Characterization of a R115777-Resistant Human Multiple Myeloma Cell Line with Cross-Resistance to PS-341

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Abstract
The farnesyl transferase inhibitor R115777 has been found to have clinical activity in diverse hematopoietic tumors. Clinical efficacy, however, does not correlate with Ras mutation status or inhibition of farnesyl transferase. To further elucidate the mechanisms by which R115777 induces apoptosis and to investigate drug resistance, we have identified and characterized a R115777-resistant human myeloma cell line. 8226/R5 cells were found to be at least 50 times more resistant to R115777 compared with the parent cell line 8226/S. K-Ras remained prenylated in both resistant and sensitive cells after R115777 treatment; however, HDJ-2 farnesylation was inhibited in both lines, implying that farnesyl transferase (the drug target) has not been mutated. Whereas many 8226 lines that acquire drug resistance have elevated expression of P-glycoprotein, we found that P-glycoprotein expression is not increased in the 8226/R5 line and intracellular accumulation of R115777 was not reduced. In fact, 8226/R5 cells were insensitive to a diverse group of antitumor agents including PS-341, and multidrug resistance did not correlate with the expression of heat shock proteins. Comparison of gene expression profiles between resistant and sensitive cells revealed expression changes in several genes involved in myeloma survival and drug resistance. Future experiments will attempt to identify genes that are directly linked to the resistant phenotype. Identification of molecules associated with R115777 and PS-341 resistance is clinically relevant because both compounds are being tested in solid tumors and hematopoietic malignancies.

Multiple myeloma is a plasma cell malignancy with no known curative therapy. RAS mutations occur frequently in myeloma (1, 2) and have been linked at least in some studies to a poor prognosis (3–5). Farnesyl transferase inhibitors (FTI) inhibit Ras function by preventing its posttranslational prenylation, a modification done by the enzyme farnesyl transferase (FTase). The FTI R115777 was designed as a highly selective inhibitor of FTase (6) and has been clinically tested in several hematopoietic tumors. This compound has shown activity in acute myelogenous leukemia, chronic myelogenous leukemia, and myelodysplastic syndrome (7–10).

Preclinical studies have reported that FTIs have antitumor activity in myeloma cell lines and primary isolates (11–13). Based on these observations, we have recently completed a phase II clinical trial testing R115777 in patients with relapsed myeloma (14). Forty-three patients with a median of four prior treatment regimens entered our study. R115777 was well tolerated and 64% of patients achieved disease stabilization. Of importance, RAS mutation and inhibition of farnesyl transferase did not correlate with clinical efficacy consistent with our prior observation that R115777 can induce apoptosis via a Ras-independent mechanism (15). In myeloma cells, R115777 activates multiple intrinsic proapoptotic cascades (15). However, the molecules and/or signaling pathways that trigger these events remain elusive.

To further elucidate the mechanisms by which R115777 induces apoptosis and to investigate drug resistance, we have established and characterized a R115777-resistant human multiple myeloma cell line (8226/R5). This line is unlike a previously described R115777-resistant colon cancer line (16) for resistance is unrelated to the prenylation activity of the enzyme FTase. This finding correlates with our observation that 8226/R5 cells are insensitive to a diverse group of antitumor agents, including PS-341. In this study, we investigate and exclude several potential mechanisms of R115777 resistance. Using comparative gene expression profiling (between sensitive and resistant cells), we have identified expression changes in several genes implicated in myeloma survival and drug resistance. Further evaluation of these genes may lead to the identification of novel FTI targets or resistance mechanisms that are clinically relevant.

Materials and Methods

Cell lines and reagents
The RPMI 8226 human myeloma cell line was obtained from the American Type Culture Collection (Manassas, VA). 8226/LR5 and
was determined by subtracting background death in untreated samples.

and propidium iodide or TO-PRO-3– positive cells. Specific cell death
doxorubicin. Cell death was calculated as the sum of Annexin V-FITC
for propidium iodide in experiments where cells were treated with
View, CA). TO-PRO-3 (Molecular Probes, Eugene, OR) was substituted
modifications of the manufacturer (BioVision Research Products, Mountain

by linear regression analysis using Excel software. Flow cytometric cell
hour (R115777) or 48-hour (PS-341) incubation period at 37
reduction assay. 8226/S and 8226/R5 cells were plated into 96-well
immunoprecipitation assay lysis buffer [150 mol/L NaCl, 1% NP40,
acid (Sigma Chemical, St. Louis, MO); staurosporine and etoposide
Pharmaceutical Research and development, LLC, Titusville, NJ). PS-341
was provided by Millennium Pharmaceuticals, Inc. (Cambridge, MA)
and FT227 from Said M. Sebti (Moffitt Cancer Center and Research
used from the following vendors: doxorubicin hydrochloride, melphanal, and perilig
acid (Sigma Chemical, St. Louis, MO); staurosporine and etoposide

Antibodies and Western blotting
Antibodies were purchased from the following vendors: anti – K-Ras-
(19; Santa Cruz Biotechnology, Inc., Santa Cruz, CA); anti-HDJ-2
(NeoMarkers, Freemont, CA); anti-hsp90, anti-hsp70, anti-hsp27, and anti –
glyceraldehyde-3-phosphate dehydrogenase (Stressgen Biotechn-
ologies, San Diego, CA); and anti –α-tubulin (BD Biosciences, San
Diego, CA). Western blotting was done as previously described (11).
For the majority of experiments, lysates were harvested using radio-
immunoprecipitation assay lysis buffer [150 μmol/L NaCl, 1% NP40,
0.5% deoxycholic acid (Sigma Chemical, St. Louis, MO); staurosporine and etoposide

Analysis of cell growth, cell cycle arrest, and cell death
Degree of resistance was determined using the tetrazolium salt 3,4,5-
dimethylazol-2-yl-2,5-diphenyl tetrazolium bromide (MTT; Sigma)
reduction assay. 8226/S and 8226/R5 cells were plated into 96-well
microtiter plates at a density of 5 × 10^5 cells/mL in 200 μL of
supplemented media. Cells were exposed to a broad range of drug
concentrations (R115777 and PS-341) in replicates of four. After a 72-
hour (R115777) or 48-hour (PS-341) incubation period at 37°C, 50 μL
of 2 mg/mL MTT was added to each well and cells were incubated for
an additional 4 hours. Plates were centrifuged for 5 minutes at 1,200
rpm in a Sorvall RT6000D table top centrifuge (Sorvall, Asheville,
NC); supernatants were removed, and water-insoluble product was dissolved
in 100 μL of 100% DMSO (Sigma). Plates were shaken for 30 seconds
and absorbance read at 540 nm on a Wallac Victor-2 1420 Multilabel
Counter (Perkin-Elmer, Torrance, CA). The number of surviving cells
was expressed as a percentage: absorbance of the experimental sample/
absorbance of the control × 100. The IC50 of the drug was calculated
by linear regression analysis using Excel software. Flow cytometric cell
cycle analysis was done after propidium iodide staining as previously
described (11). The presented histograms represent gating on live cells.
Apoptosis and cell death were evaluated by flow cytometry after
Annexin V-FITC and propidium iodide staining as per the recommenda-
tions of the manufacturer (BioVision Research Products, Mountain
View, CA). TO-PRO-3 ( Molecular Probes, Eugene, OR) was substituted
for propidium iodide in experiments where cells were treated with
doxorubicin. Cell death was calculated as the sum of Annexin V-FITC
and propidium iodide or TO-PRO-3–positive cells. Specific cell death
was determined by subtracting background death in untreated samples.

Determination of P-glycoprotein expression
8226/S, 8226/Dox40, and 8226/R5 cells were washed in PBS [0.8% NaCl, 0.02% KCl, 0.14% Na2HPO4, 0.02% KH2PO4 (pH 7.4)] and
resuspended in 200 μL of ice-cold PBS containing 0.5 μg/mL anti–P-
glycoprotein-FITC antibody (BD Biosciences) or 0.5 μg/mL isotype
control antibody anti-dansyl IgG2b-FITC (BD Biosciences). Samples
were incubated on ice for 1 hour in the dark, washed twice in PBS,
and analyzed by flow cytometry.

R115777 accumulation and efflux
To quantitate cellular accumulation of [14C]R115777, 8226/S and
8226/R5 cells were washed once in PBS and 1 × 10^6 cells were
resuspended in 1 mL supplemented media containing 5 μmol/L
R115777 (1:2.5 dilution of [14C]R115777 to cold R115777). Cells were incubated at 37°C for increasing time periods and were then
washed three times in 10 mL ice-cold PBS. Cell pellets were resuspended in 1 mL PBS and transferred to scintillation vials
containing 10 mL Scintisafe 30% scintillation fluid (Fisher Scientific,
Philadelphia, PA). Samples were analyzed on a LS5600 multipurpose
scintillation counter (Beckman Coulter, Fullerton, CA). Because drug
accumulation occurred rapidly, cells were incubated in 5 μmol/L
R115777 at decreasing dilutions (1:10, 1:2.5, 1:1) of [14C]R115777 for
1 hour and samples were processed and analyzed as described above
to confirm a dose-dependent accumulation of [14C]R115777 in each cell line. For efflux experiments, 8226/S and 8226/R5 cells were washed once in PBS and 1 × 10^6 cells were resuspended in 1 mL supplemented
media containing 5 μmol/L R115777 (1:2.5 dilution of [14C]R115777 to
cold R115777). Cells were incubated for 1 hour at 37°C and then
washed thrice in 10 mL PBS and placed in R115777-free supplemented
media for an additional hour. Samples were processed and analyzed as above to confirm a dose-
R5 lines were done in parallel.

Microarray analysis
Probe arrays. The oligonucleotide probe arrays were the Affymetrix
U133A human arrays. These arrays consist of 22,215 probe sets, which
target known and suspected genes as well as a number of suspected
splice variants. The U133A chips detect an estimated 15,000 well-
characterized human genes.

Sample processing for microarray analysis. Five micrograms of total
RNA derived from 8226/S, 8226/LR5, and 8226/R5 cells served as the
mRNA source for microarray analysis. The polyadenylate RNA was
specifically converted to cDNA and then amplified and labeled with
biotin following the procedure initially described by Van Gelder et al.
(19). Hybridization with the biotin-labeled RNA, staining, and scanning
of the chips followed the proscribed procedure outlined in the Affymetrix
technical manual and has been previously described (20).

Data analysis. Scanned output files were visually inspected for
hybridization artifacts and then analyzed using Affymetrix Microarray
Suite 5.1 software. Signal intensity was scaled to an average intensity
of 500 before comparison analysis. The MAS 5.1 software uses a statistical
algorithm to assess increases or decreases in mRNA abundance in a
direct comparison between two samples (20, 21). This analysis is based
on the behavior of 11 different oligonucleotide probes designed to
detect the same gene. The software generates a P value for the likelihood
that any perceived difference was due to chance. The P values for all
probe sets were exported to a text file and all pairwise comparisons were
then aligned in Excel. For the comprehensive analysis, P < 0.05 was
identifed as changed (increased or decreased) for each individual
comparison. Two independent samples from 8226/S, 8226/LR5, and
8226/R5 cells were collected. The samples generated from the resistant
cell line were compared with the sensitive lines in all possible
combinations. Genes were ultimately selected if they were identified
as increased in all eight comparisons or decreased in all eight comparisons.
Statistical analysis

Unless otherwise stated, statistical data are expressed as mean ± SD using descriptive statistics.

Results

8226/R5 cells are resistant to R115777-induced growth arrest and cell death. Continuous culture of 8226/S cells with increasing concentrations of R115777 established the 8226/R5 line. In cytotoxicity assays, 8226/R5 cells were at least 50 times more resistant to R115777 compared with parental 8226/S cells (Fig. 1A). The IC_{50} for the 8226/S line was 0.1 μmol/L and that for the 8226/R5 line was 5.4 μmol/L. Cell cycle analysis revealed that R115777 induced G1 growth arrest in 8226/S cells and this effect was largely abolished in the 8226/R5 line (Fig. 1B). 8226/R5 cells were also protected from apoptosis at concentrations of R115777 as high as 20 μmol/L (Fig. 1C; data not shown). The observed resistance was stable.
because cells cultured in R11577-free conditioned media for several months continued to display the resistant phenotype. Relative to doxorubicin- (8226/Dox 40) and melphalan-resistant (8226/LR5) isogenic lines, 8226/R5 cells were the most resistant to R11577 (Fig. 1C).

**Resistance to R11577 does not correlate with K-Ras prenylation or farnesyl transferase activity.** To determine whether 8226/R5 cells were cross-resistant to other compounds that inhibit prenylation, cells were exposed to inhibitors of FTase and geranylgeranyl transferase I (GGTase I). Similar to our results with R115777, 8226/R5 cells were resistant to the specific FTase inhibitor FTI 277 (22) when compared with parental 8226/S cells (Fig. 2A). In addition, resistance was maintained in the presence of pericilic acid, a compound that inhibits both FTase and GGTase I (Fig. 2B; ref. 23). 8226/S cells are known to express K-Ras and harbor a codon 12 K-RAS mutation. Similar to our prior observation in U266 cells (15), K-Ras remained prenylated in both sensitive and resistant cells after R11577 treatment (Fig. 2C). Moreover, farnesylation of HDJ-2 (a protein that can be farnesylated but not geranylgeranylated) was inhibited in both lines (Fig. 2D). These results indicate that R115777 resistance does not correlate with the prenylation status of K-Ras or HDJ-2. Furthermore, the fact that HDJ-2 farnesylation can be inhibited in 8226/R5 cells implies that mutation of FTase (the drug target) is not responsible for the development of resistance.

**Resistance is not related to decreased accumulation of R115777.** Many 8226 lines that acquire drug resistance have elevated expression of P-glycoprotein. An example of this is the 8226/Dox 40 cell line that expresses high levels of membrane P-glycoprotein that participates in the doxorubicin-resistant phenotype (18). As expected, 8226/Dox 40 cells were found to have a marked increase in surface P-glycoprotein expression when compared with parent 8226/S cells (Fig. 3A). 8226/R5 cells, however, had membrane levels that were similar to that noted for the 8226/S line (Fig. 3A). Because increased expression or activity of other membrane pumps may also produce a resistant phenotype, we investigated the influx and efflux of R115777 in both 8226/S and 8226/R5 cells. Influx of R115777 occurred rapidly in both lines and reached steady state within 15 minutes of incubation with radiolabeled R115777 (Fig. 3B). A dose-dependent increase in R115777 uptake was also observed (Fig. 3C) and both time- and concentration-dependent experiments revealed that R115777 uptake was increased in 8226/R5 cells compared with the parental line. With regard to R115777 efflux, intracellular

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**Fig. 3.** Resistance is not related to decreased influx or increased efflux of R115777. **A.** P-glycoprotein surface expression is not increased in 8226/R5 cells. 8226/S, 8226/Dox40, and 8226/R5 cells were stained with anti– P-glycoprotein (dashed line histograms) or isotype control antibody (solid line histograms) as described in Materials and Methods. Samples were analyzed by flow cytometry. Decreased accumulation and/or increased efflux of R115777 is not responsible for resistance in the 8226/R5 line (B-E). B, 8226/S and 8226/R5 cells were exposed to 5 μmol/L R115777 (1:2.5 dilution of [14C]R115777 to cold R115777) in supplemented media for increasing time periods. Cells were washed and accumulation of [14C]R115777 was quantitated by scintillation counting (see Materials and Methods). C, 8226/S and 8226/R5 cells were incubated in supplemented media containing 5 μmol/L R115777 at decreasing dilutions (1:10, 1:2.5, 1:5) of [14C]R115777 for 1 hour. Samples were processed and analyzed as in (B) to confirm a dose-dependent accumulation of [14C]R115777 in each line. D, sensitive and resistant lines were treated with 5 μmol/L R115777 (1:2.5 dilution of [14C]R115777 to cold R115777) for 1 hour, then washed and placed in R115777-free supplemented media for increasing time periods. Samples were processed and analyzed as in (B). E, 8226/S and 8226/R5 cells were treated with 5 μmol/L R115777 at increasing dilutions (1:1, 1:2.5, 1:10) of [14C]R115777 for 1 hour and then washed and placed in R115777 free media for an additional hour. Samples were processed and analyzed as in (B) to confirm a dose-dependent efflux of [14C]R115777. (A), (B), and (D) are representative of two independent experiments; (C) and (E) are representative of three independent experiments.
R115777 decreased rapidly (within 15 minutes) in both sensitive and resistant cells in time course experiments (Fig. 3D). Furthermore, a dose-dependent efflux of R115777 was noted with resistant cells retaining more R115777 compared with sensitive 8226/S cells (Fig. 3E). These results indicate that 8226/R5 resistance is not related to decreased influx or increased efflux of R115777.

8226/R5 cells display a multidrug-resistant phenotype and resistance is not associated with increased expression of heat shock proteins. In an attempt to further categorize resistance, 8226/R5 cells were exposed to agents that induce apoptosis with diverse mechanisms of action. Compounds that have been reported to promote mitochondrial dysfunction, endoplasmic reticulum stress, and nuclear stress were tested (Fig. 4A-E). Surprisingly, 8226/R5 cells were resistant to all evaluated agents when compared with the parent 8226/S line. The proteosome inhibitor PS-341 has been shown to have cytotoxic activity in several chemoresistant myeloma lines (24). In cytotoxicity assays, 8226/R5 cells were three times more resistant to PS-341 when compared with 8226/S cells with an IC₅₀ of 41.8 and 13.2 nmol/L, respectively (Fig. 5A). Unlike our findings with R115777, PS-341 induced G₂-M arrest in both sensitive and resistant cells with 8226/R5 cells being relatively protected (Fig. 5B). 8226/R5 cells were also protected from PS-341–induced apoptosis at concentrations as high as 20 nmol/L (Fig. 5C; data not shown). 8226/R5 cells maintained a PS-341–resistant phenotype when cultured in the absence of R11577 for several months, indicating stable resistance.

Heat shock proteins have antiapoptotic properties that can protect cells from stressful conditions. It has been previously reported that up-regulation of hsp70 can protect ovarian cancer cells from FTI-induced apoptosis (25). In addition, blockade of hsp27 expression has been shown to overcome PS-341 resistance in lymphoma cell lines (26, 27). To determine whether multidrug resistance was related to overexpression of heat shock proteins, sensitive and resistant 8226/S cells were evaluated for heat shock protein expression (Fig. 4F). Expression levels of hsp70 and hsp90 were similar in sensitive and resistant cells and treatment with R115777 (drug-induced stress) had no appreciable effect. Hsp27 expression was also not effected in 8226/S cells after R115777 treatment but was undetectable in the 8226/R5 line. These results indicate that resistance does not correlate with the expression of heat shock proteins hsp27, hsp70, and hsp90.

Transcriptional profile of 8226/R5 cells. To identify potential targets of R115777 and/or molecules associated with R115777 resistance, we compared gene expression profiles between sensitive and resistant variants of 8226 cells. Direct comparisons were made between the 8226/S and 8226/R5 cell lines. In an attempt to reduce the number of nonspecific gene expression changes identified, comparisons were also made between 8226/LR5 (which possessed an intermediate degree of resistance to R115777; Fig. 1C) and 8226/R5 cells. This approach produced 2,064 probe sets (representing 1,666 genes) with changes unique to the 8226/R5 line. A selected list of genes involved in cell survival and growth, maintenance of cell cytostructure, cell-microenvironment contact, cholesterol biosynthesis, and protein degradation are presented in Table 1.

Discussion

FTIs were designed as specific inhibitors of Ras intended to interfere with a crucial posttranslational processing step. Ras requires the addition of a 15-carbon farnesyl group to its carboxyl-terminal cysteine, which permits its localization to the plasma membrane (a requirement for function). This
modification is done by the enzyme FTase, the purported target of R115777. It has been well established that alternate prenylation of Ras may occur in the presence of FTIs (28). This has been most notably described for K-Ras but also applies to the N-Ras protein. These reports are consistent with our prior observation in U266 cells (N-Ras–expressing line; ref. 15) and our present finding that K-Ras remains prenylated in 8226/S cells after R115777 treatment. These results suggest a Ras-independent mechanism of cell death. This is further supported by the clinical observation that responses to R115777 do not correlate with Ras mutation status or inhibition of farnesyl transferase measured ex vivo (7, 10, 14).

We hypothesized that the isolation of a R115777-resistant human myeloma cell line (8226/R5) might provide insight concerning the potential targets of R115777 or identify novel mechanisms of FTI resistance. R115777 resistance was not associated with an increase in the surface expression of P-glycoprotein nor was it associated with decreased influx or increased efflux of R115777. In addition, the expression of heat shock proteins (hsp27, hsp70, hsp90) did not correlate with the drug-resistant phenotype. We therefore undertook molecular profiling of sensitive and resistant 8226 lines and identified expression changes in several genes implicated in cell survival and drug resistance. They included genes involved in cell signaling, cholesterol biosynthesis, and protein degradation. It remains possible that molecules associated with one of these genes is a major target of R115777 and/or directly participates in FTI resistance.

Our microarray data identified increased expression of several proteins associated with Jak-Stat signaling, including Jak2. In primary isolates, constitutive Stat3 activation has been observed in the majority of patients with multiple myeloma (29, 30). Stat3 homodimers and Stat1:Stat3 heterodimers seem to be the predominant DNA-binding forms (29). It has been reported that inhibition of Stat3 can sensitize resistant myeloma cells to chemotherapy mediated apoptosis (31). These results imply that Jak-Stat signaling may contribute to the drug-resistant phenotype. Stat3 activity has been linked to up-regulation of Bcl-XL (29) and consistent with this, Bcl-XL expression was increased in 8226/R5 cells. Elevated expression of Bcl-XL has been observed in several hematopoietic tumors, including blast crisis chronic myelogenous leukemia and non-Hodgkin’s lymphoma (32), and one study suggested that it was an indicator of chemoresistance in multiple myeloma (33). We have previously reported that R115777 can partially overcome drug resistance in U266 cells that maintain high levels of stable Bcl-XL expression (15). These findings imply that R115777 resistance may only in part be regulated by increased Jak-Stat signaling or Bcl-XL expression in 8226/R5 cells.

Our comparative gene expression profiling also identified other genes potentially involved in R115777 resistance. Phosphatidylinositol 3-kinase is a heterodimer consisting of p85 and p110 subunits (34, 35). The p110 α isoform is preferentially expressed in cells of hematopoietic origin (36, 37) and elevated expression of the catalytic α polypeptide was observed in the 8226/R5 line. An important downstream target of phosphatidylinositol 3-kinase is Akt/protein kinase B, a protein that is known to play a role in myeloma survival and drug resistance (38–40). Interestingly, it has been reported that FTI-induced apoptosis can be prevented by a constitutively activated form of Akt-2 (41). Therefore, it remains possible that phosphatidylinositol 3-kinase/Akt signaling also contributes to R115777 resistance and the multidrug-resistant phenotype. Because the p110 α subunit is mainly expressed in hematopoietic cells, it potentially represents a novel therapeutic target particularly for resistant tumors of hematopoietic origin.

Our analysis also identified increased expression of mevalonate kinase, an enzyme associated with cholesterol biosynthesis. Influx and efflux experiments suggest that R115777 is retained in 8226/R5 cells compared with the parent 8266/S line. This is relevant because an increase in cholesterol-rich microdomains, such as lipid rafts, have been associated with the development of multidrug resistance (42). R115777 is a lipophilic molecule that could potentially be...
myeloma cells (26). In 8226/R5 cells, however, hsp27 protein expression was undetectable. Our microarray analysis revealed decreased expression of the 26S proteosome subunit (the target of PS-341) in 8226/R5 cells when compared with the parental 8226/S line. This perhaps represents a novel mechanism of proteosome inhibitor resistance.

In conclusion, we have identified a myeloma cell line with resistance to both R115777 and PS-341. Further characterization of this line may lead to identification of novel drug targets or resistance mechanisms that are clinically relevant.

Table 1. Gene expression changes in 8226/R5 cells

<table>
<thead>
<tr>
<th>Expression increased</th>
<th>Expression decreased</th>
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<tbody>
<tr>
<td>Janus kinase 2</td>
<td>Interleukin-6 receptor</td>
</tr>
<tr>
<td>Signal transducer and activator of transcription 1</td>
<td>Insulin-like growth factor 2 receptor</td>
</tr>
<tr>
<td>Bcl-XL</td>
<td>Mcl-1</td>
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<tr>
<td>Phosphoinositide-3-kinase, catalytic, a polypeptide</td>
<td>Heat shock protein 27</td>
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<tr>
<td>Fibroblast growth factor receptor activating protein 1</td>
<td>Retinoblastoma 1</td>
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<td>Transforming growth factor c</td>
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<td>Tumor protein p53</td>
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<td>Farnesyl diposphatase synthetase</td>
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<td>Rab GGTTase, b subunit</td>
<td>Lamina A/C</td>
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<tr>
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<td>Ras protein activator like-1</td>
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<td>Ras and Rab interactor 2</td>
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<td>Mitogen-activated protein kinase phosphatase</td>
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<td>Cyclin D1</td>
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<tr>
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<td>Syndecan 1</td>
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<td>Actin, b</td>
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<td>Ubiquitin-specific proteases 15</td>
<td>Proteosome 26S subunit</td>
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<td>Ubiquitin-specific proteases 24</td>
<td>S100 calcium binding protein (A11 pseudogene)</td>
</tr>
<tr>
<td>Proteosome inhibitor subunit 1</td>
<td>S100 calcium-binding protein (A11 calgizzarin)</td>
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References


30. Bharti AC, Sheshodia S, Reuben JM, et al. Nuclear factor-κB and STAT3 are constitutively active in CD138+ cells derived from multiple myeloma patients, and suppression of these transcription factors leads to apoptosis. Blood 2004;103:3175–84.


