Honokiol Suppresses Epidermal Growth Factor Receptor Signaling and Enhances the Antitumor Effects of Epidermal Growth Factor Receptor Inhibitors


Abstract

Purpose: This study aimed to investigate the utility of honokiol, a naturally occurring compound, in the treatment of head and neck squamous cell carcinoma (HNSCC) as well as its ability to target the epidermal growth factor receptor (EGFR), a critical therapeutic target in HNSCC, and to enhance the effects of other EGFR-targeting therapies.

Experimental Design: Human HNSCC cell lines and the xenograft animal model of HNSCC were used to test the effects of honokiol treatment.

Results: Honokiol was found to inhibit growth in human HNSCC cell lines, with 50% effective concentration (EC50) values ranging from 3.3 to 7.4 μmol/L, and to induce apoptosis, as shown through Annexin V staining. These effects were associated with inhibition of EGFR signaling, including downstream inhibition of mitogen-activated protein kinase, Akt, and signal transducer and activator of transcription 3 (STAT3), and expression of STAT3 target genes, Bcl-XL and cyclin D1. Furthermore, honokiol enhanced the growth inhibitory and anti-invasion activity of the EGFR-targeting agent erlotinib.

Although HNSCC xenograft models did not show significant inhibition of in vivo tumor growth with honokiol treatment alone, the combination of honokiol plus cetuximab, a Food and Drug Administration–approved EGFR inhibitor for this malignancy, significantly enhanced growth inhibition. Finally, HNSCC cells rendered resistant to erlotinib retained sensitivity to the growth inhibitory effects of honokiol.

Conclusions: These results suggest that honokiol may be an effective therapeutic agent in HNSCC, in which it can augment the effects of EGFR inhibitors and overcome drug resistance.

Head and neck squamous cell carcinoma (HNSCC) is one of the most commonly occurring malignancies worldwide. Advances in therapy for HNSCC have only modestly improved the mortality rate, which has remained at 50% for the past several decades (1). Available therapies, including surgical resection, radiation, and conventional chemotherapy, are often associated with severe morbidity affecting the vital structures of the head and neck, have side effects, and are limited by therapeutic resistance. The need exists, therefore, for the discovery of new therapies, including complementary therapies that can be given in combination with available treatments to allow for lower doses of toxic drugs and to overcome drug resistance.

Honokiol is a natural compound derived from the bark of the magnolia tree and is used in traditional Chinese medicine. Studies have shown various ways in which honokiol may have a therapeutic benefit, including its ability to behave as a muscle relaxant; to have anti-inflammatory, antimicrobial, and antioxidant activity; and indications that it may be useful in protecting against hepatotoxicity, neurotoxicity, thrombosis, and angiopathy (2). Interest in the role that honokiol may play in cancer therapy began with a study showing the prevention of skin papillomas in mice (3). Subsequent studies showed the anticancer activities of honokiol in a variety of cancer cell lines (4–11) and xenograft models (4, 6, 7, 9, 10, 12–14).

In several cancer models, honokiol has been found to alter molecular targets that are known to affect tumor cell growth and survival. One of the most commonly proposed mechanisms of honokiol’s antitumor activity is inhibition of the NFκB signaling pathway. NFκB is a transcription factor that contributes to several physiologic processes (e.g., inflammation) but also regulates the expression of genes that are involved in cancer, including...
Honokiol has been shown to enhance the effects of a variety of chemotherapeutic agents and small molecule inhibitors including bortezomib (29), fludarabine (5), cladribine (29), chlorambucil (29), doxorubicin (14, 17), adriamycin (36), paclitaxel (14, 17), docetaxel (10), SAHA (14), lapatinib (33), rapamycin (33), or cisplatin (12) in different cancer models. In the current study, we hypothesized that honokiol can be used to target EGFR signaling via STAT3 in the treatment of HNSCC and may also enhance the effects of EGFR-targeting therapies, erlotinib and cetuximab.

Materials and Methods

Reagents and cells. The HNSCC cell lines Cal-33, derived from an oral squamous cell carcinoma (37), and 1483, from an oropharyngeal squamous cell carcinoma (38), were maintained in DMEM medium with 10% heat-inactivated fetal bovine serum at 37°C in a humidified incubator with 5% CO2. Cal-33 cells were provided by Dr. Gerard Milano in 2006 (Centre Anotoine-Lacassagne, Nice, France) and 1483 cells were obtained in 2007 from Dr. Gary Clayman (M.D. Anderson Cancer Center, Houston TX). 686LN and 686LNR30 cells are isogenic models of acquired EGFR TKI resistance in vitro and were obtained from Dr. Georgia Chen (Emory University, Atlanta, GA; ref. 39). All the cell lines were genotyped and validated in 2008 using the AmpFISTR Identifiler PCR Amplification Kit (Applied Biosystems). The cells are genotyped on a regular basis (twice in 2009) to ensure the validity. Honokiol is a natural product extracted from seed cone of Magnolia grandiflora as previously described (4). In treatments of cell cultures, honokiol was dissolved in 100% ethanol as a vehicle and in 20% Intralipid (Baxter Healthcare) for animal treatments. Erlotinib (Chemietek) was dissolved in 100% DMSO, as a vehicle.

Proliferation assay. HNSCC cells (1-3 × 10⁴/well) were cultured overnight and treated with honokiol, erlotinib or the corresponding vehicles, in DMEM/1% serum, the following day. After 72 hours, the cells were harvested by trypsinization, and live cells were counted after staining for trypan blue dye exclusion. Each experiment was done with triplicate samples, and the average percent survival was calculated as a comparison with cells treated with the vehicle alone. The EC₅₀ was calculated using Prism software version 4.03 (GraphPad Software Inc).

Apoptosis assay. Cal-33 cells (5 × 10⁴/well) were seeded and, the following day, treated with either honokiol (10 μmol/L) or ethanol, as the vehicle, for 72 hours. Cells were then harvested and stained with Cy3-labeled Annexin V, according to the manufacturer's instructions (Annexin V-Cy3 apoptosis detection kit, BioVision). Stained cells were imaged using a fluorescent microscope (Nikon), and the numbers of Annexin V–positive cells were counted (at least three fields per sample) using ImageJ software (NIH).

Invasion assay. 686LN cells (1.5 × 10⁵) were plated in serum-free DMEM F12 containing epidermal growth factor (EGF) alone (10 ng/mL), EGF with honokiol

Materials and Methods

Reagents and cells. The HNSCC cell lines Cal-33, derived from an oral squamous cell carcinoma (37), and 1483, from an oropharyngeal squamous cell carcinoma (38), were maintained in DMEM medium with 10% heat-inactivated fetal bovine serum at 37°C in a humidified incubator with 5% CO₂. Cal-33 cells were provided by Dr. Gerard Milano in 2006 (Centre Anotoine-Lacassagne, Nice, France) and 1483 cells were obtained in 2007 from Dr. Gary Clayman (M.D. Anderson Cancer Center, Houston TX). 686LN and 686LNR30 cells are isogenic models of acquired EGFR TKI resistance in vitro and were obtained from Dr. Georgia Chen (Emory University, Atlanta, GA; ref. 39). All the cell lines were genotyped and validated in 2008 using the AmpFISTR Identifiler PCR Amplification Kit (Applied Biosystems). The cells are genotyped on a regular basis (twice in 2009) to ensure the validity. Honokiol is a natural product extracted from seed cone of Magnolia grandiflora as previously described (4). In treatments of cell cultures, honokiol was dissolved in 100% ethanol as a vehicle and in 20% Intralipid (Baxter Healthcare) for animal treatments. Erlotinib (Chemietek) was dissolved in 100% DMSO, as a vehicle.

Proliferation assay. HNSCC cells (1-3 × 10⁴/well) were cultured overnight and treated with honokiol, erlotinib or the corresponding vehicles, in DMEM/1% serum, the following day. After 72 hours, the cells were harvested by trypsinization, and live cells were counted after staining for trypan blue dye exclusion. Each experiment was done with triplicate samples, and the average percent survival was calculated as a comparison with cells treated with the vehicle alone. The EC₅₀ was calculated using Prism software version 4.03 (GraphPad Software Inc).

Apoptosis assay. Cal-33 cells (5 × 10⁴/well) were seeded and, the following day, treated with either honokiol (10 μmol/L) or ethanol, as the vehicle, for 72 hours. Cells were then harvested and stained with Cy3-labeled Annexin V, according to the manufacturer's instructions (Annexin V-Cy3 apoptosis detection kit, BioVision). Stained cells were imaged using a fluorescent microscope (Nikon), and the numbers of Annexin V–positive cells were counted (at least three fields per sample) using ImageJ software (NIH).

Invasion assay. 686LN cells (1.5 × 10⁵) were plated in serum-free DMEM F12 containing epidermal growth factor (EGF) alone (10 ng/mL), EGF with honokiol

Materials and Methods

Reagents and cells. The HNSCC cell lines Cal-33, derived from an oral squamous cell carcinoma (37), and 1483, from an oropharyngeal squamous cell carcinoma (38), were maintained in DMEM medium with 10% heat-inactivated fetal bovine serum at 37°C in a humidified incubator with 5% CO₂. Cal-33 cells were provided by Dr. Gerard Milano in 2006 (Centre Anotoine-Lacassagne, Nice, France) and 1483 cells were obtained in 2007 from Dr. Gary Clayman (M.D. Anderson Cancer Center, Houston TX). 686LN and 686LNR30 cells are isogenic models of acquired EGFR TKI resistance in vitro and were obtained from Dr. Georgia Chen (Emory University, Atlanta, GA; ref. 39). All the cell lines were genotyped and validated in 2008 using the AmpFISTR Identifiler PCR Amplification Kit (Applied Biosystems). The cells are genotyped on a regular basis (twice in 2009) to ensure the validity. Honokiol is a natural product extracted from seed cone of Magnolia grandiflora as previously described (4). In treatments of cell cultures, honokiol was dissolved in 100% ethanol as a vehicle and in 20% Intralipid (Baxter Healthcare) for animal treatments. Erlotinib (Chemietek) was dissolved in 100% DMSO, as a vehicle.

Proliferation assay. HNSCC cells (1-3 × 10⁴/well) were cultured overnight and treated with honokiol, erlotinib or the corresponding vehicles, in DMEM/1% serum, the following day. After 72 hours, the cells were harvested by trypsinization, and live cells were counted after staining for trypan blue dye exclusion. Each experiment was done with triplicate samples, and the average percent survival was calculated as a comparison with cells treated with the vehicle alone. The EC₅₀ was calculated using Prism software version 4.03 (GraphPad Software Inc).

Apoptosis assay. Cal-33 cells (5 × 10⁴/well) were seeded and, the following day, treated with either honokiol (10 μmol/L) or ethanol, as the vehicle, for 72 hours. Cells were then harvested and stained with Cy3-labeled Annexin V, according to the manufacturer's instructions (Annexin V-Cy3 apoptosis detection kit, BioVision). Stained cells were imaged using a fluorescent microscope (Nikon), and the numbers of Annexin V–positive cells were counted (at least three fields per sample) using ImageJ software (NIH).

Invasion assay. 686LN cells (1.5 × 10⁵) were plated in serum-free DMEM F12 containing epidermal growth factor (EGF) alone (10 ng/mL), EGF with honokiol
(5 μmol/L) and/or erlotinib (5 μmol/L) or the corresponding vehicles, in a matrigel invasion chamber insert (BD Biosciences). The outer well contained DMEM F12/10% fetal bovine serum, as a chemoattractant. After 24 hours' incubation, uninvaded cells were removed and the invaded cells in the matrigel were fixed, stained with Hema 3 (Fisher Scientific), and counted under 200× magnification.

**Western blotting.** For cell lysates used in Western blots, cells were cultured in DMEM/10% fetal bovine serum for 24 hours and then in serum-free DMEM containing either honokiol or vehicle for another 24 hours. Tumor lysates from animal studies were extracted after homogenization of tumor tissue. The proteins from whole cell lysates were resolved by SDS-PAGE and transferred onto nitrocellulose membranes (Bio-Rad) by a semidry transfer apparatus (Bio-Rad). The membrane was blocked overnight with primary antibodies with 5% skim milk in TBS-T. After washing in TBS-T, membranes were incubated with secondary antibodies (antirabbit or antimouse IgG-horseradish peroxidase conjugate from Bio-Rad) and then with a luminol kit (Santa Cruz Biotechnology). Prized samples were resolved by SDS-PAGE and transferred onto nitrocellulose membranes (Bio-Rad) by a semidry transfer apparatus (Bio-Rad). Primary antibodies for STAT3, pSTAT3, pEGFR, phospho-p44/p42 mitogen-activated protein kinase (MAPK), AKT, pAKT (Cell Signaling Technology), cyclin D1, Bcl-XL (Santa Cruz Biotechnology), EGFR (BD Transduction labs) and β-actin (CalBiochem) were used to probe membranes. Relative densitometric values were calculated using DigiDoc software and normalized, for each protein, to the corresponding band representing the house-keeping gene, β-actin.

**In vivo tumor xenograft study.** Female athymic nude mice (5-6 weeks old; n = 30) were purchased from Harlan and were housed in a pathogen-free animal facility. The animal study protocols were approved by the institutional animal care and use committee. 1483 cells (2 × 10⁶/mouse) were harvested by trypsinization, washed in PBS, resuspended in saline, and s.c. injected into the flank of each mouse. After outgrowth of palpable tumors (7 days), the mice were randomized, by tumor volume, to three treatment groups (8 in the vehicle control group, 14 in the cetuximab group, and 10 in the combination group).

The cetuximab treatment group received 0.8 mg/mouse/day, by i.p. injection, twice per week. The combination treatment group received both cetuximab, twice per week, and honokiol, at 3 mg/mouse/day in 20% Intralipid (Baxter Healthcare), three times per week. Tumors were measured using digital calipers (Control Company) at least three times per week, and tumor volumes were calculated using the following formula: volume = L × (W)²/2 (L, longest diameter; W, shorter diameter). At the end of the study, the mice were euthanized and tumor tissues were harvested and frozen for analysis by immunoblot.

**Statistical analyses.** All statistical analyses of in vitro results were done using the nonparametric Mann-Whitney or Wilcoxon tests. Analysis of tumor growth rates in the xenograft model was done using a general linear model, assuming that animals are random effects. Tumor volume data were examined for the interaction between treatment group and day of observation to test whether the slopes of the growth curves were significantly different between groups.

**Results**

**Honokiol inhibits growth and induces apoptosis in HNSCC cell lines.** The in vitro growth inhibitory and pro-apoptotic activities of honokiol have been shown in several cancer cell lines (4–11). In the current study, two HNSCC cell lines, 1483 and Cal-33, were treated for 72 hours with honokiol at concentrations ranging from 0.01 μmol/L to 100 μmol/L and compared with the vehicle (ethanol) alone. 50% effective concentration (EC₅₀) values were 7.44 μmol/L for 1483 and 3.80 μmol/L for Cal-33 (Fig. 1A). These values are comparable with or lower than EC₅₀ values seen in other cancer cell types (5, 7, 10, 33).

To determine the role of programmed cell death in the growth inhibitory effects of honokiol, Cal-33 cells were treated with honokiol followed by Annexin V staining. As shown in Fig. 1B, honokiol increased the number of apoptotic cells by 7.4-fold, suggesting that...
growth inhibition, at least in part, was due to the induction of apoptosis in HNSCC (Fig. 1B; \( P = 0.03 \)).

**Honokiol inhibits the EGFR signaling pathway.** The EGFR signaling pathway, including activation of STAT3 and increased expression of STAT3 target genes, plays an important role in HNSCC. One approved HNSCC therapy (cetuximab) and various investigational therapies target EGFR and/or STAT3. By densitometric analysis of immunoblot bands, 24-hour honokiol (10 \( \mu \)mol/L) treatment resulted in an 88% decrease in EGFR in Cal-33 cells (Fig. 2A; \( P = 0.03 \)). In studies of Cal-33 cells treated with both EGF and honokiol, we found that honokiol treatment resulted in a decrease in phosphorylated EGFR to a lesser degree than the corresponding decrease in total EGFR expression levels (data not shown), which suggests that honokiol likely primarily affects levels of total EGFR.

Honokiol-induced EGFR inhibition was associated with a 61% downstream inhibition of STAT3 in Cal-33 cells (Fig. 2B; \( P = 0.03 \)), as well as 63% and 75% decreases in expression of Bcl-X\(_L\) and cyclin D1, two STAT3 target genes that inhibit apoptosis and promote the cell cycle, respectively (Fig. 2C; \( P = 0.03 \)). Honokiol treatment also resulted in decreased levels of phosphorylated p42/p44 MAPK and phosphorylated Akt, by 73% and 77%, respectively, suggesting a global effect on EGFR signaling (Fig. 2D; \( P = 0.03 \)). As EGFR signaling via STAT3 is known to play a key role in...
Honokiol enhances the activity of erlotinib in HNSCC cells. Honokiol was tested for its ability to inhibit growth in HNSCC models of EGFR inhibitor resistance. The clone 686 LNR30 was chosen for its relative resistance to erlotinib, having an EC_{50} of 134.7 μmol/L, compared with 15.13 μmol/L in the parental cell line 686 LN (39). Honokiol, on the other hand, was equally effective in inhibiting growth of the parental and erlotinib-resistant cell lines (Fig. 3A). These results suggest that honokiol is able to overcome resistance to erlotinib.

Cetuximab, which is currently FDA approved for treatment of HNSCC, is known to inhibit HNSCC growth in vitro, but has minimal effects in vivo. Therefore, in our studies of honokiol’s effect in combination with EGFR inhibition, we used erlotinib in cell culture experiments and cetuximab in animal studies. Honokiol enhanced the growth inhibition seen with erlotinib treatment in vitro. Combining honokiol at its EC_{50} in treatment of 686 LN cells (3.3 μmol/L) with the approximate EC_{50} for erlotinib (15.1 μmol/L) resulted in a 77.0% growth inhibition, compared with 52% growth inhibition for erlotinib alone, a 1.5-fold enhancement (Fig. 3B; P = 0.03), further supporting the use of honokiol in combination with EGFR blockade in HNSCC.

Honokiol has been shown to decrease the invasiveness of fibrosarcoma (40) and breast cancer cells (33). We assayed the invasiveness of 686 LN cells using matrigel invasion assays. Cells were plated in serum-free medium containing EGF (10 ng/mL) as a stimulant and either honokiol (5 μmol/L), erlotinib (5 μmol/L), a combination of both drugs, or their corresponding vehicles as controls. Medium containing 10% serum was present in the lower chamber as a chemoattractant. After 24 hours, honokiol was found to inhibit invasion, on its own, by 72.7%, compared with vehicle control (P = 0.03). Combined treatment with erlotinib plus honokiol, however, enhanced the anti-invasion activity of erlotinib but did not significantly increase the effects of honokiol alone on HNSCC invasion (P = 0.03; Fig. 3C). Neither drug alone induces significant growth inhibition at these concentrations after 24 hours, and, in combination, they decrease growth by only 23.5%, compared with an 83.3% decrease in number of invaded cells (data not shown). It is likely, therefore, that these data represent true decreases in invasiveness rather than just growth inhibition. Considering the large impact of invasion and metastasis on the clinical outcome of HNSCC, these data suggest that honokiol may have a role in HNSCC treatment, especially in combination with EGFR-targeting therapies.

Honokiol enhances the growth inhibitory activity of cetuximab and inhibits EGFR signaling in vivo. Honokiol has been found to prevent the formation of skin papillomas in vivo (3) and to inhibit growth of xenografts derived from HNSCC growth both in vitro and in vivo, these data indicate that by inhibiting both EGFR levels and STAT3 activation, honokiol may have potential utility in the treatment of HNSCC. This effect on EGFR signaling and STAT3 has recently been shown in a breast cancer cell line, albeit at much higher concentrations of honokiol (60 μmol/L compared with 10 μmol/L in our models; ref. 8).
from angiosarcoma (4), colorectal (6), prostate (7, 35), gastric (13), breast (14), lung (12), and ovarian (35) cancer xenografts, in vivo. In a pilot study of honokiol’s in vivo anticancer activity the average growth rates of HNSCC xenografts were not significantly reduced in animals treated with honokiol compared with the vehicle (14.31 mm³/day versus 19.87 mm³/day; data not shown). These data suggest that honokiol may not be effective as a single agent for in vivo treatment of HNSCC.

To determine whether or not honokiol enhances the anticancer activity of EGFR-inhibiting therapy in vivo, as was seen in vitro, female nude mice were inoculated with 1483 HNSCC cells and, after tumor outgrowth and randomization to treatment groups, treated with vehicle, cetuximab alone (0.8 mg/mouse 2 days/week), cetuximab plus honokiol (3 mg/mouse 3 days/week) on alternating days. The vehicle control group received Intralipid and saline by i.p. injection on corresponding days. Tumor measurements were done at least three times per week. B, lysates were extracted from tumors of mice in the cetuximab alone and cetuximab plus honokiol group. Selected lysates, probed by immunoblot for PSTAT3, STAT3, EGFR, and cyclin D1, are shown. C, densitometric values representing averages from lysates of tumors from all mice in each of these two groups, and which have been normalized to β-actin (P = 0.0008, 0.02, and 0.007 for EGFR, PSTAT3, and cyclin D1, respectively).

Fig. 4. The in vivo effects of honokiol in an HNSCC xenograft therapy model. A, nude mice were inoculated with 2 x 10⁶ 1483 cells, s.c., into their flanks. After tumor outgrowth mice were randomized to treatment groups, based on tumor volume. Mice were treated with either cetuximab (0.8 mg/mouse, 2 days/week), or cetuximab plus honokiol (3 mg/mouse, 3 days/week) on alternating days. The vehicle control group received Intralipid and saline by i.p. injection on corresponding days. Tumor measurements were done at least three times per week. B, lysates were extracted from tumors of mice in the cetuximab alone and cetuximab plus honokiol group. Selected lysates, probed by immunoblot for PSTAT3, STAT3, EGFR, and cyclin D1, are shown. C, densitometric values representing averages from lysates of tumors from all mice in each of these two groups, and which have been normalized to β-actin (P = 0.0008, 0.02, and 0.007 for EGFR, PSTAT3, and cyclin D1, respectively).

Leeman-Neill et al.

Clin Cancer Res; 16(9) May 1, 2010
Clinical Cancer Research

2576

Clin Cancer Res; 16(9) May 1, 2010
Clinical Cancer Research

Downloaded from clincancerres.aacrjournals.org on April 13, 2017. © 2010 American Association for Cancer Research.
Immunoblots of tumor lysates showed decreased levels of EGFR, tyrosine phosphorylated STAT3 (pSTAT3), and cyclin D1 in mice treated with cetuximab plus honokiol compared with cetuximab alone (P = 0.0008, 0.02, and 0.007 for EGFR, pSTAT3, and cyclin D1, respectively; Fig. 4B and C).

**Discussion**

The therapeutic potential, including antitumor activity, of honokiol, a natural product derived from the magnolia plant and used in traditional Chinese medicine, has been reported in various preclinical models. In the current study, we investigated honokiol’s potential utility in the treatment of HNSCC. Honokiol was found to inhibit growth and induce apoptosis in HNSCC cell lines and to enhance the growth-inhibitory and anti-invasion activities of the EGFR-targeting TKI erlotinib. Furthermore, EGFR signaling, STAT3 activity, and expression of STAT3 target genes were inhibited upon honokiol treatment. Finally, honokiol was found to enhance the efficacy of the EGFR-targeting antibody cetuximab, and inhibit EGFR signaling *in vivo*.

Many studies have focused on targeting EGFR and STAT3 with a variety of engineered molecules, including antibodies, small molecule inhibitors, oligonucleotides, peptidomimetics, and others (19, 28). Several natural compounds have also been found to inhibit STAT3, mainly via effects on upstream signaling molecules, in different models (19, 41–44). Our rationale for investigating the ability of honokiol to target EGFR and STAT3 signaling included evidence of STAT3 inhibition in honokiol treatment of a multiple myeloma cell line (29) and, more recently, inhibition of EGFR and STAT3 signaling in breast cancer cells (8). Furthermore, honokiol has been found to inhibit other signaling molecules upstream of STAT3, including Src (4) and gp130 (29), and to inhibit NF-kB (9, 11, 16, 17), which is known to experience crosstalk with STAT3. Finally, honokiol induces downregulation of various STAT3 target genes (5, 7, 17, 32, 33, 35).

Honokiol was found to decrease expression levels of total EGFR both *in vitro* and *in vivo* (Figs. 2A and 4B). Cellular exposure to various anticancer agents has been shown to affect total EGFR levels through a variety of different mechanisms, including lysosomal and proteosomal degradation (45–47), caspase-mediated degradation (48), and decreased transcription (49), and to affect surface levels through receptor internalization (47). Furthermore, a complex balance between EGFR recycling and degradation can be altered, in multiple ways, by different drug treatments and EGFR mutations (50). Honokiol’s structure contains two phenolic groups that can scavenge free radicals (2), suggesting the potential for nonspecific effects on multiple signaling pathways.

A recent study showed decreases in heat shock protein (hsp) 90 in breast cancer cells treated with high concentrations of honokiol (100 μmol/L; ref. 8). It is possible, therefore, that honokiol regulates EGFR degradation and recycling through similar mechanisms to either of the hsp90 inhibitors herbimycin A and geldanamycin (47).

Currently available treatments for HNSCC, including chemotherapy and radiation, can contribute to the morbidity and mortality associated with this disease. In addition, patients can develop resistance to standard chemotherapeutics. Critical regions of the head and neck such as the spinal cord can only tolerate a finite dose of radiation, thereby limiting the repeated clinical use of these therapies. Therefore, administration of a compound that enhances the activity of a HNSCC treatment may be a useful complementary strategy. In this study, we investigated honokiol’s ability to enhance the activity of erlotinib (Tarceva), a small molecule inhibitor that has shown promise in clinical trials in HNSCC (28). Honokiol results in decreased levels of pMAPK and pAKT as well, suggesting global inhibition of the EGFR signaling pathway, rather than a specific effect on STAT3 signaling. In addition, as honokiol has been found to inhibit Src, gp130, and EGFR, it is likely that in HNSCC cell lines, STAT3 signaling is inhibited through more than one upstream mechanism. Limited clinical responses to EGFR-targeting therapies, like cetuximab and erlotinib, may be due to activation of STAT3 through alternative signaling pathways, including Src and the IL-6 receptor. An agent that targets one of these alternative pathways, like honokiol, which inhibits Src and the IL-6 receptor, and at the same time enhances EGFR inhibition, may potentially be useful in overcoming the limited clinical responses to EGFR targeting agents seen to date in HNSCC patients. Liu et al. have shown that honokiol synergizes with lapatinib, another EGFR-targeting therapeutic, in the treatment of human epidermal growth factor receptor 2–overexpressing breast cancer cells (33).

In the current study, honokiol was found to enhance the growth inhibitory and anti-invasion activities of erlotinib *in vitro* as well as the growth inhibitory activity of cetuximab *in vivo*. To our knowledge, this is the first study showing the *in vivo* anticancer activity of honokiol in HNSCC. Our observations of the ability of honokiol to target EGFR/STAT3 signaling, to enhance the therapeutic effects of EGFR-targeting molecules, both *in vitro* and *in vivo*, and to inhibit growth of a cell line known to be resistant to other EGFR inhibitors suggest a potential role for honokiol in the treatment of HNSCC, particularly in combination with EGFR-inhibiting therapy.

**Disclosure of Potential Conflicts of Interest**

No potential conflicts of interest were disclosed.

**Grant Support**

Grants R01 CA101840, P50 CA097190 and an American Cancer Society Clinical Research Professorship (to J.R. Grandis), and F30 ES015669 (to R.J. Leeman-Neill).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received 02/09/2010; revised 03/08/2010; accepted 03/10/2010; published OnlineFirst 04/13/2010.
References

Honokiol Inhibits Epidermal Growth Factor Receptor Signaling and Enhances the Antitumor Effects of Epidermal Growth Factor Receptor Inhibitors

Rebecca J. Leeman-Neill, Quan Cai, Sonali C. Joyce, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-10-0333

Cited articles
This article cites 50 articles, 20 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/16/9/2571.full.html#ref-list-1

Citing articles
This article has been cited by 3 HighWire-hosted articles. Access the articles at:
/content/16/9/2571.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.