Modulators of Arginine Metabolism Do Not Impact on Peripheral T-Cell Tolerance and Disease Progression in a Model of Spontaneous Prostate Cancer

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Abstract

Purpose: Chronic inflammation, recruitment of myeloid-derived cells, and perturbation of the arginine metabolism have been all proposed as mechanisms favoring prostate carcinogenesis and tumor immunoevasion. Objective of this study was to evaluate whether accumulation of CD11b⁺Gr1⁺ cells, also defined myeloid-derived suppressor cells, occur in mice affected by transplantable or spontaneous prostate cancer (PC). We also investigated whether N(G) nitro-L-arginine methyl ester (l-NAME) and sildenafil, both modulators of the arginine metabolism, restrain tumor growth and restore tumor-specific immunity.

Experimental Design: Wild-type C57BL/6 mice bearing TRAMP-C1 PC and transgenic adenocarcinoma of the mouse prostate (TRAMP) mice were treated with vehicle, l-NAME or sildenafil, and evaluated for CD11b⁺ cells accumulation in the blood, several organs, and the tumor mass and for disease progression.

Results: CD11b⁺Gr1⁺, CD11b⁺Gr1low, and CD11b⁺Gr1⁺ cells differently accumulated in different organs and especially in the tumor of the two mouse models. l-NAME and sildenafil impaired the immunosuppressive function of CD11b⁺ cells in both models and restrained TRAMP-C1 growth, but they neither break tumor-specific immune tolerance nor limit tumor progression in TRAMP mice.

Conclusions: Collectively, our results emphasize substantial differences in tumor-induced alteration of myelopoiesis and sensitivity to modulators of the arginine metabolism between a transplantable and a spontaneous model of PC. They also suggest that perturbation of the arginine metabolism is dispensable for PC progression and the associated T-cell tolerance. Clin Cancer Res; 17(5); 1012–23. ©2011 AACR.

Introduction

The pathogenesis of prostate cancer (PC), the second cause of death for neoplasia among men worldwide (1), has been linked to chronic inflammation (2). PC may also represent a site of acquired immune privilege (3) where several mechanisms suppress both locally and systemically the tumor-specific immune response (4). Among these mechanisms, myeloid-derived suppressor cells (MDSC; ref. 5) have been recently the focus of intense investigation (5). MDSCs are a heterogeneous population of cells of myeloid origin that include immature macrophages, granulocytes, dendritic cells (DC), and other myeloid cells (6, 7). Recruitment of MDSCs to peripheral organs under pathologic conditions is mediated by several soluble factors, among which are interleukin (IL)-3, IL-6, IL-10, VEGF, macrophage colony-stimulating factor, and granulocyte-macrophage colony-stimulating factor (GM-CSF; refs. 5, 7, 8). In mice, MDSCs are characteristically CD11b⁺, express the Gr1 antigen at different levels, and may also express CD31, IL-4 receptor a-chain, CD115, and CD80 (5, 9). The phenotype of human MDSCs is ill defined (6, 9), and a population of CD14⁺HLA-DRlow/⁻ MDSCs has been recently observed in the peripheral blood of PC patients that significantly correlated with circulating -PSA levels (10).

MDSCs may overexpress both inducible nitric oxide synthase (iNOS) and arginase 1 (Arg1), enzymes involved in the metabolism of arginine. As reviewed in ref. 6, depletion of arginine from the microenvironment inhibits T-cell activation and proliferation and favors T-cell apoptosis. Furthermore, iNOS produces nitric oxide (NO), which interferes with IL-2 receptor signaling, leading to cell-cycle arrest. Reactive oxygen species and peroxynitrates, biproducts of arginine metabolism, contribute to T-cell inhibition. Of relevance, T cells infiltrating either human PC or the prostate of transgenic adenocarcinoma of the mouse prostate (TRAMP) mice, a primary model of spontaneous PC (11), are functionally impaired and...
Translational Relevance

Therapeutic strategies are urgently needed to overcome the immunosuppressive tumor microenvironment. Perturbation of the arginine metabolism has been proposed as one such mechanism which also favors carcinogenesis. We have investigated the therapeutic potential of modulators of the arginine metabolism, among which sildenafil that is already used in clinical trials, in realistic mouse models of prostate cancer (PC). We have found that these drugs, at concentration known to be effective in models of transplantable cancer, neither broke tumor-specific T-cell tolerance nor restrained spontaneous PC progression. Further investigation is needed to fully appreciate the therapeutic potential of modulators of the arginine metabolism in cancer.

Known, however, the in vivo effect of these treatments in models of spontaneous tumor development that more closely recapitulate the human pathology. We investigated here whether accumulation of CD11b+ cells occurs in mice bearing transplantable PC and in TRAMP mice and whether treatment with 1-NAME or sildenafil restores antitumor immunity and delays tumor growth.

Materials and Methods

Mice, cell lines, and reagents

Heterozygous C57BL/6 TRAMP mice and wild-type (WT) mice were housed and bred in a specific pathogen-free animal facility and treated in accordance with the European Union guidelines and with the approval of the Institutional Ethical Committee. Animals were typed for Tag expression by PCR-based screening assay, as described (24). TRAMP-C1 cells (25) were cultured in Dulbecco’s modified Eagle’s medium (Cambrex) supplemented with 10% FBS (Invitrogen), 150 U/mL streptomycin, and 200 U/mL penicillin (Cambrex). B6/K-0, a kidney cell line expressing Tag (26), and B6/K-1,4,5 cells, which lack the Tag epitopes I, II/III, IV, and V (27), were a generous gift of Dr. S.S. Tevethia (The Pennsylvania State University College of Medicine, Hershey, PA). RMA is an H-2b Rauscher virus–induced thymoma (28). These cell lines were cultured in RPMI-1640 (Invitrogen) supplemented with 2 mmol/L L-glutamine, 150 U/mL streptomycin, 200 U/mL penicillin, and 10% heat-inactivated FBS. Unless specified, all chemical reagents were from Sigma-Aldrich, and monoclonal antibodies (mAb) were from BD Pharmingen.

In vivo experiments

A schematic representation of the in vivo experiments is reported in Supplementary Figure 1. Six- to 8-week-old C57BL/6 male mice were challenged subcutaneously with 2.5 × 10^6 TRAMP-C1 cells. 1-NAME (added in drinking water at 1 g/L) or sildenafil (0.7 g/L; Pfizer; a generous gift of Drs. G. Da Pozzo and A. Salonia, San Raffaele Scientific Institute, Milan, Italy) were administered starting on the day of tumor challenge. As control, groups of animals were treated with vehicle (PBS) only. Water was given ad libitum. Calculation of the dosage of the drug was based on the assumption that a mouse drinks approximately 3 mL/24 h. Mice were monitored twice a week and sacrificed after 50 days. Animals were weighted and tumor size was evaluated both by measuring its weight and 2 perpendicular diameters by a caliper. Cohorts of 13- or 21-week-old TRAMP and WT mice were randomly assigned to either one of the following 3-week treatments: 1-NAME (1 g/L), sildenafil (0.13 or 0.7 g/L), or vehicle dissolved in drinking water. At the end of the second week, all mice were vaccinated intradermally with bone marrow (BM)-derived DCs (5 × 10^7/mouse; ref. 29) pulsed with 2 μmol/L of the immunodominant cytotoxic T lymphocyte epitope Tag-IV (sequence 404–411; Research Genetics; ref. 30; DC/Tag-IV). At the end of the third and last week, animals were
killed. Animals and their urogenital apparatus (UGA) were weighted. UGA were processed for histology and immunohistochemistry and scored on coded samples by a pathologist as previously described (31).

**Phenotypic characterization of cell populations**

Single-cell suspensions of tumor-draining lymph nodes (TDLN) and non–tumor-draining lymph nodes (NDLN) from tumor-challenged and -naive mice, respectively, were stained with fluorescein isothiocyanate (FITC)-labeled anti-CD4, phycoerythrin (PE)-labeled anti-CD44, and PerCP-Cy 5.5–labeled anti-CD8 mAb. Dead cells were excluded by physical parameters and/or by the addition of TOPRO5 (Molecular Probes) immediately before flow cytometric analysis. For enumeration of CD11b⁺ cells, blood and cells from spleen, BM, NDLN, TDLN, and tumor samples of naive and tumor-challenged mice were incubated with Fc-blocking mAb and stained with FITC-conjugated CD11b and APC-conjugate Gr1 mAb. Dead cells were excluded by propidium iodide. For enumeration of CD4⁺CD25⁺Foxp³⁺ cells, LN cells were stained with FITC-labeled anti-CD4, PerCP-Cy 5.5–labeled anti-CD8, and APC-labeled anti-CD25 (clone PC61) mAb, permeabilized, and, finally, stained with PE-labeled anti-Foxp3 mAb (eBioscience) according to the manufacturer’s instructions. In all experiments, cells were analyzed on a BD FacsCalibur or FacsCanto.

**In vitro cytotoxicity assay**

Splenocytes or magnetic bead-purified CD8⁺ cells (Miltenyi Biotec) were restimulated in vitro in the presence of irradiated B6/K-0 cells (10:1 ratio). Irradiated splenocytes (1:1 ratio) were added to the CD8⁺ cell culture. day 5 blasts were tested for cytolitic activity in a standard 4-hour ⁵¹Cr release assay (29). ⁵¹Cr release of target cells alone was always less than 25% of maximal ⁵¹Cr release (target cells in 0.25 mol/L SDS).

**CD11b⁺ cell purification and in vitro functional assays**

CD11b⁺ cells were purified with mouse CD11b MicroBeads (Miltenyi Biotec) following the manufacturer’s instructions. Purity of the cell population was evaluated by flow cytometry and exceeded 90%. C57BL/6 splenocytes (3 × 10⁵ cells/well) were stimulated in wells that had been coated with 3 μg/mL anti-CD3 and 2 μg/mL anti-CD28 mAb or in the presence of CD3/CD28 beads (2 μL/8 × 10⁴ splenocytes; Invitrogen). Purified splenic CD11b⁺ cells were added to the culture so as to constitute 1% to 20% of the total cells. After 3 days of incubation, cultures were pulsed with ³H-thymidine (1 μCi/well; Amersham Corp) for the last 18 hours. The incorporation of ³H-thymidine by proliferating T cells (triplicate cultures) was measured by scintillation counting.

**Arg and iNOS detection**

Immunomagnetic bead-purified CD11b⁺ cells or tumor cells were lysed for 10 minutes with lysis buffer containing 0.15 mmol/L peptatin A, 0.2 mmol/L leupeptin, and 0.4% Triton X-100. Samples were centrifuged at 20,000 × g at 4 °C for 10 minutes. The supernatant was used for Arg assay with Quantichrom Arginase Assay Kit (DARG-200; Gentaur) according to the manufacturer’s instructions. Results were normalized on 10⁶ cells. Alternatively, CD11b⁺ cells were assessed ex vivo for intracellular iNOS on stimulation with LPS (1 μg/mL) for 24 hours. Brefeldin A was added during the last 5 hours of pulsing. Cells were fixed in 2% paraformaldehyde, permeabilized, incubated with polyclonal rabbit anti-iNOS antibody (Santa Cruz Biotecology) at a 1:200 dilution for 30 minutes at room temperature, and finally analyzed by flow cytometry.

**Statistical analyses**

Statistical analyses were done using the log-rank Student’s t, ANOVA, and Newman–Keuls tests. Values were considered statistically significant for P < 0.05.

**Results**

**Effects of modulators of arginine metabolism on myelopoiesis and growth of TRAMP-C1 tumors in vivo**

We initially investigated the effect of tumor growth on myelopoiesis and recruitment of CD11b⁺ cells to peripheral organs in the TRAMP-C1 model. TRAMP-C1 is a hormone-independent PC cell line derived from a TRAMP tumor (25) and widely used to assess novel therapeutic approaches for PC. Interestingly, TRAMP-C1 tumors in vivo are infiltrated by CD11b⁺ cells and iNOS mRNA is barely detectable, whereas Arg1 mRNA levels are high and persist as tumor grows (32). Hence, tumor cells were injected subcutaneously in C57BL/6 male mice and the same-day mice were randomly assigned to a treatment group treated with either L-NAME or vehicle added to the drinking water (Supplementary Fig. 1A). We choose this treatment because it was shown to be highly effective in restraining growth of other subcutaneous tumors in C57BL/6 mice (22). Animals were killed 50 days later when the tumor mass in vehicle-treated mice had reached the dimension of approximately 150 mm². Flow cytometric analysis of the blood showed a dramatic accumulation of CD11b⁺Gr1high cells in vehicle-treated mice (Fig. 1B) when compared with naive age- and sex-matched littermates (Fig. 1A). Quantification of CD11b⁺Gr1high cells showed a statistically significant increase in tumor-bearing mice (Fig. 1D) that was marginally restrained by L-NAME treatment (Fig. 1C and D). At difference with other transplantable tumor models (22), we did not find a relevant accumulation of CD11b⁺Gr1high and CD11b⁺Gr1high cells in the blood of mice affected by TRAMP-C1 tumors (Fig. 1B and D). Because the blood may not be fully representative of myelopoiesis and MDSC accumulation during tumor growth, we quantified MDSCs in lymphoid organs and tumors of untreated and L-NAME–treated mice. An accumulation both as percentage and absolute number of CD11b⁺Gr1high cells was also evident in the spleen (Fig. 1D and Supplementary Fig. 2, respectively) and tumor samples (Fig. 1D and Supplementary Fig. 2, respectively) of tumor-bearing mice. Immunofluorescence on
TRAMP-C1 tumor sections confirmed that CD11b\(^{+}\) cells infiltrating the tumor tissue were mostly Gr1\(^{-}\) (Supplementary Fig. 3). The BM was already rich in CD11b\(^{+}\) Gr1\(^{\text{int}}\) cells that further increased in tumor-bearing mice (Fig. 1D). A marginal increase in CD11b\(^{+}\) Gr1\(^{\text{high}}\) cells was also evident in TDLN, where the mature CD11b\(^{+}\) Gr1\(^{-}\) cells dominated and further increased in mice affected by TRAMP-C1 tumors (Fig. 1D and Supplementary Fig. 2). The CD11b\(^{+}\) Gr1\(^{\text{int}}\) population was clearly enriched only in the spleen (Fig. 1D and Supplementary Fig. 2). \(\iota\)-NAME treatment did not significantly modify MDSC accrual in the organs examined except for NDLN and TDLN, where the percentage and number of CD11b\(^{+}\) Gr1\(^{\text{int}}\) cells declined (Fig. 1 and Supplementary Fig. 2).

Accumulating CD11b\(^{+}\) cells were indeed MDSCs, because CD11b\(^{+}\) cells purified from the spleen of tumor-bearing mice inhibited the \textit{in vitro} proliferation of splenocytes in a dose-dependent manner (Fig. 2A). Hence, MDSC accumulation is a characteristic also of subcutaneous hormone-independent PC. Of relevance, treatment with \(\iota\)-NAME reduced the immunosuppressive and Arg1 enzymatic activities of both CD11b\(^{+}\) splenocytes (Fig. 2A) and cells collected from the tumor mass (Fig. 2B).

Because TDLN rapidly enlarge owing to the accumulation of myeloid and lymphoid cells (33), we asked whether arginine metabolism inhibitors could alter the recruitment of T lymphocytes in TDLN. In C57BL/6 mice affected by TRAMP-C1 tumors, a 3-fold increase in cell number was evident in TDLN when compared with LN from naive mice (Supplementary Figs. 2 and 4). Flow cytometric analysis of TDLN cells showed a significant increase in the number of both CD4 and CD8 cells (Supplementary Fig. 2). As seen in other transplantable tumor models (22), the percentage of CD8 and more evidently of CD4\(^{+}\) T cells (Supplementary Fig. 4) in TDLN decreased in both \(\iota\)-NAME- and vehicle-treated mice, suggesting that in TDLN a relevant perturbation of the physiologic equilibrium among the different cell populations was undergoing that was not modified by \(\iota\)-NAME. Although MDSCs can favor induction of regulatory T cells in tumor-bearing hosts (34), in the TDLN of these

![Graphs showing flow cytometric analysis of TDLN cells](image)

Figure 1. Treatment with \(\iota\)-NAME does not alter accrual of CD11b\(^{+}\) cells in C57BL/6 mice bearing subcutaneous TRAMP-C1 tumors. C57BL/6 male mice were left untreated (naive) or challenged subcutaneously with TRAMP-C1 cells. The same-day, tumor-challenged mice were randomly assigned to either one of the following treatments: \(\iota\)-NAME (1 g/L) or vehicle added to the drinking water. Recruitment of CD11b\(^{+}\) cells in the blood and the indicated organs of naive mice and mice bearing a 50-day-old tumor were investigated by flow cytometry. Dot plot, representative results from the blood of 1 of 9 naive mice (A) and TRAMP-C1-bearing mice treated with vehicle (B; \(n=11\)) or \(\iota\)-NAME (C; \(n=16\)). The regions (R1–R3) used to define CD11b\(^{+}\) Gr1\(^{\text{int}}\) (R3), CD11b\(^{+}\) Gr1\(^{\text{high}}\) (R2), and CD11b\(^{+}\) Gr1\(^{\text{int}}\) (R1) cell populations, respectively, are depicted in the panels. D, histograms reporting the percentage of CD11b\(^{+}\) subpopulations as aggregated data (average \(\pm\) SD) for blood, spleen, BM, NDLN, TDLN, and tumor samples, respectively, collected from naive (white bars) and tumor-bearing animals treated with vehicle (black bars) or \(\iota\)-NAME (dashed bars) and analyzed as described earlier. Statistical analysis was done using the ANOVA and Newman-Keuls tests: *, 0.01 < \(P < 0.05\); **, 0.001 < \(P < 0.01\); ***, \(P < 0.001\).
mice, we found a slightly, yet not significant, increase in CD4^+CD25^+Foxp3^+ cells, which was not modified by i-NAME treatment (Supplementary Fig. 4).

Finally, subcutaneous tumors were measured and compared in vehicle- and i-NAME-treated animals. At day 35, the tumor volume and area (Fig. 2C) in i-NAME–treated mice, we found a slightly, yet not significant, increase in CD4^+CD25^+Foxp3^+ cells, which was not modified by i-NAME treatment (Supplementary Fig. 4).

Finally, subcutaneous tumors were measured and compared in vehicle- and i-NAME–treated animals. At day 35, the tumor volume and area (Fig. 2C) in i-NAME–treated animals were increased compared to vehicle-treated animals. i-NAME treatment led to a significant reduction in tumor volume and area, indicating that i-NAME has a negative impact on tumor growth.

Figure 2. Treatment with i-NAME reduces the immunosuppressive activity of both CD11b^+ splenocytes and cells from the tumor mass and restrains TRAMP-C1 tumor growth. CD11b^+ cells were magnetic bead purified from the spleen of C57BL/6 mice treated as described in the legend to Figure 1 and sacrificed at day 35. CD11b^+ cells were added at a final concentration that ranged from 20% to 1% to a mixed leukocyte culture setup with C57BL/6 splenocytes, as responders, stimulated with anti-CD3/CD28 beads (A, left). Positive and negative controls were splenocytes cultured in the absence of MDSCs and stimulated (+) or not (−) with anti-CD3/CD28 beads. Data are expressed as the counts per minute (cpm) mean ± SD of triplicates. A, right, in parallel, CD11b^+ splenocytes from mice treated with vehicle (n = 4; black bar) or i-NAME (n = 6; dashed bar) were analyzed for Arg1 enzymatic activity as described in the Materials and Methods section. Data are expressed as units/L of Arg1 activity of 10^6 cells ± SD. TRAMP-C1 tumor cells from the same mice were assessed for in vitro immunosuppressive (A) and Arg1 enzymatic activities (B, left and right, respectively) as described earlier. Data in each panel are representative of at least 3 independent experiments. In parallel, C57BL/6 mice were challenged with TRAMP-C1 tumor cells and treated with vehicle (n = 5) or i-NAME (n = 7) as described in the legend to Figure 1. B, tumor dimension at day 35 is expressed as average ± SD volume (left) and area (middle). C, right, treatment–related toxicity was evaluated by comparing the body weight of vehicle and i-NAME–treated mice. Statistical analysis was done using the Student’s t test: *, 0.01 < P < 0.05; **, 0.001 < P < 0.01. Data in each panel are representative of at least 3 independent experiments. D, additional groups (8 animals/group) of vehicle (white circles) and i-NAME–treated mice (black circles) were followed for survival and sacrificed when the tumor mass had reached the dimension of approximately 100 mm^2. Survival curves are reported in a Kaplan–Meier plot. Statistical comparison (log-rank test): P = 0.0006.
mice were half of those measured in vehicle-treated animals. To confirm this therapeutic effect, vehicle- and L-NAME–treated mice were followed in a survival experiment. As reported in Figure 2D, when all vehicle-treated mice were dead because of the tumor outgrowth, more than 50% of the tumor-bearing mice treated with L-NAME were still alive and their overall survival was significantly increased. As a measure of potential drug-related toxicity, we monitored animal weight and found no differences between drug- and vehicle-treated mice (Fig. 2C). Also, as previously reported (22), this treatment did not cause macroscopic and microscopic damage to the liver, kidney, respiratory apparatus, and esophagus (data not shown). All together, these data suggest that in mice bearing subcutaneous PC, L-NAME treatment can modulate the arginine metabolism both in periphery and at the tumor site, reduce the immunosuppressive activity of MDSCs, and restrain tumor growth.

C57BL/6 mice challenged with TRAMP-C1 cells were also treated with sildenafil (Supplementary Fig. 1B). This drug was initially used at 20 mg/kg/24 h (0.13 g/L, as suggested in ref. 23). However, this treatment did not impact on TRAMP-C1 growth (data not shown). Considering that mice have a metabolic activity higher than humans and that a dose of 100 mg/kg/24 h (0.7 g/L) is needed to reach a free plasma sildenafil concentration of approximately 10 mmol/L (35), which is well within the range found in treated humans, mice were subjected to this dose. At day 24, when the experiment ended, the tumor volume and area in sildenafil-treated mice were half of those measured in vehicle-treated animals whereas the body weight was comparable (Fig. 3A). This treatment regimen was associated with reduced iNOS expression by CD11b+ cells (Fig. 3B). More important, the effects of the drug were measurable also at the tumor site, where sildenafil treatment reduced the Arg1 enzymatic activity (Fig. 3C) and the in vitro immunosuppressive activity of the cells collected from the tumor mass (Fig. 3D).

Modification of myelopoiesis in TRAMP mice developing spontaneous PC

Several drawbacks may bias the results obtained in models of in vitro transplantation of in vitro stabilized tumor cells. Indeed, tumor cells undergo substantial genetic and epigenetic alterations during in vitro passages (e.g., ref. 36). Also, cells are usually engrafted without their stroma, therefore altering the natural development of a tumor mass (37). Finally, inflammation is generated at the site of injection (38), potentially induced also by contaminating bacterial products, known as the powerful inducers of MDSC (39).

Hence, similar experiments were conducted in the more realistic TRAMP model. TRAMP mice are transgenic for the SV40 early genes (Tag) expressed under the control of the rat probasin regulatory element. As a result, starting at puberty, male mice invariably and progressively develop spontaneous mPIN (weeks 6–12), adenocarcinoma (weeks 12–18), with lymph node and visceral metastasis (weeks 18–30; weeks 18–30; ref. 40), thus recapitulating human PC (41). Animals were first analyzed for the accumulation of CD11b+ cells during PC development and progression (Fig. 4). At 9 weeks of age, when animals are affected by scattered foci of mPIN (40), the percentage of CD11b+Gr1high, CD11b+Gr1int, and CD11b+Gr1- cells in the blood of TRAMP mice was substantially similar to that found in age-matched WT animals. In the following week, although a relevant variability among animals of the same age was evident, a significant increase in the percentage of all populations of CD11b+ cells was found in
TRAMP mice and reached a plateau after week 20. However, a similar variability and increase was found also in age-matched WT animals, as previously reported by others in 12-month-old WT mice (42).

We hypothesized that variability in disease stage in TRAMP mice of the same age could account for the variability in CD11b\(^+\) cells accrual. Hence, samples from TRAMP mice were regrouped for disease score, and compared with data from WT animals of similar age. CD11b\(^+\)Gr1\(^{high}\) cells were more abundant in the early phases of tumor development when the disease is localized to prostate lobes (Supplementary Fig. 5; disease score 2). A significant increase in the percentage of CD11b\(^+\)Gr1\(^{-}\) cells was evident in TRAMP mice affected by advanced mPIN (Supplementary Fig. 5; disease score 3). Curiously, both CD11b\(^+\)Gr1\(^{-}\) and CD11b\(^+\)Gr1\(^{high}\) cells seemed to be reduced in TRAMP mice affected by adenocarcinoma (Supplementary Fig. 5; disease score 5). No differences were found for CD11b\(^+\)Gr1\(^{int}\) cells at any disease stage (Supplementary Fig. 5).

To gain better insights on MDSC accrual in TRAMP mice when the animals are affected by adenocarcinoma, male TRAMP and WT age-matched littermates were killed at 16 weeks of age and CD11b\(^+\) cells were quantified in the blood, several organs, and the prostate. CD11b\(^+\) cells accumulation in the blood, BM, and TDNL of TRAMP mice (Fig. 5 and Supplementary Fig. 6) mimicked, although to a lesser extent what we found in mice bearing TRAMPC-1 tumors (Fig. 1 and Supplementary Fig. 2). At difference with mice bearing TRAMPC-1 tumors, however, there was no accumulation of CD11b\(^+\) cells in the spleen of TRAMP mice (Fig. 5 and Supplementary Fig. 6). Even more strikingly, spontaneously tumors were characterized by the accumulation of CD11b\(^+\)Gr1\(^{-}\) cells (Fig. 5D) whereas TRAMPC-1 tumors were infiltrated mainly by immature CD11b\(^+\)Gr1\(^{high}\) cells and to a lesser extent by CD11b\(^+\)Gr1\(^{int}\) cells at any disease stage (Supplementary Fig. 5). Immunofluorescence of TRAMP prostate sections showed that most of the CD11b\(^+\) cells were localized in the stroma of transformed acini and were Gr1\(^{-}\), therefore confirming flow cytometric analyses (Supplementary Fig. 3). These data suggest that perturbation of myelopoiesis is substantially different in mice bearing transplantable or spontaneous tumors.

To confirm that also the CD11b\(^+\) cells accumulating in TRAMP mice were "bona fide" MDSCs, splenic CD11b\(^+\) cells were purified by magnetic bead sorting and added to a culture of C57BL/6 splenocytes stimulated with anti-CD3 and anti-CD28 antibodies. Indeed, CD11b\(^+\) cells purified either from young (i.e., 9 weeks) or aged (i.e., 15 weeks) TRAMP mice could substantially reduce splenocyte proliferation (Supplementary Fig. 7).

**Figure 4.** CD11b\(^+\) cells accumulate in the blood of both aged TRAMP and WT mice. Blood cells from naive WT and age-matched TRAMP male littermates were stained for CD11b and Gr1 markers and analyzed by flow cytometry. A, dot plots depicting results from 1 of 3 WT and 15 TRAMP mice of 16 to 20 weeks of age (SSC-H, side scatter channel; FSC-H, forward scatter channel). Right, regions (R1–R3) used to define CD11b\(^+\)Gr1\(^{-}\), CD11b\(^+\)Gr1\(^{int}\), and CD11b\(^+\)Gr1\(^{high}\) cell populations, respectively. B, data from 8 independent experiments aggregated for mouse age (week) at the time of killing, in which blood samples collected from WT (white bars; 9 weeks: 3; 16–20 weeks: 9; and 24–36 weeks: 26 mice) and TRAMP animals (black bars; 9 weeks: 5; 16–20 weeks: 15; and 24–36 weeks: 33 mice) were analyzed as described earlier. Data are expressed as percentage of cells within the selected population. Statistical analysis was done using the Student’s t test: *, 0.01 < P < 0.05; **, 0.001 < P < 0.01; ***, P < 0.001.
week, all mice were vaccinated with DC/Tag-IV and sacrificed 1 week later. Body weight measurement (Supplementary Fig. 8A) and autopsy (not shown) did not evidence sign of drug-related toxicity. At microscopic examination, the prostate of most of the vehicle-treated animals was characterized by enlarged acini presenting a well-differentiated microinvasive PC (33). The average disease score for these animals was 3.5 ± 0.71 (Supplementary Fig. 8B). However, treatment with L-NAME did not modify MDSC accrual (Fig. 5 and Supplementary Fig. 6), and neither L-NAME nor sildenafil caused a significant delay in disease progression. The disease score was 3.33 ± 0.58 and 3.00 ± 0.00, respectively (Supplementary Fig. 8A).

Hypothesizing that MDSC accrual and immunosuppression could have occurred earlier during tumor progression, a similar experiment was conducted starting at 13 weeks of age and mice were sacrificed at week 16. At that age, the average disease score for vehicle-treated TRAMP animals was 2.17 ± 0.29 (Supplementary Fig. 8B). However, also in this case, treatment with neither L-NAME nor sildenafil caused a delay in disease progression and the disease score was 2.83 ± 1.04 and 3.33 ± 0.58, respectively (Supplementary Fig. 8A).

Finally, 13-week-old TRAMP mice were subjected to the dose of sildenafil that was shown to be effective in the TRAMPC-1 model (0.7 g/L; Fig. 3). Mice followed the treatment protocol described earlier and were killed at week 16. As reported in the TRAMPC-1 model (Fig. 3), sildenafil treatment inhibited the immunosuppressive activity of MDSCs (Supplementary Fig. 8C). However, the average disease score for TRAMP mice treated with this dose of sildenafil was similar to that of vehicle-treated mice [3.5 ± 0.53 (n = 8) and 2.8 ± 0.84 (n = 5), respectively; data not shown].

Modulators of arginine metabolism do not break tumor-specific tolerance in TRAMP mice

Our last goal was to investigate the effects of Arg and iNOS inhibitors on the endogenous tumor-specific immune response. Indeed, inhibitors of arginine metabolism may increase the endogenous antitumor immunity in models of transplantable tumors (22, 23). We have also

Figure 5. Treatment with L-NAME does not alter the accrual of CD11b+ cells in TRAMP mice. Thirteen-week-old TRAMP mice were randomly assigned to either one of the following 3-week treatments: L-NAME, or vehicle dissolved in drinking water. At the end of the second week, all mice were vaccinated intradermally with DC/Tag-IV. At the end of the third and last week, animals were killed. Dot plots depicting representative results from the blood of 1 of 5 WT (A) and TRAMP mice treated with vehicle (B; n = 6) or L-NAME (C; n = 6). The regions (R1–R3) used to define CD11b+ Gr10 (R3), CD11b+ Gr1int (R2), and CD11b+ Gr1high (R1) cell populations, respectively, are depicted in the panels. D, histograms report the percentage of CD11b+ subpopulations as aggregated data (average ± SD) for blood, spleen, BM, TDLN, and prostate samples, respectively, collected from naive (white bars) and TRAMP mice treated with vehicle (black bars) or L-NAME (dashed bars) and analyzed as described earlier. Statistical analysis was done using the ANOVA and Newman-Keuls tests: *0.01 < P < 0.05; **0.001 < P < 0.01.
recently reported that in TRAMP mice Tag, whose expression in prostate epithelial cells is quantitatively similarly to other prostate-associated antigens (43), causes the loss of responsiveness of low-avidity Tag-specific T cells (44), therefore mimicking the situation found for nonmutated TAA in patients with advanced PC (e.g., ref. (45). Hence, splenocytes recovered from 16-week-old TRAMP and WT mice, treated with vehicle, L-NAME, or sildenafil, were specifically restimulated in vitro and tested for their ability to recognize different targets. Blasts from WT mice selectively recognized and killed syngenic RMA lymphoma cells (28) pulsed with Tag-IV and B6/K-0 cells, which endogenously express Tag (Fig. 6A). No or marginal lysis was found against unpulsed RMA cells and B6/K-1,4,5 cells that lack Tag-IV (27). Irrespective of the treatment received, splenocytes from TRAMP mice did not significantly kill any target used (Fig. 6B). Lack of response did not seem to depend on the presence of immunosuppressive cells in the culture because CD8\(^+\) cells purified from the spleen of vehicle- and L-NAME–treated TRAMP mice and restimulated in vitro did not kill the relevant targets (Fig. 6C).

In summary, treatment with either L-NAME or sildenafil neither rescued the function of tumor-specific cytotoxic T lymphocytes nor delayed tumor progression in TRAMP mice.

**Discussion**

Strategies to overcome tumor-associated immunosuppression are essential to obtain the most rewarding clinical benefits from the application of immunotherapy to cancer patients.

Our results obtained both in a transplantable model of androgen-independent PC and in TRAMP mice at stages at which tumors are usually androgen-dependent indicate that tumor growth is associated with perturbation of myelopoiesis and accumulation of CD11b\(^+\) cells. Accrual of these cells was more noticeable in the transplantable TRAMP-C1 model, in which CD11b\(^+\)Gr1\(^{\text{int}}\) cells increased in the blood from less than 10% to almost 30%. A less dramatic but yet significant increase in CD11b\(^+\)Gr1\(^{\text{high}}\) cells was measurable in TRAMP mice. More important, CD11b\(^+\)Gr1\(^{\text{high}}\) cells represented approximately 20% of the tumor-infiltrating cells in TRAMP-C1 tumors, whereas in spontaneous PC CD11b\(^+\)Gr1\(^{\text{high}}\) cells were almost undetectable and infiltration by more matured CD11b\(^+\)Gr1\(^{\text{int}}\) cells and to a lesser extent by CD11b\(^+\)Gr1\(^{\text{int}}\) cells dominated. It is tempting to speculate that the accumulation of CD11b\(^+\) cells depends on the aggressiveness and proinflammatory activity of the tumor model used, being more pronounced in transplantable models. Indeed, mice challenged with C26-GM colon carcinoma cells producing GM-CSF or highly immunogenic RMA lymphoma cells show a far more intense accumulation of MDSCs and tumor-bearing mice have to be killed within the first 2 weeks (22). TRAMP-C1 cells have a rather indolent growth in vivo with a doubling time of 11.3 days (25), a characteristic of human PC. Also, the local differences between a transplantable subcutaneous PC and prostate microenvironment should be taken into account. Hence, it is not surprising to find a less dramatic increase of MDSCs in TRAMP mice, wherein tumors spontaneously develop in several months. In addition, it might be possible that the different androgen sensitivity of tumor cells in the 2 models...
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differently impacted on recruitment of inflammatory cells at the tumor site.

CD11b<sup>+</sup>G<sup>+</sup>Gr<sup>+</sup> cells likely represent a population of immature MDSC that can differentiate both in vitro and in vivo into Gr<sup>-</sup> cells (9). Increase in the percentage of CD11b<sup>+</sup>G<sup>+</sup>Gr<sup>+</sup> cells in TRAMP mice with disease score 2 and of CD11b<sup>+</sup>G<sup>+</sup>Gr<sup>-</sup> cells in mice with advanced mPIN (disease score 3), and especially in the tumor supports this hypothesis, and suggests that accrual of MDSCs is a dynamic process associated with a disease phase in which the growing tumor alters prostate architecture with stromal reaction and proliferation of smooth muscle cells. We anticipate that peripheral blood accrual of MDSCs should be a characteristic of PC patients with early-stage disease.

In the TRAMP-C1 model, both <i>L-NAME</i> and sildenafil inhibited the immunosuppressive function of CD11b<sup>+</sup> cells and consequently delayed tumor growth. Reduced TRAMP-C1 growth and prolonged animal survival, although statistically significant when compared with vehicle-treated mice, were limited and all mice eventually succumbed to the disease. Hence, TRAMP-C1 cells might be less dependable on arginine metabolism for growth than C26-GM cells.

In the TRAMP model, treatment of tumor-bearing mice with modulators of arginine metabolism inhibited the immunosuppressive function of MDSCs but neither modified accrual of CD11b<sup>+</sup> cells nor broke tumor-specific tolerance and restrained tumor growth. Although this is, to our knowledge, the first report on the <i>in vitro</i> effects of modulators of arginine metabolism in PC cancer and, more important, in a model of spontaneous tumor development, they seem at odds with several other reports showing the ability of these drugs in delaying tumor growth and increasing the endogenous tumor-specific immune response. In addition, it has been reported that <i>in vitro</i> Arg and iNOS inhibitors restore tumor-infiltrating lymphocyte responsiveness to PC both in humans and in TRAMP mice (12). Several considerations may reconcile this apparent discrepancy. First, in transplantable models, the drug is given at the time of tumor cell implantation and may more easily diffuse among dispersed tumor cells, blocking both the proangiogenic and immunosuppressive effects of Arg and iNOS. Second, tumor cells in TRAMP mice might be less dependable on arginine metabolism for growth, and this would explain why even if MDSCs also accumulate in this model, Arg and iNOS inhibitors are not effective in delaying tumor growth. At difference with the transplantable models, the drugs were administered in TRAMP mice once the tumor was already developed and well-vascularized (46). We are not aware of previous reports showing that such therapy is efficacious in already established tumors. This may be even more challenging in the TRAMP model, wherein prostate epithelial cells are continuously exposed to products of the oncogene and a profound state of tumor-specific immunosuppression associates with PC development.

An additional explanation for the lack of efficacy of the Arg and iNOS inhibitors in the TRAMP model, and perhaps in human PC, might be a limited penetration of the drugs into the tumor mass, due to altered vascularization, a characteristic common to tumors of different histotype (47). This would also explain why N(G)-monomethyl-L-arginine is so effective in restoring the function of PC-infiltrating lymphocytes <i>in vitro</i> (12), wherein the drug is freely available. One possibility to overcome this limitation is to combine drugs that normalize tumor vessels with chemotherapy and/or immunotherapy (47). Alternatively, this treatment might be suggested as adjuvant therapy after debulking surgery. Of relevance, whereas treatment with i-NAME would likely cause severe side effects (48), prolonged PDE-5 inhibitors have been approved for the treatment of several human pathologies and may be proposed as long-term therapy.

It might also be hypothesized that in TRAMP mice, the dominant mechanism by which MDSCs suppress the tumor-specific immune response is Arg- and iNOS-independent, therefore explaining the lack of antitumor effects of i-NAME and sildenafil in these mice. MDSCs may produce IL-10 and TGF-β, cytokines endowed with immunosuppressive functions (6, 9). MDSCs also express the zinc-based protease ADAM17 that cleaves α4-selectin on naive T cells, therefore decreasing T-cell ability to home to sites where they would be activated (49). Although the findings that the <i>in vitro</i> Arg and NOS inhibitors rescue the function of PC-infiltrating T cells in TRAMP mice (12) are strongly suggestive for an altered arginine metabolism, we cannot exclude that other MDSC-mediated mechanisms are responsible for the tumor-associated T-cell tolerance found in TRAMP mice (33, 44). This will be the focus of future work.

Several immunosuppressive mechanisms act simultaneously in cancer patients and contribute to immunoescape. Hence, acting simultaneously on more than one of the known tumor-associated immunosuppressive mechanisms will likely result in more successful therapeutic effects.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References
4.
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17.
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Modulators of Arginine Metabolism Do Not Impact on Peripheral T-Cell Tolerance and Disease Progression in a Model of Spontaneous Prostate Cancer

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