Combined MEK and VEGFR Inhibition in Orthotopic Human Lung Cancer Models Results in Enhanced Inhibition of Tumor Angiogenesis, Growth, and Metastasis

Osamu Takahashi1, Ritsuko Komaki1, Paul D. Smith5, Juliane M. Jürgensmeier6, Anderson Ryan5, B. Nebiyou Bekele2, Ignacio I. Wistuba3, Jörg J. Jacoby4, Maria V. Korshunova1, Anna Biernacka1, Baruch Erez4, Keiko Hosho1, Roy S. Herbst4, and Michael S. O’Reilly1

Abstract

Purpose: Ras/Raf/mitogen-activated protein–extracellular signal-regulated kinase (ERK) kinase (MEK)/ERK signaling is critical for tumor cell proliferation and survival. Selumetinib is a potent, selective, and orally available MEK1/2 inhibitor. In this study, we evaluated the therapeutic efficacy of selumetinib alone or with cediranib, an orally available potent inhibitor of all three VEGF receptor (VEGFR) tyrosine kinases, in murine orthotopic non–small cell lung carcinoma (NSCLC) models.

Experimental Design: NCI-H441 or NCI-H460 KRAS-mutant human NSCLC cells were injected into the lungs of mice. Mice were randomly assigned to treatment with selumetinib, cediranib, paclitaxel, selumetinib plus cediranib, or control. When controls became moribund, all animals were sacrificed and assessed for lung tumor burden and locoregional metastasis. Lung tumors and adjacent normal tissues were subjected to immunohistochemical analyses.

Results: Selumetinib inhibited lung tumor growth and, particularly at higher dose, reduced locoregional metastasis, as did cediranib. Combining selumetinib and cediranib markedly enhanced their antitumor effects, with near complete suppression of metastasis. Immunohistochemistry of tumor tissues revealed that selumetinib alone or with cediranib reduced ERK phosphorylation, angiogenesis, and tumor cell proliferation and increased apoptosis. The antiangiogenic and apoptotic effects were substantially enhanced when the agents were combined. Selumetinib also inhibited lung tumor VEGF production and VEGFR signaling.

Conclusions: In this study, we evaluated therapy directed against MEK combined with antiangiogenic therapy in distinct orthotopic NSCLC models. MEK inhibition resulted in potent antiangiogenic effects with decreased VEGF expression and signaling. Combining selumetinib with cediranib enhanced their antitumor and antiangiogenic effects. We conclude that combining selumetinib and cediranib represents a promising strategy for the treatment of NSCLC.

Introduction

Lung cancer is a global problem, and new therapies and approaches to the treatment of lung cancer are urgently needed (1, 2). The approach to the treatment of non–small cell lung carcinoma (NSCLC) is currently in transition and...
Lung cancer is a major worldwide health problem and a leading cause of cancer-related death and morbidity. The outcome for patients with lung cancer has not significantly changed in over 2 decades and more effective therapies for lung cancer are urgently needed. We evaluated combined mitogen-activated protein/extracellular signal-regulated kinase (MEK) blockade and angiogenesis inhibition directed against VEGF receptor (VEGFR) signaling in orthotopic models of human non–small cell lung carcinoma (NSCLC) that closely recapitulate clinical patterns of lung cancer progression and allow for the study of the influence of lung microenvironment upon response to therapy. MEK inhibition potentiated the effects of anti-VEGF therapy and independently inhibited lung tumor angiogenesis, VEGF production, and VEGFR signaling. These data provide a strong basis for clinical trials combining selumetinib and cediranib in lung cancer patients.

but no overall survival advantage was observed (5, 6) nor was there a benefit for overall survival when it was combined with cisplatin or carboplatin and etoposide in patients with extensive stage small cell lung cancer (7). VEGF signaling remains an important target in anticancer therapy because of its role in angiogenesis (8), which is fundamental to tumor growth and spread (9), but it has become apparent that treatment with bevacizumab alone or with systemic chemotherapy may not be sufficient for the treatment of some advanced lung cancers. Furthermore, toxicities associated with bevacizumab, when it is used with systemic chemotherapy, have been observed in clinical trials in lung cancer patients (10).

Cediranib (AZD2171) is an orally available and highly potent VEGF receptor (VEGFR)-1, 2, and 3 tyrosine kinase inhibitor with additional activity against the platelet-derived growth factor-β receptor and c-kit (11–13). Cediranib is currently being investigated in clinical trials both alone and in combination with chemotherapy for a variety of malignancies. The addition of cediranib to standard chemotherapy for metastatic colorectal cancer was associated with improved progression-free survival but did not improve overall survival in a phase III trial (14, 15). In a phase III trial of cediranib or lomustine alone and in combination in patients with recurrent glioblastoma, no statistically significant difference in median progression-free survival of 92 days for the cediranib arm or 125 days in the combination arm was observed as compared with 82 days for lomustine alone (16, 17). Progression-free survival at 6 months was 16% in the cediranib arm which was lower than the 25.8% survival at 6 months reported in the earlier phase II trial of cediranib for recurrent glioblastoma (18).

For lung cancer, a phase II/III trial (BR24) in which cediranib at a 30 mg dose was combined with carboplatin and paclitaxel for advanced NSCLC showed improved response rates and progression-free survival but was terminated early due to concerns of toxicity (19, 20). Although the clinical experience for cediranib in lung and other cancers does show evidence for activity, the results show that it may be prudent to combine it with other biologically targeted therapeutics for the treatment of lung cancer.

Targeted therapy directed against the epidermal growth factor receptor (EGFR) with gefitinib (21), erlotinib (22), or cetuximab (23) are currently being used in the clinic for the treatment of NSCLC with improvements in progression-free survival and overall survival in subsets of patients (24). EGFR tyrosine kinase inhibitors have shown single-agent activity in lung cancer patients whose tumors harbor EGFR mutations (25) but the efficacy of these agents for cancers that harbor Kras mutations is unclear (26). Although some patients will experience a durable benefit from these agents, in most cases the improvement in survival can only be measured in weeks or months. Furthermore, these agents may only be effective in a small percentage of lung cancer patients and resistance to therapy can develop (27).

To overcome some of the limitations of EGFR and other growth factor receptor inhibitors and to more broadly target lung cancer growth, therapeutic strategies to target signaling pathways that are downstream of these receptors have been investigated (28). The mitogen-activated protein kinase/extracellular signal-regulated kinase (MEK) that are situated downstream of Ras and Raf represent attractive therapeutic targets for lung and other cancers. MEK signaling is crucial in the regulation of multiple processes, including tumor cell proliferation and survival, in a variety of cancers, including lung cancer, and its inhibition offers a particularly attractive therapeutic target (29, 30). Selumetinib (AZD6244 and ARRY-142886), a potent, selective, and orally available MEK1/2 inhibitor (31, 32), has been studied clinically in a variety of cancers, including lung cancer (33) and is currently being evaluated in a phase II clinical trial in KRAS-mutant NSCLC. However, the inhibition of MEK signaling alone may not be sufficient in patients with advanced lung cancer, and feedback mechanisms in this pathway may be problematic when it is used alone (34).

To determine the efficacy of selumetinib for lung cancer growing in the lung and to investigate its use for the treatment of lung cancer with antiangiogenic therapy, we have conducted both monotherapy and combination with targeted therapy directed against VEGF signaling with cediranib in orthotopic models of human lung cancer. We report the results of this novel combination therapy in our murine orthotopic models of human lung cancer that closely recapitulate the clinical behavior of lung cancer in humans (35) using 2 different human NSCLC cell lines that harbor mutations in Kras.

Materials and Methods

Cell cultures

The human lung adenocarcinoma cell line NCI-H441 and the human large cell lung cancer cell line NCI-H460, both of which have KRAS mutations (36, 37), were obtained.
from the American Type Culture Collection. Both cell lines were molecularly characterized by The University of Texas MD Anderson Cancer Center’s Cell Line Characterization Shared Resource and determined to be free of Mycoplasma and pathogenic murine viruses. Cells were maintained in RPMI-1640 with 10% fetal bovine serum, sodium pyruvate, nonessential amino acids, l-glutamine, 2-fold vitamin solution, and penicillin–streptomycin (Invitrogen) and incubated in an atmosphere of 5% CO₂ and 95% air at 37°C.

**Orthotopic model of human NSCLC**

Six- to 8-week old male athymic nude mice (Taconic) were used for experiments in accordance with current regulations and standards of the U.S. Department of Agriculture, the U.S. Department of Health and Human Services, the NIH, and The University of Texas MD Anderson Cancer Center. Mice were anesthetized with sodium pentobarbital (50 mg/kg body weight) and placed in the right lateral decubitus position. A 5-mm skin incision overlying the left chest wall was made and the left lung was visualized through the pleura. A total of 1 × 10⁶ NCI-H441 cells or 5 × 10⁵ NCI-H460 cells (single-cell suspensions, greater than 90% viability) in 50 μl of Hank’s balanced salt solution were injected into the left lungs of the mice through the pleura with a 20-gauge needle. After tumor cell injection, the wound was stapled and the mice were placed in the left lateral decubitus position and observed until fully recovered.

**Drug preparation and treatment schedules**

Selumetinib and cediranib (provided by AstraZeneca) were formulated in vehicle consisting of either 0.5% w/v hydroxypropyl methylcellulose/0.1% w/v Tween 80 or 1% w/v polysorbate 80, respectively. Paclitaxel (Brasil-Myers Squibb) was dissolved in saline immediately before use. Fourteen days after the implantation of NCI-H441 cells or 10 days after the implantation of NCI-H460 cells, when the lung tumors were established, mice (10–13 per group) were randomly allocated to receive treatment with selumetinib (12.5 or 25 mg/kg twice daily by oral gavage), cediranib (3 mg/kg once daily by oral gavage), paclitaxel (200 μg per mouse once a week intraperitoneally), selumetinib (25 mg/kg twice a day by oral gavage) plus cediranib, or vehicle control. Selumetinib was given twice daily at 8-hour intervals and cediranib was given once daily, 4 hours after the first daily dose of selumetinib. Treatment was continued until the control mice became moribund (55 days for the NCI-H441 model or 19 days for the NCI-H460 model), at which point all mice were killed by CO₂ inhalation and assessed for lung weight, primary lung tumor volume (Π × d_long × d_short²/4), and the presence of medastinal lymph node disease or distant metastasis. For the NCI-H460 model, in which mice also develop chest wall tumors, the aggregate volume of the chest wall tumors was combined with the primary lung tumor volume to create the total tumor volume.

**Histologic preparation and immunohistochemical-immunofluorescent staining**

Primary lung tumors and adjacent lung tissues were removed from all of the mice in every treatment group and fixed with 10% formalin and embedded in paraffin or directly frozen in OCT cryoembedding compound and then sectioned and stained with hematoxylin and eosin or immunohistochemical staining for CD31 (rat anti-mouse; Pharmingen) and dual immunofluorescence staining for CD31 and activated VEGFR2-3 (rabbit anti-human; EMD Biosciences) were carried out with frozen tissues as described previously (38). Sections of formalin-fixed, paraffin-embedded tissue specimens were used to assess cleaved caspase-3 (Cell Signaling Technology), Ki-67 (rabbit a-human; Thermo Scientific), VEGF (goat anti-rat/human; R&D), VEGFR2 (rabbit a-human; Santa Cruz), and phosphorylated mitogen-activated protein kinase (MAPK) 44/42 (Erk1/2) (Thr202/Tyr204) (anti-human; SignalStain kit from Cell Signaling) as described previously (38–40).

**Quantification of microvessel density, vascular area, Ki-67, caspase-3, and activated extracellular signal-regulated kinase**

For quantification of microvessel density and vascular area in lung tumors, up to 4 random fields for each tumor section at ×100 magnification (60% center field) were captured after staining with anti-CD31 antibody. Microvessels were counted and vascular area was calculated using Image Pro software (Media Cybernetics, Inc.). Microvessel density was presented as the number of microvessels per field and as the percentage of vascular pixel area to field pixel area. The number of Ki-67- and activated extracellular signal-regulated kinase (ERK)–positive nuclei was counted regardless of the immunointensity in 4 random fields at ×100 magnification (60% center field). The number of cleaved caspase-3–positive cells was counted in similar fashion but at ×200 magnification. Ki-67 immunoreactivity was expressed as the percentage of Ki-67–positive cells to the total tumor cells per field.

**H-scoring of VEGF and VEGFR2 immunoreactivity**

For semiquantification of VEGF and VEGFR2 immunoreactivity, H-scores were independently generated by 2 of the authors (O.T. and A.B.) who were blinded as to treatment group as described previously (41), with slight modification. H-scores were based on findings from up to 4 randomly selected fields for each tumor section at ×100 magnification (60% center field). Staining intensity was graded as undetectable (0), weak (1), medium (2), or strong (3), and the percentage of positive cells per field was calculated. The intensity score and the percentage of positive cells were then multiplied to give an H-score (possible range, 0–300).

**Immunofluorescence quantification**

Dual fluorescent staining for endothelial cells (CD31, red), activated VEGFR2/3 (green), and tumor cell nuclei (blue) were completed as described above. The expression of activated VEGFR2/3 in tumor-associated endothelial cells was identified by colocalized yellow fluorescence. The pixel

---

**MEK1/2 and VEGFR Inhibition in a Mouse Model of Lung Cancer**

**Histologic preparation and immunohistochemical-immunofluorescent staining**

Primary lung tumors and adjacent lung tissues were removed from all of the mice in every treatment group and fixed with 10% formalin and embedded in paraffin or directly frozen in OCT cryoembedding compound and then sectioned and stained with hematoxylin and eosin or immunohistochemical staining for CD31 (rat anti-mouse; Pharmingen) and dual immunofluorescence staining for CD31 and activated VEGFR2-3 (rabbit anti-human; EMD Biosciences) were carried out with frozen tissues as described previously (38). Sections of formalin-fixed, paraffin-embedded tissue specimens were used to assess cleaved caspase-3 (Cell Signaling Technology), Ki-67 (rabbit a-human; Thermo Scientific), VEGF (goat anti-rat/human; R&D), VEGFR2 (rabbit a-human; Santa Cruz), and phosphorylated mitogen-activated protein kinase (MAPK) 44/42 (Erk1/2) (Thr202/Tyr204) (anti-human; SignalStain kit from Cell Signaling) as described previously (38–40).

**Quantification of microvessel density, vascular area, Ki-67, caspase-3, and activated extracellular signal-regulated kinase**

For quantification of microvessel density and vascular area in lung tumors, up to 4 random fields for each tumor section at ×100 magnification (60% center field) were captured after staining with anti-CD31 antibody. Microvessels were counted and vascular area was calculated using Image Pro software (Media Cybernetics, Inc.). Microvessel density was presented as the number of microvessels per field and as the percentage of vascular pixel area to field pixel area. The number of Ki-67- and activated extracellular signal-regulated kinase (ERK)–positive nuclei was counted regardless of the immunointensity in 4 random fields at ×100 magnification (60% center field). The number of cleaved caspase-3–positive cells was counted in similar fashion but at ×200 magnification. Ki-67 immunoreactivity was expressed as the percentage of Ki-67–positive cells to the total tumor cells per field.

**H-scoring of VEGF and VEGFR2 immunoreactivity**

For semiquantification of VEGF and VEGFR2 immunoreactivity, H-scores were independently generated by 2 of the authors (O.T. and A.B.) who were blinded as to treatment group as described previously (41), with slight modification. H-scores were based on findings from up to 4 randomly selected fields for each tumor section at ×100 magnification (60% center field). Staining intensity was graded as undetectable (0), weak (1), medium (2), or strong (3), and the percentage of positive cells per field was calculated. The intensity score and the percentage of positive cells were then multiplied to give an H-score (possible range, 0–300).

**Immunofluorescence quantification**

Dual fluorescent staining for endothelial cells (CD31, red), activated VEGFR2/3 (green), and tumor cell nuclei (blue) were completed as described above. The expression of activated VEGFR2/3 in tumor-associated endothelial cells was identified by colocalized yellow fluorescence. The pixel
areas of green, blue, red, and yellow were quantified using Image Pro Plus (Media Cybernetics, Inc.) in up to 4 random fields for each tumor section at ×200 magnification. Quantification of total activated VEGFR2/3 expression was presented as an index of green area to blue area. Activated VEGFR2/3 expression in endothelial cells was presented as an index of yellow area to red area. All of the quantification data were presented as mean ± SEM.

**Statistical analysis**

Data were analyzed by Prism5 software (GraphPad Software, Inc.). To analyze immunohistochemical and dual-fluorescent findings, the mean value for each tumor was first calculated from captured fields. The Kruskal–Wallis test was then used to assess differences among the treatment and control groups with respect to body weight, tumor volume, left lung weight, and quantitative immunohistochemical and dual-fluorescent findings within treatment groups and between treatment and control groups. We used the Kruskal–Wallis as a gate-keeping procedure (i.e., a protected testing procedure) such that we would not do any pair-wise comparisons unless the Kruskal–Wallis test was significant. This procedure controls the experiment-wise error rate against making at least one false pair-wise inference in the case that the experiment-wise null is true. In addition to this protected procedure, because multiple comparisons were anticipated but use of the Bonferroni correction for all interesting pair-wise comparisons would have resulted in too stringent a P value, we considered a P value of less than 0.007 as statistically significant to account for the multiple comparisons. Using a comparison-wise type 1 error rate of 0.007 controls the experiment-wise type 1 error rate at 0.10. Differences in the incidence of lymph node metastasis or distant metastasis were analyzed by the Fisher exact probability tests, and a P value of less than 0.05 was considered statistically significant.

**Results**

**Selumetinib and cediranib block orthotopic human lung cancer progression in the lung and thorax**

To evaluate the therapeutic efficacy of selumetinib, alone and in combination with cediranib, we used orthotopic models of lung cancer with NCI-H441 adenocarcinoma or NCI-H460 large cell human NSCLC cells in nude mice. All treatments were well tolerated, with no significant differences among groups in body weight. The incidence of tumor formation was 100% after implantation in the left lung for both models (Tables 1 and 2).

In the NCI-H441 human lung adenocarcinoma model (Fig. 1A and Table 1), lung tumors grew within the left lung and spread within the lung and then to the mediastinum. Treatment with selumetinib at both dose levels (12.5 and 25 mg/kg twice a day) inhibited the growth of primary lung tumors by 71% and 82%, respectively, compared with controls. Selumetinib, particularly at the higher dose, was also effective in reducing the incidence of mediastinal adenopathy. Cediranib monotherapy also inhibited primary lung tumor growth by 68% and the incidence of mediastinal adenopathy. The antitumor and antimetastatic effects of each agent were substantially enhanced when they were used in combination.

**Table 1. Inhibition of tumor growth and metastasis by selumetinib and cediranib in an orthotopic model of lung cancer: NCI-H441 human lung adenocarcinoma model**

<table>
<thead>
<tr>
<th>Treatment group</th>
<th>Vehicle only</th>
<th>Paclitaxel (200 μg/wk)</th>
<th>Selumetinib (12.5 mg/kg b.i.d)</th>
<th>Selumetinib (25 mg/kg b.i.d)</th>
<th>Cediranib (3 mg/kg/d)</th>
<th>Selumetinib (25 mg/kg b.i.d) + cediranib</th>
</tr>
</thead>
<tbody>
<tr>
<td>Variable</td>
<td>Body weight, g (range)</td>
<td>31.2 (28.4–25.1)</td>
<td>33.6 (29.5–37.5)</td>
<td>33.1 (21.8–36.8)</td>
<td>32 (26.3–34.5)</td>
<td>31.6 (30.2–36.2)</td>
</tr>
<tr>
<td>Tumor incidence</td>
<td>13/13</td>
<td>10/10</td>
<td>11/11</td>
<td>10/10</td>
<td>11/11</td>
<td>10/10</td>
</tr>
<tr>
<td>Left lung weight, mg</td>
<td>560 (300–710)</td>
<td>390 (50–990)</td>
<td>335 (100–710)</td>
<td>280 (100–490)</td>
<td>245 (90–610)</td>
<td>150 (90–280)</td>
</tr>
<tr>
<td>Mediastinal adenopathy</td>
<td>10/13</td>
<td>5/9</td>
<td>5/10</td>
<td>3/11</td>
<td>4/10</td>
<td>0/11</td>
</tr>
</tbody>
</table>

**NOTE:** Data are presented as medians and ranges except for incidence.

aP < 0.007.

bP < 0.001.

bP < 0.05 vs. vehicle.
were combined with a 90% reduction in median primary lung tumor volume, 73% reduction in median left lung weight, and near complete suppression of mediastinal lymph node metastasis. Treatment with paclitaxel reduced primary lung tumor volume by 74% but with only modest effects upon mediastinal adenopathy that were not statistically significant.

Similar results were observed in the NCI-H460 human large cell lung cancer orthotopic model (Fig. 1B, Table 2). In the NCI-H460 model, lung tumors grew within the left lung and spread within the lung and then to the mediastinum and also to chest wall of the left hemithorax. Paclitaxel treatment was only marginally effective in the NCI-H460 model, as compared with the NCI-H441 model. Selumetinib, at the lower dose, reduced primary lung tumor volume by 65% and the total tumor volume by 71%, as compared with control, but did not substantially reduce the incidence of mediastinal lymph node metastasis. At the higher dose, selumetinib reduced primary lung tumor volume by 90%, total tumor volume by 92%, and decreased the incidence of mediastinal lymph node metastasis. Cediranib monotherapy was also efficacious and reduced primary tumor volume by 78% and total tumor volume by 84%, but had only modest effect upon mediastinal lymph node metastasis, whereas distant metastasis was completely inhibited. The antitumor and antimitastatic effects of each agent were substantially enhanced when selumetinib and cediranib were combined with a reduction in primary lung tumor volume by 96% and total lung tumor volume by 97% and the near complete suppression of lymph node metastasis.

**Selumetinib and cediranib inhibit tumor cell proliferation and increase tumor cell apoptosis in lung tumors**

To characterize the mechanism of tumor growth inhibition observed in both of our lung cancer models by selumetinib and cediranib, lung tumors were subjected to immunohistochemical analyses. Lung tumors from each of the different treatment groups and for each of the 2 lung cancer models were assessed for evidence of tumor cell proliferation and increase tumor cell apoptosis. To determine whether selumetinib and cediranib increased tumor cell apoptosis, as determined by staining for cleaved caspase-3.

Table 2. Inhibition of tumor growth and metastasis by selumetinib and cediranib in an orthotopic model of lung cancer: NCI-H460 human large cell lung cancer model

<table>
<thead>
<tr>
<th>Treatment Group</th>
<th>Vehicle only</th>
<th>Paclitaxel (200 μg/wk)</th>
<th>Selumetinib (12.5 mg/kg b.i.d)</th>
<th>Cediranib (25 mg/kg b.i.d)</th>
<th>Selumetinib (25 mg/kg b.i.d) + cediranib</th>
</tr>
</thead>
<tbody>
<tr>
<td>Variable</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Body weight, g</td>
<td>29.9 (24.3–32.2)</td>
<td>29.5 (23.2–33.7)</td>
<td>31.0 (21.2–33.2)</td>
<td>30.6 (25.1–33.3)</td>
<td>30.0 (26.4–33.2)</td>
</tr>
<tr>
<td>(range)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tumor incidence</td>
<td>10/10</td>
<td>10/10</td>
<td>10/10</td>
<td>9/9</td>
<td>10/10</td>
</tr>
<tr>
<td>weight, mg</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>volume, mm³</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total tumor</td>
<td>805 (323–1649)</td>
<td>524 (218–1470)</td>
<td>232 (162–497)</td>
<td>66 (25–201)</td>
<td>131 (89–318)</td>
</tr>
<tr>
<td>volume, mm³</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mediastinal nodal</td>
<td>10/10</td>
<td>9/10</td>
<td>9/10</td>
<td>4/10</td>
<td>6/9</td>
</tr>
<tr>
<td>metastasis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Distant</td>
<td>7/10</td>
<td>4/10</td>
<td>0/10</td>
<td>0/10</td>
<td>0/10</td>
</tr>
<tr>
<td>metastasis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

NOTE: Data are presented as medians and ranges except for incidence.

- $P < 0.001$
- $P < 0.007$
- $P < 0.05$
- $P < 0.005$ vs. vehicle.
Figure 1. Antitumor effects of selumetinib, cediranib, and paclitaxel in orthotopic models of human NSCLC in mice. * $P < 0.007$ or ** $P < 0.001$ versus vehicle control or between groups as indicated (Kruskal–Wallis followed by Mann–Whitney U tests). A, primary tumors in the left lungs of representative mice from each treatment group after implantation of NCI-H441 human lung adenocarcinoma cells (circled), with mean primary tumor lung volumes and left lung weights. Bars, SEM. B, primary tumors in the left lungs (circled) and chest walls of representative mice from each treatment group after implantation of NCI-H460 human lung large cell cancer cells, with mean total tumor volumes (primary lung tumor + chest wall tumors) and left lung weights. Bars, SEM. The dotted horizontal line indicates the normal left lung weight.
Selumetinib and cediranib inhibit activation of VEGFR2/3 in NCI-H441 and NCI-H460 primary lung tumors and their tumor-associated endothelium

As outlined above, MEK inhibition by selumetinib results in the suppression of angiogenesis in orthotopic NCI-H441 lung adenocarcinomas and NCI-H460 large cell lung cancers that is due, at least in part, to a decreased VEGF expression in lung tumors. To further elucidate the effects of treatment upon VEGF-dependent lung tumor angiogenesis, we characterized VEGFR signaling in both tumor cells and in the tumor vasculature regardless of which lung cancer cell line was used. These data show that treatment with the MEK inhibitor selumetinib can offset VEGF expression in 2 distinct orthotopic lung cancer models and suggest that the antiangiogenic effects of treatment with selumetinib in these models is in part due to this decreased expression.

Selumetinib and cediranib inhibit activation of VEGFR2/3 in NCI-H441 and NCI-H460 primary lung tumors and their tumor-associated endothelium

As outlined above, MEK inhibition by selumetinib results in the suppression of angiogenesis in orthotopic NCI-H441 lung adenocarcinomas and NCI-H460 large cell lung cancers that is due, at least in part, to a decreased VEGF expression in lung tumors. To further elucidate the effects of treatment upon VEGF-dependent lung tumor angiogenesis, we characterized VEGFR signaling in both tumor cells and in the tumor vasculature regardless of which lung cancer cell line was used. These data show that treatment with the MEK inhibitor selumetinib can offset VEGF expression in 2 distinct orthotopic lung cancer models and suggest that the antiangiogenic effects of treatment with selumetinib in these models is in part due to this decreased expression.
Figure 2. Apoptotic and antiproliferative effects of selumetinib, cediranib, and paclitaxel in orthotopic models of human NSCLC in mice. Lung tumors were collected 2 hours after the last dose of selumetinib, 20 hours after the last dose of cediranib, and 6 days after the last dose of paclitaxel. Quantitative values were determined in 4 random fields for each tumor. Data are presented as means ± SEM for 9 to 13 samples per group. Scale bar, 100 µm. *, P < 0.007; **, P < 0.001 versus vehicle control or between groups as indicated (Kruskal–Wallis followed by Mann–Whitney U tests). A, immunohistochemical analysis of cleaved caspase 3 in tumors from mice implanted with NCI-H441 or NCI-H460 cells. Representative cleaved caspase-3 staining (brown staining in positive cells) in lung tumors viewed at ×200 magnification. Tumor cell apoptosis (number of cleaved caspase-3 positive cells per field) is shown. B, immunohistochemical analysis of Ki-67 staining in tumors from mice implanted with NCI-H441 or NCI-H460 cells. Representative Ki-67 staining of lung tumors (brown) viewed at ×100 magnification, 60% center field. Tumor proliferation (% of Ki-67–positive cells per total tumor cells) is shown.
MEK inhibition by selumetinib results in a decrease in VEGFR activation in lung tumors that is associated with an antiangiogenic effect in lung tumors in 2 distinct lung cancer models.

Discussion

The outcome for patients with advanced lung cancer has not changed substantially over the past several years but recent advances show that novel biologically targeted therapies can improve the outcomes for subsets of lung cancer patients. However, it has also become apparent that individual agents will need to be combined if the outcomes for lung cancers are to be more broadly improved. In this study, we used orthotopic models of human lung adenocarcinoma and large cell lung cancer that closely mimic clinical patterns of lung cancer spread and progression to investigate antiangiogenic therapy directed against VEGFR signaling with cediranib and molecularly targeted therapy directed against MEK signaling with selumetinib alone and in combination. To our knowledge, this is the first report of the effects of MEK inhibition with antiangiogenic therapy in murine orthotopic models of NSCLC. We found that each agent was effective for the treatment of lung cancer in these models with inhibition of lung tumor growth and, to a lesser degree, lymph node metastasis with efficacy superior to that observed for chemotherapy with paclitaxel. When selumetinib and cediranib were combined, a substantial enhancement of their individual antitumor effects was observed with improved efficacy within the lung and a near complete suppression of lung cancer progression and metastasis in both models. Our finding that the combination of these agents impacted both primary tumor and metastatic growth most effectively has direct clinical relevance. Surprisingly, MEK inhibition by selumetinib also suppressed lung tumor angiogenesis and targeted both VEGF production and VEGFR activation in lung tumors, resulting in substantial antiangiogenic effects.
MEK is an attractive therapeutic target for lung cancer treatment because it is situated downstream of Ras and Raf, which are highly activated in Kras-mutated lung cancer (43). Many Kras-mutant cancer cells have been shown to be sensitive to MEK inhibitors, (44) and Kras mutations can be detected in up to 30% of lung cancers, dependent upon histology and ethnicity (45, 46), suggesting that a subset of lung cancers would likely be highly sensitive to selumetinib. Our finding that selumetinib was effective in 2 distinct Kras mutant human lung cancer models supports and validates this hypothesis. Although monotherapy with selumetinib resulted in antitumor and some antimetastatic effects in both of our lung cancer models, the antimetastatic effects were more apparent in the NCI-H441 lung adenocarcinoma model. The increased antimetastatic efficacy observed in this model is associated with differences in the constitutive expression and activation of ERK in NCI-H460 and NCI-H441 lung tumors. Both cell lines have KRAS mutations with activation of ERK for lung tumors from both cell lines. However, activated ERK was nearly twice as high in NCI-H460 lung tumors, as compared with NCI-H441 lung tumors, and NCI-460 cells are PI3KCa and LKB1 mutant, both of which might provide a degree of resistance to MEK inhibition. In our studies, a lower dose of selumetinib inhibited ERK activation almost completely in the NCI-H441 model but by only 46% in the NCI-H460 cells. These findings underscore the importance of MEK signaling in lung cancer progression. However, additional studies are needed to determine whether molecular profiling of lung cancer specimens could be of use to select patients who might best benefit from therapy with selumetinib and to help tailor the dosing of this agent.

Selumetinib was a potent inhibitor of lung tumor angiogenesis in our orthotopic models, and the addition of selumetinib to cediranib resulted in a marked enhancement of their individual antiangiogenic effects. Interestingly, selumetinib reduced the production of VEGF in the lung tumors, particularly in the NCI-H441 model. The finding...
Figure 5. Effects of selumetinib, cediranib, and paclitaxel upon VEGF and VEGFR expression in orthotopic models of human NSCLC in mice. Lung tumors were collected 2 hours after the last dose of selumetinib, 20 hours after the last dose of cediranib, and 8 days after the last dose of paclitaxel. Quantitative values were determined in 4 random fields for each tumor. Data are presented as means ± SEM for 9 to 13 samples per group. Scale bar, 100 μm. *P < 0.007; **P < 0.001 versus vehicle control or between groups as indicated (Kruskal–Wallis followed by Mann–Whitney U tests). A, immunohistochemical analysis of VEGF in lung tumors from mice implanted with either NCI-H441 or NCI-H460 cells. Representative VEGF staining (brown cytoplasmic staining) in lung tumors and H-scores for VEGF expression are shown. B, immunohistochemical analysis of VEGFR2 in lung tumors from mice implanted with either NCI-H441 or NCI-H460 cells. Representative VEGFR2 staining (brown cytoplasmic staining) in lung tumors and H-scores for VEGFR2 expression are shown.

www.aacrjournals.org Clin Cancer Res; 18(6) March 15, 2012

MEK1/2 and VEGFR Inhibition in a Mouse Model of Lung Cancer

Published OnlineFirst January 24, 2012; DOI: 10.1158/1078-0432.CCR-11-2324

Downloaded from clincancerres.aacrjournals.org on April 14, 2017. © 2012 American Association for Cancer Research.
that MEK inhibits VEGF expression is consistent with studies showing that VEGF expression is downregulated after EGFR inhibition (47) and provides additional mechanism for this process. In vitro studies using head and neck cancer cell lines show that the VEGF expression after EGFR activation is dependent upon both PI3K and MAPK signaling (48, 49). The MEK inhibitor PD0325901 decreased the expression of the proangiogenic factors VEGF and interleukin 8 in vitro in human melanoma cells (50). Prior studies in a murine model of hepatocellular carcinoma showed that the antitumor and antiangiogenic effects of rapamycin (51) or sorafenib (52) could be enhanced by the addition of selumetinib, and that the combination of these agents was associated with modest inhibition in VEGFR signaling in liver tumor lysates with reduced circulating levels of VEGF (51). In pancreatic cancer subcutaneous xenograft murine models, MEK inhibition by selumetinib, but not rapamycin, resulted in decreased microvessel density in the subcutaneous tumors and decreased VEGF levels in tumor lysates (53).

Figure 6. Effects of selumetinib, cediranib, and paclitaxel upon VEGFR signaling in orthotopic models of human NSCLC in mice. CD31/pVEGFR2/3 dual fluorescent staining in tumors from mice implanted with either NCI-H441 or NCI-H460 cells was completed. Representative colocalized CD31/pVEGFR2/3 staining of lung tumors viewed at 200 magnification are shown. Fluorescent red indicates CD31-positive endothelial cells; fluorescent green, total pVEGFR2/3-positive cells; fluorescent yellow, pVEGFR2/3-positive endothelial cells. Quantification of total activated VEGFR2/3 expression is presented as an index of green area to blue area. Activated VEGFR2/3 expression in endothelial cells is presented as an index of yellow area to red area. All quantification data are presented as means ± SEM for 9 to 13 samples per group. Scale bar, 50 µm. **,**,* P < 0.007 or **,**,* P < 0.001 versus vehicle control or between groups as indicated (Kruskal–Wallis followed by Mann–Whitney U tests).

Tumor lysates from Calu-6 lung cancer intradermal xenografts in mice-treated selumetinib also showed decreases in VEGF levels (54). From these reports and the findings of this study, we surmise that selumetinib exerts an antiangiogenic effect in lung tumors by directly and indirectly targeting VEGF and its receptors.

The downregulation of VEGF expression that we observed in NCI-H441 lung adenocarcinomas after therapy with selumetinib also showed decreases in VEGF levels (54). From these reports and the findings of this study, we surmise that selumetinib exerts an antiangiogenic effect in lung tumors by directly and indirectly targeting VEGF and its receptors.

The downregulation of VEGF expression that we observed in NCI-H441 lung adenocarcinomas after therapy with selumetinib also showed decreases in VEGF levels (54). From these reports and the findings of this study, we surmise that selumetinib exerts an antiangiogenic effect in lung tumors by directly and indirectly targeting VEGF and its receptors.
antiangiogenic effects, and that the inhibition of MEK by selumetinib may have both direct and indirect effects upon VEGF signaling with a resultant multicentric antiangiogenic effect. Prior studies in subcutaneous tumor xenograft and in vitro organotypic angiogenesis assays have shown that the expression of dominant-negative MEK1 in the tumor vasculature results was associated with antivascular effects, and that ERK–MAPK signaling promotes endothelial cell survival sprouting with downregulation of Rho-kinase activity (55). Further investigation is needed to clarify the mechanism by which selumetinib inhibits angiogenesis, but our data show that MEK inhibition targets tumor angiogenesis with a multicentric effect.

In summary, our study is, to our knowledge, the first evaluation of therapy directed against MEK in combination with anti-VEGF therapy in orthotopic models of NSCLC. MEK inhibition resulted in potent antiangiogenic effects for lung cancers mediated by downregulation of VEGF expression and impaired VEGF signaling. We have further shown that selumetinib or cediranib can significantly inhibit tumor angiogenesis and lung cancer growth and progression with increased tumor cell apoptosis in our orthotopic models. Combining selumetinib with cediranib enhanced their antitumor and antiangiogenic effects, with near-complete suppression of lung tumor growth and metastasis. We conclude from these findings that the combination of selumetinib and cediranib represents a promising strategy for the treatment of NSCLC and provides a strong basis for the design of clinical trials for this purpose.

Disclosure of Potential Conflicts of Interest

P.D. Smith, J.M. Jurgensmeyer, and A. Ryan are or were AstraZeneca employees. R.S. Herbst and M.S. O’Reilly received research funding from AstraZeneca and formerly served on AstraZeneca Advisory Boards. The other authors disclosed no potential conflicts of interest.

Acknowledgments

The authors thank Christine F. Wogan of MD Anderson’s Division of Radiation Oncology for editorial review and comments.

Grant Support

This work was supported by the NIH through MD Anderson’s Cancer Center Support grant CA016672 and by a research grant from AstraZeneca. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received September 8, 2011; revised January 1, 2012; accepted January 5, 2012; published OnlineFirst January 24, 2012.

References


23. Takahashi et al.


Combined MEK and VEGFR Inhibition in Orthotopic Human Lung Cancer Models Results in Enhanced Inhibition of Tumor Angiogenesis, Growth, and Metastasis

Osamu Takahashi, Ritsuko Komaki, Paul D. Smith, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-11-2324

Cited articles
This article cites 55 articles, 22 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/18/6/1641.full.html#ref-list-1

Citing articles
This article has been cited by 4 HighWire-hosted articles. Access the articles at:
/content/18/6/1641.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.