MARCKS Regulates Growth and Radiation Sensitivity and Is a Novel Prognostic Factor for Glioma

John S. Jarboe1,2, Joshua C. Anderson1, Christine W. Duarte4, Tapan Mehta4, Somaira Nowsheen1, Patricia H. Hicks1, Alexander C. Whitley1, Timothy D. Rohrbach1,2, Raymond O. McCubrey4, Sherard Chiu5, Tamara M. Burleson1, James A. Bonner1, G. Yancey Gillespie6, Eddy S. Yang1, and Christopher D. Willey1,2,7

Abstract

Purpose: This study assessed whether myristoylated alanine-rich C-kinase substrate (MARCKS) can regulate glioblastoma multiforme (GBM) growth, radiation sensitivity, and clinical outcome.

Experimental Design: MARCKS protein levels were analyzed in five GBM explant cell lines and eight patient-derived xenograft tumors by immunoblot, and these levels were correlated to proliferation rates and intracranial growth rates, respectively. Manipulation of MARCKS protein levels was assessed by lentiviral-mediated short hairpin RNA knockdown in the U251 cell line and MARCKS overexpression in the U87 cell line. The effect of manipulation of MARCKS on proliferation, radiation sensitivity, and senescence was assessed. MARCKS gene expression was correlated with survival outcomes in the Repository of Molecular Brain Neoplasia Data (REMBRANDT) Database and The Cancer Genome Atlas (TCGA).

Results: MARCKS protein expression was inversely correlated with GBM proliferation and intracranial xenograft growth rates. Genetic silencing of MARCKS promoted GBM proliferation and radiation resistance, whereas MARCKS overexpression greatly reduced GBM growth potential and induced senescence. We found MARCKS gene expression to be directly correlated with survival in both the REMBRANDT and TCGA databases. Specifically, patients with high MARCKS expressing tumors of the proneural molecular subtype had significantly increased survival rates. This effect was most pronounced in tumors with unmethylated O6-methylguanine DNA methyltransferase (MGMT) promoters, a traditionally poor prognostic factor.

Conclusions: MARCKS levels impact GBM growth and radiation sensitivity. High MARCKS expressing GBM tumors are associated with improved survival, particularly with unmethylated MGMT promoters. These findings suggest the use of MARCKS as a novel target and biomarker for prognosis in the proneural subtype of GBM.

Clinical Cancer Research; 18(11); 3030–41. ©2012 AACR.
Translational Relevance

Glioblastoma multiforme (GBM) is the most common and deadly primary brain malignancy necessitating improved understanding of GBM biology. In this study, we explore the role of myristoylated alanine-rich C-kinase substrate (MARCKS) in the context of GBM. We have discovered that the MARCKS protein regulates GBM growth as well as response to radiation therapy through its effects on proliferation, senescence, and DNA repair based on our studies in cell culture and in patient-derived xenograft tumors implanted in mice. Importantly, our analysis of clinical patient data shows that MARCKS is an independent predictor for outcome in GBM patients. Indeed, high MARCKS levels promoted improved outcomes, which was consistent across our model systems. Our results suggest that MARCKS may be a biomarker for prognosis as well as a potential target for therapy in GBM.

The MGMT DNA repair mechanism is merely one of many processes that contribute to poor survival in GBM. It is well known that several different mutations in oncogenes and loss of tumor suppressors may contribute to the pathogenesis of GBM, and these aberrations differ from patient to patient. This would suggest that effective treatment regimens for GBM should be tailored toward the particular pathogenesis of that patient’s neoplasm. Over the past several years, there have been many attempts to generate molecular profiles to better understand GBM and the prognostic factors that influence survival and response to therapy. Resources such as the Repository of Molecular Brain Neoplasia Data (REMBRANDT) database and The Cancer Genome Atlas (TCGA) Research Network have provided insight into the pathogenesis of GBM through allowing researchers to correlate gene expression with clinical outcome. Recently, genomic analyses of TCGA GBM samples lead to the identification of molecular subtypes, namely classical, mesenchymal, proneural, and neural. Indeed, abnormalities in several oncogenes and tumor suppressors were identified that are characteristic of each subtype (5, 6). Moreover, treatment efficacy differs among the subtypes, indicating that future clinical approaches will depend on subtype specificity (6).

One of the most common genetic alterations observed in approximately 90% of GBM is LOH of chromosome 10q (7). This alteration often occurs in conjunction with mutation of the tumor suppressor gene PTEN in up to 60% of GBMs with LOH (8, 9). PTEN executes its tumor suppressor function by antagonizing signaling through the phosphatidylinositol 3-kinase (PI3K)/Akt pathway. Activation of the PI3K/Akt pathway begins when the phospholipid, phosphatidylinositol (4,5) bisphosphate (PIP2) is phosphorylated by PI3K to phosphatidylinositol (3,4,5)-trisphosphate (PIP3). PIP3 can then recruit the kinase Akt to the plasma membrane where it is phosphorylated and activated leading to changes in migration, invasion, angiogenesis, survival, and proliferation. PIP3 can also be converted back to PIP2 by PTEN. Therefore, mutations or deletions of PTEN in gliomas lead to an overactivation of the Akt pathway and have been associated with a worse clinical outcome (10–13). This would indicate that the availability of the precursor phospholipid PIP2 is a critical factor for modulating activation of Akt and potentially, clinical outcome. Myristoylated alanine-rich C-kinase substrate (MARCKS) is a protein that is capable of regulating PIP2 availability by sequestration via an electrostatic mechanism. MARCKS was identified more than 20 years ago as a major protein substrate for protein kinase C (PKC) in brain synaptosomes (14). Since then, its major role has been defined as an integrator of PKC and calmodulin signals into control of the actin cytoskeleton (15). It carries out this function with its polybasic effector domain that cross-links actin filaments into bundles and sequesters PIP2 with favorable stoichiometry via an electrostatic mechanism (16, 17). Phosphorylation of the effector domain by PKC or binding by Ca2+-calmodulin leads to a local release of PIP2 and actin bundles, resulting in alterations in the actin cytoskeleton. This is the mechanism by which MARCKS controls cellular structural changes such as the extension of growth cones in neurons and the promotion of chemotaxis in neutrophils (18, 19). MARCKS has also been implicated in the pathogenesis of various malignancies. A particular mutation that truncates the protein within the effector domain is common in small intestinal adenocarcinomas (20). Levels of MARCKS were found to be inversely correlated with increased proliferation in melanoma cells, suggesting that MARCKS may function as a “tumor suppressor” (21). Interestingly, a recent study on MARCKS in GBM implicated this protein as a mediator of attachment and invasion, suggesting that higher levels of MARCKS would lead to a more aggressive disease (22). It is clear that the function of this protein in the pathogenesis of malignancy is multifaceted and complex.

Due to the ability of MARCKS protein to sequester PIP2, we hypothesized that MARCKS may be able to regulate growth and radiation sensitivity in GBM through regulation of the PI3K/Akt pathway. GBM cell lines and tumors with higher levels of MARCKS would have reduced availability of PIP2 for conversion to PIP3 and thus reduced activation of the PI3K/Akt pathway. This would lead to reduced growth rates and resistance to radiation; which could in turn lead to improved clinical outcomes. We investigated this in several established GBM cell lines as well as a patient-derived tumor xenograft mouse model. We confirmed the effect of MARCKS in the regulation of proliferation and radiation sensitivity with molecular manipulation and also identified a novel role in the regulation of senescence. To more closely evaluate the MARCKS protein in a clinical context, we then correlated MARCKS gene expression with molecular subtypes and clinical outcome. We found MARCKS expression to be particularly protective in the proneural subtype of GBM, indicating that this protein may be a novel biomarker and therapeutic strategy for these patients.
Materials and Methods

Xenograft passaging and processing
All animal studies were carried out in accordance with the policies set by the UAB IACUC. Dr. C. David James (Department of Neurological Surgery, UCSF, San Francisco, CA) and Dr. Jann Sarkaria (Mayo Clinic, Rochester, MN) have developed several partially characterized human glioblastoma xenograft lines through serial passage within athymic nude mice (23–25). Eight of these human glioblastoma xenograft lines were provided to Dr. Gillespie (UAB) and have been used for this study. Athymic nu/nu mice were obtained from National Cancer Institute Mouse Repository (NCI-Frederick, MD). The following primary glioblastoma xenograft lines were used for this study: GBM 6, GBM 10, GBM 12, GBM 14, GBM 15, GBM 22, GBM 39, and GBM 59 (23–25). Tumors xenografts were serially passaged in the mice (flanks) by harvesting and mechanically disaggregating the cells before injection, as has been described before (23–25). Gross tumor is macroadsected from normal tissue and snap frozen for kinetic profiling and immunoblotting as detailed below. Intracranial tumor growth was determined by measuring the time from intracranial implantation to the day the mice begin to show neurologic impairment.

Immunoblotting
Immunoblotting was carried out by standard protocols with the following primary antibodies: rabbit anti-MARCKS, Epitomics, catalog #1944-1; rabbit anti-MARCKS pS158/162, Epitomics, catalog # 2156-1; rabbit anti-Akt, Cell Signaling, catalog #4691; rabbit anti-Akt pSer473, Cell Signaling, catalog #4060S; rabbit anti-phospho DNA-PKcs T2609, Genetex, catalog #GTX-24194; rabbit anti-phospho H2AX Ser139, Millipore, catalog #51-07-164; rabbit anti-LC3B, Novus Biologicals, catalog #NB600-1384; rabbit anti-Caspase 3, Cell Signaling, catalog #9662S; rabbit anti-GAPDH (glyceraldehyde-3-phosphate dehydrogenase), Santa Cruz, catalog #sc-25778; mouse anti-α-Tubulin, Santa Cruz, catalog #sc-5286. A detailed description of the protocol is contained in the Supplementary Methods.

Plasmid constructs
A set of 5 MARCKS short hairpin RNA (shRNA) lentiviral transfer vector plasmids (catalog # RHS4533-NM_002336) and an empty vector control plasmid (catalog # RHS4080) were obtained from Thermo Biosystems. The psPAX2 packaging plasmid (Addgene plasmid 12260) and pCMV-VSV-G envelope plasmid (Addgene plasmid 8454) were obtained from Addgene (26). Detailed description of the construction of the pLenti4-MARCKS and pLenti4-DELTA plasmids is contained in the Supplementary Methods.

Lentiviral vector production
Lentivirus was generated by cotransfection of 293FT cells with pCMV-VSVG, psPAX2, and the appropriate lentiviral transfer vector with Lipofectamine. A detailed description of the production of lentivirus is contained in the Supplementary Methods.

Cell culture and stable line selection
U87, U251, U1373, U118, D54, and D32 malignant glioma cell lines (American Type Culture Collection) and 293FT human embryonic kidney cells (Invitrogen) were cultured in Dulbecco’s modified Eagle’s medium with 10% FBS and 1% Pen-Strep at 37°C in 5% CO2. A detailed description of stable cell line selection is contained in the Supplementary Methods.

Proliferation assay
A total of 2 × 10^5 cells were seeded in 96-well plates in 100 μL of media. Cells were allowed to grow for 5 days and were then harvested with the ATPlite Luminescence Assay System (catalog # 6016949; Perkin-Elmer) or WST-1 assay System (Calbiochem) per manufacturer’s instructions. Fluorescence was measured on a Synergy H1 Hybrid plate reader (Biotek) and absorbance was read on a Vmax Kinetic Microplate Reader (Molecular Devices). All measurements were done in quadruplicate.

Clonogenic survival assay
U251 control and MARCKS knockdown cells grown to 80% confluency were trypsinized, counted with a hemacytometer, and serially diluted to defined concentrations. Three different defined numbers of cells were plated per dose of radiation in duplicate. Eight hours later, the cells were treated with 0, 3, 5, or 8 Gy radiation. Fourteen days later, cells were fixed and stained with 6.0% glutaraldehyde and 0.5% crystal violet. Colonies were counted in quadruplicate for each condition and a surviving fraction (S.F.) was calculated by using the equation (number of colonies formed/number of cells plated)/(number of colonies for sham irradiated group/number of cells plated). The results were then plotted as mean and SEM in a semilogarithmic format using Microsoft Excel software. Dose enhancement ratio is calculated as the dose (Gy) for control divided by the dose for MARCKS knockdown treated cells (normalized for plating efficiency at 0 Gy) for which a S.F. = 0.2 is achieved.

Clinical data
Clinical and gene expression data was obtained from the REMBRANDT database and TCGA Project. Gene expression values from the Affymetrix U133 Plus 2.0 Array Platform for MARCKS were downloaded for all glioma samples in the REMBRANDT database through the Advanced Search function. The clinical data were downloaded for these samples in BRB format through the Download function. For TCGA analysis, we used 192 of the 202 patients that did not have prior glioma. Gene expression data from 3 separate microarray platforms were summarized by gene across 3 different microarray platforms using a method described elsewhere (6). The gene-summarized gene expression values are contained in the Supplementary Materials in (Verhaak and colleagues, 2010; ref. 6) for 11,861 genes from the TCGA samples. Basic sample characteristics such as counts by subtype, MGMT status, and so on, are given elsewhere (6). A detailed description of the survival analysis is included in the Supplementary Methods.
Statistical analysis for in vitro data

The in vitro data were analyzed via ANOVA followed by a Bonferroni posttest using GraphPad Prism version 4.02 (GraphPad Software). Data are presented as average ± SEM.

Results

MARCKS regulates GBM growth rates in cell lines and in human xenografts

Previous studies have implicated a potential tumor suppressor role for the MARCKS protein (21). We thus hypothesized that GBM cell lines with higher levels of the MARCKS protein would have slower growth rates. To test our hypothesis, we measured MARCKS protein expression (normalized to GAPDH) in several well characterized immortalized GBM cell lines (Fig. 1A) showing a spectrum of proliferation rates (Supplementary Fig. S1). We found variable MARCKS expression levels across these cell lines and that MARCKS levels inversely correlated (R² = 0.4847) with GBM cell proliferation (Fig. 1B). Despite the extensive characterization of these cell lines, however, it has been realized that they harbor genetic alterations relative to the original tumor (27). We thus carried out a similar analysis with a more clinically relevant xenograft model.

The genetic makeup of the original tumor can be effectively preserved if the cells are maintained as xenolines in immunocompromised mice after direct implantation of patient tumor tissue (28). Human-derived GBM xenografts have been established by direct implantation of patient tumor tissue within immunocompromised mice and are serially passed within these mice as xenograft lines. It has been shown that these xenografts do not show substantial alteration of genetic changes as compared with the original tumor (28). For this reason, these xenografts exhibit a wide range of intracranial growth rates and common genetic alterations that are seen in GBM (Supplementary Table S1). We carried out similar measurements in the GBM xenografts (Fig. 1C) and correlated those levels with the average time of intracranial tumor growth for each tumor xenograft. This represents the average time from intracranial implantation to the time of passage (when mice show neurologic decline) and thus is an inverse surrogate measure for their tumor proliferation rates (Supplementary Table S1). As in the GBM cell lines, there was an inverse

![Figure 1](https://example.com/fig1.png)

Figure 1. Several GBM cell lines were grown under standard growth conditions and total protein lysates were prepared as described in the Methods and probed for MARCKS, phosphorylated Akt on serine 473 (pAkt Ser473), total Akt, and GAPDH (for protein normalization) by immunoblotting (A). Proliferation was determined for each cell line using a WST-1 cell proliferation assay (Supplementary Fig. S1) as described in Materials and Methods. Densitometric quantitation of MARCKS levels was carried out and normalized to GAPDH and relative levels of MARCKS were plotted vs. proliferation (Mean OD) with R² correlation (B). Human GBM xenografts were harvested from athymic nude mice and total protein lysates were prepared as described in Materials and Methods and probed for MARCKS, phosphorylated Akt on serine 473 (pAkt Ser473), total Akt, and GAPDH (for protein normalization) by immunoblotting (C). Densitometric quantitation of MARCKS levels was carried out and normalized to GAPDH and relative levels of MARCKS were plotted versus intracranial (IC) growth rate in days with R² correlation (D).
correlation between the tumor xenograft intracranial growth rates and MARCKS expression levels (Fig. 1D; \( R^2 = 0.4466 \)), suggesting that MARCKS is an important regulator of GBM growth.

**MARCKS silencing leads to increased growth rates and radiation resistance**

Our xenograft and cell culture data implicated MARCKS as a potential regulator of GBM growth. Therefore, we manipulated MARCKS expression in GBM cell lines to determine whether MARCKS directly affected GBM growth. We investigated MARCKS gene silencing using lentiviral mediated MARCKS shRNA knockdown in U251 cells (high basal MARCKS expression). A control cell line was generated using an empty vector control lentivirus. A stable 40% knockdown of the MARCKS protein in U251 cells (Fig. 2A) showed a 26% increase in proliferation rate (\( P < 0.01 \); Fig. 2B). Because MARCKS can sequester PIP2, we suspected that knockdown of MARCKS would be associated with increased signaling through PI3K/Akt due to increased availability of PIP2 for conversion to PIP3 by PI3K. Akt is activated by phosphorylation of serine 473 after recruitment to the membrane by PIP3. We thus assessed our MARCKS knockdown for activation of Akt by immunoblotting for phosphoserine-473. Decreases in U251 MARCKS levels produced increased activation of Akt compared with control (Fig. 2A). To show that signaling through the PI3K/Akt pathway was responsible for these increases in proliferation, we treated the MARCKS knockdown cells with the PI3K inhibitor LY294002 at 10 \( \mu \)mol/L. Treatment with LY294002 led to an 87% decreased activation of Akt as assessed by densitometry (Supplementary Fig. S2A) and subsequent decreases in proliferation (Supplementary Fig. S2B; \( P = 0.0287 \)). This suggested that decreased levels of MARCKS promote increased proliferation through enhanced signaling through PI3K/Akt.

Increased signaling through Akt has been implicated in therapeutic resistance due to increased DNA repair and decreased apoptosis (29, 30). We thus hypothesized that MARCKS knockdown would lead to increased radiation resistance. We assessed the radiation sensitivity of our knockdown cell line by a clonogenic assay and found a significant increase in radiation resistance with lower levels of MARCKS compared with control (dose enhancement ratio = 0.75; 3 Gy \( P < 0.0001 \), 5 Gy \( P = 0.0002 \), 8 Gy \( P < 0.0001 \); Fig. 2C and Supplementary Fig. S2C). Radiation induces cell death by multiple mechanisms such as apoptosis, autophagy, mitotic catastrophe, and senescence. It is thought that cell death occurs due to failure of the cell to adequately repair the DNA damage induced by irradiation. Activation of PI3K/Akt has been shown to lead to decreased apoptosis as well as increased DNA repair by a nonhomologous end joining (NHEJ) mechanism (31). We therefore investigated how knockdown of the MARCKS protein might affect these forms of cell death.

To investigate the mechanism of DNA repair, we quantified DNA double strand breaks (DSB) over time after radiation. The DSB is the most lethal lesion inflicted by radiation. Upon sensing a DSB, the cell will phosphorylate histone H2A on serine 139 (\( \gamma H2AX \)). Thus, staining for \( \gamma H2AX \) foci as a marker for DNA DSB over time is a common
method of investigating the rate of DNA repair. We noted a significantly decreased presence of DNA DSBs over time after radiation in our knockdown cell line \((P < 0.001\) at 30 minutes, 1, 4, and 8 hours; Fig. 3A). To show that this effect could be due to increased DNA repair by NHEJ, we immunoblotted for phosphorylated DNA-dependent protein kinase, catalytic subunit (DNA-PKcs) in our control and knockdown cell lines after irradiation. We found that MARCKS knockdown lead to a 4-fold increase in peak DNA-PKcs activation after 8 Gy as assessed by densitometry (Fig. 3B). Treatment of the MARCKS knockdown cell line with LY294002 at 10 \(\mu\text{mol/L}\) for 1 hour before radiation abrogated activation of DNA-PKcs, suggesting that this increase in DNA repair is mediated through PI3K/Akt

Figure 3. Quantification of \(\gamma\text{H2AX}\) foci was carried out at the indicated time points after 8 Gy radiation \(***, P < 0.001; \text{A})\). DNA repair by NHEJ was assessed by immunoblotting for phosphorylated DNA-PK at the indicated time points after treatment with 8 Gy (B). The distribution of the cell cycle at 8 and 24 hours after 8 Gy was assessed by propidium iodide staining and flow cytometry. Representative histograms for the control and MARCKS knockdown cell line at each time point are shown (C). Mean values for each phase of the cell cycle at each time point and \(P\) values for control versus MARCKS knockdown are available in Supplementary Fig. S3B and S3C.
signaling (Supplementary Fig. S3A). This increased DNA repair mechanism would suggest that manipulation of MARCKS levels would alter the various forms of cell death induced after irradiation. We therefore assessed the effect of MARCKS knockdown on cell cycle, mitotic catastrophe, autophagy, senescence, and apoptosis. Cell-cycle analysis was done in the control and knockdown cell line after 8 Gy. We found no difference in the basal cell-cycle distribution; however, knockdown of the MARCKS protein lead to a more rapid recovery toward the basal cell-cycle distribution at 24 hours after 8 Gy (Fig. 3C and Supplementary Fig. S3B and C). This was consistent with more rapid DNA repair and recovery from cell-cycle arrest. Mitotic catastrophe was assessed after 8 Gy at 24, 48, and 72 hours by 4’,6-diamidino-2-phenylindole (DAPI) staining and quantification of the number of multinucleated cells per field (40×; *, P < 0.05; A). An example of mitotic catastrophe is available in Supplementary Fig. S4A. Autophagy was quantified at 72 hours after 8 Gy radiation by immunoblotting for conversion of LC3-I to LC3-II with normalization to α-tubulin (B). Apoptosis was quantified after 8 Gy radiation by immunoblotting for cleaved caspase 3 at 12 and 24 hours (C), and by quantification of pyknotic nuclei per field (40×) at 24, 48, and 72 hours (*, P < 0.05; D). An example of a pyknotic nucleus is available in Supplementary Fig. S4B.

**MARCKS overexpression leads to decreased growth rates and senescence**

Because a reduction in MARCKS levels led to increased growth and resistance to radiation, we hypothesized that MARCKS overexpression would lead to decreased growth and increased sensitivity to radiation. MARCKS overexpression was achieved in U87 GBM cells (low basal MARCKS expression) using lentiviral infection, with an empty vector serving as a control. Expectedly, 6.3-fold overexpression of the MARCKS protein within U87 GBM cells (Fig. 5A) showed a 52% decrease in proliferation rate compared with control (P = 0.0003; Fig. 5B). We attempted to generate a stable overexpressing cell line, but we were unable to do so as these cells did not increase in confluence over a period of 21 days, although the control cell line was passaged 3 times during the same period. Because these cells did not proliferate, the effect on radiation sensitivity due to MARCKS overexpression could not be assessed.
We noted from imaging that the MARCKS overexpressing cells were adopting a flattened morphology and had a 50-fold increase in size compared with the control cell line (\( P < 0.0001 \); Fig. 5C, D, and E). These morphologic changes are consistent with senescence, a state in which normal diploid cells lose their ability to divide (34, 35). The MARCKS overexpressing U87 cells were stained for SA-\( \beta \)-gal to confirm senescence (Fig. 5C and D). The percent of cells staining positive for SA-\( \beta \)-gal was quantified by counting the percent of cells staining positive for SA-\( \beta \)-gal at \( 10 \times \) magnification (\( ***, P < 0.0001 \); F).

Because SA-\( \beta \)-gal is a relatively insensitive marker of senescence, we also quantified the percent of cells showing the morphologic changes consistent with senescence. A total of 200 cells were imaged and 79.5% of the MARCKS overexpressing cells showed these changes compared with 1% of the control cells. We also noted the presence of multinucleated giant cells, which are morphologically similar to those found in a previously identified giant cell GBM subtype (ref. 36; central panel, Fig. 5D). The multinucleated giant cells in giant cell GBM are halted in the early phases of mitosis and lack proliferative capacity (37). These results confirmed MARCKS as a regulator of GBM growth and suggested a novel role in regulating senescence.
MARCKS expression is a favorable prognostic factor for glioma patients

In light of our cell culture and xenograft model data, we anticipated that overexpression of MARCKS would be a favorable characteristic for survival. Thus we sought to evaluate MARCKS expression more closely in terms of clinical outcomes. We thus analyzed MARCKS gene expression in all glioma samples (low grade and high grade) within the REMBRANDT database for survival probability. As shown in Fig. 6A, we compared the survival of glioma patients with MARCKS expression above and below the median expression and found a statistically significant survival benefit for those patients with high MARCKS expression (HR, 0.55; 95% CI, 0.43–0.69; \( P < 0.001 \) by Cox model). This suggested that upregulation of MARCKS is a favorable prognostic factor for patients with glioma.

To confirm and expand the REMBRANDT data, we carried out similar analyses using the TCGA database that was used to define GBM molecular subtypes (6). We analyzed MARCKS gene expression within the TCGA dataset of 192 GBM patients that had no prior glioma with defined molecular subtype information. We fit a Cox proportional hazards model as a function of age and MARCKS expression and discovered a protective influence of increased MARCKS expression on survival outcome (\( P = 0.044 \), Supplementary Table S2). We also carried out survival analyses for each subtype (classical, mesenchymal, neural, and proneural) and found that only the proneural subtype retained statistical significance when age adjusted (\( P = 0.003594 \); Supplementary Table S2). This analysis showed that the influence of MARCKS expression on survival outcome shows high subtype specificity with very strong effect in the proneural subtype and almost no effect in the other subtypes, thus the full sample analysis effectively showed a dilution of the effect in the proneural subtype. To observe the linearity of the effect of MARCKS gene expression on survival within proneural individuals, we carried out survival regression in which we categorized MARCKS gene expression within proneural individuals.
expression by quartiles (4 levels; Supplementary Table S2). The quartile-based analysis showed the effect of MARCKS to be highly nonlinear, with the fourth quartile (highest MARCKS expression) showing a strong protective influence compared with other quartiles (HR, 0.28, \( P = 0.01 \)). In further analysis, we chose to dichotomize MARCKS at the fourth quartile for maximum discrimination (median survival of 47.2 months vs. 12.2 months, HR, 0.21; 95% CI, 0.09–0.48; \( P < 0.0002 \)) as shown in Fig. 6B.

**MGMT** methylation status and age are the 2 most common factors in determining prognosis for GBM. We therefore carried out survival analysis after adjusting for both **MGMT** status and age using the MARCKS quartile expression data (Fig. 6C and Supplementary Table S2). The **MGMT** term did not show significance (HR, 1.48, \( P = 0.24 \)) as a main effect, but we suspected a possible interaction with MARCKS. We then stratified by **MGMT** status (methylated and unmethylated) and found that MARCKS protection was highly significant in the unmethylated individuals (median survival of 65.3 months vs. 10.7 months, HR, 0.11; 95% CI, 0.03–0.39; \( P < 0.001 \)), but not significant in the methylated individuals (HR, 0.39, \( P = 0.19 \)). This indicated that the patients that benefit to the highest degree from elevated MARCKS expression in GBM are those of the proneural subtype with unmethylated **MGMT** status.

**Discussion**

Our analysis of survival outcomes in the REMBRANDT and TCGA databases with respect to MARCKS expression in all samples indicated a direct correlation with patient survival (Fig. 6A and B, Supplementary Table S2). However, we suspect that the importance of MARCKS may ultimately depend on the particular cell signaling cascades that are active within the tumor. This is consistent with our data showing molecular subtype specificity in the TCGA analysis. Indeed, when we looked at MARCKS impact on survival based on molecular subtype, only the proneural subtype retained significance (Supplementary Table S2). An even more striking result is observed when MARCKS expression is broken down into quartiles. Within the proneural subtype, the highest quartile (fourth quartile: highest MARCKS levels) shows a significant survival benefit compared with the lower 3 quartiles; with 50% of patients surviving 3 years or more (Fig. 6B). Surprisingly, the importance of MARCKS was particularly evident in unmethylated **MGMT** tumors of the proneural subtype. Patients with unmethylated **MGMT** are expected to suffer from especially poor outcomes. However, we noted that unmethylated **MGMT** tumors with high MARCKS expression had a much better prognosis than those with low **MGMT** expression (Fig. 6C). Therefore, MARCKS expression has protective influence and prognostic value above and beyond the 2 most commonly used predictive factors for survival, age, and **MGMT** status (HR, 0.18, \( P = 0.001 \)) for those patients with GBM tumors of the proneural subtype. However, the mode of action may be complex, as the protective influence seems to interact with **MGMT** status. Our results predict that individuals with unmethylated **MGMT**, which are considered of poorer prognostic status due to their decreased sensitivity to therapy, benefit to a much greater extent from higher MARCKS expression than their methylated counterparts. To our knowledge, this is the first prognostic factor for individuals with tumors with unmethylated **MGMT** that would outperform the methylated group in terms of survival.

Our results suggest that this survival benefit may be in part due to the ability of the MARCKS protein to regulate proliferation and radiation sensitivity. This likely occurs through modulation of PIP2-dependent signaling pathways such as cell division and the PI3K/Akt pathway, as well as a potential role in regulating senescence. PI3K mutations are a common occurrence in the proneural subtype and thus MARCKS expression would have a direct impact on signaling through this pathway in these tumors (6). PDGFR mutations are also frequent, and MARCKS expression could modulate this pathway by controlling the availability of PIP2 to cleavage by phospholipase C or conversion to PIP3 by PI3K. A link between MARCKS and other mutations such as IDH1 and TP53 that are characteristic of this subtype is less clear.

It should be noted that MARCKS expression did not have an impact on survival in the classical subtype (Supplementary Table S2), for which EGFR mutations are relatively common and aberrant phospholipid signaling would be expected. A potential explanation for this might be the increased phosphorylation of MARCKS that was observed in EGFRVIII expressing cells that showed changes in invasion (22). In these cells a greater proportion of MARCKS is phosphorylated and thus unable to sequester PIP2 from participating in these signaling cascades. In the context of this cellular background, high MARCKS expression may have a reduced impact on modulating these pathways. It is also interesting that the U87 cell line enters a senescent state with overexpression of MARCKS, but U251 cells possess high levels of MARCKS and continue to proliferate. Senescence is known to be dependent on p53 and pRB DNA damage signaling pathways. U251 and U87 differ with respect to p53 status (U251 is mutant, U87 is wild type; ref. 38), so it is also likely that the differing cell signaling backgrounds are responsible for this effect. This supports the concept that the effect of high MARCKS expression on clinical outcome would be highly dependent on the particular cell signaling background.

Our results have also identified a potential novel role for the MARCKS protein in promoting senescence and the formation of multinucleated giant cells. The large, flattened, multinucleated giant cells (central panel, Fig. 5D) found upon MARCKS overexpression are morphologically similar to the giant cell GBM subtype (36). This giant cell subtype of GBM strikes younger patients, has a better prognosis, and genetically resembles secondary disease (i.e., high incidence of TP53 mutations and a low incidence of **EGFR** aberrations; refs. 9, 39, 40), similar to the recently identified proneural subtype (6). It has been proposed that the formation of these multinucleated giant cells is the product of abnormal centrosome duplication resulting from the high incidence.
of TP53 mutations that is characteristic of the giant cell GBM subtype (39, 40). As mentioned above, the wild-type p53 status of U87 cells (38) suggests that high levels of the MARCKS protein may interfere with the function of the p53 protein leading to the formation of multinucleated giant cells. However, the formation of these cells may also be a result of the role of the MARCKS protein in regulating the actin cytoskeleton, as it has been previously shown that experimental sequestration of PIP2 leads to defects in cytokinesis and the formation of multinucleated cells (41, 42). We also noted that these giant cells were in a senescent state, indicated by positive staining for SA-β-gal (Fig. 5C and D). High levels of MARCKS have been previously associated with a quiescent state in fibroblasts (43), however, to our knowledge this is the first evidence that the MARCKS protein can induce a senescent state. The induction of such a state in GBM has obvious clinical implications.

To our surprise, the increases in PI3K/Akt signaling following MARCKS knockdown in U251 were not mirrored by MARCKS overexpression in U87 as we found no commensurate decrease in Akt activation. This is likely the result of changes in signaling that occurs with senescence. It has been previously shown that Akt activity increases with adoption of the senescent state (44). Thus, any decreases in signaling through PI3K/Akt from MARCK overexpression may be eliminated by a compensatory upregulation of Akt activity. These data suggest that alternative cell signaling pathways, regulated by the MARCKS protein, should not be excluded and warrant further investigation. For example, as a major substrate for PKC, higher levels of MARCKS might lead to decreased signaling through alternate PKC substrates. As a binding partner for Ca2+-calmodulin, higher levels of MARCKS might lead to decreased signaling through alternate calmodulin pathways. Thus, the impact that MARCKS expression has on these cell signaling pathways, and therefore patient survival, is likely due to a balance of the predominant cell signaling pathways that are characteristic of that particular disease background. As such, our results indicate that MARCKS is a novel biomarker and therapeutic target for a specific group of GBM patients, particularly those with proneural tumors with unmethylated MGMT promoters. Efforts are currently underway to identify therapies that increase MARCKS expression in the context of this cellular background, as this may provide a significant survival benefit for these patients.

Disclosure of Potential Conflicts of Interest
The authors declare that there are no conflicts of interest pertaining to the contents of this article. Nevertheless, occasional honoraria from Bristol-Myers Squibb, ImClone Systems, Inc. and Eli Lilly (J.A. Bonner) and Varian (C.D. Willey) are disclosed.

The content of this publication does not necessarily reflect the views or policies of the Department of Health and Human Services, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. Government.

Acknowledgments
The authors thank Didier Trono and Bob Weinberg for providing the psPAX2 and pCMV-VSV-G plasmids, respectively, via Addgene. Plasmid constructs were sequenced at the UAB Center for AIDS Research DNA Sequencing Core, supported by Core grant P30-017767. D54 cells were obtained from Harald Sontheimer (UAB). U87 and U251 were obtained from Tika Benveniste (UAB). U373, U118, and D52 were obtained from Donald Buchsbaum (UAB), American Society for Radiation Oncology Junior Faculty Research Training Award (to C.D. Willey); University of Alabama Health Services Foundation General Endowment Fund Scholar Award (to C.D. Willey); IMPACT Award from the Department of Radiation Oncology, University of Alabama at Birmingham Comprehensive Cancer Center, University of Alabama at Birmingham School of Medicine (to E.S. Yang).

Grant Support
This project has been funded in whole or in part with Federal funds from the National Cancer Institute, NIH, in the form of grant P50 CA 097247 (to C.D. Willey) and the associated UAB Brain SPORE pilot project program (to C.D. Willey).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received December 5, 2011; revised March 28, 2012; accepted March 29, 2012; published OnlineFirst May 22, 2012.

References
MARCKS Regulates GBM Growth and Is Prognostic in Patients


MARCKS Regulates Growth and Radiation Sensitivity and Is a Novel Prognostic Factor for Glioma


Updated version
Access the most recent version of this article at: doi:10.1158/1078-0432.CCR-11-3091

Supplementary Material
Access the most recent supplemental material at: http://clincancerres.aacrjournals.org/content/suppl/2012/04/05/1078-0432.CCR-11-3091.DC1

Cited articles
This article cites 46 articles, 24 of which you can access for free at: http://clincancerres.aacrjournals.org/content/18/11/3030.full.html#ref-list-1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.