Profiles of Genomic Instability in High-Grade Serous Ovarian Cancer Predict Treatment Outcome

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Abstract

Purposed: High-grade serous cancer (HGSC) is the most common cancer of the ovary and is characterized by chromosomal instability. Defects in homologous recombination repair (HRR) are associated with genomic instability in HGSC, and are exploited by therapy targeting DNA repair. Defective HRR causes uniparental deletions and loss of heterozygosity (LOH). Our purpose is to profile LOH in HGSC and correlate our findings to clinical outcome, and compare HGSC and high-grade breast cancers.

Experimental Design: We examined LOH and copy number changes using single nucleotide polymorphism array data from three HGSC cohorts and compared results to a cohort of high-grade breast cancers. The LOH profiles in HGSC were matched to chemotherapy resistance and progression-free survival (PFS).

Results: LOH-based clustering divided HGSC into two clusters. The major group displayed extensive LOH and was further divided into two subgroups. The second group contained remarkably less LOH. BRCA1 promoter methylation was associated with the major group. LOH clusters were reproducible when validated in two independent HGSC datasets. LOH burden in the major cluster of HGSC was similar to triple-negative, and distinct from other high-grade breast cancers. Our analysis revealed an LOH cluster with lower treatment resistance and a significant correlation between LOH burden and PFS.

Conclusions: Separating HGSC by LOH-based clustering produces remarkably stable subgroups in three different cohorts. Patients in the various LOH clusters differed with respect to chemotherapy resistance, and the extent of LOH correlated with PFS. LOH burden may indicate vulnerability to treatment targeting DNA repair, such as PARP1 inhibitors. Clin Cancer Res; 18(20); 5806–15. ©2012 AACR.
alleles. We previously used single nucleotide polymorphism (SNP) arrays to profile allelic imbalance (AI) in breast cancer, and found correlations with subtypes classified by expression of the estrogen receptor (ER), progesterone receptor (PR), and the HER2 receptor–like protein, and by histologic grade (6, 7). ER-positive or HER2-positive breast tumors may derive from luminal epithelial progenitors (luminal breast cancers), whereas tumors negative for ER, PR, and HER2 (triple-negative breast cancer, TNBC) may have a different histogenesis (8–10). TNBCs share features with HGSC. Both TNBC and HGSC frequently have p53 mutations and both display chromosomal instability and are observed in women inheriting a disease-associated mutation in BRCA1 (3, 9, 11–13). Platinum-containing drugs are very active in ovarian cancer, particularly in HGSC, and platinum drugs alone or in combination have been used successfully in TNBC (14–18). Finally, the genomes of sporadic HGSC and TNBC harbor common loss of single parental alleles, detected as LOH or AI, within or encompassing whole chromosomes (6, 7, 19, 20). We reasoned that the burden and pattern of AI may distinguish subtypes of HGSC, and genomic subtypes might predict response to initial treatment as well as overall survival. Analysis of AI in HGSC may also provide further insights into molecular mechanisms between these diseases. Our findings provide biomarkers of response to treatment in HGSC, which may aid treatment planning with therapies targeting DNA repair, such as PARP1 inhibitors.

Translational Relevance

High-grade serous cancer (HGSC) of the ovary is a highly lethal disease that is frequently characterized by chromosomal instability, defects in homologous recombination repair (HRR), and sensitivity to DNA-damaging agents, especially platinum-based therapies. However, not all HGSC show defects in HRR and currently the ability to predict response to first-line treatment is limited. We show HGSC can be reproducibly separated into groups based on the pattern of LOH, and multivariate analysis finds a subgroup with low chemotherapy resistance. The extent of LOH correlated with progression-free survival. Extensive LOH was also seen in triple-negative breast cancer (TNBC) relative to other breast cancer subtypes. Although some chromosomal regions affected by LOH differed between TNBC and HGSC, finding extensive LOH in both cancers suggests common molecular mechanisms between these diseases. Our findings provide biomarkers of response to treatment in HGSC, which may aid treatment planning with therapies targeting DNA repair, such as PARP1 inhibitors.

Materials and Methods

Subjects and tissues from Boston

Ovarian cancer tissue was collected at the Dana-Farber Cancer Institute and Brigham and Women’s Hospital (Boston, MA) along with treatment and outcome information. All tumors were serous, grade 2 or 3; grade 1 tumors were excluded. Forty-seven cases passed quality control and were analyzed (Supplementary Table S1). Fifty Bloom-Richardson grade 3 breast cancer cases were from a larger study of breast cancer, described elsewhere (21). All tissue samples and clinical information were identified and collected under a Dana-Farber/Harvard Cancer Center Institutional Review Board-approved protocol.

HGSC were considered platinum resistant if there was progressive disease during initial chemotherapy or evidence of progression within 6 months after completion of chemotherapy, and platinum sensitive if disease did not recur or progress within 6 months after completion of initial chemotherapy (15, 22, 23). All patients underwent debulking surgery before platinum- and taxane-based chemotherapy. Optimal debulking resulted in 1 cm or less of residual tumor and suboptimal debulking left more than 1 cm; 23 patients had optimal debulking surgery. Most cases (73%) were stage III, 18% stage IV, and 9% stage II (Supplementary Table S1).

Enrichment of tumor cells and DNA preparation

Ovarian and breast tumor cells were enriched by needle microdissection to remove stromal components from 7 μm frozen sections lightly stained with hematoxylin and eosin. Tumor cells were isolated from ascites by affinity enrichment with anti–EPCAM-conjugated beads as described (24). The preparation of DNA and RNA was described previously (6).

SNP and gene expression array analysis

Genomic DNA digested with NspI was hybridized to Affymetrix GeneChip Human Mapping 250K NspI arrays using recommended procedures (Affymetrix, Inc.). The data have been submitted to and will be posted in GEO (GSE39130). 10K SNP data from 50 breast cancers have been submitted to and will be posted in GEO (GSE19594). LOH was determined by comparing genotype calls (heterozygous or homozygous) from tumor and matched normal cells in blood or morphologically normal breast tissues. For tumors without a paired normal sample from the same patient, LOH was defined by an LOH prediction algorithm in dChip software (25, 26). We compared patterns and overall frequency of LOH derived from 10K and 250K SNP arrays and the results were similar (Supplementary Fig. S1).

LOH-based hierarchical clustering was done based on significant LOH patterns using algorithms running on

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dChip software as described (27). Significantly frequent LOH regions were identified by a permutation test carried out across the entire cohort (permutation \( P < 0.05 \)). These significant regions were used for hierarchical clustering. The distance between any 2 individuals was defined as the proportion of discordance among significantly frequent LOH regions. The average-linkage algorithm is divisive and merges samples into clusters, based on similarity of LOH patterns (27). Differences between 2 adjacent specimens are represented by the length of the vertical lines in the clustering dendrogram.

To estimate and minimize the effect of stromal contamination, we used major copy proportion (MCP) analysis to measure AI, an alternative allele copy ratio–based approach for LOH analysis (28). MCP is the ratio of the copy number of the major allele divided by the copy number for both major and minor alleles. The MCP scores for heterozygous and homozygous loci are 0.5 and 1.0, respectively. In a homogeneous tumor sample, an MCP score of 1.0 at a heterozygous locus represents AI with complete allelic loss of one parental copy (LOH). MCP scores will shift down from 1.0 when normal heterozygous alleles contaminate the tumor samples (28). More than 20% contamination shifts MCP scores to less than 0.7 in regions of AI that possess an absolute copy number decrease. Such samples were excluded. Samples containing regions of AI with absolute copy number decrease and MCP scores of 0.7 or more were considered acceptable and included in this study.

To find the correlation coefficient between FLOH levels and resistance rates, and \( \chi^2 \) was used to evaluate the difference among groups of HGSC. Logistic regression was used for multivariate analysis. Kaplan–Meier analysis was used to evaluate clinical outcome after chemotherapy. The difference between Kaplan–Meier curves was evaluated by the log-rank test.

Results

Genomic profiling HGSCs of ovary

We conducted genome-wide LOH analysis in 47 HGSC specimens from Boston. Heterogeneity of LOH patterns in HGSC was observed, and 2 top clusters were identified and separated using LOH-based hierarchical clustering as described in Materials and Methods (Fig. 1A). A major cluster of 29 cases (62%) harbored massive intra- and whole-chromosomal LOH across each tumor genome (cluster Hi; Fig. 1A). This major cluster harbored a mean FLOH of 36.3% and contrasted with a minor cluster with a low frequency of LOH events (cluster Lo, mean FLOH = 12.8%; Fig. 1A and B). Hierarchical clustering further separated the major cluster Hi into 2 subclusters (HiA and HiB; Fig. 1A). The subclusters of the Hi cluster showed similar levels of LOH (mean FLOH = 37.9% and 32.7%; Fig. 1B), but differed in the regions of LOH involved. Loss or near loss of 1 parental copy of the acrocentric chromosome 13 was seen almost exclusively in subcluster HiA, and LOH of 5q and 17p was more frequent in subcluster HiA (Fig. 1A and Supplementary Table S4).
Nonmalignant cells could interfere with proper genotype calls from tumor-derived DNA. All cases were microdissected and the proportion of tumor cells to normal cells was estimated to equal or exceed 90%. To minimize the influence of normal contamination that could contribute to the appearance of cluster Lo, with low FLOH, we calculated AI using major copy proportion (MCP; ref. 28). The number of genomic regions with AI were significantly lower in cluster Lo, and the overall pattern was consistent with the presence of distinctive groups of HGSC defined by LOH profiles (Fig. 1C and Supplementary Fig. S2).

We compared the genomic patterns of HGSC with high-grade breast cancers by conducting LOH-based clustering in a group of 50 breast tumors. These cancers contained 15 high-grade ER-positive, 10 HER2-positive, 20 sporadic TNBC, and 5 additional cases from women with inherited BRCA1 mutations. One major cluster was dominated by TNBC and BRCA1-associated tumors. A second cluster was dominated by HER2-positive and high-grade ER-positive cancers (Supplementary Fig. S3A). Sporadic TNBC and BRCA1-associated breast cancers possessed a significantly higher median FLOH ($P < 0.0001$) and number of AI regions per genome ($P < 0.0002$) compared with HER2-positive and high-grade ER-positive tumors (Supplementary Fig. S3B and S3C). Therefore, HGSC and TNBC share high levels of LOH.

**LOH patterns and copy number alterations**

We examined qualitative patterns of LOH and copy number gains and losses in LOH-defined clusters of HGSC. In HGSC-Hi, regions of common LOH reached a prevalence of $\geq 50\%$ in 15 chromosomes. Uniparental loss of the entire, or nearly the entire chromosome 17 was seen in all but one case of HGSC-Hi and less common in HGSC-Lo.

Permutation testing defined a threshold level of significance ($P \leq 0.05$) for LOH regions on chromosomes 4q, 5q, 6q, 13q, 17p, and 17q in HGSC-Hi (Fig. 2A). Similar to allelic loss, copy number loss was a prominent feature of HGSC-Hi, and less common in HGSC-Lo (Fig. 2B). In contrast to the allelic and copy loss, copy gains were common and remarkably similar in both cluster Hi and cluster Lo of HGSC (Fig. 2B). Allelic loss may be either an absolute loss of DNA content, or copy neutral loss of a parental allele. In HGSC-Hi, allelic loss was approximately equally divided between hemizygosity and isodisomy. Loss, particularly allelic loss, seems to define subclusters of HGSC.

In TNBC, regions of common LOH reached a prevalence of 50% on 15 chromosomes; however, permutation testing with a threshold of $P \leq 0.05$ revealed significant LOH on chromosomes 5q, 11q, 14q, 17p, and 17q. In high-grade ER-positive and HER2-positive breast cancer, significant regions of LOH were seen only on chromosomes 8p and 17p, and patterns were different than observed in TNBC. When comparing HGSC-Hi to TNBC, with the exception of chromosome 17 and 5q, patterns of significant LOH regions were quite different (compare Fig. 2A to Supplementary Fig. S4). Copy number gains were commonly seen in TNBC and high-grade luminal breast cancers (ER-positive and HER2-positive; Supplementary Fig. S4).

**BRCA1 promoter methylation**

BRCA1 methylation was detected in 9 of 44 HGSC (20%) and 3 of 18 sporadic TNBC tumors (17%). More tumors in HGSC-Hi than HGSC-Lo possessed methylated BRCA1 promoter regions (30.7% vs. 5.5%, respectively, $P < 0.042$; Fig. 3A). FLOH was higher and BRCA1 transcript levels were lower in HGSC tumors with BRCA1 promoter methylation than in those without this event ($P < 0.019$ and
These results are consistent with the notion that BRCA1 promoter methylation suppresses BRCA1 expression and contributes to chromosomal instability.

Validation of LOH-based clustering in independent datasets of HGSC

LOH-based hierarchical clustering of ovarian cancer was validated using datasets from the AOCS and TCGA. Normal stromal cells in tumors from the AOCS were removed by needle microdissection, as was done with the Boston cases. The TCGA required tumors have at least 80% tumor cells in blocks used for analysis. Hierarchical clustering based on LOH separated 85 cases from the AOCS into 2 clusters: a minor subset (Cluster Lo) of 18 tumors with relatively lower LOH levels (mean FLOH = 5.1%) and a major cluster (Cluster Hi) with 67 tumors (Fig. 4A). As observed in the Boston dataset, cluster Hi could be divided into 2 subclusters (HiA and HiB), which had mean FLOH levels of 31.5% and 21.6%, respectively.

Clustering 116 HGSC in the TCGA produced a similar topology of subclusters (Fig. 4B). Again, subcluster HiA was associated with loss of chromosome 13q, whereas subcluster HiB retained both parental copies of 13q in both the AOCS and TCGA samples (Fig. 4A-B and Supplementary Table S4). Loss of one parental copy of chromosome 17 was a striking feature of tumors in Cluster Hi from both the AOCS and TCGA, and was less common in Cluster Lo tumors. The 3 major clusters (Cluster Lo, and subclusters HiA and HiB) were remarkably reproducible in all 3 cohorts studied.

Tothill and colleagues described 4 gene expression-based molecular subtypes of HGCS (4). These subtypes were reproduced in data from the TCGA (data not shown). We found a trend toward the association of cases in molecular subtype C1 with our LOH-based cluster HGCS-Lo in the AOCS cohort (P = 0.063), but not in the TCGA (data not shown).

LOH and impact on treatment resistance

To explore the relationship between the LOH-defined subclusters of HGSC and clinical outcome, we compared chemotherapy-resistant rates for patients pooled from all 3 cohorts. The frequency of platinum-resistant disease (progression or recurrence within 6 months after completion of primary platinum-based chemotherapy) was different for each subcluster (Lo, HiA, and HiB). In patients with stage III disease and optimal debulking, the HiA subcluster, which had the highest FLOH, had the lowest resistance rate compared with the 2 other subclusters (Fig. 5A and B and
Supplementary Table S5A). The Lo cluster, which harbored tumors with the lowest FLOH, had the highest chemotherapy-resistance rates. These trends generally remained same when data from all patients irrespective of stage and debulking status were analyzed (Fig. 5C and D and Supplementary Table S5B). When the 3 cohorts were analyzed individually, HiA had the lowest resistance rates and cluster Lo had the highest rates of chemotherapy resistance (Supplementary Fig. S5A and S5B). Resistance rates were inversely correlated to the mean FLOH of subclusters when the results from the 3 cohorts were plotted together (Pearson correlation, coefficient = −0.6, P < 0.041; Supplementary Fig. S5C).

Because at least 20% of HGSC harbor either germline or somatic mutations in BRCA1/2 (3, 33), we questioned whether BRCA1 and BRCA2 germline mutations account for the better initial response to chemotherapy in patients within the HiA group. Data were available for a subset of 39 AOCS patients, including 10 with either BRCA1 or BRCA2 germline mutations. Tumors from BRAC1/2 mutation carriers were concentrated in subclusters HiA and HiB (Supplementary Table S6). Tumors from mutation carriers had a significantly higher FLOH than tumors in HGSC-Lo and HiB from noncarriers, but were similar to tumors from noncarriers in subcluster HiA (Supplementary Fig. S6A). After removing the mutation carriers, the patients with tumors in the HiA subcluster still had low rates of chemotherapy resistance (Supplementary Fig. S6B). The same was true for the HiA subcluster when we combined AOCS and TCGA patients (Supplementary Fig. S6C). In the combined data, there was a tendency for tumors with BRCA mutations to have a lower resistance rate to initial chemotherapy, but this tendency did not reach statistical significance. HiA was further tested against other covariates, including BRCA gene status and FLOH, in a multivariate analysis with platinum therapy resistance (Supplementary Fig. S6B). The same was true for the HiA subcluster when we combined AOCS and TCGA datasets, although the differences were not statistically significant (Supplementary Table S8).

We evaluated LOH burden in tumors, irrespective of the subclusters and clinical outcome for patients with stage III HGSC and residual disease of 1 cm or less in the Boston and AOCS cohorts. Patients were separated by quartiles of FLOH in their tumors. HGSC patients in the fourth quartile with the highest FLOH had longer PFS; however, the relationships were nonmonotonic. Except for patients with early disease progression, patients in the first quartile with the lowest FLOH also enjoyed a longer PFS; whereas patients whose tumors were in the second quartile with intermediate FLOH had the shortest PFS (Fig. 6A and B). Similar trends were seen when the cases with stage II and IV disease and suboptimal debulking were included (Supplementary Fig. S7A and S7B). The relationship between FLOH quartiles and PFS was not clearly seen in TCGA, which may be due to interinstitutional variation in PFS across TCGA. After removing BRCA1/2-mutation carriers from the AOCS...
dataset, the same nonmonotonic relationship between tumor FLOH and patient’s PFS remained (Supplementary Fig. S8). The PFS for BRCA1/2 mutation carriers was similar to noncarriers with tumors in the highest FLOH quartile. Ovarian cancer with germline BRCA1/2 mutations have a higher mean FLOH level, and are known to be platinum sensitive. In this analysis, evidence emerges that high FLOH without BRCA1/2 mutation may also mark tumors more sensitive to platinum-based chemotherapy and with better outcomes.

Discussion

HGSC is the most common subtype of epithelial ovarian cancer and has distinct molecular features, including p53 mutations, genetic instability, and mutations in BRCA1 or BRCA2 (1–3, 11). This study identified remarkable genomic heterogeneity in HGSC otherwise considered histologically homogeneous. Two dominant groups of HGSC were uncovered by hierarchical clustering based on common regions of LOH; the major group possessed high-level LOH/AI (HGSC-Hi) and the minor group had lower levels of LOH/AI (HGSC-Lo). HGSC-Hi was further separated into 2 subclusters (HiA and HiB) based on specific regions of chromosomal loss. The HiA, HiB, and Lo pattern was nearly identical in 3 independent studies of HGSC (Figs. 1A and 4). To compare ovarian and breast cancer, we selected a cohort of exclusively high-grade breast cancers containing HER2-positive, high-grade ER-positive, and TNBC. HGSC-Hi and TNBC harbor similar amounts of copy loss and allelic loss, and more than seen in HGSC-Lo and in other breast cancer subtypes. These findings imply shared mechanism(s) of failed DNA repair have a similar overall impact on HGSC and TNBC genomes. However, cell lineage–specific factors may select for specific regions of LOH in the 2 different cancers. The concept of “BRCAness” has been advanced to describe the phenotypic and molecular relatedness of certain breast and ovarian cancers to tumors without germline inactivation of BRCA1 and BRCA2.
BRCA1 with methylation of the responses to chemotherapy and overall survival, patients somatic or germline mutations in mechanisms of FLOH. TCGA compared clinical outcome with different genes may contribute to genomic instability marked by high BRCA1 mutations, defects in other DNA repair–associated significant (Supplementary Fig. S9). In the cases without mutations in TCGA, but this relationship was not transcripts and high FLOH in tumors without BRCA1 in TCGA (data not shown). There was a trend toward low with high FLOH; however, this relation was not significant toward cohort, methylated BRCA1 promoter was associated with high FLOH; however, this relation was not significant in TCGA (data not shown). There was a trend toward low BRCA1 transcripts and high FLOH in tumors without BRCA1 mutations in TCGA, but this relationship was not significant (Supplementary Fig. S9). In the cases without BRCA1 mutations, defects in other DNA repair–associated genes may contribute to genomic instability marked by high FLOH. TCGA compared clinical outcome with different mechanisms of BRCA1 inactivation. Although patients with somatic or germline mutations in BRCA1 had improved responses to chemotherapy and overall survival, patients with methylation of the BRCA1 promoter had similar survival characteristics to patients without obvious defects in the BRCA pathway (3). Tumor genomes are complex and methylation of the BRCA1 promoter may be part of global methylation that affects the expression of multiple genes and will have a complex impact on the tumor genome and treatment outcome.

Intrinsic defects in HR in sporadic HGSC and TNBC may force the execution of "error prone" pathways that repair double-strand DNA breaks, otherwise more faithfully repaired by HR. One of these error prone pathways is nonhomologous end joining (NHEJ). Stephens and colleagues sequenced both ends of randomly generated DNA fragments in breast cancer cell lines and tissue to detect somatic genetic recombination (37). Recombination was common in breast cancer, particularly within TNBC, and from tumors or cell lines containing germline BRCA1 mutations. Recombination was recognized by discordant sequences on either end of the DNA fragment spanning the breakpoint. The fusion junction commonly contained regions of microhomology and small insertions of non-templated DNA. These features are considered "signatures" of NHEJ and frequent in BRCA1-associated and TNBC (37, 38). A high burden of LOH within a tumor is a hallmark for this type of chromosomal aberration. Tumors in the HGSC-Hi cluster bear this hallmark of defective HR and these cancers may be forced into NHEJ and accumulate allelic loss.

HGSC-Lo, with a lower burden of LOH, may represent a distinct subset of HGSC with apparently intact DNA repair capability. Recent studies involving ex vivo DNA damage of tumor samples has shown that about one third of HGSC retain the ability to form RAD51 foci and therefore seem to have intact HR pathways (39). HGSC-Lo may represent those tumors with intact HR pathways, perhaps evolved from low-grade serous tumors carrying RAS, PI3K, or BRAF mutations (1, 2, 40), or the result of variants in which defects in HR are partially corrected or compensated for during tumor progression (41). Impaired HR may also sensitize HGSC to repair pathways that depend on the multifunctional enzyme poly (ADP-ribose) polymerase 1 (PARP1) (42, 43). Clinically, HGSC-Lo is a subgroup of ovarian cancer that may not be sensitive to PARP1 inhibitors, whereas the cancers in subcluster HiA may be more sensitive.

Membership in HiA was the only predictor of platinum-sensitivity surviving multivariate analysis that included BRCA status and FLOH. In addition, excluding BRCA mutation carriers and somatic BRCA mutations from analysis, tumors in subcluster HiA remain the most sensitive to platinum treatment. Allelic loss of chromosome 13 is a signature of membership in HiA. Expression of critical genes may be lost or reduced as a result losing one parental copy of chromosome 13. The endosome copper-transporters ATP7A and ATP7B are the 2 major proteins that pump
platinum out of cells and away from the nucleus (44). Genes for ATP7B and ATP7A are located on chromosomes 13 and X, respectively. Growing evidence suggests increased expression of ATP7B is particularly associated with platinum resistance in ovarian cancer (45). A nuclear excision repair gene ERCC5 on 13q33.1, critical for removing platinum-caused DNA adducts, is a prognostic biomarker in ovarian cancer (46). Both RB1 and BRCA2 are located on 13q, and may be affected by allelic loss of a parental copy of chromosome 13. It is possible that genes on chromosome 13q contribute to chemotherapy sensitivity in ovarian cancer, in addition to the burden of LOH and impaired DNA repair. Sensitivity to genotoxic chemotherapy is predicted by HiA class membership; however, outcomes such as PFS may depend on other factors, and may be more complicated.

This study found a nonmonotonic relationship between content of LOH and PFS, with shorter PFS in tumors with an intermediate content of LOH (Fig. 6 and Supplementary Fig. S6). A similar “J-shaped” nonmonotonic relationship was described between recurrence-free survival and quartiles of a gene expression signature reflecting chromosomal instability (CIN70) in 13 publically available cohorts of chemotherapy-treated patients. In that report, highest and intermediate CIN70 signatures were associated with the longest and shortest recurrence-free survival after adjuvant multidrug chemotherapy, respectively, in multiple cancer types including TNBC and ovarian cancer (47). We observed a paradoxical longer PFS for patients whose tumors contained the lowest quartile of FLOH, except for those having an early disease progression. This result may reflect the more indolent biology of tumors with a low burden of LOH, or in part because of the effects of taxanes used in combination with platinum.

Evidence suggests relative resistance to taxanes in breast and ovarian cancers with BRCA1 deficiency (48). Ovarian cancers with the higher CIN70 signatures were more taxane resistant in a clinical trial of taxane monotherapy (49). In metastatic breast cancer, taxane monotherapy was less effective in carriers of BRCA1 mutations compared with patients without mutations in this gene (50). It is possible that HGSC with less LOH, a lower CIN70 signature and more stable genomes may be more sensitive to taxanes. Increased sensitivity to taxanes may explain the longer PFS in our patients whose ovarian cancers harbored the lowest quartile of LOH. Patients with tumors that are BRCA1/2 mutation associated or those in HGSC-HiA (with the highest levels of LOH), may benefit by genotoxic chemotherapy, alone or with newer agents such as PARP inhibitors that target DNA repair.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

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References


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