Targeted Inhibition of Inducible Nitric Oxide Synthase Inhibits Growth of Human Melanoma In vivo and Synergizes with Chemotherapy

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Abstract

Purpose: Aberrant expression of inflammatory molecules, such as inducible nitric oxide (NO) synthase (iNOS), has been linked to cancer, suggesting that their inhibition is a rational therapeutic approach. Whereas iNOS expression in melanoma and other cancers is associated with poor clinical prognosis, in vitro and in vivo studies suggest that iNOS and NO can have both protumor and antitumor effects. We tested the hypothesis that targeted iNOS inhibition would interfere with human melanoma growth and survival in vivo in a preclinical model.

Experimental Design: We used an immunodeficient non-obese diabetic/severe combined immunodeficient xenograft model to test the susceptibility of two different human melanoma lines to the orally-given iNOS-selective small molecule antagonist N6-(1-iminoethyl)-L-lysine-dihydrochloride (L-nil) with and without cytotoxic cisplatin chemotherapy.

Results: L-nil significantly inhibited melanoma growth and extended the survival of tumor-bearing mice. L-nil treatment decreased the density of CD31+ microvessels and increased the number of apoptotic cells in tumor xenografts. Proteomic analysis of melanoma xenografts with reverse-phase protein array identified alterations in the expression of multiple cell signaling and survival genes after L-nil treatment. The canonical antiapoptotic protein Bcl-2 was downregulated in vivo and in vitro after L-nil treatment, which was associated with increased susceptibility to cisplatin-mediated tumor death. Consistent with this observation, combination therapy with L-nil plus cisplatin in vivo was more effective than either drug alone, without increased toxicity.

Conclusions: These data support the hypothesis that iNOS and iNOS-derived NO support tumor growth in vivo and provide convincing preclinical validation of targeted iNOS inhibition as therapy for solid tumors. Clin Cancer Res; 16(6); 1834-44. ©2010 AACR.

Upregulation of proinflammatory molecules by tumor cells is a poorly understood phenomenon, which can play a role in both the induction and maintenance of certain human cancers (1). One such molecule, inducible nitric oxide (NO) synthase (iNOS) is constitutively overexpressed in many cancers, including melanoma and gastric (2), breast (3), colon (4), and head and neck (5, 6) carcinomas. iNOS and its product NO have wide-ranging and varied effects on cellular physiology, signal transduction, and cell survival. At high levels, such as produced by activated macrophages during inflammatory responses to pathogens, NO alone or in combination with reactive oxygen species can have a direct cytotoxic effect on pathogens or tumor cells (7). At lower levels, NO can affect signal transduction pathways by interacting with metal ligands of proteins (8) or covalently modifying proteins through nitration and nitrosylation (9, 10). These protein...
Thus, both clinical and in vitro evidence supports the hypothesis that targeted inhibition of iNOS and iNOS-derived NO may be an effective therapeutic approach for melanoma and other iNOS-expressing tumors.

In the present study, we tested the effect of iNOS inhibition with small-molecule antagonists on human melanoma in vivo using a xenograft model. We found that the iNOS selective antagonist N<sup>ε</sup>-(1-iminoethyl)-l-lysine dihydrochloride (L-nil; ref. 22) inhibited iNOS-dependent NO production by human melanoma cells in a dose-dependent fashion in vitro and strongly suppressed melanoma growth in vivo without evident toxicity. Growth suppression was associated with a decrease in tumor microvessels, downregulation of the antiapoptotic gene Bcl-2, increased number of intratumoral apoptotic cells, and enhanced efficacy when L-nil treatment was combined with cisplatin in vivo. These data suggest that iNOS-selective small molecule inhibitors, alone or in combination with conventional cytotoxic chemotherapy, are a promising approach to therapy of melanoma and other solid tumors.

Materials and Methods

Western blots. Cell lysates were separated by size using SDS-PAGE and transferred to a membrane. Membranes were incubated with a primary antibody against iNOS (sc-651, Santa Cruz Biotech), Bcl-2 (Pharmingen), or extracellular signal-regulated kinase 2 protein levels.

Relative signal intensities for Bcl-2 were validated by Western blot, and normalized densitometry values were expressed as ratio of Bcl-2 to extracellular signal-regulated kinase 2 protein levels.

Tumor cell lines and transient transfection. The human melanoma cell lines melan cells 624 and 528 were a kind gift from Dr. J. Wunderlich, Surgery Branch, National Cancer Institute, NIH. The A375 human melanoma line was purchased from American Type Culture Collection. The human colon cell line WiDR was a kind gift of J. Hodge, Laboratory of Tumor Immunology and Biology, National Cancer Institute, NIH. WiDR cells were transfected with Lipofectamine according to the manufacturer’s (Invitrogen) protocol. At 24 h after transfection, cells were replated for use in the indicated assays. Plasmid expressing the gene encoding full-length human iNOS protein was a generous gift from Keping Xie and has been previously described (23). All melanoma lines were submitted to the Characterized Cell Line Core of the M.D. Anderson Cancer Center Support Grant. Fingerprinting analysis is done, and cells are validated every 2 y; the lines in this study were validated in March 2008.

Detection of NO and in vitro inhibitor experiments. Cell lines were cultured in phenol-free DMEM supplemented
with 10% fetal bovine serum, penicillin, and streptomycin at 37°C in 5% CO₂. Cells (2 × 10⁵) were cultured in a 24-well plate with the indicated concentration of L-NIL for 72 h before assay for cell death or NO levels. Cumulative NO production was measured in vitro by the nitrite reconversion method with the Apollo 4000 bioanalyzer (World Precision Instruments) as previously described (24). In cytotoxicity experiments, the 2,3-bis[2-methoxy-4-nitro-5-sulfaphenyl]-2H-tetrazolium-5-carboxanilide inner salt (XTT) cell proliferation assay (Cayman Chemical) or Annexin/propidium iodide cell death assay (BD Biosciences) was used according to the manufacturer’s directions.

For in vivo NO measurements, C57BL/6 mice received 0.1% L-nil in drinking water or plain water control for 48 h before i.p. injection of 250 μg lipopolysaccharide (LPS). Mice were sacrificed at 7 h postinjection, and serum was prepared from whole blood by centrifugation. Serum was then clarified by passage through a 10-kDa cutoff spin-filter (Millipore), and nitrite levels were determined by Griess assay (25).

L-nil was obtained from A.G. Scientific. Griess reagent, 1,3-PBIT, LPS, and aminoguanidine were obtained from Sigma Aldrich Corp.

**Intracellular Bcl-2 staining.** Staining with anti-Bcl-2 antibody was modified from the published protocol (26). In brief, cells were permeabilized using cytotoxicytoperm kit (BD Pharimgen) and stained with Bcl-2-FITC antibody or matched isotype control (BD Pharimgen).

**Animals and in vivo tumor experiments.** Female non-obese diabetic (NOD)/severe combined immunodeficient (SCID) mice (ages 6–8 wk) were purchased from the Animal Production Area of the National Cancer Institute-Frederick Cancer Research and Development Center. Female C57BL/6 mice (ages 6–8 wk) were purchased from JAX labs. The mice were bred and maintained in a pathogen-free environment and fed irradiated mouse chow and autoclaved reverse osmosis-treated water. All of the animal procedures were done in accordance with a protocol approved by the Institutional Animal Care and Use Committee of the University of Texas M.D. Anderson Cancer Center. In xenograft experiments, animals were injected with human melanoma tumor cell lines (either mel624 or A375) at initial concentrations of 5 × 10⁶ or 7.5 × 10⁶ in 100 μL PBS, respectively. Animals were randomized after tumors established and assigned to experimental groups (10 per group unless otherwise noted). Experimental animals received 0.15% or 0.2% L-NIL or 0.2% PBIT in their drinking water, replaced every 2 to 3 d, beginning 3 d after tumor implantation. Control animals received plain drinking water. In some experiments, animals received cisplatin given at the indicated dose and time point via i.p. injection. Animals were weighed regularly and monitored for signs of toxicity. Tumor growth was measured thrice per week using electronic calipers, and tumor volumes were determined by multiplying the longest axis by its perpendicular. Mice were humanely sacrificed when tumors reached >250 mm², ulceration occurred, or mice become moribund.

In some tumor experiments, representative animals were evaluated by magnetic resonance imaging (MRI) using a 7.0T Biospec USR small animal imaging system (Bruker Bispin MRI). A linear volume resonator with 35 mm inner diameter was used for signal excitation and detection. Axial T₁-weighed (TE/TR 8.5 ms/900 ms, two averages, 156 nm × 156 nm × 1 mm resolution) and T₂-weighed images (TE/TR 65 ms/5,000 ms, RARE factor 12, three averages) with matching slice prescriptions were collected for tumor visualization.

**Immunohistochemistry.** Paraffin-embedded sections of excised human melanoma tumor xenografts were examined for nitrotyrosine and iNOS expression by immunohistochemistry using an anti-nitrotyrosine polyclonal antibody (Upstate Biotechnology) or anti-iNOS monoclonal antibody (BD-Transduction Laboratories). Preimmune normal rabbit IgG (Vector Laboratories) and anti-vimentin antibody (BioGenex Laboratories) were used as negative and positive controls, respectively. Tissue sections were deparaffinized and rehydrated, then placed in Antigen Unmasking Solution (Vector Laboratories), and microwave intermitently for a total of 10 min to maintain boiling temperature. After cooling, the slides were placed in 3% H₂O₂ in cold methanol for 15 min and then 0.05% Triton X-100 (Sigma) for 15 min. An avidin-biotin-peroxidase complex kit (Vectastain, Vector Laboratories) was then used for antigen detection according to the manufacturer’s instructions, and immunolabeling was developed with the chromogen 3-aminio-9-ethylcarbazole for 10 min.

For detection of CD31, frozen tumor sections were fixed in cold acetone for 10 min before immunostaining with anti-CD31 primary antibody (Pharimgen) for 1 h at room temperature, detection with peroxidase-conjugated secondary antibody, and counterstaining with hematoxylin. Terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) assay was done on paraformaldehyde-fixed tissues with an apoptosis detection kit (Promega) according to the manufacturer’s directions. CD31/TUNEL double-staining was done by subjecting the same tissues to sequential CD31 immunostaining with Alexa Fluor 594–conjugated secondary antibody and TUNEL assay with fluorescein-dUTP.

**Quantification of nitrotyrosine, iNOS, microvessel density, apoptotic tumor, mean vessel diameter, and endothelial cells.** Immunofluorescence microscopy images were obtained with a Leica DMLA microscope (Leica Microsystems) equipped with a Hamamatsu 5810 cooled CCD camera (Hamamatsu Corp.) and ImagePro Plus 6.0 software (Media Cybernetics). Photomontages were prepared using Adobe Photoshop software (Adobe Systems, Inc.). For the quantification analysis, four slides were prepared for each group. The percentage of positively-staining cells in each group was calculated and compared. For quantification of TUNEL expression, the cells positively stained were counted in five random 0.04-mm² fields at 200× magnification per slide. To quantify microvessel density, areas containing higher numbers of tumor-associated blood vessels were identified at low microscopic power (100×).
Vessels completely stained with anti-CD31 antibodies were counted in five random 0.159-mm² fields at 100× magnification per slide. For comparison of vessel diameter between control and experimental groups, mean diameter in pixels was calculated for the stained blood vessels in 10 random 0.159-mm² fields at 100× magnification.

**Reverse-phase protein array.** Reverse-phase protein array (RPPA) as done has been described previously (27) and was used to quantify protein and phosphorylated protein expression of 59 cancer-related and signal transduction proteins, including Bcl-2, phosphorylated PDK1 (Ser241), phosphorylated AMPK (Thr172), 4EBP1, P70S6K, S6, TSC2, PTEN, mammalian target of rapamycin (mTOR), and others. Comparison antibodies were from Cell Signaling (PTEN, mTOR, and all phosphorylated-specific antibodies), Epitomics, Inc. (total p70S6K antibody), and DAKO (Bcl-2).

**Statistical analysis.** The quantifications of the immunohistochemical expression of nitrotyrosine, iNOS, TUNEL, microvessel density, and mean vessel diameter were compared by the paired Student’s t test, with significance at \( P < 0.01 \). For in vitro experiments, Student’s t test and a significance threshold of \( P < 0.05 \) was used. For in vivo survival experiments, Kaplan-Meir survival curves were generated, and significant differences were determined using the log-rank test. In vivo tumor growth curves were compared using the nonparametric method of Koziol et al. to allow comparisons despite unequal survival of mice between groups (28).

**Results**

*Human melanoma lines express iNOS and release NO, which is inhibited by the selective antagonist L-nil.* Western blot analysis was used to screen melanoma cell lines for iNOS protein expression (Fig. 1A), using the previously confirmed iNOS-negative colon cancer cell line WIDR (4) as a negative control. iNOS protein was strongly expressed in the melanoma lines mel526 and A375 and expressed less strongly in mel624. iNOS expression was maintained in vivo, as A375 tumor xenografts were strongly positive for both iNOS and the stable NO end product nitrotyrosine by immunohistochemistry (Fig. 1B). The presence of nitrotyrosine shows that iNOS is expressed and catalyzes NO production in human melanoma cells.

![Fig. 1. Human melanoma lines constitutively express iNOS and make NO in vitro and in vivo.](image-url)

A, human melanoma lines mel244, A375, and mel526 and the iNOS-negative colon cancer line WIDR were assessed by Western blot for iNOS protein expression. B, \( 5 \times 10^5 \) A375 melanoma cells were injected s.c. into recipient NOD/SCID mice and allowed to form 0.5-cm tumors. Xenografts were then harvested, and sections were stained by immunohistochemistry for iNOS and the stable NO reaction product nitrotyrosine. Representative MRI (tumor in false color) and immunohistochemistry results.
As basal NO production by melanoma lines was consistently at the lower limit of detection of the relatively insensitive Griess assay, further NO measurements were made with the TBI 4100 electrochemical NO probe (WPI Instruments), which is capable of measuring nanomolar levels of NO. All iNOS-positive melanoma lines produced nanomolar levels of NO (Fig. 2A). Basal NO levels in media from WIDR cells (which do not express iNOS mRNA; ref. 29) were just above those of medium alone; after transfection with an iNOS-encoding expression plasmid, WIDR/iNOS cells released NO at levels comparable with melanoma cells. The iNOS-selective competitive antagonist L-nil, when added to culture medium, inhibited NO release by WIDR/iNOS and melanoma cells in a dose-dependent fashion. The IC50, calculated from WIDR/iNOS cells (for which the only significant source of measurable NO is exogenously-supplied iNOS), was 595 μmol/L (Fig. 2B). The decrease in NO production by L-nil was not due to cytotoxicity, as viability of melanoma cells was not significantly affected by L-nil after up to 72 hours incubation (Fig. 2D).

To test whether L-nil could reach a biologically relevant and therapeutic concentration in vivo, C57BL/6 mice received L-nil (0.15%) in drinking water before i.p. challenge with LPS, a strong inducer of iNOS-dependent NO production from resident macrophages. L-nil treatment resulted in a strong decrease in serum nitrite levels after LPS challenge (Fig. 2C), showing the ability of orally given L-nil to inhibit iNOS activity in vivo.

Targeted inhibition of iNOS antagonizes human melanoma growth in vivo and extends survival of tumor-bearing mice. We provided L-nil (0.15% in drinking water) or plain drinking water control to NOD/SCID mice bearing mel624 xenografts beginning on day 3 after tumor implantation. L-nil suppressed tumor growth for as long as the inhibitor was supplied in drinking water (4 weeks; Fig. 3A). Whereas the tumor resumed growth after discontinuation of treatment, L-nil–treated mice survived significantly longer than untreated mice (Fig. 3B). Another iNOS antagonist, 1,3-PBIT, had a similar effect on mel624 growth when given in drinking water (Fig. 3C), suggesting that the antitumor effect of L-nil is due to iNOS inhibition rather than non-specific cytotoxicity or activity against non-NOS targets. Neither L-nil nor 1,3-PBIT caused any outward signs of toxicity in mice (behavioral changes, reduction in food or water intake, ruffled fur, hunched posture) after 4 weeks at the indicated concentration. There was no significant difference in body weight among control, L-nil, and 1,3-PBIT groups (Fig. 3D) at the indicated doses; however, wasting and behavioral changes were seen at higher doses of 1,3-PBIT.
iNOS inhibition results in decreased intratumoral microvessel density, downregulation of Bcl-2, and increased intratumoral apoptosis in vivo. As expected, treatment with L-nil did not appreciably alter in vivo expression of iNOS protein levels in melanoma xenografts, but reduced staining for nitrotyrosine confirmed the inhibition of intratumoral NO levels (Fig. 4). Because NO is known to have angiogenic activity via induction of vascular endothelial growth factor expression, we examined microvessel density in mel624 xenografts by immunohistochemistry for the vascular endothelial marker CD31 (Fig. 4). After 19 days of L-nil treatment, there was a significant ($P = 0.006$) decrease in density of CD31+ microvessels in L-nil–treated mice, although the vessels’ diameter was significantly greater (insert; average of 27 ± 2 pixels versus 10 ± 1 pixels in diameter). Because NO has been suggested to inhibit apoptosis in human melanoma cells in vitro (21), we next examined the density of apoptotic (TUNEL+) cells in xenografts from L-nil–treated and control-treated mice (Fig. 5A). Tumors from L-nil–treated mice contained ~3-fold more TUNEL+ cells than tumors from control mice, suggesting that the decreased growth of xenografts is at least in part due to a higher rate of cell death. Staining of serial sections with TUNEL and the vascular marker CD31 did not reveal consistent colocalization of TUNEL staining and CD31 (Fig. 5A), as would be expected if endothelial cells were undergoing frequent apoptosis. Distribution of apoptotic cells was also not restricted to areas lacking vessel growth.

To further explore the possible mechanisms of tumor inhibition by L-nil, we measured the protein and phosphorylated protein expression of 59 proteins involved in signal transduction and cell survival pathways by RPPAs (Supplementary Table S1). Two tumors were analyzed for each treatment, and multiple samples from each tumor were analyzed to control for intratumoral heterogeneity. This analysis showed consistent decreases in multiple proteins, including several components of the phosphoinositide 3-kinase/AKT signaling pathway, although we did not observe changes in phosphorylated AKT itself (Supplementary Fig. S1). A potential link between iNOS inhibition and increased apoptosis in tumors from L-nil–treated mice was suggested by a marked decrease in Bcl-2 expression levels, which was confirmed by Western blotting analysis (Fig. 5B). Bcl-2 is not only a critical regulator of cell death in response to a variety of stimuli, but its posttranslational stability is also known to be regulated by NO via nitrosylation of key cysteine residues (14). This mechanism is supported by the observation that prolonged treatment of A375 melanoma cells...
with 1 mmol/L L-nil almost completely abolished Bcl-2 expression (Fig. 5C).

Combination therapy with cisplatin + L-nil is effective against human melanoma cell lines in vivo. NO is known to affect signal transduction pathways regulating apoptosis (Bcl-2, caspase III, p53, and other survival and death pathways), and depletion of NO with the chemical NO scavenger PTIO has been shown to enhance sensitivity of human melanoma cells to cisplatin in vitro (21). Because we found that L-nil downregulated Bcl-2 expression in melanoma cells in vivo, we hypothesized that L-nil treatment would potentiate cytotoxic therapy of melanoma. Because resistance to conventional cytotoxic agents is a common clinical problem, we used both a relatively cisplatin-sensitive human melanoma line (A375) and the line mel624, which is 3-fold to 4-fold more resistant to cisplatin (Fig. 6A). Whereas neither cell line was sensitive to L-nil treatment alone (Fig. 2C) in vitro, pretreatment of A375 cells with L-nil for 5 days increased sensitivity of cells to cisplatin-induced killing (Fig. 6B).

We then tested combination therapy with L-nil plus cisplatin against human melanoma in vivo. Whereas cisplatin or L-nil alone only partially suppressed the growth of established mel624 and A375, combination therapy with L-nil and cisplatin inhibited growth of mel624 and A375 more efficiently than either drug alone (Fig. 6C) without additional toxicity. Although mice received only a single three-dose course of cisplatin treatment, continued treatment with L-nil alone was sufficient to suppress tumor growth.

Discussion

Whereas inflammation plays a well-established role in the initiation and progression of certain cancers, the ability of proinflammatory molecules to support the persistence of established cancer is a relatively recent observation. The present data confirm the hypothesis that iNOS is one such inflammatory mediator capable of promoting the survival and proliferation of human melanoma cells in vivo. More importantly, we show that iNOS can be readily targeted in melanoma without overt toxicity and that iNOS inhibition reverses the chemoresistance of human melanoma.

Our data are consistent with previous studies, mostly in vitro, which show an effect of NOS inhibition on tumor growth, survival, or both (30–33). Our study is among the first to show dramatic antitumor activity of iNOS-selective inhibitors in vivo and feasibility of combination treatment of cancer combining targeted iNOS inhibition and cytotoxic chemotherapy. There are several potential mechanisms by which iNOS antagonists may inhibit tumor growth, either directly or by sensitizing cells to other forms of stress, such as hypoxia or reactive oxygen species–mediated stress. Mechanisms by which iNOS inhibition may affect the resistance of cancer cells to apoptosis include interference with phosphoinositide 3-kinase/AKT–mediated overexpression of survivin (34, 35) or reversal of caspase inactivation (15) or Bcl-2 stabilization mediated by S-nitrosylation of these proteins (14). Indeed, we observed that L-nil induced downregulation of Bcl-2 protein expression in vivo and in vitro, suggesting that this is one mechanism by which iNOS inhibition may enhance susceptibility of cancer cells to apoptosis. L-nil’s relative lack of direct cytotoxicity in vitro (Fig. 2C), despite causing markedly increased tumor apoptosis in vivo, may reflect the relative absence of stressful stimuli (hypoxia, acidosis, cytokines, etc.) in vitro, which might force melanoma cells to depend on Bcl-2 expression for survival.

NO has well-established proangiogenic properties, and stimulation of angiogenesis is one proposed mechanism...
by which iNOS expression may support tumor growth (36, 37). We observed a nearly 2-fold decrease in tumor microvessel density in tumors from L-nil–treated mice, although the average caliber of the remaining vessels was greater (see inset, Fig. 4, bottom). As both suppression of vascularization and increased apoptosis may lead to a diminished overall rate of tumor growth, they may act additively or synergistically to cause the observed significant decrease in tumor growth in vivo.

Another potential mechanism of tumor growth suppression is interference with the NO-dependent proliferation of tumor cells. Whereas relatively few studies have attempted to disentangle the dual effect of NO on tumor cell survival and proliferation, NO has been shown to increase proliferation of human breast cancer cells in vitro via stimulation of the AKT/mTOR pathway and downstream upregulation of cyclin D1 (38). It is interesting that, of the nearly 60 proteins and phosphorylated proteins we examined by RPPA, multiple proteins in the mTOR pathway, including P70S6K, and mTOR were significantly and coordinately downregulated by L-nil treatment in vivo (Supplementary Fig. S1). We are currently investigating the effect of iNOS/NO inhibition on melanoma proliferation and regulation of the AKT/mTOR and other proliferation/survival pathways.

When we tested the in vivo antitumor efficacy of iNOS inhibition, alone and in combination with cisplatin, both lines tested exhibited nearly complete sensitivity to combination treatment with doses of L-nil and cisplatin that were only partly therapeutic as single agents. Because melanoma quickly acquires resistance to conventional chemotherapy in the clinical setting, restoring the efficacy of cytotoxic chemotherapy by adding a well-tolerated targeted agent to the regimen is an exciting possibility that deserves testing in clinical trials. It is notable that although cisplatin was given for only a single three-dose course, the beneficial effect of combination therapy persisted for significantly longer, so long as L-nil was continued. This suggests the potential for therapeutic regimens, which use up-front or periodic combination chemotherapy/targeted therapy followed by maintenance therapy with a relatively nontoxic targeted iNOS inhibitor.

The use of L-nil and related compounds in vivo seems promising because of the magnitude of tumor inhibition...
(by a factor of 3-fold to 5-fold at 4 weeks) and its favorable toxicity profile. Whereas it important to note that we did not monitor changes in blood pressure, a potential side effect of chronic treatment with NO active compounds, we did not observe any overt L-nil toxicity at doses of up to 0.2% (7.7 mmol/L in drinking water) when given for up to 4 weeks. This is despite our observation that L-nil is a relatively weak inhibitor in vitro, with an IC₅₀ of nearly 600 μmol/L (Fig. 2A and B). This is significantly higher than the published IC₅₀ values for L-nil in cell-free enzyme assays (22) and likely reflects the need for active transport of the molecule across the cell membrane (39) as well as degradation in aqueous solution over the course of 24-hour culture (Cayman Chemical product information). Variations in the efficiency of transport may also contribute to differences in the efficacy of L-nil–mediated NO suppression among our panel of cell lines. In our in vitro experiments, the predicted concentration of L-nil required to suppress >90% NO production varied from 1 to >3 mmol/L. This is quite similar to the theoretical concentration of L-nil in tissues in vivo during treatment (assuming complete absorption of the drug in a drinking water volume of 6 mL/d), which would be 2.13 mmol/L. Because complete drug distribution into the bloodstream after oral administration is unlikely, the tumor suppressive effects observed in vivo are likely due to partial, rather than complete, inhibition of iNOS; this is also supported by the partial inhibition of intratumoral nitrotyrosine deposition (Fig. 4) and LPS-induced nitrite production in vivo (Fig. 2D) after treatment with 0.1% L-nil. This suggests that it may be possible to more efficiently inhibit iNOS by escalating the dose of L-nil, potentially improving its antitumor efficacy. As L-nil and similar NOS inhibitors have previously been the subject of noncancer human clinical trials (40–42), these findings have strong translational potential.

Our results do not exclude the possibility that the antitumor activity of L-nil is enhanced by inhibition of iNOS in other (host) cell types in vivo. This is particularly relevant because there is little direct cytotoxic effect of L-nil in vitro. iNOS can be expressed by host stromal, endothelial, and bone marrow–derived cells present in tumors in vivo, and inhibition of NO production by any of these cell types could indirectly affect tumor growth and viability. We are currently working to determine what role, if any, host-derived iNOS plays in the antitumor effect of L-nil on melanoma xenografts in vivo. Another limitation of our experimental system is reliance on small molecule iNOS-selective inhibitors to implicate iNOS-derived NO in the growth and resistance to cytotoxicity of tumor cells. No chemical inhibitor is completely selective, and in fact many so-called “targeted” agents can act on a relatively broad set of molecular targets (43). However, the similar antitumor activity of two structurally dissimilar iNOS inhibitors, L-nil and 1,3 PBIT (Fig. 3C), suggests that iNOS is indeed the primary target of these treatments. Finally,

**Fig. 6.** L-nil enhances sensitivity of human melanoma cells to antitumor effect of cisplatin in vitro and in vivo. A, A375 and mel624 cells were treated with the indicated concentrations of cisplatin for 48 h, and proliferation was measured by XTT assay. Data are presented as mean ± SEM and are representative of three independent experiments. B, 2×10⁶ A375 melanoma cells were cultured in the indicated concentration of L-nil for 5 d. On day 3, the medium was discarded, and cells were incubated for another 48 h in fresh media with the indicated concentration of L-nil and/or cisplatin. Cells were then trypsinized, and cell death was assessed by Annexin V staining and flow cytometry. Data are presented as mean ± SEM and are representative of three independent experiments. *P = 0.012; **P = 0.003. C, SCID-NOD mice were injected s.c. with 5×10⁶ A375 or mel624 cells and treated with 0.2% L-nil for the indicated time, starting on day 3. Cisplatin was given as three i.p. injections of 2.5 mg/kg (A375) or 6 mg/kg (mel624) spaced 3 d apart, beginning on day 13 (mel624) or day 25 (A375). The numbers to the right of each curve give the fraction of surviving mice. For A375 P < 0.001 for CDDP + L-nil versus CDDP alone; P = 0.009 for CDDP + L-nil versus L-nil alone. For mel624 P < 0.001 for CDDP + L-nil versus CDDP alone; P = 0.010 for CDDP + L-nil versus L-nil alone on day 30.
although NO has been shown to suppress T-cell–dependent immune responses, our immunodeficient human/mouse xenograft model is not ideal for studying effects of iNOS inhibition on tumor-mediated immunosuppression and interference with endogenous antitumor immunity.

In summary, this study clearly shows the robust antitumor effect of iNOS-selective small molecule inhibitors in a preclinical human melanoma model and their ability to synergize with conventional cytotoxic chemotherapy in vivo. The lack of appreciable toxicity associated with chronic L-nil treatment and its ability to enhance the efficacy of cisplatin suggest that iNOS inhibition, alone or in combination with cytotoxic chemotherapy, deserves evaluation in clinical trials for melanoma and other solid tumors. More generally, the regulation of inflammatory mediators in cancer is a promising approach to targeted cancer therapy and reversal of chemoresistance.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

References

Acknowledgments
We thank Dr. Nina Chinosomwatana (Department of Otolaryngology, Mount Sinai School of Medicine) for assistance in performing experiments (which contributed to Fig. 6B), Ajish George (Department of Biomedical Sciences, School of Public Health, SUNY Albany) for helpful advice regarding molecular pathway analysis, and the M.D. Anderson Small Animal Imaging Facility for the MRI analysis (supported by M.D. Anderson Cancer Center Support Grant CA016672).

Grant Support
P50 CA05349 (PP-DRP5 and PP-CDP5), University of Texas M.D. Anderson Cancer Center Specialized Programs of Research Excellence in Melanoma (E.A. Grimm, W.W. Overwijk, A.G. Sikora); NIDCD National Research Service Award Institutional Training Grant T32 D007367 (A. Gelbard, A.G. Sikora); American Head and Neck Society/American Academy of Otolaryngology Young Investigator Award (A.G. Sikora); and NCI/CA16672 supporting the M.D. Anderson Cancer Center Characterized Cell Line Core and RPPA Core Laboratories.

This work was partially supported by VENI Grant 916.046.014 from the Netherlands Organization for Scientific Research (NWO) to W.W.O. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received 11/25/2009; revised 01/22/2010; accepted 01/22/2010; published OnlineFirst 03/09/2010.
Clinical Cancer Research

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doi:10.1158/1078-0432.CCR-09-3123

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