**RRM2 Regulates Bcl-2 in Head and Neck and Lung Cancers: A Potential Target for Cancer Therapy**

Mohammad Aminur Rahman1, A.R.M. Ruhul Amin1, Dongsheng Wang1, Lydia Koenig1, Sreenivas Nannapaneni1, Zhengjia Chen1, Zhibo Wang2, Gabriel Sica3, Xingming Deng4, Zhuo (Georgia) Chen1, and Dong M. Shin1

**Abstract**

**Purpose:** Ribonucleotide reductase subunit M2 (RRM2) plays an active role in tumor progression. Recently, we reported that depletion of RRM2 by systemic delivery of a nanoparticle carrying RRM2-specific siRNA suppresses head and neck tumor growth. The aim of this study is to clarify the underlying mechanism by which RRM2 depletion inhibits tumor growth.

**Experimental Design:** siRNA-mediated gene silencing was carried out to downregulate RRM2. Immunoblotting, reverse-transcriptase PCR, confocal microscopy, tissue fractionation, gene overexpression and knockdown were employed to analyze critical apoptosis signaling. Conventional immunohistochemistry and quantum dot-based immunofluorescence were applied to detect RRM2 and Bcl2 expression and localization in tissue samples from patients and mice.

**Results:** Knockdown of RRM2 led to apoptosis through the intrinsic pathway in head and neck squamous cell carcinoma (HNSCC) and non-small cell lung cancer (NSCLC) cell lines. We showed that Bcl-2 is a key determinant controlling apoptosis, both in vitro and in vivo, and that RRM2 depletion significantly reduces Bcl-2 protein expression. We observed that RRM2 regulates Bcl-2 protein stability, with RRM2 suppression leading to increased Bcl-2 degradation, and identified their colocalization in HNSCC and NSCLC cells. In a total of 50 specimens each from patients with HNSCC and NSCLC, we identified the colocalization of Bcl-2 and RRM2 and found a significant positive correlation between their expression in HNSCC (R = 0.98; P < 0.0001) and NSCLC (R = 0.92; P < 0.0001) tumor tissues.

**Conclusions:** Our novel findings add to the knowledge of RRM2 in regulating expression of the antiapoptotic protein Bcl-2 and reveal a critical link between RRM2 and Bcl-2 in apoptosis signaling. *Clin Cancer Res; 19(13); 3416–28. ©2013 AACR.*

**Introduction**

Ribonucleotide reductase (RR) is a potential therapeutic target for cancer because it catalyzes the conversion of ribonucleoside 5’-diphosphates into their corresponding 2’deoxyribonucleoside 5’-triphosphates that are necessary for DNA replication and repair (1). Human RR comprises two subunits, RRM1 and RRM2. The cellular RRM1 protein level is relatively stable through the entire cell cycle, whereas RRM2 is only expressed during the late G1–early S-phase when DNA replication occurs (2, 3). Because the RRM2 subunit is the primary regulator of RR enzymatic activity (1), studies of its influence on the biologic activities of the RR protein are highly sought. RRM2 was reported to play an active role in tumor progression (4, 5) and was identified as an indicator of poor patient outcome in several cancer types (6, 7). RRM2 is known to be involved in chemoresistance. Suppression of RRM2 can sensitize colon and pancreatic cancer cells to gemcitabine and DNA damaging agents for which RRM2 plays a role in the relevant DNA synthesis and repair mechanisms (3, 8). RRM2 serves as a prognostic biomarker of many cancer types (7, 9–11) and regulates several oncogenes that control malignant potential (12).

In recent years, several inhibitors (i.e., triapine, gemcitabine, and GTI-2040) have been studied as potential inhibitors of RRM2 activity and entered in clinical trials. Shao and colleagues reviewed several potential inhibitors of RR (13). Triapine and GTI-2040, potent inhibitors of RR, have been evaluated in phase I clinical trials (14, 15). However,
Bcl-2 activity is regulated by various mechanisms, including therapy, and bias toward cancer cell survival (24–28). Typically, Bcl-2 overexpression is associated with poor prognosis for patients with cancer. In this study, we unravel the mechanistic details linking RRM2 to apoptosis and provide vital clues for the development of novel cancer therapies. We show a new molecular regulatory pathway wherein RRM2 contributes to the accumulation and stabilization of Bcl-2. RRM2 was found to be significantly involved in regulating Bcl-2, and a reciprocal regulation between these proteins in apoptosis signaling was observed. Furthermore, expression of both proteins was positively correlated in tumors from patients with both HNSCC and NSCLC. Our results have broader implications, suggesting that suppression of Bcl-2 by targeting RRM2 could provide an effective therapeutic strategy, especially for Bcl-2-mediated apoptosis-resistant cancer types. Given the correlation between RRM2 and Bcl-2 expression levels, the expression level of RRM2 may serve as a predictive marker of Bcl-2 level. We present RRM2 as an attractive interventional target to downregulate Bcl-2, resulting in the induction of mitochondria-mediated intrinsic apoptosis. This finding provides a strong framework for translation of this approach to the clinic.

**Translational Relevance**

Elevated levels of ribonucleotide reductase subunit M2 (RRM2) are correlated with poor prognosis for patients with cancer. In this study, we uncover a novel molecular mechanism in which Bcl-2 is a key determinant controlling intrinsic apoptosis triggered by RRM2 depletion, both in vitro and in vivo. To clinically validate our novel findings, we used quantum dot–immunofluorescence (QD–IHF) and conventional immunohistochemistry (IHC) techniques to stain RRM2 and Bcl-2 proteins in tumor tissue samples from patients with HNSCC and NSCLC. We confirmed the colocalization of Bcl-2 and RRM2 and found a significant positive correlation between their expressions in these tumor tissues. Our results show a new function for RRM2 in regulating expression of the antiapoptotic protein Bcl-2 and reveal a critical link between RRM2 and Bcl-2 in apoptosis signaling.

**Materials and Methods**

**Cell lines**

HNSCC cell lines Tu212 and Tu686 (provided by Dr. Gary L. Clayman, University of Texas MD Anderson Cancer Center) and NSCLC cell lines A549, H1299, and H358 were kindly provided by Dr Shi-Yong Sun (Emory University, Atlanta, GA). sh p53 in lentivirus vector was a generous gift from Dr. Didier Trono, Ecole Polytechnique Federale de Lausanne, Switzerland, and A549-expressing sh p53 cell line was established as described (33). The dominant-negative p73 plasmid generation and validation is described elsewhere (34). pWZL-NeoAKT plasmid (constitutively active) was obtained from Addgene, and pLNCX-Bcl-2 plasmid was obtained from the laboratory of Dr. M.W. Jackson (Case Western Reserve University, Cleveland, Ohio). Bcl-2 overexpressing Tu212 and Tu686 (pool) cell lines were generated by retroviral transduction of Bcl-2 and selection by G418. Constitutively active Akt was overexpressed in A549 and Tu686 cell lines (G418 selected pool).

**Western blotting, coimmunoprecipitation, and cell fractionation**

Western blot was incubated with primary followed by secondary antibodies and detected using an enhanced chemiluminescence system as described (35). For coimmunoprecipitation, cells were lysed in a Chaps cell extract buffer (Cell Signaling Technology). Lysates were centrifuged at 15,000 rpm for 20 minutes at 4°C, and supernatants were incubated with primary antibodies for 4 hours and coupled to protein A/G-agarose (Pierce) for 2 hours at 4°C. Immunoprecipitates were washed 3 times with Chaps buffer and subjected to immunoblotting. Cytosolic and mitochondrial fractionation was carried out using a kit (Clontech). Primary antibodies were anti-RRM2, anti-GAPD, anti-p53, anti-Bcl-2, anti-Bcl-XL, anti-Mcl-1, and anti-Bax from Santa Cruz Biotech; anti-phospho Akt, anti-caspase 9, anti-caspase 3, and anti-poly ADP-ribose polymerase (anti-PARP) from...
Cell Signaling Technology; anti-p73 from Bethyl Laboratories; and anti-β-actin from Sigma Aldrich. Secondary antibodies were from Santa Cruz Biotech. Western band quantification was carried out using Image-Quant TL software (GE/Amersham Biosciences).

siRNA duplexes
Control siRNA (siC) was bioinformatically designed to minimize potential for targeting any human gene (Dharmacon). Unmodified RNA duplex siR2 (RRM2 siRNA) was a gift from Calando Pharmaceuticals. siR2-1 and siR2-2 RNA duplexes were purchased from Qiagen. The sequences are as below:

Control siRNA: 5′-UAGCGACUAAACACAUCAAUU-3′

siR2: 5′-GCGAUUUAGCCAAGAAGUUCA-3′

siR2-1: 5′-GCGAUUUAGCCAAGAAGUUTT-3′

siR2-2: 5′-GGGAUUAAACAGUCCUUUATT-3′

Lipofectamine transfection
Cells were seeded in 6-cm plates 24 hours before transfection in medium containing 10% FBS, so that they reached 30% to 50% confluency. siRNA was complexed with Lipofectamine 2000 (Invitrogen) according to the manufacturer’s instructions and applied to each plate. Transfection media was removed and replaced with new media after 5 hours.

Apoptosis assay
Cells were transfected with different concentrations of siR2 or siC, and cells were collected after 72 hours by trypsinization, washed with cold ×1 phosphate-buffered saline (PBS), and stained with Annexin V-PE and 7-AAD (BD PharMingen) for 15 minutes at room temperature. The samples were measured using a fluorescence-activated cell sorting (FACS) caliber bench-top flow cytometer (Becton Dickinson). FlowJo software (Tree Star) was used for apoptosis analysis.

Reverse-transcriptase PCR
RNA was collected by direct lysis in TRIzol (Invitrogen) and reverse-transcribed using SuperScriptIII reverse transcriptase. Two microliters of prepared sample cDNA was used for PCR. A Bd-2 PCR primer set was purchased from...
Figure 2. Knockdown of RRM2 induces apoptosis via the mitochondria-mediated intrinsic pathway. A. Tu212 (left) and A549 (right) cells were transfected with siC or siR2. After 72 hours, cell lysates were analyzed by Western blotting with the indicated antibodies. A single set of blots is pictured from three independent experiments. B. Tu212 (top) and A549 (bottom) cells were transfected with siC or different siRNAs against RRM2, siR2, siR2-1, and siR2-2. After 72 hours, cell lysates were analyzed by Western blotting with the indicated antibodies. A single set of blots is pictured from three independent experiments. C. mitochondrial fraction (MF) and cytosolic fraction (CF) of Tu212 and A549 cells were isolated 72 hours after transfection with 5 nmol/L siR2, 5 nmol/L siC, or no treatment (NT). Western blotting was conducted to detect Cytochrome C and Cox-4.

Immunohistochemistry

To detect intracellular localization and expression levels of RRM2 and Bcl-2, we used goat anti-human R2 polyclonal antibody and mouse anti-human Bcl-2 monoclonal antibody (Santa Cruz Biotechnology) respectively, as primary antibodies, then conjugated to secondary antibody, and cell nuclei were counterstained using 4,6-diamidino-2-phenylindole (DAPI, Invitrogen). Mouse and rabbit immunoglobulin G (IgG) were used as negative controls. All the tissue slides were evaluated and scored by a qualified pathologist. All the tissue slides were evaluated and scored by a qualified pathologist.

Confocal microscopy

Cells were grown on glass coverslips (Lab-Tek II chamber slide, Nunc International), fixed in 4% paraformaldehyde for 15 minutes, and permeabilized with 0.5% Triton X-100 in PBS for 5 minutes. The cells were exposed consecutively to primary antibodies (anti-RRM2 and anti-Bcl-2) and rhodamine-conjugated or fluorescein isothiocyanate (FITC)-conjugated secondary antibodies, before mounting with ProLong Gold antifade reagent with DAPI (Invitrogen). A Zeiss LSM 510 confocal scanning microscope was used to collect the images.

Protein half-life

Tu212 cells in 100-mm dishes were transfected with siC or siR2. After 24 hours, the cells were reseeded into 60-mm dishes and allowed to attach for 24 hours. The cells were treated with 100 μg/mL cyclohexamide 5 minutes before starting the time course (0, 1, 3, 6, 9, and 12 hours), and cell lysates were collected at the indicated time points.

Tissue specimens

This study was approved by the Institutional Review Board at Emory University, and analyses were conducted using de-identified data in compliance with the Health Insurance Portability and Accountability Act. Fifty HNSCC and 50 NSCLC tissues (30 adenocarcinoma, 18 squamous cell carcinoma, and 2 others) were randomly collected for the study from surgical specimens of patients who were diagnosed with HNSCC or NSCLC at Emory University Hospital and whose initial treatment was surgery without prior treatment with radiation and/or chemotherapy. This was a retrospective study which used tissue samples from surgical specimens dated before April 14, 2003.

QD-based immunohistofluorescence

QD staining was conducted on paraffin-embedded formalin-fixed NSCLC and HNSCC tissue sections overnight at 4°C as described previously (36). Primary antibodies were mouse anti-human Bcl-2 at 1:50 dilution and goat anti-human R2 at 1:100 (both from Santa Cruz Biotechnology) in a mixed solution. PBS alone was used as a negative staining control. Tissues were incubated with QD-secondary antibody conjugates [QD 605 goat F(ab)2 anti-mouse IgG; QD 655 goat F(ab)2 anti-goat IgG, 1:50 dilution, Invitrogen] in a mixed solution at 37°C for 2 hours. Cell nuclei were counterstained using DAPI.
QD spectral imaging and signal quantification

QD imaging and quantification procedures were described previously (36). All cubed image files were collected from patient tissue slides at 10-nm wavelength intervals from 450 to 750 nm, with an auto exposure time per wavelength interval at ×200 magnification. Taking the cube with a long wavelength band pass filter allowed transmission of all emission wavelengths above 450 nm. Both separated and combined QD images were obtained after establishing the QD spectral library and unmixing the image cube. For each slide, 5 cubes were taken. The background signal was removed for accurate quantification of the QD signals. The average signal of each QD signal was obtained by selecting 4 to 10 tumor areas in each cube for measurements by the Nuance imaging software (Caliper/PerkinElmer). Measurements were averaged from the 5 cubes from each slide to obtain a total average signal count on each slide for both QD signals.

Statistical analysis

All results represent the average of at least three separate experiments and are expressed as mean ± SD. Statistical analysis was conducted using t test. P less than 0.05 was considered statistically significant. Spearman correlation coefficient was estimated to measure the relationship between Bcl-2 and RRM2. A linear regression was used to plot their relationship with 95% CI bound.

Results

Knockdown of RRM2 promotes apoptosis in HNSCC and NSCLC cells

Our previous observation of cell growth inhibition by RRM2-targeting siRNA (siR2) (16) prompted us to further investigate the mechanistic details of RRM2’s action. Upon knockdown of RRM2 in Tu212 (HNSCC) and A549 (NSCLC) cells (Fig. 1A and B, Inset), we observed...
apoptosis induction in a dose-dependent manner. After 72 hours, 5 nmol/L siR2 induced apoptosis (Tu212 ~43% and A549 ~60%; Fig. 1A and B), whereas no significant apoptotic effects were observed at an early time (24 to 48 hours; data not shown). The target specificity of siR2 mRNA was confirmed previously (16, 37–39). Two other RRM2 siRNAs, siR2-1 and siR2-2, also effectively knocked down RRM2 and induced apoptosis in Tu212 and A549 cell lines after 72 hours (Fig. 1C and D), arguing against any nonspecific effects. In all experiments, nontargeting siRNA (siC) was used as a negative control.

Knockdown of RRM2 promotes apoptosis by inducing the mitochondria-mediated intrinsic apoptosis pathway in both HNSCC and NSCLC cells

We explored whether intrinsic or extrinsic apoptosis occurs following siR2-mediated depletion of RRM2 in Tu212 and A549 cell lines. Our results showed the efficient triggering of the apoptotic cell signaling cascade in Tu212 and A549 cell lines following siR2 treatment (Fig. 2A). Western blot analysis revealed that suppression of RRM2 induced apoptotic signaling molecules mainly involved in intrinsic apoptosis (17, 40–42), such as cleaved caspase 9 (initiator caspase) and cleaved caspase 3 (effector caspase), followed by cleavage of substrate PARP. We did not detect any induction of extrinsic apoptosis signaling molecules (DR5, caspase 8) by RRM2 suppression (data not shown). We observed caspase 3 activation and PARP cleavage in both Tu212 and A549 cells following treatment with siR2-1 and siR2-2 siRNAs but less effectively than with siR2 (Fig. 2B). In the intrinsic apoptosis pathway, highly controlled mitochondrial outer membrane permeabilization is the crucial event driving initiator caspase activation and apoptosis (17). The release of cytochrome C from mitochondria has been considered a key step in the early stages of apoptosis (17, 41). We isolated mitochondrial and cytoplasmic fractions from cell lysates of Tu212 and A549 cells transfected with siR2 or siC and observed cytochrome C release from the mitochondria to cytoplasm only in the siR2-treated group (Fig. 2C). Taken together, our results show that knockdown of RRM2 promotes apoptosis via the mitochondria-mediated intrinsic apoptosis pathway.

Depletion of RRM2 promotes apoptosis in antiapoptotic protein Bcl-2-dependent manner

Evasion of apoptosis has proven to be critical for the development and sustained growth of cancers. Members of the Bcl-2 protein family are critical regulators of apoptosis (40). One approach to inhibiting the apoptotic pathway is through the upregulation of antiapoptotic Bcl-2 family members (43). We determined that RRM2 suppression by siR2, siR2-1, and siR2-2 significantly reduced Bcl-2 expression in Tu212 and A549 cells (Fig. 3A and B), whereas levels of other antiapoptotic Bcl-2 family proteins, Bcl-xL and Mcl-1 remained unchanged in both cell lines following siR2
treatment (Fig. 3A). To explore whether Bcl-2 status could affect the regulation of apoptosis by RRM2, we established a Bcl-2–overexpressing Tu212 cell line (Tu212/Bcl-2; Supplementary Fig. S1A, top). We showed significant resistance to apoptosis upon suppression of RRM2 in the Tu212/Bcl-2 cell line (Fig. 3C). In addition, apoptosis marker molecules such as cleaved caspase 3 and cleaved PARP were triggered by suppressing RRM2 in Tu212 cells but not in Bcl-2–overexpressing cells (Fig. 3D). To confirm our observation, another cell line Tu686/Bcl-2 (Supplementary Fig. S1A, bottom) was tested to determine the importance of Bcl-2, and we similarly found that RRM2-mediated apoptosis was Bcl-2 dependent (Supplementary Fig. S1B). Taken together, our results suggest that the antiapoptotic protein Bcl-2 plays a critical role in siR2-mediated apoptosis induction.

Figure 5. RRM2 regulates Bcl-2 through direct protein–protein interaction and partially influences Bcl-2 mRNA levels 72 hours after transfection of Tu212 and A549 cells with 5 nmol/L siC or siR2. RRM2 was normalized to GAPDH levels within the same sample. B, Tu212 cells were stained with anti-RRM2 (green) and anti-Bcl-2 (red), and nuclei were counterstained with DAPI (blue). A z-stack of optical sections was created at 0.59-μm intervals using a confocal microscope (LSM 510; Carl Zeiss MicroImaging, Inc.). Inset, top shows magnification of white box in merged figure. Inset, bottom shows z-axis reconstructions along the bars indicated in inset top. Bar, 50 μm. C, Tu212 and (D) A549 cells were stained with anti-RRM2 (green), anti-Bcl-2 (red), and nuclei (blue) 48 hours after transfection with 5 nmol/L siC or siR2. Bar, 50 μm. E, Co-immunoprecipitation (IP) with anti-RRM2 (left) or anti-Bcl-2 (right) and immunoblotting with anti-Bcl-2 or anti-RRM2 in Tu212 and A549 cells. IM, immunoblotting.
Apoptosis induction by suppression of RRM2 is p53, p73, and Akt independent

We then extended our study to identify any other signaling molecules involved in the regulation of siR2-mediated apoptosis, particularly p53 and p73, as these are potential candidates in apoptosis induction (44). An elevated level of p53 was observed upon RRM2 suppression in A549 cells (wild-type p53), whereas a sustained level was seen in Tu212 cells [mutant p53 (P151S; ref. 45); Fig. 4A]. To explore the role of p53, we established p53-knockdown A549 cells (A549/sh p53) using siRNA (Fig. 4B). By knocking down RRM2, we found cleaved caspase 3 and PARP induction in both WT-A549 and p53 knocked-down cells (Fig. 4B), suggesting that apoptosis induction by siR2 in A549 cells is p53 independent. To assess the involvement of p73, we determined the percentage of apoptosis and apoptotic marker proteins in the H1299 lung cancer cell line and in two clones of dominant-negative p73-expressing H1299 cells (H1299/dN p73 Cl-7, and Cl-10; Supplementary Fig. S2A, top). We found similar apoptosis induction (Fig. 4C) or PARP modification (Fig. 4D) in all cell lines upon suppression of RRM2. Another cell line, H358/dNp73 (Supplementary Fig. S2A, bottom), was also tested for apoptosis upon RRM2 knockdown, and we observed a similar effect as in H1299 cells (Supplementary Fig. S2B), suggesting that apoptosis induction is also p73 independent. Akt plays a vital role in antiapoptotic signaling (46), and we observed that Akt activation was significantly reduced upon suppression of RRM2 in both Tu212 and A549 cells (Fig. 4E). The role of Akt in RRM2-mediated
apoptosis was evaluated by its overexpression in A549 cells (A549/Akt; Supplementary Fig. S3A, top), and no significant differences in apoptosis or PARP modification were seen compared with unmodified A549 cells (Fig. 4F and G). In the Tu686/Akt cell line (Supplementary Fig. S3A, bottom), we found that knockdown of RRM2-mediated apoptosis is Akt independent (Supplementary Fig. S3B). Taken together, our results suggest that apoptosis induction by silencing of RRM2 is independent of p53, p73, and Akt.

RRM2 regulates Bcl-2 partially at the transcription level, and the two proteins colocalize and interact with each other

To determine whether Bcl-2 is regulated by RRM2 at the transcriptional or the protein level, we transfected cells with siR2 or siC and collected RNA samples after 72 hours for reverse-transcriptase PCR (RT-PCR). RRM2 levels were normalized to GAPDH levels within the same sample. The mRNA expression of Bcl-2 was suppressed to approximately 50% (Fig. 5A), whereas the protein level was suppressed to more than 90% (Fig. 3A) following RRM2 knockdown, indicating that RRM2 most likely regulates Bcl-2 not only at a transcriptional level but also at the posttranslational level. To investigate whether these two proteins colocalize, Tu212 cells were stained with anti-RRM2 (green), anti-Bcl-2 (red), and DAPI (blue). A z-stack of 32 optical sections was created at 0.59-μm intervals using a confocal microscope (LSM 510; Carl Zeiss MicroImaging, Inc.), shown in a gallery of merged images in a single display (Supplementary Fig. S4). We found that RRM2 and Bcl-2 proteins were colocalized with each other (Fig. 5B, yellow color on merge). The white box in the merged figure was magnified (Fig. 5B, inset, top) and the inset lower panel shows the z-axis reconstructions along the bars indicated in the inset top. To assess the impact of RRM2 depletion, Tu212 cells (Fig. 5C) and A549 cells (Fig. 5D) were transfected with siC or siR2, fixed after 48 hours, and stained with anti-RRM2

Figure 7. RRM2 and Bcl-2 proteins colocalize, and their expression is positively correlated in tumor tissues from patients with HNSCC and NSCLC. A and D, staining of RRM2 and Bcl-2 in paraffin-embedded formalin-fixed HNSCC (A) and NSCLC (D) tissue sections using primary antibodies with QD-secondary antibody conjugates. A representative QD-image is shown (∼400 magnification). B and E, quantification of QD signals. Average signals of RRM2 and Bcl-2 expression in each HNSCC (B) and NSCLC (E) patient sample were plotted.
and anti-Bcl-2 (red). Upon silencing of RRM2, Bcl-2 protein was significantly suppressed, confirming our Western blot data showing that RRM2 regulates Bcl-2 protein stability. To show the interaction between these two proteins, coimmunoprecipitation using anti-RRM2 and anti-Bcl-2 antibodies in Tu212 and A549 cell lysates revealed that the proteins indeed existed as a complex (Fig. 5E).

Depletion of RRM2 regulates Bcl-2 protein expression and stability by reducing its half-life

To understand the effect of RRM2 depletion on Bcl-2 protein stability, we transfected Tu212 cells with siC or siR2 for 24 hours. Then, cells were reseeded and allowed to attach for another 24 hours. Cells were treated with cyclohexamide to block protein synthesis, and Bcl-2 expression level was assessed at multiple time points to determine its half-life. As shown in Fig. 6A, knockdown of RRM2 significantly increased the rate of Bcl-2 protein degradation, reducing its half-life from approximately 9 hours to approximately 2 hours. These results show that knockdown of RRM2 causes significant reduction in Bcl-2 protein stability. It is known that Bcl-2 can be cleaved and degraded by activated caspase 3 (47, 48). Thus, the observed Bcl-2 degradation in earlier findings might be a secondary effect due to activation of caspase 3 triggered by siR2. To confirm this, we assessed caspase 3 activation in all samples shown in Fig. 6A. We did not find any caspase 3 activation at early time points (Fig. 6B) of siR2 treatment, whereas Bcl-2 degradation was observed from 1 hour after blockade of de novo protein synthesis in the siR2-transfected group (Fig. 6A). This finding suggests that Bcl-2 degradation triggered by siR2 is not due to caspase 3 activation. To confirm the involvement of the ubiquitin-proteasome pathway in Bcl-2 degradation, we blocked proteasome function by using the proteasome inhibitor MG132 in Tu212 and A549 cell lines. We observed that proteasome inhibition protected Bcl-2 degradation upon RRM2 knockdown, suggesting that RRM2 knockdown leads to Bcl-2 degradation through proteasome (Fig. 6C). We found significant reduction of Bcl-2 protein in tissue samples from mice treated with siR2-nanoparticle (CALAA-01) compared with mice treated with control.
siRNA-nanoparticle (siCON1; n = 8; Fig 6D). Taken together, the findings from our study provide functional evidence that RRM2 regulates Bcl-2 protein stability.

RRM2 and Bcl-2 proteins colocalize, and their expression is positively correlated in tumor tissues from patients with HNSCC and NSCLC

We extended the study to examine the correlation between RRM2 and Bcl-2 protein levels in tissue samples from patients with cancer. A total of 50 cases each of HNSCC and NSCLC (30 adenocarcinoma, 18 squamous cell carcinoma, and 2 others) were analyzed. Clinicopathologic features of the HNSCC and NSCLC patient groups are shown in Supplementary Table S1. QD-IHF analysis allows quantification of multiple biomarkers simultaneously in the same tissue slide (36, 49). In our study, we quantified the expression levels of RRM2 and Bcl-2 in the same tissue slide by using secondary antibody-conjugated QDs with two different emission wavelengths (655 nm and 605 nm). Both separated and combined QD images were established after determining the QD spectral library and unmixing the cube. Representative QD images from patients with HNSCC and NSCLC are shown in Fig. 7A and D, respectively. RRM2 and Bcl-2 were found to be highly colocalized in both types of cancer. Average signal counts (A.U.) of each QD signal were obtained from 5 image cubes of each tissue slide using Nuance imaging software (Supplementary Table S2 and S3). QD signals were plotted by de-identified patient ID.

We extended the study to examine the correlation between RRM2 and Bcl-2 expression in both HNSCC and NSCLC samples (HNSCC: RRM2 and Bcl-2 expression in the same tissue slides from patients with HNSCC and NSCLC tumors. Interestingly, expression of both proteins was positively correlated in both HNSCC and NSCLC tumors.

Because p53 is one of the most commonly mutated genes in cancer and inhibition of RRM2 is known to activate DNA damage (4, 8), we chose cell lines containing wild-type p53 (A549), mutant p53 (Tu212, Tu686), and no p53 (H1299, H358). These cell lines also have variable expression levels of p-AKT and Bcl-2 (54). The Bcl-2 family proteins regulate cell fate through the control of mitochondrial outer membrane potential. Dysregulation of Bcl-2 proteins is common in cancer. The upregulation of antiapoptotic Bcl-2 proteins serves to inhibit proapoptotic Bcl-2 proteins to ensure the survival of abnormal cells. Thus, many malignant cells are primed for cell death and will undergo apoptosis if the function of antiapoptotic proteins is therapeutically impaired (55). Overexpression of Bcl-2 is a marker of resistance to chemotherapy or radiotherapy in squamous cell carcinoma and other types of cancer (24, 26, 27). Targeting cellular processes that regulate Bcl-2 at posttranslational levels is crucial. Our novel findings suggest that targeting RRM2 is a promising approach to reduce Bcl-2 expression and to initiate apoptosis. Only Bcl-2 was identified as a key determinant controlling intrinsic apoptosis triggered by RRM2; we did not observe an increased rate of degradation of other antiapoptotic Bcl-2 family member proteins. Bcl-2 is known to be regulated through posttranslational modifications (56). Here, we show a new molecular regulatory pathway wherein RRM2 contributes to the accumulation and stabilization of Bcl-2. Knockdown of RRM2 significantly reduced the half-life of Bcl-2. We observed that depletion of RRM2 reduced Bcl-2 expression and eventually initiated apoptosis. Bcl-2 protein expression was assessed at multiple time points upon RRM2 knockdown (Fig. 6A). We found that RRM2 knockdown significantly increased Bcl-2 protein degradation (Fig. 6A) but failed to induce caspase-3 activation at those time points but activation was observed only at 72 hours (Fig. 6B). Moreover, we confirmed that RRM2 knockdown leads to Bcl-2 degradation through the proteasome pathway. These observations clearly suggest that Bcl-2 downregulation by RRM2 siRNA precedes caspase-3 activation. In contrast, overexpression of Bcl-2 protected cells from apoptosis triggered by RRM2 knockdown. Our results have broader implications, suggesting that suppression of Bcl-2 by targeting RRM2 could provide an effective therapeutic strategy for Bcl-2-mediated apoptosis.
resistant cancer types. Given the correlation between RRM2 and Bcl-2 expression levels, it is most likely that the expression level of RRM2 could also predict the Bcl-2 level that is involved in cancer cell survival and resistance to chemotherapy. This finding provides a strong framework for translation of this approach to the clinic.

To determine the correlation between RRM2 and Bcl-2, we employed two different techniques, QD-IHF and conventional IHC, for staining proteins in tumor specimens from patients with HNSCC and NSCLC. QD-IHF allows multiple biomarker analysis in the same tissue slide (36, 49), facilitating observation of the colocalization of multiple proteins and also correlations between their expression levels. We found that RRM2 and Bcl-2 were highly colocalized, and a positive correlation was observed by both QD-IHF and conventional IHC techniques.

Knockdown of RRM2 induces DNA damage (1, 4, 8) that could be the cause of p53 and p73 induction. RRM2 knockdown initially inhibits cellular growth, which might be the cause of downregulation of Akt activation. However, we excluded the possibility that activation of p53 or p73 and inhibition of p-AKT are involved in apoptosis using appropriate genetic approaches. Pathways mediating cell death in the absence of p53 are of interest for therapeutic intervention because p53 is mutated in one half of human cancers and inactivated by indirect mechanisms in a large percentage of the remainder (34, 57). Therefore, one particularly interesting indication of our study, that RRM2 siRNA may suppress cancer cell proliferation regardless of p53 status, may be relevant for RRM2-targeted cancer therapy, especially for cancers with mutant p53.

In summary, our data show that alteration of RRM2 induces apoptosis by modulating Bcl-2 expression. RRM2 suppression contributes to the instability of Bcl-2 or leaves Bcl-2 unprotected from degradation. We present RRM2 as an attractive interventional target to downregulate Bcl-2, resulting in the induction of mitochondria-mediated intrinsic apoptosis. Moreover, we confirmed the therapeutic potential of siRNA targeting RRM2 in HNSCC and NSCLC.

We have provided insights into the molecular understanding of tumor progression by RRM2 that could be fundamental in developing rational therapeutic approaches against cancer. Further clinical studies are needed to elucidate the clinicopathologic significance of RRM2 and Bcl-2 expression in HNSCC and NSCLC.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors' Contributions
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): M.A. Rahman, A.R.M. Ruhul Amin, D. Wang, L. Koering, S. Nannapaneni, G.L. Sica, D.M. Shin
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): M.A. Rahman

Acknowledgments
The authors thank Dr. Mark Davis and Dr. Yun Yen for supplying reagents. The authors also thank Dr. Anthea Hammond for her critical and editorial review of this article.

Grant Support
This work was supported by NIH/NCI grant U54 CA119338-04, P50 CA128613 (Head and Neck Cancer SPORE). Drs. D.M. Shin and Z.G. Chen are recipients of Georgia Cancer Coalition (GCC) Distinguished Scholar Awards.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received January 9, 2013; revised April 29, 2013; accepted May 16, 2013; published OnlineFirst May 29, 2013.

References


RRM2 Regulates Bcl-2 in Head and Neck and Lung Cancers: A Potential Target for Cancer Therapy


Updated version

Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-13-0073

Supplementary Material

Access the most recent supplemental material at:
http://clincancerres.aacrjournals.org/content/suppl/2013/05/29/1078-0432.CCR-13-0073.DC1

Cited articles

This article cites 57 articles, 22 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/19/13/3416.full.html#ref-list-1

Citing articles

This article has been cited by 6 HighWire-hosted articles. Access the articles at:
/content/19/13/3416.full.html#related-urls

E-mail alerts

Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions

To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions

To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.