Cancer Therapy: Preclinical

Antitumor Efficacy of PKI-587, a Highly Potent Dual PI3K/mTOR Kinase Inhibitor

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Abstract

**Purpose:** The aim of this study was to show preclinical efficacy and clinical development potential of PKI-587, a dual phosphoinositide 3-kinase (PI3K)/mTOR inhibitor.

**Experimental Design:** In *vivo* class 1 PI3K enzyme and human tumor cell growth inhibition assays and in *vivo* five tumor xenograft models were used to show efficacy.

**Results:** In *vivo*, PKI-587 potently inhibited class I PI3Ks (IC50 vs. PI3K-α = 0.4 nmol/L), PI3K-α mutants, and mTOR. PKI-587 inhibited growth of 50 diverse human tumor cell lines at IC50 values of less than 100 nmol/L. PKI-587 suppressed phosphorylation of PI3K/mTOR effectors (e.g., Akt), and induced apoptosis in human tumor cell lines with elevated PI3K/mTOR signaling. MDA-MB-361 [breast; HER2+, PIK3CA mutant (E545K)] was particularly sensitive to this effect, with cleaved PARP, an apoptosis marker, induced by 30 nmol/L PKI-587 at 4 hours.

In *vivo*, PKI-587 inhibited tumor growth in breast (MDA-MB-361, BT474), colon (HCT116), lung (H1975), and glioma (U87MG) xenograft models. In MDA-MB-361 tumors, PKI-587 (25 mg/kg, single dose i.v.) suppressed Akt phosphorylation [at threonine(T)308 and serine(S)473] for up to 36 hours, with cleaved PARP (cPARP) evident up to 18 hours. PKI-587 at 25 mg/kg (once weekly) shrank large (>1,000 mm3) MDA-MB-361 tumors and suppressed tumor regrowth. Tumor regression correlated with suppression of phosphorylated Akt in the MDA-MB-361 model. PKI-587 also caused regression in other tumor models, and efficacy was enhanced when given in combination with PD0325901 (MEK 1/2 inhibitor), irinotecan (topoisomerase I inhibitor), or HKI-272 (neratinib, HER2 inhibitor).

**Conclusion:** Significant antitumor efficacy and a favorable pharmacokinetic/safety profile justified phase 1 clinical evaluation of PKI-587. *Clin Cancer Res;* 17(10); 3193–203. ©2011 AACR.

Introduction

Class 1 phosphoinositide 3-kinases (PI3K) play a key role in the biology of human cancer. The gene encoding the PI3K-α isoform (PIK3CA) is amplified or mutated in a wide range of cancers (1, 2). Aberrantly elevated PI3K/Akt/mTOR pathway signaling has been implicated in poor prognosis and survival in patients with lymphatic, breast, prostate, lung, glioblastoma, melanoma, colon, and ovarian cancers (1–6). In addition, PI3K/Akt/mTOR pathway activation contributes to resistance of cancer cells to both targeted anticancer therapies and conventional cytotoxic agents (5, 6). An effective inhibitor of the PI3K/Akt/mTOR pathway could prevent cancer cell proliferation and induce programmed cell death (apoptosis; refs. 1, 2, 5). Many pharmaceutical companies now have substantial PI3K/mTOR signaling pathway inhibitor programs. Examples of advanced dual PI3K/mTOR inhibitors include: BEZ-235 (Novartis), BGT-226 (Novartis), XL765 (Exelixis), SF1126 (Semafore), and PKI-402 (Wyeth/Pfizer) (7–11). Examples of advanced inhibitors selective for class 1 PI3Ks include: GDC-0941 (Genentech), XL147 (Exelixis), SF1126 (Semafore), and PKI-402 (Wyeth/Pfizer) (7–11). Examples of advanced inhibitors selective for class 1 PI3Ks include: GDC-0941 (Genentech), XL147 (Exelixis), BKM120 (Novartis), GSK1059615 (Glaxo), CAL101 (Calistoga), and PX-866 (Oncotherapy) (7, 8, 12–14). BEZ-235 and BGT-226 are in phase 2, whereas most other compounds are undergoing phase 1 clinical evaluation (7, 8).

The Wyeth PI3K inhibitor discovery project identified PKI-587, an exceptionally potent, selective, ATP-competitive, and reversible PI3K/mTOR inhibitor for clinical development (15). In *vivo*, PKI-587 displayed antitumor activity (i.v. route) in breast (MDA-MB-361, BT474), colon (HCT116), glioma (U87MG), and non–small cell lung cancer [(NSCLC) H1975] xenograft models. PKI-587 caused tumor regression in some models and its favorable...
Translational Relevance

PKI-587, a potent pan-class I phosphoinositide 3-kinase (PI3K)/mTOR inhibitor, showed single-agent efficacy in multiple preclinical tumor models. Tumor regression was observed in several models. This effect was most pronounced against MDA-MB-361 (breast), which has elevated HER2 levels and mutant PI3K-α. Preclinical data suggest utility of PKI-587 in the treatment of cancers with elevated PI3K/mTOR signaling, including those resistant to agents that target HER2 or epidermal growth factor (EGF) receptors (EGFR).

PKI-587 efficacy was enhanced when combined with a MEK1,2 kinase inhibitor (PD0325901), or irinotecan in a colon tumor model (HCT116) with mutant K-Ras. PKI-587 showed single-agent efficacy against a non-small cell lung cancer model (H1975) with mutant EGFR (L858R/T790M), and this activity was also enhanced when combined with the irreversible HER2 kinase inhibitor, HKI-272. These preclinical data suggest strategies for clinical targeting and combination uses of PKI-587.

efficacy, pharmacokinetic, and safety profile advanced it to phase 1 clinical evaluation.

Materials and Methods

Enzyme assays

Enzyme assays were done in fluorescent polarization format as previously described (11, 16, 17). PKI-587 selectivity was evaluated in the Invitrogen 236 human kinase panel, at [ATP] = K_m for each enzyme.

Cell culture, growth inhibition, and translocation assays

All cell lines, except U2OS, were from American Type Culture Collection. Mutational status (Table 1) of various oncogenes in cell lines was from the Wellcome Trust Sanger Institute site (18). PKI-587 was tested in additional human tumor cell lines by Caliper Life Sciences (Supplementary Table S1).

U2OS cells, engineered to monitor FOXO1-GFP cellular translocation, were from Thermo Scientific. Cell growth inhibition and FOXO1-GFP translocation assays were done as previously described (17).

Cell lysis and Western blotting

Cells were exposed to PKI-587 for 4 hours (unless indicated otherwise in the following text). Cell lysis and lysate handling were done as previously described (11, 17). Antibodies were from Cell Signaling Technology. Inhibition of protein phosphorylation was quantified from Western blots, using the BioRad Fluor-S Multimager with Quantity One Analysis software.

Caspase activation assay

Cellular caspase 3/7 activity was measured by the Caspase-Glo 3/7 Luminescent Assay (Promega). Cells were exposed to PKI-587 for 4 to 24 hours, and assay format and data collection were as previously described (17).

Establishment of xenograft tumors, efficacy studies, and biomarker analysis

In vivo methodology was carried out as previously described (11, 17). PKI-587 or vehicle (5% dextrose, water, pH 3.5) was administered by i.v. route in various regimens: daily or intermittent (days 1, 5, 9, etc.). In vivo studies were conducted under an approved Institutional Animal Care and Use Committee protocol. Significant (statistically, Student’s t test) reduction in the tumor growth of treated groups compared with controls (vehicle) was defined as a value of P < 0.05.

Pharmacokinetic and pharmacodynamic (biomarker) measurements were done on tumor-bearing female nude mice administered PKI-587. Tissue samples were processed and probed with the various antibodies as described previously (11, 17).

Results

Enzyme assays

PKI-587 (Fig. 1), an ATP-competitive triazine scaffold compound (Supplementary Table S2), potently inhibited PI3K-α (IC_50 = 0.4 nmol/L). Mutant forms of PI3K-α with elevated lipid kinase activity (19) were inhibited by PKI-587 at concentrations equivalent to the IC_50 for wild-type PI3K-α. PI3K-β, δ, and γ isoforms were inhibited by PKI-587 at concentrations approximately 10-fold higher than that observed for PI3K-α. mTOR kinase inhibition (IC_50 = 1 nmol/L) indicated that PKI-587 was an equipotent PI3K-α/mTOR inhibitor.

PKI-587 showed a highly selective profile when tested against 236 human protein kinases (IC_50 > 10 μmol/L, Supplementary Table S3; Invitrogen). Only wild-type and mutant (V600E) B-Raf s were inhibited by PKI-587 at IC_50 values of 10 μmol/L.

Cell growth inhibition assay

PKI-587 was a potent cell growth inhibitor with IC_50 values of 50 nmol/L or less in 19 of 23 human tumor cell lines (Table 1). PKI-587 was also a potent cell growth inhibitor (IC_50 < 100 nmol/L) in 37 of 43 tumor cell lines (from the NCI-60) assayed by Caliper Life Sciences (Supplementary Table S1). PKI-587 suppressed phosphorylation of downstream effectors of PI3K signaling (e.g., Akt at T308) at concentrations that closely matched growth inhibition IC_50 values in MDA-MB-361, BT474, HCT116, H1975, U87MG, and A498. Phosphoblot data for BT474 and U87MG, as well as A498 and 786-0 with/without 10 μmol/L verapamil (see later), are shown in Supplementary Figure S1A and B.

Higher IC_50 values against A498, 786-0, H1299, and DLD1 (IC_50 range = 267–433 nmol/L, Table 1) were
investigated. Drug efflux capacity sensitive to the L-type calcium channel and P-glycoprotein (multidrug resistance) inhibitor verapamil (20) was reported for these cell lines (21, 22). PKI-587 growth inhibition in these cells was reevaluated in the presence of 10 μmol/L verapamil, which blocks P-glycoprotein function but does not affect cell growth or viability. A498, 786-0, H1299, and DLD1 exposed to PKI-587 with 10 μmol/L verapamil had 4- to 9-fold lower growth inhibition IC₅₀ values (Supplementary Table S4). Therefore, A498, 786-0, H1299, and DLD1 "resistance" to PKI-587 was related to verapamil-sensitive compound efflux and not a PI3K/Akt/mTOR signaling pathway-independent mechanism.

**PKI-587 in vitro profile in biomarker, caspase activation, and FOXO1-GFP translocation assays**

**PKI-587 effect on phosphorylation of PI3K and mTOR effector proteins, and activation of caspase 3/7 in MDA-MB-361 [HER2⁺, PIK3CA (E545K)].** The effect of PKI-587 on a group of PI3K/mTOR effector proteins in MDA-MB-361 after 4 hours of exposure is shown in Fig. 2A. This linked PKI-587 enzyme inhibition to cellular antiproliferative effects. Suppression of cellular PIP3 by PKI-587 was indirectly shown by potent (IC₅₀ < 3 nmol/L) suppression of phosphorylated Akt (p-Akt) at T308 (Fig. 2A). Full activation of Akt kinase occurs when the mTOR TORC2 protein complex phosphorylates Akt at S473. PKI-587 caused potent (IC₅₀ < 10 nmol/L) suppression of p-Akt at S473 (Fig. 2A). Examples of PKI-587 mTOR TORC1 complex inhibition were suppression of 4EBP1 and p70S6 kinase (70S6K) phosphorylation at IC₅₀ values of less than 3 nmol/L (Fig. 2A).

PKI-587 suppression of p-Akt caused consequent effects on Akt effectors such as PRAS40 (proline-rich Akt substrate, 40 kDa), ENOS (endothelial nitric oxide synthase), and GSK3 (glycogen synthase kinase 3). Akt phosphorylation of
PRAS40 at T246 was suppressed at an IC_{50} < 10 nmol/L (Fig. 2A). Akt phosphorylation of ENOS at S1177 and GSK-3a/GSK-3b at S9/S21 was suppressed by PKI-587 at IC_{50} values of less than 3 nmol/L (Fig. 2A).

In MDA-MB-361 cells, the effect of PKI-587 on the induction of cPARP, an indicator of cell apoptosis (23), was evident. Complete PKI-587 suppression of p-Akt in MDA-MB-361 correlated with detectable cPARP at 30 nmol/L PKI-587 (Fig. 2A). Cleaved PARP was detected in MDA-MB-361 within 1 hour after exposure to PKI-587 (Supplementary Fig. S2). PKI-587 did not affect the overall Akt level in MDA-MB-361 cells at concentrations tested (Fig. 2A).

Because caspase 3 is a critical mediator of apoptosis, associated with proteolytic cleavage of many key proteins, including the nuclear enzyme PARP (24, 25), we examined PKI-587 effect on this enzyme in MDA-MB-361. Figure 2B shows that PKI-587 caused a dose-dependent increase in caspase 3/7 activity at 4 and 24 hours. The increased caspase 3/7 activity caused by PKI-587 exceeded that caused by a highly selective mTOR kinase inhibitor, MTI-178 (26). In addition, only 6% of MDA-MB-361 cells exposed to 100 nmol/L PKI-587 for 24 hours remained viable (Fig. 2C).

**FOXO1-GFP translocation in U2OS.**

FOXO1 activity is regulated by Akt-mediated phosphorylation (27). Akt-phosphorylated FOXO1 is sequestered in the cytosol by 14–3-3 protein, and unphosphorylated FOXO1 locates to the cell nucleus. Figure 2D showed that PKI-587 suppression of p-Akt caused FOXO1-GFP translocation to cell nuclei in U2OS cells (Thermo Scientific). The IC_{50} value was 43 nmol/L for PKI-587 effect on FOXO1-GFP (Fig. 2D).

**In vivo biomarker profile and efficacy of PKI-587 in MDA-MB-361 (breast) tumor xenografts**

Initially, the biomarker targets p-Akt and cPARP were used to assess PKI-587 activity in vivo. Figure 3A shows that PKI-587 at 2 mg/kg (single dose) suppressed p-Akt.
PKI-587, a Dual PI3K/mTOR Kinase Inhibitor

Figure 3. PKI-587 efficacy against MDA-MB361 xenografts. A, suppression of p-Akt (T308 and S473) and induction of cPARP in MDA-MB-361 tumor tissue from nude mice after single-dose exposure to 2 (top) or 25 (middle and bottom) mg/kg PKI-587. B, pharmacokinetic data from MDA-MB-361 tumor-bearing nude mice given PKI-587 at 3 and 25 mg/kg (single dose). PKI-587 plasma half-life values at 3 and 25 mg/kg were 4.9 and 14.4 hours, respectively. Supplementary Table S3 shows a pharmacokinetic /safety summary for PKI-587.

Dose response for PKI-587 (0.5–10 mg/kg) in the MDA-MB-361 model for 5 days (d/C25, 3 rounds, 2-day intervals between rounds). PKI-587 induced regression of large (~1,000 mm³) MDA-MB-361 tumors (Fig. 3D) when given at 25 mg/kg once weekly or at 10 mg/kg dx5 (1 round). With an intermittent dosing schedule (days 1, 5, 9), minimal effective dose (≥50% tumor growth inhibition) was 3 mg/kg against MDA-MB-361 tumors (Supplementary Fig. S3A). PKI-587 at 20 mg/kg (day 1, 5, 9 regimen) had greater efficacy against MDA-MB-361 than taxol at 60 mg/kg (i.p., once; Supplementary Fig. S3B). PKI-587 also had significant efficacy in the BT474 [breast; HER2⁺, PIK3CA (K111N)] xenograft model when given at 5 and 10 mg/kg (day 1, 5, 9 regimen; Supplementary Fig. S4A and B). PKI-587 was well tolerated at all dosing levels described here and later. Single maximum tolerated dose (MTD) that affected animal viability was 30 mg/kg.
In vivo efficacy of PKI-587 in HCT116 (colon) tumor xenografts

In vivo regimens to test PKI-587 efficacy in the HCT116 [K-Ras, PIK3CA (H1047R)] model were guided by in vitro data. In vitro, PKI-587 inhibited HCT116 growth (IC$_{50}$ = 8 nmol/L) and suppressed p-Akt (T308) at 30 nmol/L or more after 18 hours of exposure (Fig. 4A), but induction of cPARP was only evident at 3 μmol/L. To enhance PKI-587 in vitro efficacy against HCT116, we combined it with the cancer chemotherapeutics taxol, cisplatin, and camptothecin. Only camptothecin (topoisomerase I inhibitor) enhanced PKI-587 efficacy against HCT116. This effect occurred at a camptothecin concentration (320 nmol/L) that did not affect caspase 3/7 activity or induce cPARP. This combination increased caspase 3/7 activity (Supplementary Fig. S5A) and lowered cPARP induction to 3 μmol/L PKI-587 (Fig. 4A).

Against large (~1,000 mm$^3$) HCT116 tumor xenografts irinotecan (a semisynthetic camptothecin analogue) at 40 mg/kg, or PKI-587 at 12.5 mg/kg (day 1, 5, 9 regimen for both), only attenuated HCT116 tumor growth (Fig. 4B). Combined, irinotecan (40 mg/kg) and PKI-587 (12.5 mg/kg), day 1, 5, 9 regimen, prevented HCT116 tumor size increase in a 13-day study (Fig. 4B).

Because HCT116 cells have both PIK3CA and K-Ras mutations, we tested PD0325901 [MEK1/2 inhibitor; ref. 28] ability to enhance the PKI-587 in vitro effect on cPARP induction. The IC$_{50}$ value of PD0325901 for in vitro growth inhibition of HCT116 was 230 nmol/L, and IC$_{50}$ for phosphorylated mitogen-activated protein kinase [(p-MAPK) T202/Y204] suppression was 50 nmol/L (24 hours). Only minimal cPARP was detected in HCT116 after 24 hours of exposure to 3.0 μmol/L PD0325901. When PKI-587 and PD0325901 (100 nmol/L) were combined, increased...
caspase 3/7 activity was detected (Supplementary Fig. S5B) and cPARP was induced at 30 nmol/L or more PKI-587 (Fig. 4C).

Against small HCT116 tumor xenografts (300 mm³) either PKI-587 (25 mg/kg) or PD0325901 (50 mg/kg) alone (day 1, 5, 9 regimen) were ineffective (Fig. 4D). In combination, statistically significant (P < 0.01) antitumor efficacy was observed (Fig. 4D).

PKI-587 antitumor efficacy against H1975 (NSCLC) and U87MG (glioma) tumor xenografts

Efficacy of PKI-587 in H1975 (NSCLC) tumor xenografts. In a mouse transgenic lung tumor model driven by mutant, activated epidermal growth factor receptor (EGFR; L858R and T790M), the combined antitumor effect of the irreversible HER2(neu)/EGFR inhibitor HKI-272 and the mTOR inhibitor rapamycin was greater than that observed for either compound alone (29). HKI-272 overcomes resistance the L858R/T790M EGFR mutant displays against reversible inhibitors such as Iressa and Tarceva (30). We therefore tested the efficacy of PKI-587 alone, or with HKI-272, in a human NSCLC tumor line equivalent to the mouse lung tumor model. We used H1975 that has L858R and T790M mutant EGFR (30).

In vitro, PKI-587 inhibited H1975 growth (IC₅₀ = 14 nmol/L), suppressed p-Akt (T308; IC₅₀ = 30 nmol/L), induced caspase 3/7 activity at 300 nmol/L or more (Supplementary Fig. S5C), caused cPARP induction at 1 μmol/L (Fig. 5A), and did not affect p-MAPK (Supplementary Fig. S9B). HKI-272 alone also suppressed p-Akt (T308; IC₅₀ = 350 nmol/L), suppressed p-MAPK (IC₅₀ = 250 nmol/L; Supplementary Fig. S9A), and induced caspase 3/7 and cPARP at 3 μmol/L. Exposure (24 hours) of H1975 to combined PKI-587 and HKI-272 (250 nmol/L)
markedly increased suppression of p-Akt (T308; IC₅₀ < 30 nmol/L), increased caspase 3/7 activity (Supplementary Fig. 5C), and induced cPARP at 30 nmol/L or more (Fig. 5A).

These data suggested that in vivo combined PKI-587 and HKI-272 could be more effective than either compound alone. When PKI-587 (5 mg/kg, d×5 once) was given in combination with HKI-272 (40 mg/kg, po daily) significantly (P < 0.02), more antitumor effect than either compound given alone was observed in a 14-day study (Fig. 5B). In the combination group, tumors were 35% smaller at day 14 than with PKI-587 or HKI-272 alone, and tumor size in the combination group was 54% smaller than untreated controls. In addition, at all measurement times, combination treatment outperformed either compound given alone.

PKI-587 also showed single-agent efficacy in the H1975 model, including tumor regression at early time points after continuous dosing at more than 5 mg/kg (Supplementary Fig. S6). Furthermore, PKI-587 showed antitumor activity in an orthotopic version of the H1975 xenograft model. Nude mice with H1975 cells injected into their pleural cavity were given 25 mg/kg PKI-587 weekly. Only 1 mouse in the treatment group (10 mice per group) died, but the death was not tumor or compound related (Fig. 5C). In contrast, at day 40, all untreated animals were dead. PKI-587 at 20 mg/kg in an intermittent (day 1, 5, 9) regimen was equally effective in the H1975 orthotopic lung cancer model (Supplementary Fig. S7).

**Efficacy of PKI-587 in U87MG (glioma) tumor xenografts.** PKI-587 efficacy was tested in the U87MG glioblastoma multiforme ([GBM] PTEN negative) xenograft model, which has rapid, aggressive growth. At 25 mg/kg, daily for 2 days (d×2), PKI-587 caused U87MG tumor regression (Fig. 5D). PKI-587 at 25 mg/kg (d×2) caused mice to become pale and lethargic, but all animals recovered subsequent to compound cessation. PKI-587 tested at 1.56 to 12.5 mg/kg (d×5, dose–response regimen) caused significant (P < 0.01) tumor growth inhibition at all dosing levels. PKI-587 at 12.5 and 6.25 mg/kg resulted in 78% and 86% reduction in tumor growth, respectively, at day 10 compared with control group (Supplementary Fig. S8). PKI-587 at both 6.25 and 12.5 mg/kg had a cytostatic effect on U87MG tumors.

**Discussion**

PKI-587 was the most potent Wyeth PI3K/mTOR inhibitor advanced to clinical development. Its preclinical profile places it among the most potent dual PI3K/mTOR inhibitors reported to date (7, 8). It had subnanomolar IC₅₀ values against wild-type and mutant forms of PI3K-α, and IC₅₀ values of less than 100 nmol/L in 50 of 59 tumor cell lines tested (Table 1, Supplementary Table S1). In vitro, PKI-587 suppression of p-Akt closely correlated with tumor cell growth suppression. PKI-587 potency translated into a broad range of in vivo efficacy in MDA-MB-361, BT474, H1975, U87MG, and HCT116 tumor models. These models have reported genetic changes that aberrantly upregulate PI3K signaling including PIK3CA mutation (E545K, H1047R, K111N), receptor tyrosine kinase (RTK) over-expression (HER2*), RTK mutation (EGFR; L858R, T790M), PTEN phosphatase inactivation, or K-Ras mutation (2, 5–8, 18). PKI-587 caused regression in MDA-MB-361 [HER2*/PIK3CA (E545K)], H1975 (EGFR, L858R/T790M), and U87MG (PTEN) models. MDA-MB-361 was the most sensitive to PKI-587, whereas HCT116 [K-Ras, PIK3CA (H1047R)] was less responsive. The PKI-587 antitumor efficacy gradient was as follows: MDA-MB-361, BT474 > H1975, U87MG > HCT116.

In MDA-MB-361 cells, PKI-587 activated caspase 3/7, induced cPARP, and caused cell death after relatively short exposure times (Fig. 2A–C; Supplementary Fig. S2A). Furthermore, PKI-587 caused much more rapid and extensive activation of caspase 3/7 and cPARP induction in MDA-MB-361 than the highly selective mTOR inhibitor MTI-178 (Fig. 2C; Supplementary Fig. S2B). The effects of PKI-587 or MTI-178 on caspase 3/7 activation and induction of cPARP coincided with PKI-587 suppression of p-Akt at T308 and not p-Akt at S473 (Supplementary Fig. S2B). This strongly indicates that some PIK3–sensitive components of the PI3K/Akt portion of the PI3K/Akt/mTOR signaling pathway regulate MDA-MB-361 cell survival. More sophisticated analysis (e.g., phosphoproteomics, expression profiling) may reveal what factor(s) is responsible for acute MDA-MB-361 sensitivity to PKI-587 and possibly point to novel drug discovery target(s).

In the MDA-MB-361 xenograft model, PKI-587 suppressed Akt phosphorylation (T308 and S473), induced cPARP, and when given at sufficient levels, caused tumor regression (Fig. 3D; Supplementary Fig. S3B). PKI-587 at 2 mg/kg suppressed p-Akt and induced cPARP at 1-hour postadministration, but this effect dissipated at 8 hours (Fig. 3A). PKI-587 at equivalent doses (1–3.125 mg/mL, Fig. 3A; Supplementary Fig. S3) only attenuated tumor growth. PKI-587 at 25 mg/kg (single dose) suppressed p-Akt (T308 and S473) for up to 36 hours and cPARP was evident for up to 18 hours (Fig. 3A). PKI-587 at 25 mg/kg (once weekly) rapidly shrank large MDA-MB-361 tumors (tumor volume of ~1,000 mm³ reduced to ~230 mm³; Fig. 3D) and suppressed tumor re-growth. Data from the MDA-MB-361 model indicate that tumor regression correlated with durable p-Akt suppression, a result of the potency and long plasma half-life of PKI-587.

Not surprisingly, the HCT116 colon model with mutant K-Ras and PI3K-α, was refractory to PKI-587. This was consistent with data from various studies evaluating PI3K inhibitor efficacy in tumor models with mutant K-Ras. Combining PKI-587 with a MEK inhibitor in the HCT116 model was an obvious choice because such combinations mitigate resistance to PI3K inhibitors found in tumor cell lines harboring mutant K-Ras (31–34). In vitro the proapoptotic effects of PKI-587 were greatly enhanced when it was combined with PD0325901 (Fig. 4C), and...
in vivo the antitumor effect of combined PKI-587 and PD0325901 exceeded that achieved by either compound alone. Positive outcome from this combination study in HCT116 suggests that PKI-587 along with Ras/MAPK signaling inhibitors may achieve clinical response in cancers driven by mutant K-Ras.

We also tested PKI-587 in combination with cisplatin, taxol, and camptothecin in the HCT116 model. We chose these agents because they are standard-of-care colon cancer treatments (35). Only the camptothecin/PKI-587 (in vitro) or irinotecan (camptothecin analogue)/PKI-587 (in vitro) combinations showed enhanced efficacy. In cells, camptothecin causes covalent topoisomerase I–DNA complexes (36), which reportedly convert into DNA double-strand breaks on collision with the replication fork (36). This induces p53-mediated DNA damage response and cell-cycle arrest at G₂/M (36, 37). In HCT116, Akt activation may allow these cells to bypass DNA damage–associated G₂ arrest (36, 37). Inhibition of Akt activation by PKI-587 may account for enhanced in vivo antitumor effects of the PKI-587/irinotecan combination against HCT116. Data in Figure 4B suggests that our hypothesis may be correct, but only further experimental data will confirm this.

PKI-587 showed single-agent efficacy in both xenograft and orthotopic versions of the H1975 [NSCLC; EGFR (L858R/T790M)] model. In H1975 xenografts, continuous dosing of PKI-587 (at >5 mg/kg) caused early time point tumor regression. In the H1975 orthotopic model, 25 mg/kg PKI-587 (weekly) kept (9 of 10) treated mice alive, whereas all control mice (10 of 10) were dead by day 40. This suggests that PKI-587 could be used against lung tumors that have acquired resistance to EGFR inhibitors such as Iressa or Tarceva. PKI-587 may also be effective in lung tumors that have acquired resistance to HER2/EGFR inhibitors by MET amplification or IGF-IR (insulin-like growth factor I receptor) or AXL activation (38, 39). PKI-587 efficacy in tumors driven by these RTKs will hinge on their dependence on PI3K/mTOR signaling for growth and survival. Because these RTKs activate both PI3K/mTOR and Ras/MAPK signaling pathways, PKI-587 in combination with targeted agents that inhibit Ras/MAPK signaling (e.g., PD0325901, HKI-272, herceptin) should be tested. Indeed, the antitumor efficacy of concomitant inhibition of the PI3K/mTOR and Ras/MAPK signaling pathways has been shown in NSCLC, breast, and colon tumor models (31–34, 40, 41). In H1975, HKI-272 suppression of p-MAPK (Fig. 5A) enhanced PKI-587 in vivo efficacy. This combination improved PKI-587–mediated caspase 3/7 activation and cPARP induction in vivo and, more importantly, improved PKI-587 antitumor efficacy in vivo (Supplementary Fig. SSC; Fig. 5A and B). Clinical outcome for NSCLC is especially bleak (42), and further investigation of PKI-587 alone and in combination with Ras/MAPK signaling inhibitors in NSCLC models should help direct PKI-587 clinical development.

In the aggressive glioma model, U87MG [PTEN negative], 25 mg/kg PKI-587 for 2 consecutive days caused tumor regression. However, this occurred near the limit of tolerated exposure in nude mice. Lower dosing regimens (1.56–12.5 mg/kg, d × 5) were well tolerated and effectively suppressed U87MG tumor growth (Supplementary Fig. S8). Patients with PTEN-negative GBM tumors in general have a poor prognosis (43). In clinical settings, gliomas are resistant to EGFR inhibitors, radiotherapy, and most alkylating agents (44). In GBM, PKI-587 may be efficacious either as a single agent or in combination with cytostatic or cytotoxic drugs. Good rationale for testing such combinations derives from data showing that the Novartis PI3K/mTOR inhibitor BEZ235 combined with temozolomide caused U87MG tumor xenograft regression (7).

A key question about PI3K/mTOR signaling inhibitors has been: Can they cause tumor regression in preclinical models? Some compounds (e.g., PKI-402, GDC-0941) have positively answered this question (23, 45). But here we show how the very potent PKI-587 profile caused regression (e.g., U87MG model) at lower and/or less frequent dosing regimens than those reported for other PI3K pathway signaling inhibitors. An example of this is the comparison of PKI-587 with PKI-402, a PI3K/mTOR inhibitor we previously reported (23). In the aggressive U87MG glioma tumor model, used as a critical potency test, PKI-402 at 100 mg/kg (d × 5, near MTD) only attenuated U87MG xenograft tumor growth (23). PKI-587 at just 25 mg/kg (d × 2) caused U87MG tumor regression (Fig. 5D). Less efficacy by PKI-402 in the U87MG model correlated with a shorter 3.5 hour plasma half-life than that of the 14.4 hours for PKI-587 (25 mg/kg single dose, both). The compelling efficacy profile of PKI-587 will be more firmly established by direct experimental comparison of PKI-587 with other PI3K/mTOR signaling inhibitors.

Finally, unlike cytostatic PI3K inhibitors that cause tumor cell G₀/G₁ arrest (45–47), potent PKI-587 inhibition of class 1 PI3Ks can fully inhibit Akt activation and cause apoptosis induction (e.g., MDA-MB-361 model). This is the desired outcome against cancer cells. PKI-587 antitumor efficacy and its favorable drug safety profile in toxicology studies enabled it to enter phase 1 clinical evaluation in December 2009.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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