist (49), a situation highlighted by the observation that patients who relapse after previous response to tamoxifen can subsequently respond to aromatase inhibitors (6, 7). Whether resistance is defined as agent-selective, pan, intrinsic or acquired, there is compelling evidence that up-regulation of signal transduction pathways (exemplified by increased ErbB2 signaling in tamoxifen-resistant breast cancer cells) plays a major role in resistance to endocrine therapies. Thus, clinical trials are ongoing, to examine combinations of endocrine agents with ErbB receptor inhibitors (49). Although estrogen deprivation may be more effective than antagonizing endocrine agents with ErbB receptor inhibitors (49), the use of mTOR inhibitors (such as RAD001) in combination with letrozole provides a rational approach not only in breast tumors sensitive to both agents alone (as shown here) but may also have potential as an approach to circumvent/comb endocrine resistance in the clinic.

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Anne Boulay, Joelle Rudloff, Jingjing Ye, et al.


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