The Topoisomerase I Poison CPT-11 Enhances the Effect of the Aurora B Kinase Inhibitor AZD1152 both In vitro and In vivo

Jayasree S. Nair,1 Elisa de Stanchina,2 and Gary K. Schwartz1

Abstract Purpose: AZD1152 is an Aurora B kinase inhibitor currently in clinical trials. As the topoisomerase I poison CPT-11 induces a G2 arrest, a mechanistic understanding of the cell cycle interactions between these agents may prove critical for combination therapy.

Methods: AZD1152 was tested in vitro and in vivo with SN-38 and CPT-11 against HCT-116 cells. Inhibition of clonogenicity, induction of apoptosis, effects on polyploidy, and tumor growth were examined.

Results: AZD1152 alone induced polyploidy of HCT-116 cells at low nanomolar concentrations. The induction of apoptosis required prolonged exposure (48 hours) and higher concentrations of drug. When SN-38 was given before or concomitantly with AZD1152, SN-38 blocked the AZD1152 effect by arresting cells in G2 and inhibiting cells from undergoing polyploidy. With the reverse combination (AZD1152 followed by SN-38), there was a significant induction of polyploidy and apoptosis, even with shorter exposure (24 hours) of AZD1152. In vivo, AZD1152 alone suppressed HCT-116 xenograft tumor growth in a dose-dependent manner with target inhibition of phosphoH3, induction of multinucleated giant cells, but without induction of apoptosis. In combination, both sequences in vitro (CPT->AZD, AZD->CPT, P = 0.008, AUC/d) proved superior to either single agent therapy. However, AZD->CPT still showed a greater increase in apoptosis and greater suppression of tumor regrowth than CPT->AZD (P = 0.02, AUC/d).

Conclusions: The results from these studies indicate a promising therapeutic strategy for combining AZD1152 with CPT-11, and suggest that the sequence of drug administration is pivotal when an Aurora B kinase inhibitor is administered with a topoisomerase I poison.

Aurora kinases are mitotic serine threonine kinases. They play an important role in chromosome alignment, segregation, and cytokinesis. The Aurora kinase family consists of three members, Aurora A, B, and C kinases (1). Aurora A and Aurora B are highly homologous in sequence and structure, but share few regulators or substrates and are different in their subcellular distribution (2). Aurora A localizes to the spindle poles where it is involved mainly in centrosome separation, mitotic entry, and spindle assembly. Aurora B, which moves to centromeres from prometaphase to metaphase and relocates to the midbody until cytokinesis is completed, is important in chromosome alignment, microtubule-kinetochore attachment, spindle checkpoint, and cytokinesis (3–5). The activity of Aurora kinases is strictly regulated during mitosis, with Aurora A activity peaking at the G2-M transition, whereas activity of Aurora B persists from metaphase to the end of mitosis (2). Besides recent reports showing Aurora C to be a chromosomal passenger protein that binds directly to the inner centromere protein and survivin in vitro, not much is known about this member of the Aurora kinase family (6, 7).

Inappropriate levels of Aurora kinases are shown to be linked to genetic instability (8). Aurora kinases are overexpressed in a wide range of human cancers. Aurora A is commonly amplified in primary breast tumors, colorectal cancers, and in cancer cell lines of different origin including breast, ovarian, colon, prostate, neuroblastoma, and cervix (9–11). Aurora B is also expressed at high levels in primary human colorectal cancers and other tumor cell lines (12). Therefore, Aurora kinases have become promising targets for cancer therapy. Considerable effort has been devoted in developing new small molecule inhibitors. ZM447439 (Astra Zeneca), Hesperadin (Boehringer Mannheim), VX-680 (Vertex Inc.), and AZD1152 are a few of the several small molecule inhibitors that have been evaluated (13, 14). The common cellular phenotype induced by inhibition of Aurora B kinase is failure of cytokinesis with the induction of polyploidy and multinucleated giant cells. This is in contrast to inhibition of Aurora A, which induces mitotic arrest followed by apoptosis. Interestingly, drugs that inhibit both Aurora kinases A and B induce multinucleated cells, indicating that the cellular phenotype associated with Aurora B inhibition is favored when both kinases are inhibited concurrently.
AZD1152 is a dihydrogen phosphate prodrug of a pyrazoloquinazoline Aurora kinase inhibitor, which is rapidly converted to the active species AZD1152-hydroxyquinazoline pyrazol anilide (HQPA) in plasma. AZD1152 has been shown to be a potent and selective inhibitor of Aurora B with a Ki of 0.36 nmol/L, as compared with Aurora A, with a Ki of 1,369 nmol/L (14, 15). It was shown that AZD1152, as a single agent, potently inhibits tumor xenograft growth in vivo (14), and, when used in combination, AZD1152 enhances the radiation response in tumor cell lines in vitro and in vivo (16). In human leukemia cells, AZD1152 was effective in inhibiting growth and showed an additive effect when used in combination with vincristine and daunorubicin both in vitro and in vivo (17).

In the present study, we used AZD1152, the prodrug, for in vitro studies and AZD1152-HQPA, the active drug, for in vivo studies against the human colon carcinoma cell line HCT-116. These studies were conducted both as a single agent, and in combination with the topoisomerase I poison CPT-11, as well as with its active metabolite, SN-38 (18). As a single agent, AZD1152 suppressed tumor growth in athymic mice bearing HCT-116 tumors in a dose-dependent manner without evidence of toxicity. In vitro studies showed antagonism when SN-38 was given first or concurrently, by arresting the cells in G2 and inhibiting AZD1152-HQPA–induced polyploidy. In the reverse sequence, however, AZD1152-HQPA followed by SN-38, cells underwent polyploidy and apoptosis. These combination effects were reproduced in another colon cancer cell line, Colo-205, indicating that the effect we observe in HCT-116 is not exclusive to HCT-116 and is a rather more generalized effect (Supplement 1 and 2). In vitro, CPT-11 enhanced the effect of AZD1152 in HCT-116 tumor xenografts independent of sequence. However, the regrowth of tumors was most effectively suppressed when AZD1152 was given before CPT-11. This effect seemed to be due to the added effect of apoptosis on the polyploid cells, indicating the importance of the sequence of drug administration when combining an Aurora B kinase inhibitor with a topoisomerase-I poison.
histones H3 (Cell Signaling Technology; 1:250) and subsequently with Alexa Fluor-conjugated secondary antibody (Molecular Probes). DNA was labeled with DAPI at a final concentration of 25 μg/mL for 10 min.

**Cell extraction, immunoprecipitation, and immunoblotting.** Cell lysates were prepared as described by lysis of both floating and adherent cells in whole cell lysis buffer (50 mmol/L Tris, pH 8, 250 mmol/L NaCl, 0.5% NP-40, 0.2 mmol/L EDTA, 2 mmol/L EGTA, 10% Glycerol, 0.1 mmol/L phenylmethylsulfonyl fluoride, 0.1 mmol/L Na3VO4, 1 mmol/L DTT, 1 μg/mL leupeptin, 2 μg/mL aprotinin, 1 μg/mL pepstatin), allowed to lyse on ice for 10 min, syngred, and cleared by centrifugation in a microcentrifuge at 13,000 rpm for 10 min at 4°C (20). Fifty micrograms of protein were fractionated by SDS-PAGE and transferred onto Immobilon membranes (Millipore). Equal protein loading was confirmed by Amido black staining (Bio-Rad). After blocking with 5% nonfat milk, membranes were probed with primary antibodies. The following antibodies were used in this study: mouse monoclonal to poly (ADP-ribose) polymerase (PARP) was from BD Biosciences; mouse monoclonal anti-p53 was from Santa Cruz Biotechnology; mouse monoclonal anti-p21 (1:1000) was from Oncogene; rabbit polyclonal to phospho Ser 10 H3, rabbit polyclonal to cleaved β-tubulin was from Upstate Biotechnology. Bound primary antibodies were detected with horseradish peroxidase–conjugated secondary antibodies (ICN/Jackson ImmunoResearch) and visualized by enhanced chemiluminescence reagent (Amersham Pharmacia).

**Xenograft studies.** Athymic mice bearing HCT-116 tumors (7 mice/ cohort) of 200 mm3 diameter were treated with specified dose of AZD1152 (25 to 100 mg/kg/d) i.p. as a single agent, 2 d/wk for 3 wk. Twenty-four hours after each treatment on days 8 and 22, and 3 wk after the last treatment on day 43, one animal from each cohort was sacrificed and the tumors examined for H&E, Ki67 staining, and TUNEL. Tumors were measured every 2 to 3 days. Mice in the control groups were given vehicle (0.3 mol/L Tris at pH 9) alone. Twenty-four hours after the last treatment (day 22), two animals from each cohort were sacrificed and the tumors examined for H&E, Ki67 staining, and terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL). For combination studies 7 mice/cohorts received AZD1152 alone i.p. (100 mg/kg) daily ×2, CPT-11 alone i.p. (100 mg/kg, the maximum tolerated dose) ×1, or the same doses i.p. of AZD1152 on days 1 and 2 and CPT-11 on day 3 or CPT-11 on day 1 followed by AZD1152 for 2 d. Each treatment condition was repeated weekly for 3 wk. Mice in the control groups were given vehicle (0.3 mol/L Tris at pH 9) alone. Twenty-four hours after the last treatment (day 22), two animals from each cohort were sacrificed and the tumors examined for H&E, Ki67 staining, and TUNEL. Tumors were measured every 2 to 3 d with calipers, and tumor volumes were calculated by the formula 4/3 x (larger diameter + smaller diameter)/4. The percentage of tumor regression was calculated as the percentage ratio of difference between baseline and final tumor volume to the baseline volume. Toxicity was monitored by weight loss. These studies were done in accordance with the Principles of Laboratory Animal Care (N.I.H. Publication No. 85-23, released 1985).

**Histopathology.** For immunohistochemistry analysis, representative sections were deparaffinized, rehydrated in graded alcohols, and subjected to antigen retrieval by microwave oven treatment using the published technique (21) and sections were incubated with Avidin-Biotin Complex Elite ( Vectastain ABC Elite Kit; Vector Labs) diluted 1:25 in PBS for 30 min followed by staining with 3,3’-diaminobenzidine (Sigma) for 3 min. The percentage of TUNEL-positive cells from tumor sections was determined by counting at least 100 cells each from at least three randomly selected fields. For Ki-67 and phospho histone H3 immunohistochemistry, after blocking in 10% normal goat serum in a humid chamber for 30 min, sections were incubated in primary antibodies, Ki-67 (1:10; Vector Labs) and phospho H3 (1:100; Cell Signaling Technologies) overnight at 4°C followed by secondary antibody (Biotinylated Anti Rabbit IgG; Vector Labs) for 30 min at room temperature.

**Statistical analysis.** All in vitro experiments were carried out at least three times. The statistical significance of the experimental results was determined by the two-sided t test and was adjusted for multiple comparisons by using Bonferroni correction. We chose P = 0.05 as statistically significant in individual comparisons. This was corrected to 0.0125 after the Bonferroni adjustment for four statistical comparisons. For in vivo studies, the two-sided t test and the area under the time-volume curve was used as a summary measure for each mouse, as previously described (22). Tumor volume was compared between groups of mice at various points in time based on the experiment or by calculating AUC/d over a range of days. This area was calculated using the trapezoidal rule. This method takes the longitudinal aspect of the data into account and does not require assumptions of linear growth (or decay), which are clearly violated for our data. P values were calculated using the Wilcoxon Rank Sum test.

**Results**

AZD 1152-HQPA induces growth arrest, apoptosis, and polyplody in HCT116 cells. HCT-116 colon carcinoma cells were treated with varying concentrations of AZD1152-HQPA to determine its target specific and its biological effects. As shown in Fig. 1A, AZD1152-HQPA, by both Western blot analysis and immunofluorescence, inhibited histone H3 phosphorylation, a known substrate of Aurora B, at concentrations of 20 nmol/L. As seen with DAPI staining, this corresponded to the induction of polyploid cells (cells with large, multilobed nuclei). Its effect on cellular proliferation was determined by clonogenic assay and the IC50 value was determined to be 20 nmol/L (Fig. 1B). However, there was no induction of apoptosis at this concentration. With higher concentrations (100-200 nmol/L), and with prolonged drug exposure (48 hours), induction of apoptosis could be detected by the appearance of a modest sub G1 peak using flow cytometry (Fig. 1C) and PARP cleavage by Western blot (Fig. 1D). AZ1152-HQPA showed no difference in the kinetics of mitotic entry or exit as compared with the untreated controls (not shown). However, cells treated with AZD1152-HQPA showed an increased DNA content from 2N to 4N to 8N despite the induction of p53 and p21 (Fig. 1C and D).

Single agent AZD1152 inhibits the growth of HCT-116 tumor xenografts. Because AZD1152-HQPA showed a very potent inhibition of proliferation in vitro, its efficacy was next tested in vivo with the prodrug AZD1152. Preliminary studies indicated that i.v. push for two days, i.p. injections for two days, and continuous infusion of AZD1152 for 48 hours using a mini-pump, gave identical results (data not shown). However, cells treated with AZD1152-HQPA showed an increased DNA content from 2N to 4N to 8N despite the induction of p53 and p21 (Fig. 1C and D).
changes in nuclear morphology by H&E. Figure 2B shows only 10% inhibition of Aurora B kinase activity with 25 mg/kg. However, there was 70%, 80%, and 90% inhibition of phospho histone H3 at doses of 50, 75, and 100 mg/kg, respectively (P = 0.03; 50 mg/kg versus 75 and 100 mg/kg). Proliferation of tumor cells was similarly inhibited in a concentration-dependent manner, as determined by staining with Ki-67 (P = 0.01; 50 mg/kg versus 75 and 100 mg/kg; Fig. 2B). As shown in Fig. 2C (top, middle panel, arrow H&E staining), the cells were polyploid with enlarged nuclei, presumably due to the inhibition of cytokinesis and continued endoreduplication, a known effect of Aurora B inhibition. This was associated with decreased Ki-67 staining (Fig. 2C, bottom, middle panel), indicating a significant decrease in cellular proliferation. There was minimal induction of apoptosis by TUNEL under any condition tested (data not shown), suggesting that the decrease in tumor volume (Fig. 2A) with single agent AZD1152 was related to inhibition of cellular proliferation. In order to determine whether the growth inhibition was reversible, the tumor volume was monitored for 3 weeks after the last drug administration. At the end of 3 weeks (day 44), tumor volumes had increased back to baseline (data not shown). Tumors were then collected and stained for Ki-67, H&E, and TUNEL. As shown in Fig. 2C, Ki-67 staining showed an increased proliferation rate. H&E staining showed a restoration of mononuclear cells with a decrease in the giant multinucleated cell population that had been observed at the end of treatment (day 23). These results suggest that inhibition of tumor growth by an Aurora B kinase inhibitor requires sustained induction of the polyploid phenotype in order to maintain the decrease in cellular proliferation that is associated with inhibition of tumor growth. Interestingly, when rechallenged with AZD1152, a decrease in tumor growth was observed in these animals, indicating that the tumors did not acquire resistance to the effects of Aurora B inhibition (data not shown).

SN-38 in combination with AZD1152-HQPA. CPT-11 is a topoisomerase I poison and is used as a standard part of chemotherapy for the treatment of metastatic colon cancer (23). Because our long-term goal is to combine AZD1152 with cytotoxic chemotherapy for the treatment of colon cancer, we elected to test AZD1152-HQPA (the active agent) in vitro with SN-38, the active metabolite of CPT-11, against the HCT-116 cells. As these drugs have independent effects on the cell cycle, we elected to test these drug combinations in a sequence-specific manner. The combination was first tested for its effect on clonogenicity and cell viability. SN-38 alone has been shown to have a profound effect on inhibition of clonogenicity, even at concentrations as low as 1 nmol/L, due to induction of p53 and p21 and a resulting G2 arrest (22). In view of this, differences in clonogenicity are generally difficult to show with SN-38 in combination therapy. Nevertheless, as shown in Fig. 3A, the inhibition of clonogenicity favored AZD->SN (P = 0.0001, AZD->SN versus SN->AZD, and all other comparisons showed a P < 0.005). Even a modest effect was observed with AZD->SN over AZD+SN (P = 0.0007). This could be due to the escape of cells from G2 arrest, induced by SN-38 at the concentration used (1 nmol/L). Because multiple comparisons have been applied for AZD->SN, the Bonferroni adjustment was applied and all comparisons were shown to be statistically significant. However, inhibition of cell viability...
using Dojindo cell proliferation kit, was significantly greater for AZD->SN than AZD, SN, or SN->AZD (P > 0.001, 0.002, and 0.01) and for any of the combinations tested, including AZD+SN (P = 0.03). In this assay, after Bonferroni correction, all statistical comparisons were statistically significant except for the concurrent treatment, which, though individually significant, fell on the borderline of 0.03. This same sequence dependency was observed in the Colo-205 cells, such that AZD->SN had the greatest effect on inhibiting cell growth (Supplement 1A).

We next examined the effects of the combinations on cell cycle distribution by flow cytometry. As predicted (Fig. 3B), SN-38 alone (SN) at a pharmacologically relevant concentration (20 nmol/L) induced a permanent G2 (4N DNA) arrest. This effect was sustained for at least another 24 hours even when these cells were placed in drug-free media (22). This was in contrast to AZD1152-HQPA (AZD, 100 nmol/L) which induced endoreduplication, resulting in polyploidy (8N DNA). However, when SN-38 was given prior to (SN->AZD) or concomitantly with AZD1152-HQPA (AZD+SN), SN-38 blocked induction of polyploidy by AZD1152-HQPA. This was due to the induction of permanent G2 arrest by SN-38, thereby inhibiting cells from entering mitosis where AZD1152-HQPA is active. A small 8N peak was observed with AZD+SN, suggesting those cells that were out of G2 and were first affected by AZD1152-HQPA. On the other hand, when AZD1152-HQPA was given prior to SN-38 (AZD->SN), the 8N peak was comparable with AZD alone, and there was a large increase in the sub G1 (<2N) population, reflecting an enhancement of apoptosis (15 ± 1% with AZD alone to 30 ± 3% with AZD->SN). Of particular note, with SN->AZD a significant reduction in the sub G1 peak was observed (8 ± 1%), compared with AZD alone (P < 0.01).

The induction of apoptosis by SN-38 and AZD1152-HQPA is sequence-specific. In order to further understand the mechanism for these sequence-dependent effects, we next elected to
examine this further by quantitative fluorescent microscopy (DAPI) and Annexin V labeling in the HCT-116 cells. As shown in Fig. 4A, examination of the DAPI-stained nuclear morphology by fluorescence microscopy showed enlarged interphase nuclei with the SN, SN->AZD, and the SN+AZD combinations. In contrast, AZD treatment alone resulted in large multilobed nuclei, whereas AZD->SN showed decondensed apoptotic nuclei (arrow heads), besides the multilobed nuclei, indicating both polyploidy and apoptosis. The induction of apoptosis increased from 15 ± 2% with AZD alone to 29 ± 4% with AZD->SN (P = 0.013), an effect that was quite similar to that observed with analysis of the sub G1 peak. Annexin V staining (Fig. 4B), which detects both early and late stages of apoptosis, further confirmed the increase in apoptosis with the AZD->SN combination, when compared with the other conditions tested. Again, using both quantitative fluorescent microscopy and Annexin V, the reverse sequence of SN->AZD showed a partial reversal in the induction of apoptosis induced by AZD alone. In addition, Western blot analysis also showed a large increase in PARP cleavage with AZD->SN, which is not observed with the other combinations (Fig. 4C). As determined by quantitative fluorescent microscopy, sub-G1 peak, and PARP cleavage, the effect of enhanced apoptosis with the sequential AZD->SN was also observed in Colo-205 cells (Supplement 1B to D). Taken together, these data indicate that cells which have undergone AZD1152-HQPA–induced endoreduplication are susceptible to apoptosis with subsequent SN-38 therapy. In contract, SN-38 can induce a prolonged G2 arrest, which is sufficient to prevent the induction of both polyploidy and apoptosis by subsequent AZD1152-HQPA therapy. This finding also strongly indicates that the sequence of drug administration is critical to achieve the full combination effect.

**CPT-11 enhances the effect of AZD1152 in vivo.** Based on the increased efficacy of the AZD->SN-38 sequence combination, we chose to compare AZD1152 in sequence with CPT-11
with the other treatment conditions in HCT-116 xenografts. As AZD1152 at 100 mg/kg induced at least 90% inhibition of phospho-histone H3 without weight loss, we selected this dose for combination therapy. All animals were treated on days 7, 14, and 21. As shown in Fig. 5A, all treatment conditions produced comparable levels of growth suppression, but the time of onset of tumor regrowth and the percent increases in tumor volume after completion of therapy were different between groups. Tumor regrowth was noted as early as day 29 for CPT-11 alone, day 36 for AZD1152 alone, day 43 for CPT-11→AZD, and day 50 for AZD→CPT-11. In terms of overall increases in tumor volume, both sequential combinations at day 60 showed greater inhibition of tumor regrowth when compared with either AZD1152 alone (P = 0.06, AZD versus CPT-11→AZD; P = 0.008, AZD versus AZD→CPT-11) or CPT-11 alone (P = 0.008, CPT-11 versus CPT-11→AZD; P = 0.008, CPT-11 versus AZD→CPT-11). Furthermore, a comparison of the two sequential combinations indicates that the overall increase in tumor volume up to day 71 was significantly less for AZD→CPT-11 than for CPT-11→AZD (P = 0.02). There was no associated weight loss with any condition tested.

These data suggest that AZD1152 followed by CPT-11 is superior to CPT-11 followed by AZD1152, but the differences between the sequential combinations were not as dramatic as expected from the in vitro studies, in which polyploidy induced by the Aurora B kinase inhibitor was essentially inhibited when SN-38 was given before AZD1152-HQPA. This could be due to the less effective in vivo conversion of a prodrug to its active metabolite, but a more likely explanation could be due to cell cycle interactions between the two agents. Therefore, the tumor samples were evaluated by H&E for morphology, Ki-67 for proliferation, and TUNEL for apoptosis 24 hours after the last treatment. As shown in Fig. 5B, AZD1152 alone again induced large numbers of giant, multinucleated cells. However, with AZD→CPT-11 there were fewer appreciable tumor cells present, and those that were present were bizarrely shaped (right, top panel). In addition, when compared with the other treatment conditions, apoptosis by TUNEL increased from 19 ± 5% with AZD alone to 48 ± 5% with AZD→CPT-11 (Fig. 5C, P = 0.036).
Nevertheless, with Ki-67 staining, some cells continued to proliferate, suggesting an explanation for the modest degree of tumor regrowth. Interestingly, Ki-67–positive cells could still be detected after CPT-11 and CPT-AZD treatment, indicating that cells had escaped from G₂ arrest, and continued to proliferate. Therefore, it seems that even at the maximum tolerated dose of CPT-11 (100 mg/kg), some tumor cells are still able to escape from G₂ cell cycle arrest, making them susceptible to the subsequent effect of AZD1152 therapy. This would explain the increase in apoptosis observed with sequential CPT-AZD as compared with CPT alone (Fig. 5C).

AZD1152 followed by CPT-11 can now be considered. By CPT-11. Based on these results, sequential combinations of specific drug sequencing, particularly when individual drugs (even conventional cytotoxic agents) have specific cell cycle effects. In fact, use of the wrong sequence of administration result in a process termed “cell-cycle mediated drug resistance,” such that the cell cycle interactions can result in antagonism (27). Similar effects have been reported with other cell cycle inhibitors when combined with chemotherapy (28). In the current studies, the in vivo effects did not exactly mirror the in vitro effects. This finding does suggest that the most efficient sequence of drug administration would be AZD1152 followed by CPT-11. Based on these results, sequential combinations of AZD1152 followed by CPT-11 can now be considered.

**Disclosure of Potential Conflicts of Interest**

G. Schwartz, advisory board, AstraZeneca.

---

**References**

11. Sen S, Zhou H, White RA. A putative serine/threonine...
kinase encoding gene BTAK on chromosome 20q13 is amplified and overexpressed in human breast cancer cell lines. Oncogene 1997;14:2195 – 2199.


The Topoisomerase I Poison CPT-11 Enhances the Effect of the Aurora B Kinase Inhibitor AZD1152 both *In vitro* and *In vivo*

Jayasree S. Nair, Elisa de Stanchina and Gary K. Schwartz


<table>
<thead>
<tr>
<th>Updated version</th>
<th>Access the most recent version of this article at: <a href="http://clincancerres.aacrjournals.org/content/15/6/2022">http://clincancerres.aacrjournals.org/content/15/6/2022</a></th>
</tr>
</thead>
<tbody>
<tr>
<td>Supplementary Material</td>
<td>Access the most recent supplemental material at: <a href="http://clincancerres.aacrjournals.org/content/suppl/2009/03/13/1078-0432.CCR-08-1826.DC1">http://clincancerres.aacrjournals.org/content/suppl/2009/03/13/1078-0432.CCR-08-1826.DC1</a></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Cited articles</th>
<th>This article cites 28 articles, 13 of which you can access for free at: <a href="http://clincancerres.aacrjournals.org/content/15/6/2022.html#ref-list-1">http://clincancerres.aacrjournals.org/content/15/6/2022.html#ref-list-1</a></th>
</tr>
</thead>
<tbody>
<tr>
<td>Citing articles</td>
<td>This article has been cited by 2 HighWire-hosted articles. Access the articles at: /content/15/6/2022.html#related-urls</td>
</tr>
</tbody>
</table>

**E-mail alerts**  Sign up to receive free email-alerts related to this article or journal.

**Reprints and Subscriptions**  To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

**Permissions**  To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.