The FLIP-Side of Fas Signaling
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Fas (CD95/APO-1) is a 45-kDa type I transmembrane protein belonging to the tumor necrosis factor superfamily of receptors (1, 2). Fas was identified in 1989 as a cell death inducer of malignant human cancer and leukemia cell lines (3, 4). Fas contains a classic “death domain” within its cytosolic tail, typical of a branch of the tumor necrosis factor family implicated in apoptosis induction. Indeed, for many tumor cell lines, cross-linking Fas with agonistic anti-Fas antibodies leads to apoptosis (5, 6). Nonapoptotic roles for Fas have also been found, however, including stimulating proliferation of several types of normal cells under some conditions, including T cells, thyomocytes (7), fibroblasts (8–10), and hepatocytes (11). This dual role of Fas complicates the biology of this tumor necrosis factor family receptor from the standpoint of predicting the role of Fas in tumor biology. An article in a previous issue of Clinical Cancer Research by Mitsiades et al. (12) illustrates the conundrum of Fas by showing a proliferative rather than apoptotic role for this “death receptor” in thyroid carcinoma cell lines and also providing some evidence that Fas expression is associated with more aggressive thyroid cancers.

The findings of Mitsiades et al. beg the question of how cross-linking Fas with an antibody (specifically CH-11) that is well known for its ability to induce apoptosis instead triggers proliferation of thyroid cancer cell lines. This is the first report to show that Fas can induce cell proliferation in thyroid cancer cells, but similar observations have been made previously for epithelial cancer, melanoma, myeloma cell lines, and primary chronic lymphocytic leukemia B cells (13–15). What makes these malignancies different from the many other tumor cell types where CH-11 anti-Fas antibody triggers apoptosis?

Previous work by Mitsiades et al. has shown that Fas stimulation failed to induce apoptosis of papillary thyroid carcinoma cell lines (16). In the current report, Mitsiades et al. (12) expand their analysis and show that Fas resistance does not seem to result from diminished Fas or Fas-associated death domain protein (FADD) expression. FADD is a unique cytosolic adapter protein that contains a death domain and a death effector domain (DED) and is an integral component of the death-inducing signaling complex formed upon activation of Fas (see Fig. 1). The death-inducing signaling complex is comprised of Fas, FADD, the DED-containing cell death proteases caspase-8 and/or caspase-10 (which bind the DED of FADD). Mitsiades et al. (12) used immunoblotting and immunohistochemistry methods to show the presence of Fas and FADD expression in thyroid cancer cell lines and tissue specimens. Furthermore, the investigators found that the death domain of Fas was free of mutations, unlike some malignancies (17, 18), excluding this as a possible explanation for the Fas-resistant phenotype. Despite apparent integrity of Fas, FADD, and procaspase-8 expression, these investigators previously showed that treatment of thyroid cancer cell lines with agonistic antibody does not induce recruitment of procaspase-8 to Fas (16), showing a defect in death-inducing signaling complex assembly.

An important clue to the mechanisms modulating Fas activity in thyroid cancer cell lines was uncovered when Mitsiades et al. found the protein synthesis inhibitor cycloheximide sensitized these cells to Fas-induced cell death, suggesting the presence of a short-lived inhibitory protein. One logical candidate for such an inhibitor is the Flice-like inhibitory protein (FLIP), a DED-containing antiapoptotic protein that associates with caspase-8 and caspase-10 at the death-inducing signaling complex and is known to have a short half-life (19–21). The flip gene encodes two principal proteins, a more abundant longer isoform (FLIP-L) that contains tandem DEDs resembling the prodomains of caspase-8 and caspase-10 followed by a caspase-like domain and a less prevalent shorter protein (FLIP-S) composed of only the DEDs. Although FLIP-S is exclusively a caspase inhibitor, the longer FLIP-L has a dual function as either a caspase inhibitor or activator, depending on a variety of issues including the ratio of FLIP-L to caspase-8 (22). The investigators show that agonistic anti-Fas antibody CH-11 induces FLIP-L association with Fas in thyroid cancers, using coimmunoprecipitation experiments. Consistent with an antiapoptotic role for FLIP in these thyroid cancer cell lines, FLIP antisense oligodeoxynucleotides sensitized to Fas-mediated apoptosis, thereby implicating FLIP-L in the Fas resistance mechanism. These findings are enlightening, given that FLIP-L can display either proapoptotic or antiapoptotic phenotypes, depending on the cellular context (22–24).

But, what about the proliferative actions of Fas in thyroid cancers? Does FLIP explain that phenomenon? It has been suggested that FLIP may provide the molecular switch allowing Fas to promote cell proliferation as opposed to apoptosis (reviewed by refs. 25, 26). For example, flip−/− murine T cells are defective in proliferation induced by stimulation through the T-cell antigen receptor complex (27, 28). Although FLIP antisense was interrogated for its effects on apoptosis, Mitsiades et al. were unable to explore whether experimentally reducing FLIP expression alters proliferation due to induction of apoptosis.

In addition to proliferation, the investigators observed that Fas induces signal transduction events, such as mitogen-activated protein/extracellular signal-regulated kinase kinase and extracellular signal-regulated kinase phosphorylation and nuclear factor-κB (NF-κB) and activator protein 1 activation in thyroid cancer cells. In this regard, it has been shown previously that FLIP can induce activation of extracellular signal-regulated

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kinases (apparently by binding the upstream kinase Raf-1), at least when overexpressed (29), raising the possibility of FLIP involvement in this Fas-induced signaling event observed in thyroid cancers. FLIP also reportedly activates NF-κB when overexpressed in HEK293 cells, and the two DED domains are sufficient for the activation (30). The role of FLIP in NF-κB activation, however, is complicated. When FLIP-L is overexpressed in Jurkat or HEK293 cells, and those cells are then stimulated with Fas ligand, NF-κB activation is suppressed (31, 32). Moreover, reducing FLIP using small interfering RNA also enhances Fas ligand–induced apoptosis in the human epidermal KB cell line rather than inhibiting it (31). Similarly, in endothelial cells, whereas gene transfer–mediated increases in FLIP protected against apoptosis, they suppressed NF-κB activation induced by lipopolysaccharide. Conversely, reducing FLIP by small interfering RNA in endothelial cells increased lipopolysaccharide-induced NF-κB activity (33). Thus, FLIP seems to be a NF-κB enhancer in some contexts and an inhibitor in others.

One possible explanation for this dual phenotype can be envisioned if FLIP-L participates in a multiprotein complex involving three or more proteins that engender NF-κB activation, such that the proper stoichiometry of the individual subunits is achieved only if the right amount of FLIP protein is present relative to the other components. In that case, either too much or too little FLIP-L would result in diminished signaling. Hints of a candidate complex that might link FLIP-L to proliferation are found in observations from knock-out mice that ablation of the genes encoding FADD, caspase-8, and FLIP phenocopies each other, and that all three of these proteins are required for T-cell proliferation, and all three of these proteins interact (reviewed by ref. 19). Thus, like FLIP-L, other components of the death-inducing signaling complex (i.e., FADD and caspase-8) lead surprising dual lives in both apoptosis regulation and signal transduction, apparently inducing apoptosis in some cellular contexts and proliferation in others.

Precisely how do FADD, caspase-8, and FLIP-L all collaborate in inducing NF-κB and other signaling events that promote cell proliferation? Many details remain unclear, but data collected from several prior studies suggest a model in which FADD recruits both caspase-8 and FLIP-L (presumably as a heterodimer), resulting in activation of caspase-8 and proteolytic cleavage of both FLIP and caspase-8. The cleavage of caspase-8 in
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