Human Cancer Biology

Fyn Is Downstream of the HGF/MET Signaling Axis and Affects Cellular Shape and Tropism in PC3 Cells

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Abstract

Purpose: Fyn is a member of the Src family of kinases that we have previously shown to be overexpressed in prostate cancer. This study defines the biological impact of Fyn inhibition in cancer using a PC3 prostate cancer model.

Experimental Design: Fyn expression was suppressed in PC3 cells using an shRNA against Fyn (PC3/FYN-). Knockdown cells were characterized using standard growth curves and time-lapse video microscopy of wound assays and Dunn Chamber assays. Tissue microarray analysis was used to verify the physiologic relevance of the HGF/MET axis in human samples. Flank injections of nude mice were performed to assess in vivo growth characteristics.

Results: HGF was found to be sufficient to drive Fyn-mediated events. Compared to control transductants (PC3/Ctrl), PC3/FYN- showed a 21% decrease in growth at 4 days (P = 0.05). PC3/FYN- cells were 34% longer than control cells (P = 0.018) with 50% increase in overall surface area (P < 0.001). Furthermore, when placed in a gradient of HGF, PC3/FYN- cells showed impaired directed chemotaxis down an HGF gradient in comparison to PC3/Ctrl (P = 0.001) despite a 41% increase in cellular movement speed. In vivo studies showed 66% difference of PC3/FYN- cell growth at 8 weeks using bidimensional measurements (P = 0.002).

Conclusions: Fyn plays an important role in prostate cancer biology by facilitating cellular growth and by regulating directed chemotaxis—a key component of metastasis. This finding bears particular translational importance when studying the effect of Fyn inhibition in human subjects. Clin Cancer Res; 17(10); 3112–22. ©2011 AACR.

Introduction

The Src-family kinases (SFKs) have long been recognized as key players in cancer biology. Currently, the known members of the SFKs include Src, Lck, Fyn, Yes, Fgr, Lyn, Hck, Blk, and Yrk and all these family members share a common structure and pattern of activation. The existence of a unique domain provides high variability and impacts protein–protein interactions that confer specific physiologic and pathophysiologic function for each member.

Fyn is a ubiquitous SFK that has been previously demonstrated by our group to be overexpressed in prostate cancer (1). Fyn is localized to the inner cytoplasmic leaflet of plasma membrane; a process is driven by posttranslational fatty acid acylation of amino acids in the SH4-domain, typically with myristic and palmitic acids, as well as methylation of lysine residues (2, 3). Activation of Fyn results in tyrosine phosphorylation of a variety of target proteins resulting in downstream signaling of a number of pathways. The role of Fyn has been studied in a variety of cellular processes including T-cell and B-cell receptor signaling, oligodendrocyte, keratinocyte and natural killer cell differentiation, platelet activation, integrin and growth factor-mediated signaling, and cell–cell adhesion and cell migration (4–8). These interactions are mediated by signaling partners such as FAK and paxillin that are overexpressed in prostate cancer concurrent with Fyn (1).
Given our findings, we have made efforts to delineate the causal relationship between Fyn overexpression and prostate cancer progression. Our previous studies demonstrated a concurrent upregulation of FAK and paxillin in prostate cancer tissues suggesting that Fyn may act as a regulator of shape and motility (1). We thus hypothesized that disruption of Fyn activity would result in impaired cellular motility and alternation of cell shape in prostate cancer. Further, upstream of Fyn, there are a number of growth factors and receptors whose downstream signaling may be mediated by the activity of SFKs. Serum concentrations of HGF have been shown to be elevated in men with prostate cancer (9). Furthermore, MET expression has been described in several studies suggesting that this axis may be active in stimulating biochemical events related to disease progression (10); however, the role of the HGF/MET axis has not been well characterized in prostate cancer. In this study, we show that Fyn strongly impacts cellular tropism and shape and that this behavior can be driven by activation of the HGF/MET signaling axis in prostate cancer cell line.

Methods

Cells and Fyn knockdown

PC3 cells were a generous gift of Dr. Carrie Rinker-Schaeffer. Cells were propagated and maintained in RPMI 1640 media (Gibco BRL) supplemented with 1% streptomycin/penicillin (Cellgro) and 10% fetal calf serum (Cellgro) at 37°C in humified air at 5% CO₂, except where noted.

Suppression of Fyn expression was achieved using MISSION shRNA Lentiviral transduction particles (Sigma-Aldrich). Transduction conditions were optimized with a GFP containing construct from Sigma using the same lentiviral transduction system. In the presence of hexadimethrine bromide at 8 mcg/mL, PC3 cells were transduced with shRNA against Fyn or a nontargeting (control) shRNA named PC3/FYN- and PC3/Ctrl, respectively. Knockdown cell lines were propagated in media containing 0.25 mcg/mL puromycin (Sigma Chemical Co.) as the construct contained a puromycin resistance vector. Immunoblots for Fyn were performed in conjunction with all studies to ensure continued Fyn suppression.

Antibodies

Anti-Fyn antibody for use in immunoblotting, immunohistochemistry (IHC), and immunofluorescence (IF) was purchased from Upstate Biotechnology, Inc. Rhodamine-labeled phalloidin and fluorescein isothiocyanate-conjugated anti-mouse and rhodamine-conjugated anti-rabbit antibodies for use as secondary antibodies for IF were obtained from Molecular Probes. Total MET antibody was obtained from Zymed Laboratories. Two phospho-MET antibodies were utilized for IHC (pY1003 and pY202/3/4, Biosource). HGF antibody was obtained from R&D systems.

Preparation of cell lysates and immunoblotting

Cell lysates were prepared using lysis buffer containing 20 mmol/L Tris, pH 8.0, 150 mmol/L NaCl, 10% glycerol, 1% Nonidet P-40, and 0.42% NaF containing inhibitors (1 mmol/L sodium orthovanadate, 1 mmol/L HALT phosphatase inhibitor cocktail (Thermo Scientific). Cell lysates were separated using a 7.5% Tris-HCl gel with SDS-PAGE under reducing conditions. Protein was transferred to polyvinyl chloride membranes and processed for immunoblotting using established methods with enhanced chemiluminescence techniques (GE Healthcare).

Quantitative PCR for FYN

RNA from cell lines was extracted using an RNAqueous kit (Ambion) according to the manufacturer’s recommendations. Samples were stored at −80°C until processed. Customized primers for Fyn were prepared by Integrated DNA Technologies (Coralville). The left primer was: 5'-ATG GAA ACA CAA AAG TAG CCA TAA A-3'; and the right primer: 5'-TCT GTG AGT AAG ATT CCA AAA GAC C-3'. Data were calibrated to the expression of glyceraldehyde phosphate dehydrogenase. Quantitative PCR was performed using SYBR Green dye on an ABI 7700 (Applied Biosystems).

Time-lapse video microscopy and image analysis

All time-lapse experiments were performed using an inverted Olympus IX71 microscope with an attached QImaging Retiga EXi camera. Cells were maintained on a heated stage at 37°C (Omega CN9000A) with a constant flow of 5% CO₂. Image capture was achieved using IPLab version 3.65a (Scanalytics, Inc.). Analysis of still images was performed using the ImageJ software package from the NIH (http://rsb.info.nih.gov/ij/).

Wound-healing assay

Cells were plated onto either 60-mm plates or 6-well plates at a concentration of 1 × 10⁶ cells/cm² and allowed to attach overnight. Cells were allowed to grow to approximately 80% confluence by visual inspection prior to scratch assay. At the time of the scratch, cells were washed 3 times with PBS and starved in serum-free RPMI 1640 for 3 hours. A linear wound was then made with a 10 μl plastic pipette tip. After washing 3 times with serum free media, the cells were stimulated with media containing fetal calf serum or HGF. Wound width was measured at 3 randomly chosen sites using ImageJ. Growth factors used included hepatocyte growth factor (HGF), epidermal growth factor (EGF), and basic fibroblast growth factor (bFGF; Cell Signaling). Wound closure was quantified by parallel assessments of wound length at 4 fixed positions over time and expressed as a percentage of baseline wound distance at that point.

Single-cell shape and motility assay

Cells were plated onto 35-mm plates at a concentration of 1.5 × 10⁵ cells/cm² and allowed to attach for 48 hours to approximately 20% confluence. The cells were then washed 3 times with PBS and starved for at least 3 hours. Cells were
then stimulated with 10 to 50 ng/mL HGF and recorded as described earlier using time-lapse video microscopy (TLVM). Cell movements were tracked using the Meta-morph 7.6 software package (Molecular Devices). Using this software package, 15 to 25 cells per sample were identified and tracked over a 12-hour period. The tracking data were fed in to the IBIDI cell tracker tool in ImageJ yielding analysis of velocity and path length. Data provided represents an average of the cells tracked. Shape characterizations (area, circularity, and length) were performed by manual measurements using ImageJ no less than 20 cells. Cellular area refers to a 2-dimensional projection of the cell onto an XY plane. For membrane ruffling, cellular perimeter was manually measured to determine the fraction of total membrane perimeter involved in ruffling in ImageJ.

**Dunn chamber assay**

Cell chemotaxis was studied using a Dunn chamber assay as previously described (11). In brief, a Dunn Chamber is a modified Zigmond chamber in which a diffusion gradient of a chemotactic factor was made by creating a liquid bridge across 2 wells: one containing media with a high concentration of a chemotactic factor and the other well containing media alone. This creates a diffusion gradient across the area where the 2 are connected. Glass coverslips were placed at the bottom of a 35-mm plate and to this was added 1.5 x 10^5 cells in RPMI supplemented with 10% FCS. Cells were allowed to attach over 24 hours then placed under serum-starved conditions with RPMI for 3 hours. The coverslip was then inverted onto a Dunn chamber (Hawksley) filled with media (no serum). The coverslip was then sealed on the outer edges with hot VALAP mixture (1:1 vaseline, beeswax, and paraffin). The outer chamber of the Dunn apparatus was subsequently evacuated and refilled with media supplemented with HGF at a concentration of 10 ng/mL. Cell chemotaxis was then captured by video microscopy over 3 hours. Analysis of motility was completed as described earlier.

**Immunofluorescence**

PC3 cells were plated onto a glass coverslip in a 6-well plate at a concentration of 1.5 x 10^5 cells/well and allowed to attach over 48 hours in media supplemented with 10% fetal calf serum. Cells were then fixed with 4% paraformaldehyde, and permeabilized with 0.1% Triton X100-PBS before blocking with 3% bovine serum albumin in TBST. The coverslips were then incubated with primary antibody in TBST at 100:1 dilution for 1 hour. Cells were subsequently washed 3 times in TBST before incubating with secondary antibody and/or Rhodamine-phalloidin in TBST at 50:1 dilution for 1 hour. Cells were washed once again in TBST before mounting onto coverslips using ProLong Gold antifade mounting medium with DAPI (Invitrogen Molecular Probes). Images were analyzed using ImageJ after deconvolution using a Huygen’s algorithm. Colocalization was detected using the JACOP plugin for ImageJ.

**Human tissue source**

All human tissue samples used in this study were obtained from the University of Chicago. Utilization of tissue was performed under an institutional review board approved protocol requiring that all samples were kept anonymous to the primary investigational team. Tissue was analyzed in the form of 2 tissue microarrays previously fabricated by the Department of Pathology at the University of Chicago. Microarray fabrication has been described elsewhere (12). In short, the arrays used contained specimens from 45 patients planned to have triplicate representation on the array. Each array element was 1.5 mm in diameter. Tissue samples included primary tumor from prostate cancer patients with Gleason scores of 6 to 9. When possible both normal and tumor elements were scored on a section. The identity of patients was kept blinded to the primary analytic group. A patient’s sample was only considered usable if represented at least twice on the array.

**Immunohistochemistry**

For IHC, stained TMA sections were analyzed by a dedicated urological pathologist (H.A.A. or K.D.H.). Results were reported semiquantitatively on a scale of 0 to 3 for intensity, where 0 was negative, 1 was weak, 2 was moderate, and 3 was strong. The percentage of tumor staining was reported as 0% to 100% in increments of 10%. A composite score was formed using the product of the intensity and percentage of glands staining. Staining was performed at the following antibody concentrations: MET 1:100, MET-Y1003 at 1:20, MET-Y1202/3/4 at 1:25, and HGF at 25 µg/mL. Each TMA contained on-slice controls of lymph node tissue to ensure absence of artifacts contributing to differential staining reported.

**Mice**

Eight-week-old nude (nu/nu) mice (Strain code: 088, Charles River Laboratories) were kept in a specific pathogen free colony, in microisolator cages, and were fed sterile rodent chow and sterile water ad libitum. All protocols were approved by the University of Michigan Animal Care and Use Committee. Tumors were harvested at necropsy and preserved in formalin. Staining was performed using standard hematoxylin and eosin (H&E) as well as CD31 (Abcam, 1:50 dilution) to assess for microvessels. H&E sections were analyzed for mitotic index [per 10 high powered field (HPF) counting up to 30 HPFs], coagulative tumor necrosis (% tumor volume), lymphoid aggregates within the tumor (per 10 HPF), ratio of epithelioid: spindled cells, and neovessel density (CD31+ with luminal formation by pathologists review).

**In vivo growth assay**

PC3/FYN- and PC3/Ctrl cells were harvested by trypsinization washed twice with PBS and resuspended at a density of 2 x 10^6 cells in 100 µL PBS for each injection site. Mice were monitored for tumor growth and when detected by palpation, measurement of the tumors began. Tumor
volumes were calculated by the formula: volume = \((\text{minimum measurement})^2 \times (\text{maximum measurement})\)/2 as described by Smith (13). Tumors were measured weekly until volume exceeded 1 mm^3. Each mouse was given 2 subcutaneous doses of PC3/FYN- (right flank) and PC3/Ctrl (left flank). Alternatively, a cross product of the longest tumor diameter and one orthogonal to the longest diameter were calculated and compared. At the conclusion of the study, all mice were sacrificed, and tissue samples were collected.

**Statistical analyses**

All analyses were performed using SPSS version 17.0 for Windows. A general linear model (GLM) was used to compare the effects of FYN expression and HGF stimulation and their interaction on cell morphology parameters from baseline to 12 hours and to compare changes in cell length between baseline and 12 hours. To evaluate the differences of cellular shape and growth between groups (PC3/FYN- and PC3/Ctrl, serum starved and HGF stimulated), the independent-samples T test was used for the data based on specific distributional assumptions such as the normal distribution. If the data was of a non-normal distribution, then Mann–Whitney test to assess was appropriately used. The Watson–Williams test was performed for the equality of mean angle of cellular motility using calculating angular movements from a relative origin (14). Comparisons of quantified IHC data were performed using a Wilcoxon signed ranks test. For in vivo growth studies, tumor volume and cross product data were logarithmically transformed so that a paired-samples T test could be used to assess differences of PC3/FYN- cell growth compared to PC3/Ctrl. A 2-sided P < 0.05 was used as a threshold for declaring statistical significance.

**Results**

**Generation of PC3 Fyn knockdown cell line**

To characterize the effect of Fyn variation in vitro, we generated a knockdown line using PC3 cells and lentiviral transduction particles containing shRNA constructs targeted specifically against Fyn (PC3/FYN-). Using a multiplicity of infection of 2, 5 constructs (Supplementary Table S1) were tested. The construct leading to maximal Fyn suppression by immunoblot with minimal effect on non-Fyn SFKs, such as Src, was labeled PC3/FYN- and advanced for further study. A control cell line was developed (PC3/Ctrl) using a nontargeting shRNA construct. Both lines were maintained under continuous selection of puromycin. Fyn mRNA expression was measured with a comparative RT-PCR using PC3/Ctrl as a reference and protein by immunoblot (Fig. 1A and B). Both assays revealed that Fyn expression was decreased by at least 60%. Minimal off target effects were seen as evidenced by the lack of change in Src expression (Fig. 1B). The knockdown effect was stable through serial passage under these conditions.
Fyn knockdown results in impaired \textit{in vitro} growth and motility

Under standard propagation conditions, PC3/FYN- cells showed a small but statistically significant diminishment in growth rate (Fig. 1C). After 4 days under standard conditions, PC3/FYN- cells grew at only 79% the rate of PC3/Ctrl cells ($P = 0.05$).

Wound healing assays in concert with TLVM were performed to characterize motility (Fig. 2A and B and Supplementary movies 1 and 2). Virtually no wound closure was seen in the absence of serum or other mitogens. However, in serum-replete conditions, PC3/Ctrl cells showed near complete closure within 12 hours whereas PC3/FYN- cells failed to show complete closure.

HGF stimulates PC3 motility

Isolated growth factors were utilized to further understand growth and motility. HGF, EGF, and bFGF were selected given overexpression or increased serum concentrations in men with advanced prostate cancer (9, 15, 16). Wound closure was observed using TLVM under conditions of media with a single growth factor at 10 ng/mL. HGF was determined to be the optimal motility stimulus for PC3 cells from this pool as it produced the highest rate of wound closure per unit time (Supplementary Fig. S1).

Fyn knockdown results in altered cell shape and increased cell speed with impaired directional motility

A series of single cell motility experiments with serum or HGF stimulation were performed to measure the impact of Fyn knockdown on morphology and speed of movement.

Cell area. Area was measured as a measure of size and cell spreading. Differential effects of HGF stimulation on cell shape are summarized in Figure 3. At baseline, a small but statistically significant difference was seen between PC3/FYN- and PC3/Ctrl cells. Under serum-starved conditions, the PC3/FYN- cells were 33% larger than the
Figure 3. A, representative images of PC3/FYN- and PC3/Ctrl cells on polystyrene plates stimulated by HGF. PC3/FYN- cells were found to have greater cell length, area, and ruffling than PC3/Ctrl cells. B, immunofluorescence of actin and MET in PC3 sublines. Overlap of actin and MET is shown in yellow and demarcated by arrows in the rightmost panels. These areas are most consistent with focal adhesion plaques. As seen in the lower right panel, there is no focal accumulation of both MET and actin to suggest plaque formation. B, rose plots showing direction of movement of PC3 sublines relative to an HGF gradient (source on right). A rose plot is a circular histogram in 10° increments showing cumulative motion in any given direction in 360° without regard to the magnitude of movement. C, graphical representation of quantified morphologic variations between PC3/FYN- and PC3/Ctrl cells.
PC3/Ctrl cells \((p = 0.018)\), and after 12 hours of stimulation with serum there was a 50% increase in surface area \((p < 0.001)\) for the PC3/FYN- cells compared to PC3/Ctrl. Comparing Feret’s (longest) diameter, the PC3/FYN- cells were 34% longer at baseline \((p < 0.001)\) and 56% longer after 12 hours of serum stimulation \((p < 0.001)\). Membrane ruffling was not significantly different between the 2 cell lines before or after serum stimulation. The effect of HGF was similar on both cell lines; in comparing PC3/FYN- to PC3/Ctrl cells after 12 hours of stimulation we found a 43% increase in area \((p = 0.016)\), and 17% increase in Feret’s diameter \((p = 0.026)\) with no change in ruffling.

Cell length. A similar trend was also observed for cell length as determined by longest cell axis under the same experimental conditions (Fig. 3A and C). After 12 hours of stimulation with HGF, PC3/FYN- cells were 31% longer than PC3/Ctrl cells \((p = 0.003)\). In fact, PC3/Ctrl cells showed 17% increase in cell length \((p = 0.010)\) after 12 hours in HGF, whereas the PC3/FYN- showed no statistically significant change.

Cell morphology. In response to HGF stimulation, PC3/FYN- acquired a broader distribution of various cell shapes and morphologies (Fig. 3A and C). PC3/FYN- cells also produced a larger number of filopodia compared to PC3/Ctrl cells. Using representative images of subconfluent cellular monolayers, we manually counted the percentage cells with filopodia. In the absence of HGF supplementation, PC3/Ctrl cells underwent a decrease of the percentage of cells with filopodia from 11% to 4.5%, a 59% decrease over 12 hours. Under the same conditions, the percentage of PC3/FYN- cells with filopodia only decreased 20%, from 40.5% to 32.5%. Under HGF stimulation, PC3/Ctrl cells had a 9% increase in cells with filopodia whereas PC3/FYN- cells had a 12% decrease over 12 hours. There were variations in the percentage of cell perimeter ruffling. PC3/Ctrl cells had a greater degree of ruffle formation in response to HGF supplementation (a 16% increase in percentage of cell circumference with ruffles upon HGF stimulation) whereas PC3/FYN- cells were not observed to have any change in ruffling in the presence of HGF. Cell circularity [as defined by \(4\pi(area/\text{perimeter}^2)\)] was calculated as an assessment of cell shape symmetry. There was an increase in circularity under serum-starved conditions over 12 hours for PC3/Ctrl cells of 12%, whereas there was no significant increase observed for PC3/FYN- cells. Under HGF supplementation, no changes in circularity were observed for either cell line.

To test the hypothesis that the variations in shape were related to alterations in the actin cytoskeleton, phalloidin staining with MET immunostaining were performed (Fig. 3B). In response to HGF stimulation, PC3/Ctrl cells showed retraction of small hair-like projections in favor of forming larger cellular extensions. PC3/FYN- cells continued to show these small hair-like projections despite HGF stimulation.

To quantify the effect of Fyn knockdown on focal adhesion formation and MET distribution, colocalization of MET and actin was analyzed by detecting signals above a threshold level to exclude background beyond the cellular membrane—given this colocalization pattern suggests focal adhesion formation. At threshold, signal overlap between actin and MET were expressed as Mander’s coefficients expressing degree over overlap. In the setting of HGF stimulation, 28% of the actin signal was associated with MET predominately at focal points along the cell surface and predominately at the tips of filopodia, which would represent focal adhesion plaque formation. Conversely, only 2% of actin was associated with HGF stimulation in PC3/FYN- cells.

Cell speed. Representative films of cellular movement of PC3/FYN- and PC3/Ctrl cells on glass bottom plates are shown in supplemental videos 3 and 4. Although PC3/FYN- cells exhibited decreased ability to migrate over monolayer defects, analysis of single cells showed a relative increase in cellular speed when no gradient was present. Both PC3/Ctrl and PC3/FYN- cells showed decreased cell speed in the absence of HGF stimulation, 24% and 31% from baseline respectively. However, PC3/FYN- cells exhibited increased movement speed compared to PC3/Ctrl with a 41% increase in average cell speed over a 12-hour period of HGF stimulation.

Directed chemotaxis. Given the dichotomy between elevated cell speeds but decreased rates of wound closure, we hypothesized that Fyn regulates vectorial velocity or more specifically, directed chemotaxis. To test this hypothesis we employed a Dunn chamber to allow for the direct observation of cell migration under a chemotactic gradient. Unlike a Boyden Chamber assay, this allows for direct visualization of cells in transit. Individual cells were observed under TLVM for a period of 3 hours. More than 20 cells were tracked to create a composite map of movement represented as a rose plot in \(10^\circ\) increments (Fig. 2). PC3/Ctrl cells moved toward the HGF source whereas PC3/FYN- cells were either insensitive to or moved away from the HGF source. We performed a comparison of direction relative to the vector defined by the initial and final location (Euclidean distance, ED) of the each cell (Table 1 and Supplementary Fig. S2). In brief, this directionality coefficient is a ratio of the shortest linear distance between the

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<th>Table 1. Summary of motility parameters from Dunn Chamber assay</th>
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Abbreviations: AD, accumulated (total) distance; ED, Euclidean distance; FMI, forward motion index. Supplementary Figure S2 shows a graphical representation of the above parameters.
In vivo growth assay

To measure the impact specifically on growth variation in a biologically relevant system, subcutaneous flank injections were performed on 5 nude mice with each mouse receiving 2 injections of PC3/FYN- and 2 injections of PC3/Ctrl. After approximately 8 weeks of growth, mice were sacrificed and tumors measured. Four of the mice grew visible tumors. Figure 4A shows a representative mice bearing PC3/Ctrl (left or top) and PC3/FYN- (right or bottom) tumors as well as the tumors removed at necropsy. In comparing PC3/FYN- to PC3/Ctrl in each mouse (Fig. 4B), an average 33% difference was seen in the size of the PC3/Ctrl and PC3/FYN- tumors (mean volume day 43: 712 vs. 120 mm³; P = 0.014; day 57: 1299 vs. 449 mm³; P = 0.071). Alternatively, comparing the cross product of the longest diameter and an orthogonal diameter, there was a 66% difference (average cross product day 57: 570 mm² vs. 193 mm²; P = 0.002). Histomorphological and immunohistochemical analysis of the tumor samples did not show significant changes in cellular morphology or tumor neovessel formation as evidenced by CD31 staining (Fig. 4C). There was a trend however, to an increase in mitotic index, tumor necrosis, and tumor infiltration by lymphocytes (as evidenced by intratumoral lymphoid aggregates) favoring the PC3/Ctrl tumors without meeting significance criteria. Neovessel density was also not significantly different but favored the PC3/FYN- tumors (Fig. 4D).

HGF and MET expression in prostate cancer tissue samples

The relevance of the HGF/MET signaling axis in human disease was validated by immunohistochemical analysis of a tissue microarray composed of 40 patient samples. Representative sections are shown in Supplementary Figure S3. HGF was up-regulated 1.3-fold in cancer compared to normal (mean composite score 140.74 vs. 179.72, P = 0.035). No significant variations in MET, pMET-Y1003, or pMET-Y1349 were found in the sample population between malignant and nonmalignant glands (data not shown).

Discussion

These studies show that focused reduction of Pyn expression apart from other SFKs results in a notable variation in the in vitro phenotype. This phenotype is characterized by a change in growth kinetics that is durable over time with a significant impact on cellular shape and motility. The in vitro growth studies showed a small but statistically significant separation in growth that became more pronounced during in vivo studies due to the significant increase in experimental duration. Further studies of multi-SFK inhibition would potentially be warranted for the future. Analysis of the tumor specimens from the mouse experiment did not show significant differences between the 2 conditions outside of tumor necrosis as this likely reflected the difference in size of the tumors. The lack of differences in mitotic index and neovessel formation could have been affected by tumor size and necrosis as well as loss of antigen retrieval during prolonged formalin fixation.

Our Fyn knockdown cells in general showed an accelerated speed but more importantly, aberration of directional response in PC3 cells. This change in function is accompanied by a change in macroscopic and microscopic cellular structure as shown by the variation in the arrangement of cytoskeletal elements and focal adhesions. In particular, the changes in colocalization of MET with actin points to variation in focal adhesion formation—key step in the metastatic process. This presentation is particularly focused on the PC3 model given its extensive use in both in vitro and in vivo studies. Interestingly, we have found the HGF/MET axis to be a sufficient activator of Fyn-driven events. Elevation of serum HGF is well recognized in prostate cancer. Our finding of increased HGF expression in malignant prostatic glands is certainly consistent with this previous observation (9). Despite this, the expression of the MET in both total and active forms was stable to diminished. The limited sample size restricted our ability to detect more subtle differences in MET expression and activation, but if a difference was present, it would likely only represent a small variation. The role of MET in this system and in prostate cancer in general continues to be an important and exciting area of investigation that exceeds the scope of this particular study.

The SFKs are among the first oncogenes recognized in cancer biology. SFKs have remained of great interest given the central role they play in mediating extracellular stimuli to the nucleus. Although the majority of research in Src biology has focused on the prototypical member of this family, c-Src, emerging data suggests that the various SFKs may affect cancer cells differently. Fyn is known to be ubiquitously expressed under normal physiologic conditions and its functions are typically ascribed to mediating T-cell response and neuronal development (17). Fyn knockout mice have been characterized as having a subtle phenotype in neuropsychological development and T-cell function with little phenotypic characteristics otherwise (18). This has been attributed to the high degree of...
homology of the family members that may allow for compensation for the loss or absence of another family member. There is now growing evidence, however, that non-Src SFKs play an important role in tumor progression. In a study by Park and colleagues, inhibition of Lyn expression with RNAi resulted in altered cellular growth apart from migration whereas Src knockdown resulted in impaired metastatic capability (19). Similarly, our group has previously demonstrated that Fyn is up-regulated in prostate cancer (1). Therefore, we have made efforts to define the role that this up-regulated kinase may play in cancer biology.

![Figure 4. A (left), nude mouse with subcutaneous injection of PC3/Ctrl and PC3/FYN- cells at day 57; (right) tumor recovered at necropsy. B, table of mouse tumor measurements showing both cross products (length × width) and volumetric approximations based on maximal and minimal tumor dimensions. C, immunoistochemical analysis of tumors from in vivo growth study. PC3/Ctrl (upper) and PC3/FYN- cells (bottom) were injected into the flanks of nu/nu mice and collected at necropsy. H&E and CD31 staining is shown at 1× (left), 4.2× (middle), and 20× (right). No significant change in cellular morphology or neovessel formation was seen between the 2 conditions. D, table of quantification of mitoses, tumor necrosis, lymphoid aggregate formation, ratio of epithelioid to spindled cells, and neovessel density. No significant differences were seen.](image-url)
Fyn is a member of the SFKs up-regulated and germane to prostate cancer progression. It functions to promote not only growth but more importantly, directed chemotaxis, a finding that makes Fyn a putative target for metastasis-directed therapy in prostate cancer and other malignancies where it is overexpressed.

Disclosure of Potentially Conflicting Interests

No potential conflicts of interest were disclosed.

Acknowledgments

The authors would like to thank Ms. Can Gong for her assistance in the preparation of the tissue microarrays for pathology review. Dr. Christine Labno and the University of Chicago microscopy core facility for assistance with cell tracking and immunohistochemistry analysis. We also would like to thank Dr. Karen Kaul for helping to coordinate tissue samples from the Chicago SPORE tissue bank and Ms. Leslie Martin for assistance with TMA sections from the Department of Pathology at the University of Chicago. The authors would also like to express their thanks for the generous support provided by the University of Chicago Auxiliary Board, The Riviera Club, and the Arlington Million Ladies. Without their generosity, this project could not have started.

Grant Support

(EP) DOD Physician Research Training Award W81XWH-08-1-0470; Prostate SPORE Career Development Award; University of Chicago Auxiliary Board; Rivera Club; Arlington Million Ladies; [EJ] P01093900; [YN] R01 HL 073936; [RS] 5R01CA125541-02; 3R01CA125541-03S1; 5R01CA129501-02S1.

Received May 13, 2010; revised January 25, 2011; accepted February 15, 2011; published OnlineFirst March 1, 2011.

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Clin Cancer Res 2011;17:3112-3122. Published OnlineFirst March 1, 2011.