Omental Adipose Tissue–Derived Stromal Cells Promote Vascularization and Growth of Endometrial Tumors

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Abstract

Purpose: Adipose tissue contains a population of tumor-tropic mesenchymal progenitors, termed adipose stromal cells (ASC), which engraft in neighboring tumors to form supportive tumor stroma. We hypothesized that intra-abdominal visceral adipose tissue may contain a uniquely tumor-promoting population of ASC to account for the relationship between excess visceral adipose tissue and mortality of intra-abdominal cancers.

Experimental Design: To investigate this, we isolated and characterized ASC from intra-abdominal omental adipose tissue (O-ASC) and characterized their effects on endometrial cancer progression as compared with subcutaneous adipose-derived mesenchymal stromal cells (SC-ASC), bone marrow–derived mesenchymal stromal cells (BM-MSC), and lung fibroblasts. To model chronic recruitment of ASC by tumors, cells were injected metronomically into mice bearing Hec1a xenografts.

Results: O-ASC expressed cell surface markers characteristic of BM-MSC and differentiated into mesenchymal lineages. Coculture with O-ASC increased endometrial cancer cell proliferation in vitro. Tumor tropism of O-ASC and SC-ASC for human Hec1a endometrial tumor xenografts was comparable, but O-ASC more potently promoted tumor growth. Compared with tumors in SC-ASC–injected mice, tumors in O-ASC–injected mice contained higher numbers of large tortuous desmin-positive blood vessels, which correlated with decreased central tumor necrosis and increased tumor cell proliferation. O-ASC exhibited enhanced motility as compared with SC-ASC in response to Hec1a-secreted factors.

Conclusions: Visceral adipose tissue contains a population of multipotent MSCs that promote endometrial tumor growth more potently than MSCs from subcutaneous adipose tissue. We propose that O-ASCs recruited to tumors express specific factors that enhance tumor vascularization, promoting survival and proliferation of tumor cells. Clin Cancer Res; 18(3); 771–82. ©2011 AACR.

Introduction

Endometrial cancer is the most common gynecologic malignancy in the United States and is strongly linked with obesity (1, 2). Women with a body mass index (BMI, calculated as kg/m²) greater than 40 have a greater than 6-fold relative risk of death from uterine cancer than lean women (2, 3). Furthermore, estimates suggest that 39% of cases of endometrial cancer can be attributed to obesity (4). The relationship between obesity and endometrial cancer has generally been attributed to estrogen and other paracrine factors secreted by adipose tissue (5).

However, a more direct relationship between adipose tissue and malignant tumors is suggested by recent studies showing that adipose tissue contains a population of mesenchymal progenitor cells that migrate into tumors and can facilitate tumor progression (6). These adipose stromal cells (ASC) have many features of bone marrow–derived mesenchymal stromal cells (BM-MSC), including cell surface marker expression, plastic adherence, and the capacity to differentiate into cells of mesenchymal lineage: osteoblasts, chondrocytes, and adipocytes (7, 8). Accumulating evidence indicates that cancer-associated fibroblasts (CAF), which provide a trophic tumor microenvironment, are, at least to some extent, derived from MSCs (9–13). MSCs have been reported to impact tumor progression, increasing tumor initiation and promoting metastasis in some models (14–19). This evidence suggests that the strong link between...
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SC-ASC from a cancer-free female donor (BMI stably transfected using Lipofectamine, with pEF-Luc. 
lin–streptomycin. Hec1a and WI38 cells were obtained
assays.

ex vivo (SC-ASC), and control fibroblasts by using 
bone marrow–derived MSC (BM-MSC), subcutaneous ASC 
of omental-derived ASC (O-ASC) was then compared with 
isolated a population of ASCs from the omentum, the major 
tissue could explain this relationship. To investigate this, we 
isolated and characterized adipose stromal cells (ASC) from the omentum of two patients with gynecologic cancer (O-ASC). Using an 
endometrial cancer xenograft model, we compared the 
effects of O-ASC and subcutaneous ASC (SC-ASC) on cancer progression. O-ASC more potently facilitated 
tumor growth and enhanced tumor vascularization. Our 
data indicate that O-ASC and SC-ASC are equally effi- 
cient in homing to tumors but are markedly different in 
their function. These findings suggest that excess visceral 
adipose tissue promotes malignancy by providing mes-
enchymal stromal cells to intra-abdominal tumors and thus may represent a new therapeutic target.

Translational Relevance
Both incidence and progression of endometrial cancer are positively associated with obesity. Preferential dis-
tribution of adipose tissue in the abdomen further 
increases the risk of endometrial cancer, as well as the 
risk and mortality of other intra-abdominal cancers. We 
 hypothesized that visceral adipose tissue contains a 
population of tumor-promoting stromal progenitors to 
account for the link between visceral obesity and malign-
nancy. To investigate this, we isolated and characterized adipose stromal cells (ASC) from the omentum of two patients with gynecologic cancer (O-ASC). Using an 
endometrial cancer xenograft model, we compared the 
effects of O-ASC and subcutaneous ASC (SC-ASC) on cancer progression. O-ASC more potently facilitated 
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cient in homing to tumors but are markedly different in 
their function. These findings suggest that excess visceral 
adipose tissue promotes malignancy by providing mes-
enchymal stromal cells to intra-abdominal tumors and thus may represent a new therapeutic target.

Materials and Methods

Cell culture
Cells were grown in α-minimum essential medium 
(α-MEM) containing 20% FBS, l-glutamine, and penicil-
lin–streptomycin. Hec1a and WI38 cells were obtained 
from American Type Culture Collection. Hec1a cells were 
stably transfected using Lipofectamine, with pEF-Luc. 
SC-ASC from a cancer-free female donor (BMI = 33.0) were 
reported previously (22). BM-MSCs were isolated from 
normal individuals undergoing bone marrow harvest for 
allogenic transplantation following informed consent, 
according to institutional guidelines under the approved 
protocol, as described previously (23). Briefly, mononuclear 
cells were separated by centrifugation over a Ficoll-Hypaque 
gravity (Sigma-Aldrich), suspended in α-MEM/10% FBS, 
and plated on 180 cm² dishes. After 3 days, nonadherent 
cells were removed by washing with PBS, and monolayers of 
adherent cells were cultured until they reached confluence. 
Cells were then trypsinized (0.25% trypsin with 0.1% 
EDTA) and subcultured at densities of 5,000 to 6,000 
cells/cm². Cell passages 3 to 10 were used for the experi-
ments (24). Cultured cells were routinely tested for viability 
with trypan blue exclusion and maintained high viability 
(>95%). For in vivo experiments, the minute fraction of dead 
cells was predominantly removed by washing before tryp-
sinization. For flow cytometric analysis, cells were routinely 
pre-gated to exclude cell clumps, contaminating polymor-
phonuclear cells, red blood cells, platelets, endothelial 
microparticles, debris, as well as dead cells based on 7-AAD 
staining.

Isolation of omental ASC
Under an Institutional Review Board-approved protocol, 
grossly normal appearing human omentum was obtained 
from the staging procedures. O-ASC1 was isolated from a 
patient (BMI = 25.3) with recurrent adenocarcinoma of 
endometrium and ovary (omentum not involved). O-ASC2 
was isolated from a patient (BMI = 30.4) with recurrent 
granulosa cell tumor of the ovary (omentum involved).
ASCs were isolated according to published protocols (24). 
Tissue was subjected to mechanical disruption, followed by 
0.5 mg/mL collagenase type I (Worthington Biochemical) 
and 50 U/mL dispase (Becton Dickinson) digestion accord-
ing to published protocols (6). Digested adipose tissue was 
centrifuged at 1,000 rpm. Supernatant containing adipo-
cytes was moved and cell pellet was resuspended in EGM-
2MV (EBM-2 media with EGM-2MV growth factor supple-
mentation from Lonza). Resuspended cells were filtered 
through 100 μm cell strainer (Becton Dickinson) and 
followed by 40 μm cell strainer. Red blood cells were lysed 
using in 5 mL of RBC Lysis Buffer (eBioscience) with 
icubation in 37°C water bath for 10 minutes. Remaining 
mononuclear cells were resuspended and plated in EGM-
2MV medium.

Characterization of O-ASC lines
After isolation, cells were expanded in vitro, and then 
characterized with flow cytometry to evaluate cell surface 
marker expression. O-ASC1 was characterized at passage 3 
and O-ASC2 at passage 5 with antibodies against the fol-
lowing markers: CD34, CD44, CD45, CD29, CD90, EpCam 
(from Becton Dickinson), and CD105 (from BioLegend). 
Cell differentiation studies were conducted as described 
previously (25, 26). For osteoblast differentiation, confluent 
cells in 6-well plates were cultured in NH OsteoDiff 
Media (Millenyi) for 3 weeks with the media changed 2 
times a week. After 3 weeks, media were removed, cells were 
ashed 3 times with PBS and fixed with precooled 100% 
ethanol. Cells were stained with Alizarin Red S solution 
for 2 to 5 minutes at room temperature to visualize orange 
red calcium deposits, or with von Kossa. For adipocyte 
differentiation, confluent cells in 6-well plate were cultured 

obesity and endometrial cancer may be attributed, at least in 
part, to increased availability of ASCs which are recruited to 
form a supportive tumor microenvironment.

Excess intra-abdominal adipose tissue further increases the 
risk, and in some cases, mortality of intra-abdominal 
cancers such as prostate, colon, pancreatic, and endometrial 
cancer (3, 20, 21). We hypothesized that visceral adipose 
tissue could explain this relationship. To investigate this, we 
isolated a population of ASCs from the omentum, the major 
depot of visceral intra-abdominal adipose tissue. The effect 
of omental-derived ASC (O-ASC) was then compared with 
bone marrow–derived MSC (BM-MSC), subcutaneous ASC 
(SC-ASC), and control fibroblasts by using ex vivo and in vivo 
assays.

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in adipogenic induction media, Dulbecco’s Modified Eagle’s Media (DMEM; Mediatech) supplemented with 10% FBS, 1% penicillin/streptomycin, 10 μg/mL insulin, 500 μmol/L 3-isobutyl-1-methylxanthine (Sigma-Aldrich), 1 μmol/L dexamethasone (Sigma-Aldrich), and 200 μmol/L indomethacine (Sigma-Aldrich). After cells were maintained in induction media for 48 to 72 hours, media were changed to adipogenic maintenance media supplemented with 10% FBS, 1% penicillin/streptomycin, and 10 μg/mL insulin for 24 hours. After 24 hours, media were changed back to induction media and cycle was repeated 3 times. After the third induction step, cells were in maintenance media for 10 days changing media twice a week. Cells were then fixed in 10% formalin for 1 to 2 hours followed by 60% isopropanol. Oil Red O (Sigma-Aldrich) stain was applied to visualize red lipid vacuoles. For chondrocyte differentiation, one million cells in a 15 mL conical were pelleted and resuspended in 2 mL of chondrocyte media; DMEM 1× (Mediatech) supplemented with 1% penicillin/streptomycin, 50 μg/mL ascorbic acid (Sigma-Aldrich), 100 nmol/L dexamethasone (Sigma-Aldrich), and 10 ng/mL TGF-β3 (Invitrogen). Cells were then incubated under 5% CO₂ as a pellet at 37°C with cap loosened to allow for gas exchange. TGF-β3 was added daily to a concentration of 10 ng/mL. Media were changed 3 times a week for 21 days. Cell pellet was rinsed in PBS and fixed with Gendre’s Fluid, after which paraffin-embedded blocks were sectioned and stained with antibodies against collagen II.

**Proliferation assay**

Luciferase-labeled Hec1a cells were plated at a density of 1,000 cells per well in a BD Falcon 96-well plate alone or in 1:1 coculture with O-ASC1, O-ASC2, BM-MSC, SC-ASC, or WI38. Cocultures were conducted in either α-MEM (Mediatech) with 250 μg/mL G418. After 24 hours, luciferase expression was measured in 0.15 mg/mL of luciferin. After 1 hour incubation at 37°C, luminescence was measured with spectrophotometer (FLUOstar Omega, BMG Labtech). Media were then replaced and measurements were repeated every other day until cells reached confluence.

**Generation of luciferase-GFP- and RFP-expressing cells**

Cell lines were labeled with the GFP using a lentiviral vector, pFUGW (27) that was generously provided by Dr. Baltimore (Caltech). The titers of virus stocks were determined by calculating the percentage of GFP-positive (GFP⁺) 293T cells transduced with serially diluted virus suspensions. For transduction, the O-ASC and SC-ASC cell lines were seeded at moderate density overnight. Two hours before transduction, the medium was changed, and then transductions were carried out for 24 hours in the presence of 8 μg/mL polybrene (Sigma-Aldrich). The cells were then fluorescence-activated cell sorted and further expanded for injections. The MSCs were labeled similarly but with a red fluorescent protein (RFP) generously obtained from Tsien’s laboratory (UCSD) that was cloned into pFUGW construct in place of GFP.

**Migration assay**

Stromal cell migration including O-ASC, WI38, BM-MSC, and SC-ASC were assayed in conditioned media (CM) generated from Hec1a cells. CM was collected from a 6-well plate which was plated with 300,000 cells per well incubated in serum-free media for 48 hours. Migration assays were conducted in a 24-well Transwell (Corning Inc.) with 8 μm pores. CM was placed in lower portion of Transwell and cell lines of interest on the upper portion of Transwell. All samples were assayed at least in quadruplicate. Twenty-four hours later, the number of cell migrated to the bottom of the Transwell was measured by staining membranes with Hema 3 Stat Pack (Thermo Fisher Scientific) and mechanically removing residual cells on upper membrane. The number of cells on the membrane was then quantitated per high-power field.

**In vivo studies**

Animal experimentation was approved by the Animal Welfare Committee of the University of Texas, Houston, TX. Ten-week-old mice N1/N1-foxn1tm1 (Jackson Laboratory) were injected subcutaneously with 1.5 x 10⁶ Hec1a cells in 150 μL of PBS into the right flank (5 mice per group). WI38 fibroblasts, MSC, O-ASC2, and SC-ASC were grown in α-MEM containing 20% FBS, 1-glutamine before injection. O-ASC1 was grown in either α-MEM containing 20% FBS or EGM-2MV. Starting the next day, mice were injected with 1.5 x 10⁶ corresponding cells (or PBS) every third day onto lower back, alternating the side of injection. Tumor size was measured with a caliper and volume was calculated as length x width² x 0.52. Before tumors reached 1 cm in size, mice were euthanized and tumors were recovered from anesthetized mice after heart perfusion with 10 mL PBS. Immunofluorescence on formalin-fixed, paraffin-embedded tissue sections was carried out after antigen retrieval, washing with 0.2% Triton X-100, blocking in Serum-Free Protein Block (DAKO), followed by incubation with primary antibodies (4°C, 12 hours) and secondary antibodies (room temperature, 1 hour) in PBS containing 0.05% Tween-20. The primary antibodies used were as follows: goat anti-GFP from GeneTex (1:100), rabbit anti-RFP from Abcam (1:100), rabbit anti-Ki-67 from Thermo Scientific (1:200), rabbit (1:200), or goat (1:100) anti-CD31 from Santa Cruz Biotechnology, and rabbit anti-desmin from Abcam (1:200). Secondary donkey FITC-conjugated (1: 150) and Cy3-conjugated (1:300) IgG were from Jackson ImmunoResearch. Nuclei were stained with Hoechst 33258 (Invitrogen). Masson’s Trichrome staining of tumor sections was carried out at UTHealth histology CORE facility. Images were acquired with Olympus IX70 inverted fluorescence microscope/MagnaFire software.

To assess tumor composition, 3 random internal tumor field views proximal to the capsule were inspected for each tumor at 100× magnification. In each field views, the proportion of the area positive for necrosis (Mason’s tri-chrome anuclear pink staining), proliferation (containing Ki-67⁺ cells), vascularization (containing CD31⁺ vessels), and mature vasculature (containing desmin⁺ vessels) was
scored by two investigators blinded to the experimental group of the samples. Data were averaged from individual field views for each group.

Statistical analysis was conducted with unpaired Student t test. Normal distribution of data was confirmed by the Kolmogorov–Smirnov test.

Results

Characterization of O-ASC

Cells were isolated from omental adipose tissue of 2 patients with gynecologic cancer: one with localized adenocarcinoma of endometrium and ovary and the other with granulosa cell tumor of the ovary infiltrating the omentum. Tissue was subjected to mechanical disruption, followed by collagenase and dispase digestion (6). Adipocytes were removed with centrifugation and remaining mononuclear cells were plated and cultured in standard MSC expansion conditions. Morphologically, O-ASC from patients 1 and 2 (O-ASC1 and O-ASC2) exhibited a mesenchymal phenotype, similar to that previously reported for BM-MSC and SC-ASC (Fig. 1A).

To characterize cell surface marker expression on O-ASC, we expanded O-ASC1 and 2 and compared cell surface marker expression with flow cytometry to BM-MSC (Fig. 1B). O-ASC expressed CD29, CD44, CD73, CD90, and CD105, which are characteristically expressed on MSCs (7, 28). BM-MSCs expressed slightly higher levels of CD44 and CD105 than O-ASC, whereas O-ASC expressed slightly higher levels of CD29 than BM-MSCs. O-ASCs were negative for an epithelial marker EpCam, a monocyte marker CD11b, and for CD34 and CD45. This excludes malignant, monocyte, and endothelial cells as a possible origin of O-ASC.

To test whether O-ASCs exhibit multipotency typical of MSCs (7) in vitro, we subjected them to differentiation assays (Supplementary Fig. S1). On adipogenic induction, O-ASC formed lipid droplets, which were detected with Oil Red O stain showing lipogenesis typical of adipocyte differentiation. Similar to BM-MSCs, O-ASC1 cells were capable of accumulating large lipid droplets, whereas O-ASC2 cells tended to uniformly accumulate smaller lipid droplets. Following osteogenic induction, O-ASC1, and to a lesser degree O-ASC2, formed mineral deposits detected with Alizarin red (Supplementary Fig. S1) and expressed alkaline phosphatase indicating osteoblastic differentiation. On chondrogenic induction, Toluidine Blue staining of sections from pellets grown in suspension indicated the capacity of O-ASC to differentiate into cartilaginous lineage. Expression of collagen II, although detectable in O-ASC1 and O-ASC2, was lower than in BM-MSC differentiation cultures, suggesting that O-ASC formed primarily fibrocartilage. Combined, these results confirm that O-ASCs are indeed MSCs from the omentum.

ELISA for cytokines

Stromal cells (O-ASC1, BM-MSC, SC-ASC, and WI38) were cultured in 10% FBS α-MEM media. Media were removed at intervals from 1 to 7 days, centrifuged and frozen. ELISA assays were then done on supernatants by using kits from R&D Systems.

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**Figure 1.** Characterization of O-ASC. A, morphology of adherent cells showing similarity of O-ASCs to SC-ASCs and BM-MSCs. B, flow cytometric characterization of O-ASC based on surface marker expression indicates their similarity to BM-MSC. Light gray lines, isotype controls.
Homing of O-ASC to tumors and in vivo effects on tumor growth

To evaluate in vivo effects of O-ASC on endometrial cancer growth, immunodeficient nude mice with human endometrial adenocarcinoma Hec1a xenografts in the upper flank were injected into the lower flanks with low doses \((10^5\text{ cells \times 3 times weekly})\) of MSCs. In this metronomic model based on our previous study (6), the mesenchymal cells are remote from the xenografted tumor and must thus be recruited into the tumors, simulating the off-site recruitment of ASC from regional adipose tissue in human tumors. The impact of O-ASC1 and O-ASC2 on Hec1a xenograft growth was then compared with SC-ASC and BM-MSC as well as control lung fibroblasts (WI38) and a PBS control group in this metronomic model. Stromal cells injected into lower flank were labeled with GFP or RFP to track engraftment of cells into Hec1a tumors.

Weekly tumor volume measurements showed that Hec1a xenograft growth was significantly \((P < 0.05)\) accelerated by administration of O-ASC1, O-ASC2, and BM-MSC, as compared with PBS controls (Fig. 2A). In contrast, tumors grown in mice injected with SC-ASC or WI38 did not grow significantly more rapidly than PBS-injected control tumors.

To determine whether the effect of O-ASC on tumor growth could be attributed to more efficient migration of stromal cells into the tumor microenvironment, we evaluated tissue sections to assess biodistribution of injected cells with immunofluorescence to RFP and GFP. Consistent with published studies (6), no GFP\(^+\) or RFP\(^+\) cells were detected in liver, pancreas, spleen, and other control organs (data not shown). In contrast, comparable numbers of GFP\(^+\) or RFP\(^+\) cells were detected in tumors of mice administered with O-ASC1, O-ASC2, BM-MSC, and SC-ASC but not WI38. The marker-expressing cells were detected both in the internal zone of the tumor and, more commonly, in the stromal capsule at the tumor periphery (Fig. 2B). On the basis of the numbers of cells counted in the tumor section area, and the inferred number of such sections composing each tumor, we estimated that on the average 10,000 to 15,000 cells were detectable within a tumor, which corresponds to 10\% to...
15% of each injection dose. These numbers, consistent with those reported previously for MSC tumor homing (29), indicate efficient recruitment of administered cells by tumors but not by normal organs. Lack of tumor growth promotion by SC-ASC despite their capacity for tumor homing indicate that properties unique for O-ASC and BM-MSC are responsible the tumor growth advantage observed upon recruitment of these cells.

We tested that the administered cells contribute to the tumor vasculature. Immunofluorescence of tumor sections with antibody recognizing human and mouse CD31 show that engrafted GFP+ cells are CD31-negative, although often associated with CD31-positive endothelium (Supplementary Fig. S2). This indicates that mesenchymal cells do not transdifferentiate into endothelial cells in this experimental setting. We also did not detect expression of CD34 in engrafted GFP+ cells by using antibodies recognizing human CD34. This is consistent with in vitro flow cytometry analysis of O-ASC showing they were negative for CD34 (Fig. 1).

**Effect of O-ASC on tumor matrix organization and cell viability**

To investigate the mechanism through which O-ASC promote tumor growth, we analyzed morphology of fixed tumors recovered in the end of the study. Masson’s Trichrome staining for ECM proteins revealed differences in organization of tumors with recruited O-ASC1, O-ASC2, and BM-MSC as compared with the other groups and control tumors in PBS-injected mice (Fig. 3). The former had thicker peripheral tumor capsule with defined extracellular matrix (ECM) representation (blue) as well as dense inner tumor mass all the way to the center of the tumor core. In contrast, tumors in mice injected with PBS had a thinner peripheral tumor capsule (Fig. 3). Histologic analysis also showed that tumors in mice injected with O-ASC1 and O-ASC2 had no detectable necrosis at the tumor periphery and only limited areas of necrosis in the center (Fig. 3). In contrast, the other groups had tumors displaying vast areas of necrosis (detectable as pink areas missing nucleated cells) both at the periphery and particularly in the tumor center. The percentage of tumor containing necrotic areas was quantified to be significantly lower (average of 22% and 9% necrotic tumor area in Hec1a xenografts recruiting O-ASC1 and O-ASC2 as compared with 48% necrotic tumor area in PBS control tumors, \( P < 0.05 \); Supplementary Fig. S3). These results indicate that O-ASC support the formation of the fibrous tumor capsule and decrease the extent of intratumoral necrosis.

**Effect of O-ASC on cancer cell proliferation**

To determine whether O-ASCs promote malignant cell proliferation, we carried out immunofluorescence to detect Ki-67, a protein expressed in dividing cells. Immunofluorescence showed that malignant cells throughout the tumors of mice recruiting O-ASC1 and O-ASC2 were almost uniformly positive for Ki-67, whereas in WI38 and PBS control groups, proliferative tumor cells were limited to the subcapsular region (Fig. 4A). Quantification of tumor section area containing proliferating cells revealed a higher percentage of proliferative tumor mass in O-ASC1 and O-ASC2 groups, as compared with PBS-injected control mice (average of 56% and 88% proliferative tumor mass in O-ASC1 and O-ASC2 recruiting tumors respectively as compared with 22% for control PBS tumors, \( P < 0.05 \); Supplementary Fig. S3). The frequency of proliferating cells for tumors recruiting SC-ASCs was not significantly higher than PBS control tumors (Fig. 4A and Supplementary Fig. S3).

To test whether the cells recruited by tumors have a direct mitogenic effect on malignant cells, we analyzed proliferation of Hec1a culture ex vivo (Supplementary Fig. S4). In this experiment, luciferase-labeled Hec1a cells were grown with and without O-ASC1, O-ASC2, BM-MSC, SC-ASC, or WI38 fibroblasts and tumor cell proliferation was monitored with luciferase imaging. Although coculture with either O-ASC1 or O-ASC2 significantly increased HEC1a cell proliferation, SC-ASC and WI38 control cells had a comparable effect, whereas BM-MSCs on the contrary inhibited Hec1a proliferation. This lack of correlation between the ex vivo and in vivo effects of administered cells...
on Hec1a suggested that increased proliferation in tumors was likely because of indirect effects of O-ASC.

**Effect of O-ASC on tumor vascularization**

Blood vessels are essential for tumor oxygenation and nutrient supply, and their deficiency or inefficiency can limit tumor perfusion and growth (30). We therefore investigated whether enhanced angiogenesis could account for the effect of O-ASC on xenograft growth. Interestingly, immunofluorescence with antibodies against CD31 (PECAM-1) or VCAM-1 revealed only weak level of expression of these endothelial makers on many Hec1a xenograft structures that morphologically appeared as vasculature. Nevertheless, CD31 immunofluorescence signal was sufficient for us to detect higher density of vasculature in tumors observed for mice administered with O-ASC1, O-ASC2, and BM-MSC as compared with controls and the other groups (Fig. 4). According to quantification studies, in O-ASC2 groups, the percentage of tumor area containing CD31⁺ blood vessels was significantly higher than in PBS controls, and there was also a trend for increased vascularization in the O-ASC1 and MSC groups (Supplementary Fig. S3). Although large tortuous blood vessels were numerous both at the periphery of the tumor closer to the capsule (Fig. 4A) and in the core of the tumor (Fig. 4B) for the O-ASC1, O-ASC2, and BM-MSC groups, virtually no CD31⁺ vessels were detected in the tumor center for SC-ASC, WI38, and PBS control groups. Similar results were obtained by immunohistochemistry with antibodies against CD34, which stain both endothelial and some of the perivascular/stromal cells (data not shown).

![Figure 4. Tumor cell proliferation and vascularization.](image-url)

According to the histologic evaluation of tumors, the tumors that recruited O-ASC1, O-ASC2, and BM-MSC had a more extensive fibrovascular network and appeared to contain more mature vasculature. The mesenchymal cells within this network are known to be a mixed population of leukocytes and mesenchymal cells, some of which functionally integrate into tumor blood vessels as pericytes and...
contribute to vascular perfusion capacity (31). To compare the extent of vasculature maturation in tumors recruiting different types of MSCs, we analyzed tumor sections for expression of desmin, a marker of pericytes (32). Anti-desmin immunofluorescence revealed robust pericyte coverage throughout the tumor vasculature in mice administered with O-ASC1 and O-ASC2, and to a slightly less extent with BM-MSC and SC-ASC (Fig. 5). Quantification analysis enumerating both perivascular and stromal desmin-positive cells indicated a significant increase in their recruitment, for all groups receiving cell injections, as compared with PBS-injected mice. Mice that received O-ASC1 and O-ASC2 injections had significantly more desmin-positive cells in tumors (Supplementary Fig. S3), consistent with their increased vascularization. Desmin expression was abundant on perivascular cells in large tortuous blood vessels both in subcapsular layers (Fig. 5A) and in the center (Fig. 5B) of the tumors in mice injected with O-ASC1, O-ASC2, or BM-MSC. In contrast, tumors in mice injected with for SC-ASC and WI38 contained predominantly disseminated desmin-positive cells, whereas only some areas of the tumor contained rare desmin-positive cells in the PBS group (Fig. 5A and B). Because no clear endothelial networks were revealed with CD31 (Fig. 4) or VCAM-1 (not shown) immunofluorescence in these 3 groups, it is not clear whether these individual desmin-positive cells are associated with abnormal microvasculature weakly expressing endothelial markers or are simply disseminated throughout the stroma. Combined, these observations suggest that O-ASCs promote formation of pericyte coverage and, therefore, likely functional blood vessels throughout the tumor upon recruitment, which explains improved cell survival and proliferation.

**Cross-talk between O-ASC and tumor cells**

To investigate the potential differences in distinct ASC types to secrete factors known to promote cancer progression, we carried out ELISA assays (Supplementary Fig. S5). CM were analyzed for the concentration of secreted VEGF-A, fibroblast growth factor (FGF)-2, and hepatocyte growth factor (HGF).
factor (HGF). SDF1-α secretion was analyzed in parallel, as HEC1a tumors secrete CXCR4 and had been shown to respond to SDF1-α. Our data show that significantly higher levels of VEGF, FGF, and SDF1-α are secreted by BM-MSC and O-ASC, as compared to SC-ASC and WI38. These findings suggest that O-ASCs and BM-MSCs promote tumor growth above the level observed for SC-ASC because of the combined activity of several factors that can directly or indirectly enhance angiogenesis.

Finally, to determine whether O-ASCs exhibit enhanced in vitro migration in response to Hec1a-secreted factors, we carried out an ex vivo migration assay. The effect of Hec1a-conditioned medium on the motility of cells used in the study was tested by using the Boyden chamber Transwell assay (Supplementary Fig. S6). O-ASC1, O-ASC2, and BM-MSC, but not SC-ASC and WI38 fibroblasts, exhibited significant in vitro migration activation in response to Hec1a-secreted factors (P < 0.05). Because the cells responding to tumor signals also appear to be comparatively more efficient in supporting tumors, our data collectively suggest that the crosstalk between malignant and MSCs is an important component of cancer progression. To gain insight into the potential mechanisms of ASC trafficking to the tumor, we profiled chemokine secretion by tumor cells. Among a panel of 31 human chemokines tested, we identified IL8 and CXCL1, as chemotactic proteins secreted by Hec1a cells (Supplementary Fig. S7). To determine whether these factors could be mechanistically involved in O-ASC migration, we investigated expression of the IL-8 and CXCL1 receptors, CXCR1 and CXCR2, on stromal cells. High levels of CXCR1 expression were detected on O-ASC1, O-ASC2, and BM-MSC, but not on SC-ASC or WI38 fibroblasts (Supplementary Fig. S8) whereas CXCR2 was expressed on all stromal cells investigated. These findings suggest that IL-8 and CXCL1 secretion by Hec1a cells may trigger chemotaxis of stromal cells toward tumors via CXCR1 and CXCR2. Because all cell types displayed tumor homing, we propose that the ubiquitously expressed CXCR2 is responsible for engraftment of the injected cells in our experiment.

Discussion

**Mechanism of endometrial tumor growth promotion by O-ASC**

Activation and proliferation of stromal cells recruited by tumors leads to their conversion into myofibroblasts/reactive stroma (12, 33, 34). The resulting remodeling of the ECM executed by mesenchymal tumor cells and the resulting desmoplasia is an integral component of cancer progression (35). However, the origins of these stromal cells, which compose the tumor microenvironment and their individual roles in cancer progression, are not well understood. This is due largely to the absence of any specific marker to identify the origins of stromal cells within tumors.

Studies in animal models have showed that on tumor infiltration, MSCs contribute to the CAF pool and establish a trophic microenvironment (9, 11). In previous studies by us and others, MSCs have been shown to traffic to tumors (19, 29) and participate in forming the supportive fibrovascular network (13, 32, 36, 37). Although there is evidence that cells composing tumor stroma can be recruited from the bone marrow, their possible extramedullary origins have been less well explored. Adipose tissue is particularly rich in MSCs and could thus serve as primary MSC source for tumors (7, 38). Although systemic mobilization of MSC from adipose tissue (ASC) has been proposed (25, 26, 39) and recruitment of ASC from remote depots is conceivable and has been shown to occur in mouse cancer models (6), it is likely that adipose tissues adjacent to the tumor may be a more significant contributor of ASC. Moreover, the differences reported for ASCs residing in subcutaneous and visceral adipose depots (40) suggest that ASCs from the omentum could have a distinct role in the progression of intraperitoneal cancers.

Here, we analyzed omental adipose tissue, which is the primary intraperitoneal reservoir of adipose tissue which is routinely removed during staging procedures for ovarian and some endometrial cancers (41). This is the first study investigating the effects of O-ASCs on cancer progression. We found that O-ASCs from different patients exhibit morphology, cell surface marker expression, and differentiation capacity comparable with those observed for BM-MSCs and previously reported for SC-ASCs (42). O-ASC, SC-ASC, and BM-MSC exhibited tumor tropism when injected into tumor-bearing mice distantly from the endometrial xenograft. Despite these similarities, we found that O-ASCs had more potent effect than SC-ASCs on endometrial cancer cell proliferation and survival in vivo, thus resulting in expedited tumor growth. O-ASCs and SC-ASCs were isolated from donors with comparable BMI, suggesting that the differences in their cancer promotion capacity cannot be attributed to obesity-associated changes in ASC properties. Consistent with promoted angiogenesis being the foundation of increased tumor cell mass (30), O-ASC recruitment correlated with higher abundance of both small and large tortuous blood vessels suggesting that O-ASC tumor homing promotes the vascularization of endometrial caners. In addition, we observed increased pericyte coverage of vasculature in tumors recruiting O-ASCs. Accumulating evidence indicates that ASCs serve as pericytes in adipose tissue (43), which could be their function uncoupled from multipotency and capacity to serve as progenitor cells. Although our previous studies show that ASCs can acquire perivascular localization upon tumor homing, the numbers of administered O-ASCs observed in the tumors cannot account for the increased numbers of desmin-positive cells. Therefore, O-ASCs may play a role in recruiting endogenous pericyte progenitors that contribute to the formation of functional tumor vessels.

Increased microvascular density (MVD) has been associated with advanced stage and poor prognosis in endometrial cancer (44, 45). In our study, there was a clear correlation between the capacity of an administered mesenchymal cell type to enhance tumor vascularization and its miogenic...
We identified interleukin (IL)-8 as a potential chemotactic signaling pathway involved in O-ASC recruitment. Chemokine profiling showed that CXCL8 (IL-8) and CXCL1 are secreted by Hec1a endometrial carcinoma cells. The IL-8 receptor, CXCR1, was detected on O-ASC1, O-ASC2, and BM-MSC but not SC-ASC or WI38 cells, suggesting a role for IL-8 signaling in O-ASC recruitment and possibly tumor promotion. IL-8 has previously been reported to support MSC recruitment in a glioma model and to mediate radiation-enhanced tumor tropism of MSC (47–49). Future studies will investigate the therapeutic potential of inhibiting O-ASC recruitment by suppressing signaling through chemotactic pathways such as IL-8.

In summary, our results show that the omentum contains a population of MSCs which resemble BM-MSCs and SC-ASCs with regard to cell surface marker expression and multipotency but have a unique effect on tumor progression, suggesting their role in etiology of human intraperitoneal cancers. Our combined data indicate that O-ASCs support tumor vascularization, which leads to enhanced survival and proliferation of malignant cells and may contribute to O-ASC effects on tumor growth. We propose that the cross-talk between the tumor and the surrounding adipose tissue could contribute to ASC “education” and their adaptation for the tumor benefit, thus resulting in a “vicious circle” driving the progression of cancer. Our findings suggest a novel mechanism accounting for the increased risk and mortality of endometrial cancer in women with excess visceral adipose tissue. Preventing migration of O-ASCs into endometrial cancers by disrupting interactions of O-ASCs with malignant cells, tumor endothelium, and other tumor stromal cells may be a novel treatment strategy.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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