Chordomas are uncommon, slow-growing, malignant neoplasms that are thought to originate from notochordal remnants along the axial skeleton in the sacrococcygeal/sacral, sphenoccipital/clivus, and spinal regions (1). Chordomas are usually sporadic, but a few families have been reported with multiple family members affected by chordoma (2–6), suggesting that inherited variations may predispose to these tumors. Intriguingly, several investigations identified chordomas in tuberous sclerosis complex (TSC) patients (7–11). We previously reported two cases of TSC with concomitant sacrococcygeal chordomas and identified somatic inactivation of the TSC genes, thus providing the first evidence of a pathogenic role of the TSC genes in sacrococcygeal chordomas (12).

TSC is a tumor suppressor syndrome characterized by abnormal tissue growths, known as hamartomas, in many organs (13). TSC is caused by inactivating mutations in tumor suppressor genes TSC1 or TSC2, which encode hamartin (TSC1) and tuberin (TSC2), respectively. Of note, somatic mutations in these genes are also found in sporadic lymphangiomyomatosis (14, 15). TSC proteins form a heterodimeric TSC1-TSC2 complex and function to inhibit mTORC1 signaling, which critically regulates cell growth and proliferation (16, 17). Activation of the phosphoinositide-3-kinase (PI3K)/Akt or Ras/mitogen-activated protein kinase (MAPK) pathway by growth factors leads to TSC2 phosphorylation and inhibition of the TSC protein complex by Akt, extracellular signal-regulated kinase (ERK), and p90 ribosomal S6 kinase 1 (RSK1) kinases (Fig. 1A). Another important negative regulator of the Akt/mTOR pathway is PTEN (phosphatase and tensin homologue deleted on chromosome ten), a lipid phosphatase that decreases the effective concentration of phosphatidylinositol 3,4,5-triphosphate in cells. In cells lacking TSC1, TSC2, or PTEN, mTORC1 is hyperactivated, resulting in constitutive phosphorylation of S6K, S6, and 4EBP1, which is potently inhibited by the mTOR inhibitor rapamycin. Importantly, the TSC/mTORC1 pathway is dysregulated in several hamartoma syndromes as well as in many cancers (16–18).
Hyperactivated mTORC1 signaling in Sporadic Chordomas

Translational Relevance

Inhibitors of mammalian target of rapamycin complex 1 (mTORC1), such as rapamycin and its analogues, are currently being tested in clinical trials for tuberous sclerosis complex as well as many human cancers, which display hyperactivated mTORC1 signaling. mTORC1 has emerged as a critical integrator of signals from growth factor, nutrient, oxygen, and energy to regulate cell growth and proliferation. This study shows for the first time that mTORC1 signaling is aberrantly hyperactivated in primary chordoma tumors/cell lines, and PTEN (phosphatase and tensin homologue deleted on chromosome ten) deficiency may be frequently associated with sporadic chordomas. Furthermore, we show that the mTOR inhibitor rapamycin suppresses mTORC1 signaling and proliferation of chordoma-derived cell line. Therefore, this study not only reveals the pathogenic mechanisms of chordomas, but also provides a rationale for initiating clinical trials of Akt/mTORC1 inhibition in patients with sporadic chordomas.

Based on the roles of the TSC genes in the pathogenesis of TSC-associated chordomas, we hypothesized that dysregulation in the TSC/mTORC1 pathway may be associated with sporadic chordoma. Here, we show that aberrant hyperactivation of the Akt/mTORC1 pathway is commonly found in sporadic sacrococcygeal chordomas, and rapamycin treatment strongly inhibits mTOR activation and proliferation of chordoma-derived U-CH1 cells. In addition, we have observed either reduced or lack of expression of PTEN in many of the sporadic sacral chordomas examined, which may explain the activation of Akt/mTORC1 signaling. These findings suggest the potential efficacy of Akt/mTOR inhibitors as a possible therapeutic approach for this malignant tumor.

Materials and Methods

Tissue samples. Ten cases of sporadic chordoma and one case of TSC-associated chordoma tumors were available through the Department of Pathology at Massachusetts General Hospital under the approval of its Institutional Review Board.

Cell culture, antibodies, and reagents. The human sacrococcygeal chordoma-derived U-CH1 cell line was maintained as described with minor modification (19). U-CH1 cells were grown in plates coated with 0.005% collagen (Sigma) in a 4:1 mixture of Iscove’s modified Dulbecco’s medium (Invitrogen) and RPMI-1640 (Sigma) media containing 10% fetal bovine serum, 2 mmol/L glutamine, 100 U/mL penicillin G, and 100 µg/mL streptomycin. Human prostate tumor cell lines PC3 and DU145, human embryonic kidney 293, and human chordoma-derived Ch1 cell lines were grown in DMEM (Invitrogen) containing 10% fetal bovine serum, 2 mmol/L glutamine, 100 U/mL penicillin G, and 100 µg/mL streptomycin. All antibodies used in this study, except for anti-TSC2 (20), anti-GAPDH (Millipore), and anti-pPRAS40 (Invitrogen), were purchased from Cell Signaling Technology. Wortmannin and DMSO (Sigma), rapamycin (Calbiochem), and Dulbecco’s PBS (Invitrogen) containing glucose and pyruvate were also utilized.

Western analysis. Preparation of cell lysates and Western analysis were done as described previously (20).

Immunohistochemistry. Human tissues were fixed in formaldehyde and embedded in paraffin. Immunohistochemistry was carried out using anti-PTEN (1:100), anti-pPRAS40 (T246, 1:750), anti-pS6 (S240/244, 1:100), and anti-p4EBP1 (T37/46, 1:100) antibodies, employing methods described earlier (21).

Proliferation assay of U-CH1 cells. On day 0, U-CH1 cells were plated at a density of 2 × 10^4 cells per well in a 24-well plate. Each experimental group was carried out in triplicate. On days 1, 3, 5, and 7, cells were fed with fresh growth media containing vehicle DMSO (0.1%) or rapamycin (1, 5, 10, 25, and 50 nmol/L). On days 3, 6, and 9, cells were collected by trypsin treatment, and cell numbers were determined using a hemacytometer.

For 5-bromo-2-deoxyuridine (BrdUrd) incorporation to detect S-phase cells, U-CH1 cells were plated in collagen-coated coverslips, treated with DMSO or varying concentration of rapamycin as described above, and incubated with 10 µmol/L BrdUrd for the final 24 h. BrdUrd incorporation was detected using a fluorescein-conjugated anti-BrdUrd antibody.
antibody in In Situ Cell Proliferation Kit, FLUOS (Roche). BrdUrd-positive cells were counted in 5 randomly chosen fields per condition using a TCS SP5 confocal microscopy (Leica) and the percentage of BrdUrd-positive cells was calculated.

Statistical analysis. For comparisons of cell numbers in vehicle versus rapamycin-treated groups, a paired two-tailed Student’s t-test was used.

Results

mTORC1 signaling is activated in U-CH1 cells. We first examined mTORC1 signaling in the sacral chordoma-derived U-CH1 cell line, which has nearly identical immunohistochemical, morphologic, and cytogenetic aberrations found in its parental primary sacral tumor (19). We tested whether mTORC1 signaling in U-CH1 cells is constitutively active in the absence of growth factors or amino acids, by monitoring the level of phosphorylated S6 (Ser235/236), which serves as a readout for mTORC1 signaling. The human prostate tumor–derived, PTEN-negative PC3 and PTEN-positive DU145 cells were employed as positive and negative controls for mTORC1 signaling, respectively (21, 22). S6 phosphorylation in both U-CH1 and another chordoma-derived Ch1 cell line was persistently elevated, even under serum-starved conditions, in the same way as in PC3 cells, whereas S6 phosphorylation was significantly reduced in serum-starved 293 and DU145 cells (Fig. 1B). In contrast to serum deprivation, amino acid depletion for 1 hour resulted in complete termination of S6 phosphorylation in U-CH1 cells, which was also observed in amino acid–starved 293 cells (Fig. 1C). These results indicate that mTORC1 signaling in chordoma-derived cell lines is deregulated in response to growth factor deprivation, but remains sensitive to amino acid availability.

mTORC1 signaling is abnormally hyperactivated in sporadic chordomas. We next examined mTORC1 signaling in 10 sporadic sacral chordoma tumors and one TSC-associated chordoma by immunohistochemical staining to detect the levels of pS6 (S240/244) and p4EBP1 (T37/46) proteins, which serve as readouts for mTORC1 signaling. Previously, we reported that TSC-associated angiomyolipomas, tubers, and subependymal giant cell astrocytomas displayed highly elevated levels of mTORC1 signaling (21). Consistent with these results, the TSC-associated chordoma stained strongly positive for pS6 (S240/244; Fig. 2A). Strikingly, all 10 sacral chordomas examined in this study also exhibited varying degrees of positivity for pS6 (S240/244), compared with nonneoplastic neighboring normal cells (Fig. 2B and C and Table 1). Furthermore, all the tumors also showed high immunoreactivity to p4EBP1 (T37/46) antibody (Fig. 2 and Table 1). These results indicate that mTORC1 signaling is hyperactivated in sporadic sacral chordomas.

Constitutively activated Akt signaling and PTEN loss in U-CH1 cells. To determine the mechanism of constitutively elevated mTORC1 signaling in U-CH1 cells, we examined Akt/TSC and MAPK signaling pathways, which are well-known upstream regulators of mTORC1 signaling (Fig. 1A). Activated Akt, ERK, and RSK1 kinases phosphorylate and inactivate the tumor suppressor TSC2, thus relieving its inhibitory role on mTORC1. Akt also phosphorylates and inactivates a novel mTOR-binding partner PRAS40 (proline-rich Akt substrate 40 kDa), a negative regulator of mTORC1 (Fig. 1; ref. 23). Similar to PC3 cells, serum-starved U-CH1 cells exhibited high levels of pAkt,

<table>
<thead>
<tr>
<th>Case no.</th>
<th>pS6</th>
<th>p4EBP1</th>
<th>PTEN</th>
<th>pPRAS40</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>++</td>
<td>++</td>
<td>-</td>
<td>++</td>
</tr>
<tr>
<td>2</td>
<td>+++</td>
<td>++</td>
<td>-</td>
<td>+++</td>
</tr>
<tr>
<td>3</td>
<td>+++</td>
<td>+++</td>
<td>+</td>
<td>++</td>
</tr>
<tr>
<td>4</td>
<td>+++</td>
<td>+++</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>++</td>
<td>++</td>
<td>+</td>
<td>++</td>
</tr>
<tr>
<td>6</td>
<td>+++</td>
<td>+</td>
<td>-</td>
<td>+++</td>
</tr>
<tr>
<td>7</td>
<td>+++</td>
<td>+</td>
<td>-</td>
<td>++</td>
</tr>
<tr>
<td>8</td>
<td>+++</td>
<td>+</td>
<td>+</td>
<td>ND</td>
</tr>
<tr>
<td>9</td>
<td>+++</td>
<td>+</td>
<td>-</td>
<td>ND</td>
</tr>
<tr>
<td>10</td>
<td>++</td>
<td>+</td>
<td>-</td>
<td>ND</td>
</tr>
</tbody>
</table>

Table 1. Summary of immunohistochemical staining in 10 cases of sporadic chordomas and one case of TSC-associated chordoma

NOTE: Two independent observers blindly scored staining semi-quantitatively: -, negative; +, weak; ++, medium; ++++, strong; ND, not determined.
pTSC2, and pPRAS40, compared with serum-starved DU145 cells (Fig. 3A). These results indicate that Akt signaling in U-CH1 cells is also constitutively activated in a growth factor–independent manner. Interestingly, ERK phosphorylation was also high in serum-starved U-CH1 cells.

We next examined expression of the tumor suppressor PTEN, which is a negative regulator of Akt signaling. Strikingly, PTEN expression was not observed in U-CH1 cells and significantly reduced in Ch1 cells. PTEN-negative PC3 and PTEN-positive DU145 cells were used as negative and positive controls for PTEN expression.

**Fig. 3.** Constitutively activated Akt signaling and PTEN deficiency in chordoma-derived cell lines and sporadic sacral chordoma tumors. **A,** constitutive phosphorylation of Akt, TSC2, PRAS40, and ERK in U-CH1 cells. Cells were serum-starved for 20 h before lysates were prepared for Western analysis. **B,** PTEN expression was not observed in U-CH1 cells and significantly reduced in Ch1 cells. PTEN-negative PC3 and PTEN-positive DU145 cells were used as negative and positive controls for PTEN expression. **C and D,** representative images of two sporadic chordomas showing negative staining with anti-PTEN and positive staining with anti-pPRAS40 (T246) antibodies. N, nonneoplastic normal cells; T, tumor. Magnification: ×400 (left column and insets in right column); ×200 (right column). **E and F,** PTEN-negative PC3 cells (E) and PTEN-positive DU145 (F) cells showing pPRAS40 and PTEN staining. Magnification: ×400.

**Loss of PTEN expression in sporadic chordomas.** We next examined PTEN expression by immunohistochemical staining in sporadic sacral chordomas and a TSC-associated chordoma. Surprisingly, 4 of 10 sporadic cases stained rather weakly and
6 of 10 sporadic cases were negative for PTEN staining, whereas a TSC-associated chordoma stained positive (Table 1). Results from two representative sporadic chordomas are shown (Fig. 3C and D). Supportive of these results, 5 of 7 sporadic chordomas stained positive for pPRAS40 (T246) and 2 of 7 cases displayed weak signal (Fig. 3C and D and Table 1). As expected, PTEN-positive DU145 control cells stained strongly for PTEN and revealed negative staining for pPRAS40. PTEN-negative PC3 control cells exhibited no staining for PTEN and showed a strong staining for pPRAS40 (Fig. 3E and F). These results suggest that partial or complete deficiency of PTEN could be responsible for hyperactivation of mTORC1 signaling in at least a subset of chordomas.

**Rapamycin-sensitive mTORC1 signaling and proliferation of U-CH1 cells.** In order to characterize constitutively high, growth factor–independent mTORC1 signaling in U-CH1 cells, the effect of the mTOR-specific inhibitor rapamycin was examined. Upon treatment with rapamycin, S6 phosphorylation in serum-deprived U-CH1 cells was completely eliminated at concentrations as low as 1 nmol/L, but phosphorylation of Akt and ERK was not down-regulated (Fig. 4A). These results indicate that constitutive S6 phosphorylation in U-CH1 cells requires mTOR activity.

We next examined the effect of the upstream PI3K and MEK inhibitors wortmannin and U0126, respectively. Wortmannin significantly diminished phosphorylation of Akt and TSC2 at a low concentration of 10 to 25 nmol/L, and gradually inhibited S6 phosphorylation in serum-deprived U-CH1 cells (Fig. 4B).
However, the MEK inhibitor U0126 was unable to suppress S6 phosphorylation, although it completely abolished ERK phosphorylation as expected (Fig. 4C). These results indicate that hyperactive PI3K/Akt activity, and not ERK/MAPK activity, is responsible for constitutive mTORC1 signaling through TSC2 inactivation in U-CH1 cells.

In order to assess the effect of rapamycin on proliferation of U-CH1 cells, we kept track of cell counts with increasing dosage of rapamycin over the course of 9 days. Compared with the control group treated with vehicle DMSO, rapamycin treatment resulted in a decrease in cell number even at 1 nmol/L concentration, which was observed as early as day 3 with no significant increase in cell numbers through day 9 (Fig. 5A). Furthermore, rapamycin treatment as low as 5 nmol/L resulted in a significant decrease of BrdUrd incorporation into U-CH1 cells, compared with DMSO-treated control group (Fig. 5B). Therefore, these data show that inhibition of mTORC1 signaling may be effective in repressing the proliferation of U-CH1 cells.

**Discussion**

The pathogenic mechanism(s) of chordomas remain unclear, and besides surgery there is no effective treatment for patients with chordomas. We show here for the first time that hyperactivity of Akt and mTORC1 signaling is common in sporadic sacral chordomas. We also observed lack of PTEN expression in approximately 60% (6 of 10) of chordomas examined as well as the chordoma-derived U-CH1 cell line, suggesting that inactivation of PTEN may be responsible for Akt and mTORC1 activation and consequent development of at least a subset of sporadic chordomas. Genomic DNA analyses by comparative genomic hybridization revealed loss of chromosome 10 in 3 of 16 sporadic chordomas where the PTEN gene is located (19, 24). PTEN mutations are frequently found in glioblastoma, hepatocellular carcinoma, lung carcinoma, melanoma, endometrial carcinoma, and prostate cancer (25, 26). Further genetic investigations are essential in order to confirm PTEN inactivation in a larger panel of sporadic chordomas. However, other possibilities including inactivating mutations in TSC genes or activating mutations in PI3K/Akt may also explain the mTOR dysregulation in sacral tumors. In addition, it will be interesting to extend these studies to sphenoid/occipital/clivus chordomas to understand whether Akt/mTORC1 signaling is also aberrantly regulated in these tumors. Deregulation of cellular signaling pathways involving amplification/activating mutations in PI3K, Akt, S6K, or loss/inactivating mutations in tumor suppressor including PTEN, NF1, TSC1, TSC2, VHL, and LKB1 all result in aberrant mTOR activation commonly seen in several hamartoma syndromes and other human cancers (16, 18, 27). Due to their ability to block cell proliferation and angiogenesis, the mTOR inhibitor rapamycin and its analogues, such as RAD001 (Novartis) and CCI-779 (Wyeth), are currently being tested in clinical trials on a wide range of tumors, including those associated with TSC as well as lymphangioleiomyomatosis (28). Many reports show the effectiveness of rapamycin in the treatment of several cancers, such as renal cell carcinoma (29), TSC-associated acute myelogenous leukemia (30, 31), TSC-associated astrocytomas (32), Kaposi’s sarcoma (33), acute myeloid leukemia (34), and mantle-cell lymphoma (35). In addition, PTEN-negative cells have enhanced sensitivity to mTOR inhibitors (36–38). Taken together, our findings imply that mTOR inhibition may provide clinical benefits to chordoma patients. In addition, combined treatment with PI3K inhibitor and mTORC1 inhibitor may be more potent, because hyperactivation of PI3K/Akt activity is known to critically contribute to dysregulation in cell proliferation.

In summary, we found that PTEN deficiency and hyperactivation of Akt/mTORC1 signaling are strongly associated with sporadic sacral chordomas. mTORC1 inhibition is effective in suppressing the proliferation of chordoma-derived cell line. Therefore, this study adds sporadic chordoma to the growing list of mTORC1-hyperactivated tumors, and provides a rationale for initiating clinical trials of Akt/mTOR inhibition in patients with sporadic chordomas. However, given the low incidence of chordomas, it would be necessary to build national/international collaborations and/or partner with other sarcoma initiatives in order to have sufficient number of patients for clinical trial.

**Disclosure of Potential Conflicts of Interest**

No potential conflicts of interest were disclosed.

**Acknowledgments**

We thank Dr. Peter Möller (University Hospitals of Ulm, Germany) for providing U-CH1 cell line and the Chordoma Foundation for their support.

**References**

15. Carsillo T, Astrinidis A, Henske EP. Mutations in the
Aberrant Hyperactivation of Akt and Mammalian Target of Rapamycin Complex 1 Signaling in Sporadic Chordomas

Sangyeul Han, Carolyn Polizzano, Gunnlaugur P. Nielsen, et al.


Updated version Access the most recent version of this article at: [http://clincancerres.aacrjournals.org/content/15/6/1940](http://clincancerres.aacrjournals.org/content/15/6/1940)

Cited articles This article cites 37 articles, 10 of which you can access for free at: [http://clincancerres.aacrjournals.org/content/15/6/1940.full#ref-list-1](http://clincancerres.aacrjournals.org/content/15/6/1940.full#ref-list-1)

Citing articles This article has been cited by 4 HighWire-hosted articles. Access the articles at: [http://clincancerres.aacrjournals.org/content/15/6/1940.full#related-urls](http://clincancerres.aacrjournals.org/content/15/6/1940.full#related-urls)

E-mail alerts Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link [http://clincancerres.aacrjournals.org/content/15/6/1940](http://clincancerres.aacrjournals.org/content/15/6/1940). Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.