The cyclin-dependent kinase inhibitor p21 (G1P1, WAF1, CDKN1A) is thought to be the main executor of p53-induced growth arrest (1–4). Transcriptionally induced by p53, the p21 protein binds and inhibits the cdk2-cyclin B/E complexes but increases stability and nuclear localization of cyclin D, stimulating assembly of the cdk4-cyclin D complex (3, 5–13). Interestingly, a second p53-induced transcript from the p21 gene, p21B (Fig. 1) was recently identified and found to encode a protein expressing proapoptotic activity (14), suggesting a role for the p21 gene in both cell cycle arrest and apoptosis.

We recently described p21/p21B status in 73 locally advanced breast cancers (15), observing no promoter methylation or somatic mutations. We did, however, observe two novel polymorphisms, p21BT35C (Leu12Pro), in addition to the previously known p21BT35C (Ser31Arg). Although several groups have reported an increased frequency of the p21G251A polymorphism in cancer patients (16–20), others reported a similar distribution of the polymorphism in both cancer and healthy individuals (21–25). The frequencies of patients harboring the A allele of p21G251A or the C allele of p21BT35C were 8.2% and 16.4%, respectively. Although we are not aware of other analyses of the p21B coding sequence, the fact that the p21G251A polymorphism had not been reported previously made us hypothesize that it may occur at low frequency in non–breast cancer individuals. Analyzing a large set of healthy individuals and breast cancer patients, we here report an increased frequency of the p21G251A polymorphism among breast cancer patients. Most interestingly, this
increased frequency was limited to the subgroup of breast cancer patients diagnosed with locally advanced tumors.

**Materials and Methods**

**Patients.** In this study, we analyzed samples from several cohorts of patients and healthy controls. Ninety-one and 35 patients all suffering from locally advanced breast cancer enrolled in prospective studies exploring resistance to treatment with doxorubicin (26) or mitomycin and 5-fluorouracil (27), respectively, were included. In addition, we analyzed an extra group of six patients with locally advanced breast cancer that were excluded from the mitomycin and 5-fluorouracil study for reasons other than tumor status and 60 patients from a prospective study regarding resistance to paclitaxel. Seventy-three of the locally advanced breast cancers were analyzed in our previous work on p21 (15). In addition, we analyzed tumor samples from 201 patients suffering from metastatic breast cancer and blood DNA from 170 breast cancer patients (132 stage I-II and 38 stage III) collected at the Norwegian Radiumhospital during the period 1984 to 1990 under an institutional review board–approved protocol.

As controls, we analyzed blood DNA from a total of 1,017 healthy postmenopausal Norwegian females recruited from the national mammographic program into other studies described elsewhere (28, 29).

**RNA purification.** RNA was purified by Trizol (Life Technologies, Gaithersburg, MD) extraction from snap-frozen tissue samples as per the manufacturer’s instructions. After extraction, the RNA was dissolved in 100 μL DEPC-treated double-distilled water. CDNA was synthesized by reverse transcription-PCR from 4 μL RNA solution using either Superscript II reverse transcriptase (Invitrogen, Carlsbad, CA) or Transcriptor reverse transcriptase (Roche, Indianapolis, IN).

**DNA purification.** Genomic DNA was prepared from lymphocytes using chloroform/phenol extraction followed by ethanol precipitation (Nucleic Acid Extractor 340A; Applied Biosystems, Foster City, CA) according to standard procedure for the samples from the Norwegian Radiumhospital, or by QIAamp DNA Mini kit (Qiagen, Chatsworth, CA) according to the manufacturer’s instructions. Before the latter procedure, lymphocytes were prepared by cultivating 0.4 mL blood at 37°C for 3 days in 10 mL Medium 199 (Life Technologies) containing 18% FCS (Life Technologies).

**Genotyping of p21 and p21B.** The p21 gene was analyzed on cDNA regarding patients from the doxorubicin, mitomycin and 5-fluorouracil, paclitaxel, and metastatic protocols. Results were verified on genomic DNA from blood when available. The remaining patients and all controls were analyzed on DNA from blood. Amplification of p21 and p21B, both from cDNA and genomic DNA, were carried out as described earlier (15).

One microliter of the PCR product was used as template for sequencing, and the sequencing reaction was done with the Big Dye terminator mix (Applied Biosystems), as described (15). Primers for the sequencing reactions were the same as those used for PCR amplification. Capillary gel electrophoresis, data collection, and sequence analysis were done on an automated DNA sequencer (ABI 3700).

**Haplotype analysis.** Haplotypes were first constructed using the program PHASE 2.1 (30, 31). PHASE 2.1 was also used to perform permutation testing to determine if there was a difference in the distribution of haplotypes between cases and controls. The permutation test examines the null hypothesis that the case and control haplotypes are a random sample from a single set of haplotype frequencies versus the alternative hypothesis that cases are more similar to other cases than to control haplotypes. The significance was calculated giving the P for a permutation test of the null hypothesis. The algorithm was run on 10 permuted data sets for 10 separate times to check for consistency. The final run lasted 10 times longer than the other runs to increase the number of interactions. The final output was the run with the best average value for the goodness of fit variables. The extent of linkage...
frequencies of the p21C93A sequence in 100 healthy controls (29) indicated that the allele p21 (Fig. 1). Preliminary screening of the (17). Similarly, all patients in our study carrying the p21 downstream of the samples. However, we found the A allele of the novel polymorphism that were analyzed for the region encompassing this association would remain statistically significant. Fisher’s exact test). Based on these preliminary findings, we used to ensure that the genotypes are in Hardy-Weinberg equilibrium, to exclude technical genotyping errors.

Statistical analysis. Each 2 × 2 test was done using the Fisher’s exact test. When three or more groups were compared, analysis was done by the chi-square test. All computations were done using the SAS System (SAS Institute, Inc., Cary, SC); Ps reported are two – sided.

Results

In our previous study analyzing locally advanced breast cancers (15), we reported one known (p21C515T) and two novel (p21G251A and p21B74G) polymorphisms of the p21 gene (Fig. 1). Preliminary screening of the p21 and p21B coding sequence in 100 healthy controls (29) indicated that the allele frequencies of the p21C93A and the p21B74G polymorphisms were equally distributed in normal individuals and tumor samples. However, we found the A allele of the novel p21G251A polymorphism in only 1 out of 100 normal samples compared with 6 of 73 individuals suffering from breast cancer (P = 0.043, Fisher’s exact test). Based on these preliminary findings, we increased the number of patients and controls to test whether this association would remain statistically significant.

In addition to the three polymorphisms reported in our original study (15), we observed two other p21 polymorphisms (Fig. 1). The polymorphism p21C515T is located 20 bp downstream of the p21 stop codon and rarely analyzed for but is always reported together with the p21C93A polymorphism (17). Similarly, all patients in our study carrying the p21C93A polymorphism that were analyzed for the region encompassing nucleotide 515 (n = 27) also carried the p21C515T polymorphism. The G allele of the previously reported p21T74G (Val25Gly) polymorphism (35) was recorded once, in a patient suffering from locally advanced breast cancer (Fig. 1).

Our observations on p21C93A, p21G251A, and p21B74G are summarized in Tables 1, 2 and 3 respectively. Homozygosity for the rare allele is infrequent (Tables 1-3), and all further statistical analyses thus concern wild-type individuals versus the combined groups of heterozygous and homozygous for the rare allele. The p21C93A and p21B74G polymorphisms occur at similar frequencies among healthy controls and cancer patients. (Frequency of presence of the rare allele was 9-11% for each polymorphism in each group; Tables 1 and 3). p21G251A is more infrequent, present in only in 1.9% of the samples (Table 2). Although presence of the rare allele is observed more frequently in patients (2.7%) than in controls (1.0 %), the difference is not statistically significant (P = 0.076, Fisher’s exact test). Dividing our patients into two subgroups based on tumor stage, we found the p21G251A variant in 4.5% of our locally advanced breast cancers versus 1.5% among individuals diagnosed with stage I and II tumors (P = 0.0526, Fisher’s exact test). Comparing the frequency of p21G251A among healthy controls, stage I/II patients, and locally advanced patients, a clear difference between the three groups was detected (P = 0.007, \( \chi^2 \)). Similarly, comparing stage III cancer patients with either stage I/II patients and healthy controls together, or healthy controls alone, we found a statistically higher frequency of carriers of the A allele of the p21G251A polymorphism among stage III patients (P = 0.0049 and P = 0.0057, respectively, Fisher’s exact test).

The haplotype structure, considering all the five coding polymorphism in the p21, gene was determined. Overall, six different haplotypes were observed for the combined cases and controls data set. PHASE calculated six different haplotypes for the case group and four for the controls. Two infrequent haplotypes were found in the cases only, whereas no control-specific haplotypes were observed. A case-control permutation test did not show a significantly different distribution of haplotypes in cases compared with controls (P = 0.47).

<table>
<thead>
<tr>
<th>Table 1. Observations of p21C93A</th>
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<tbody>
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<td><strong>No. patients analyzed</strong></td>
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</tr>
<tr>
<td>Healthy controls</td>
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<tr>
<td>Stage I/II breast cancers</td>
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<table>
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<th>Table 2. Observations of p21G251A</th>
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Discussion

In this article, we report the presence of previously identified as well as novel polymorphisms of p21/p21B among 474 Norwegian breast cancer patients and 1,017 healthy individuals.

The polymorphisms are rarely found together (with the exception of the linkage between p21C93A and p21C515T), and when two are observed together, they are always heterozygous. This indicates that no haplotype containing more than one of the polymorphism p21T74G, p21G251A, p21B T35C; or both p21C93A and p21C515T are present among patients and controls analyzed. This constitutes five of the six PHASE reported haplotypes (including the haplotype wild type in all positions). The final haplotype predicted by PHASE is disregarded, as it results from estimation of status for one of the polymorphisms, and thus may not be counted among our observed haplotypes. We here refer to the haplotypes as wild type or by the polymorphism defining the haplotype, as no haplotype carries two or more of the p21T74G, p21G251A, p21B T35C, or p21C93A polymorphisms.

Expanding on our previous results (15), we observe the novel p21 polymorphism, p21B T35C, as well as the previously defined p21C93A, to occur with a similar frequency among breast cancer patients and non-breast cancer individuals. Although the p21G251A polymorphism was observed more frequently among breast cancer patients compared with controls, this difference did not reach statistical significance. In contrast, we found this polymorphism to be significantly associated with stage III (locally advanced) breast cancer. Although locally advanced breast cancer seems to present cDNA microarray profiles resembling the profile of breast tumors in general (36, 37), recent studies suggest a potential difference in biology between breast cancer seems to present cDNA microarray profiles resembling the profile of breast tumors in general (36, 37), recent studies suggest a potential difference in biology between breast cancer and BRCA2 carriers in general reveal a biology resembling the biology of tumors in patients with no germ line mutation, the opposite is the case for tumors among BRCA1 carriers (41–43).

In conclusion, we find the novel p21G251A polymorphism to be associated not to breast cancers in general but to a subgroup of the disease characterized by aggressive growth.

References

7. Gu Y, Turk CW, Morgan DO. Inhibition of Cdk2

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Table 3. Observations of p21B T35C

<table>
<thead>
<tr>
<th>No. patients analyzed</th>
<th>No. observations</th>
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<td>Locally advanced breast cancers</td>
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<td>Homozygous (TT) 191</td>
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The Novel $p21$ Polymorphism $p21^{G251A}$ Is Associated with Locally Advanced Breast Cancer

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