Identification of Lethal microRNAs Specific for Head and Neck Cancer

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Abstract

Purpose: The prognosis of head and neck squamous cell carcinomas (HNSCC) remains disappointing and the development of novel anti-cancer agents is urgently awaited. We identified by a functional genetic screen microRNAs that are selectively lethal for head and neck cancer cells but not for normal cells. We further investigated the genes targeted by these microRNAs.

Experimental Design: A retroviral expression library of human microRNAs was introduced in HNSCC cell lines and normal oropharyngeal keratinocytes to identify tumor-selective lethal microRNAs. Potential downstream gene targets of these microRNAs were identified by gene expression profiling and validated by functional assays.

Results: We identified six microRNAs that selectively inhibit proliferation of head and neck cancer cells. By gene expression profiling and 3'-untranslated region (UTR) luciferase reporter assays, we showed that the ataxia telangiectasia mutated (ATM) gene is a common target for at least two and likely three of these microRNAs. Specific inhibition of ATM resulted in a similar tumor-specific lethal effect, whereas the phenotype was reverted in rescue experiments.

Conclusions: These six microRNAs might be developed as novel anti-cancer agents and highlight ATM as an interesting novel therapeutic target for head and neck cancer. Clin Cancer Res; 19(20); 5647–57. ©2013 AACR.

Introduction

Head and neck squamous cell carcinoma (HNSCC) develops in the mucosal linings of the upper aerodigestive tract and contributes to approximately 5% of all cancers in the Western world (1, 2). Well-known risk factors are tobacco smoking, excessive consumption of alcohol containing beverages, and infection with the human papillomavirus (HPV; refs. 3–5). About one third of the patients present with early-stage tumors and receive single modality treatment, either surgery or radiotherapy. The 5-year survival rates for patients with early disease stages are more than 90%. Unfortunately the majority of patients with HNSCC present with advanced disease stages. These patients are treated with either a combination of surgery and radiotherapy or chemoradiation, the concurrent application of systemic cisplatin chemotherapy combined with locoregional radiotherapy. Patients with advanced disease stages frequently develop locoregional recurrences, distant metastasis, and/or second primary tumors, which results in 5-year survival rates of less than 60% (1). Therefore, the development of novel anti-cancer agents to improve outcome is urgently awaited.

MicroRNAs (mi-RNAs) are about 22 nucleotides long, noncoding RNAs that are able to regulate the expression of multiple target genes (6). Classically, mi-RNAs regulate the expression of target genes through sequence-specific complementarity between the mi-RNA seed sequence and the 3'-untranslated region (UTR) of the target mRNAs (7). Perfect complementarity between the mi-RNA and the mRNA will generally target the mRNA for degradation, whereas imperfect complementarity between the mi-RNA and the mRNA will preferentially repress mRNA translation (8). However, insights on the interaction of mi-RNA with their target genes evolve continuously. Through this regulation at the post-transcriptional level, mi-RNAs are able to modulate the expression of numerous genes simultaneously, thereby regulating individual signaling pathways at multiple levels (9).

Several studies have shown the importance of mi-RNAs in cancer, including HNSCCs. Altered mi-RNA expression profiles have been observed in both HNSCC cell lines and...
tumors when compared to the normal counterpart tissues (10, 11). The overexpression of miR-21 is described in many tumor types, including HNSCCs. Moreover, overexpression of miR-21 also increased with a higher grade of neoplastic lesions during malignant progression (10). A number of these differentially expressed mi-RNAs were shown to be associated with poor prognosis, such as miR-21 and miR-211 (12, 13). Even more interestingly, recent studies have revealed that mi-RNAs may act as tumor suppressors by targeting oncogenes. For example, the miR-16 family inhibits cell-cycle progression and induces apoptosis via the silencing of BCL2 (14), described in multiple tumor types, including HNSCCs. In a similar manner, ectopic expression of miR-181a resulted in decreased proliferation by targeting the oncogene KRAS (15).

In the present study, we examined the potential of mi-RNAs for treatment of HNSCCs. We hypothesized that mi-RNAs might cause tumor-specific cell death in HNSCCs by targeting genes that might show a synthetic lethal interaction with one or more inactivated cancer genes. Several tumor-suppressing routes are inactivated in HNSCCs among which the p53 and pRB pathways. Other genes or signaling routes may take over part of the lost functions and show a synthetic lethal interaction when inhibited (16). We used a human mi-RNA expression library in retroviral vectors to conduct a functional genetic screen to specifically identify mi-RNAs that cause cell death of tumor cells and not of primary keratinocytes. We further investigated, by expression array analysis, the target genes of these mi-RNAs as functional inhibition of the target genes may elicit the same lethal phenotype and could be conducted by small molecules.

Materials and Methods

Cell culture
Normal oral or oropharyngeal keratinocytes were isolated and cultured as previously described (17). Conditionally immortalized oropharyngeal keratinocytes (ciOKC) were generated by transformation of primary oropharyngeal keratinocytes with a temperature-sensitive SV40 large T-antigen. Primary keratinocytes and ciOKCs were cultured in keratinocyte serum-free medium (KSF-M; Invitrogen) supplemented with 0.1% bovine serum albumin, 25 mg bovine pituitary extract, 2.5 μg human recombinant EGF, 250 μg amphotericin B (MP Biomedicals) and 250 μg gentamycin (Sigma-Aldrich) at 32°C (18). VU-SCC-120, VU-SCC-OE, UM-SCC-6, SiHa, MCF7, HT29, U87, and Phoenix cells were cultured in Dulbecco’s Modified Eagles’ Medium (DMEM), 5% fetal calf serum, 2 mmol/L L-glutamine, 50 U/mL penicillin, and 50 μg/mL streptomycin at 37°C and 5% CO2. The HNSCC cell lines used were all negative for the human papillomavirus and were sequenced for TP53 mutations. Cell line UM-SCC-6 was TP53 wild-type, VU-SCC-120 contained 2 missense mutations (c.181G>T and c.527G>A) and VU-SCC-OE a truncating deletion (c.1191del). Cell lines are authenticated regularly by their morphologic characteristics and analysis of TP53 mutations and genetic markers.

Functional screen of human mi-RNA library
Amphotropic retroviral supernatants were produced for 370 annotated and putative mi-RNAs included in the human mi-RNA expression library (mi-R-Lib) with miR-Vec-Ctrl (scrambled sequence) as negative control (18, 19).

RNA isolation from formalin-fixed, paraffin-embedded tissue
Normal oropharyngeal mucosa was derived from 3 formalin-fixed, paraffin-embedded (FFPE) specimens from patients who underwent uvulopalatopharyngoplasty. In addition, FFPE tumor biopsies were obtained from 5 patients with HNSCCs. The mucosal epithelium was microdissected from sections of the uvula specimen as previously described (21). Likewise, neoplastic areas were microdissected from tumor samples. Microdissected tissues were treated with 1 mg/mL of proteinase K for 17 hours at 56°C in buffer containing 100 mmol/L Tris-HCl (pH 9.0), 10 mmol/L NaCl, 1% SDS, and 5 mmol/L EDTA (pH 8.2). Nucleic acids were isolated by phenol/chloroform
extraction and precipitated by sodium acetate and ethanol according to standard protocols using glycogen as carrier. After the nucleic acids were washed with 70% ethanol, they were redissolved in RNase-free water.

**Lentiviral short hairpin RNA ATM transduction**

Lentiviral vectors with short hairpin RNA (shRNA) sequences targeting ATM transcripts were obtained from the MISSION short-hairpin library of The RNAi Consortium (Sigma-Aldrich) that is available at VU University Medical Center. Viral supernatants were produced by cotransfection of HEK239T cells using FuGENE 6 (Roche Diagnostics) with the pLKO.1 short-hairpin vector together with the packaging and envelope vectors. Both ciOKC and VU-SCC-120 (HNSCC cell line) were transduced with lentiviruses at 2 following days for 4 hours in the presence of 3 μg/mL polybrene (Sigma-Aldrich). In total 48 hours after transduction, cells were subjected to puromycin selection. For ciOKC, this was 2 days of 5 μg/mL and subsequently 5 days of 10 μg/mL puromycin (Sigma-Aldrich), and for VU-SCC-120, the selection was conducted with 1 μg/mL for 7 days.

**Quantitative reverse transcription PCR**

Total RNA was isolated using the mirVana mi-RNA Isolation Kit (Ambion) according to the instructions of the manufacturer with the only modification that columns that were eluted with 2 × 25 μL elution buffer. Expression of hsa-miR-181a, hsa-miR-323, hsa-miR-326, hsa-miR-342, hsa-miR-345, and hsa-miR-371 was analyzed by TaqMan microRNA assays following the instructions of the manufacturer (Applied Biosystems). ATM expression was analyzed by TaqMan gene expression assay. Relative expression was calculated via the comparative Ct method using the small nuclear RNA transcript RNU44 (for microRNA analysis) or beta-glucuronidase (BGUS: for ATM analysis) as internal references (22). Quantitative reverse transcription-polymerase chain reactions (qRT-PCR) without reverse transcriptase were carried out in parallel for each RNA sample to exclude signal by contaminating genomic DNA.

**Gene expression profiling**

The retroviral clones with miRNA genes miR-181a, miR-323, miR-326, miR-342, miR-345 and miR-371, and negative control miR-Vec-Ctrl were transiently transfected in VU-SCC-120 by FuGENE 6 (Roche Diagnostics). Total RNA was isolated 72 hours after transfection using the mirVana mi-RNA Isolation Kit (Ambion). Microarray hybridization using the Agilent Low Input Quick Amplification Labeling Kit and 4 × 44K Whole Human Genome Arrays was carried out according to the manufacturer (Agilent Technologies). Normalization of the gene expression data was conducted within the R statistical software using the Limma package and comprised RMA background correction, loess within-array normalization, and A-quantile between-array normalization. Then, missing values were imputed using the impute-package (impute.knn with default settings). Finally, the slide and dye effects were removed by gene-wise linear regression using the log intensity values.

The log fold changes between the reference group and each treated group were used to cluster the 6 treatments. This was carried out by means of hierarchical clustering with ward linkage, and the similarity defined both as the Euclidean distance and as one minus the absolute value of the Spearman rank correlation measure. The grouping from hierarchical clustering was verified by means of principal component plots. Within each cluster, the differences in gene expression between reference and treated samples were evaluated by means of a t test. The multiplicity problem (many genes were tested) was addressed by application of the Benjamini–Hochberg procedure to the raw P values to control the false discovery rate (FDR). Data are accessible under GEO number GSE34881.

**ATM inhibitor treatment**

Both ciOKC and VU-SCC-120 were subjected to a concentration range of 40 to 0.075 μmol/L ATM inhibitor CP466722 (Axon Medchem). After 72 hours, cell viability was assessed with the CellTiter-Blue Cell Viability Assay.

**ATM 3′UTR luciferase reporter assay**

CiOKC cells were transiently cotransfected with a luciferase reporter construct containing the 3′UTR sequence of ATM (GeneCopoeia Inc.) and one of the retroviral vectors containing the miR-181a, miR-323, miR-326 genes, or negative control miR-Vec-Ctrl by FuGENE 6. In total 72 hours after transfection, firefly and Renilla luciferase activity was measured using the LucPair miR Dual Luciferase Assay Kit according to the instructions of the manufacturer (GeneCopoeia Inc.). Luciferase activity was measured using the Infinite 200 Plate Reader (Tecan Group Ltd.).

**Lethal phenotype rescue**

VU-SCC-120 cells were transiently transfected with either pcDNA3.1(+)Flag-His-ATMwt (wild-type ATM cDNA sequence, Addgene plasmid 31985) or pcDNA3.1(+) Flag-His-ATMkd (kinase-dead ATM cDNA sequence, Addgene plasmid 31986; ref. 23). Amphotropic retroviral supernatants were produced for miR-181a, miR-326, miR-345, and miR-Vec-Ctrl (scrambled sequence) as negative control. VU-SCC-120-ATMwt and VU-SCC-120-ATMkd cells were transduced with the microRNA expressing retroviruses at 2 following days for 4 hours in the presence of 3 μg/mL polybrene. After 72 hours, cell viability was assessed by the CellTiter-Blue Cell Viability Assay.

**Results**

**Identification of mi-RNAs lethal for HNSCCs**

To identify miRNAs that are lethal for head and neck cancer cells, we introduced a human miRNA expression library (mir-Lib) in HNSCC cell line VU-SCC-120 and close to normal ciOKCs (Fig. 1A). The ciOKC cells have been immortalized by a temperature-sensitive SV40 large T antigen. When cultured at 32°C, they are immortalized, while they become senescent when shifted to 39°C as a result of the inactivation of the large T-antigen. They behave, also at
32°C, as normal keratinocytes except for the immortalized phenotype (18).

The majority of mi-RNAs did not influence the survival of either tumor cells or ciOKC cells, or the lethal effect was similar in both models. However, a subset of 19 mi-RNAs (5.4%) specifically affected the head and neck cancer cell line, whereas the ciOKCs remained unaffected (Supplementary Table S1; Fig. 1B). To verify the tumor-specific lethal effect of these mi-RNAs, we screened all 19 mi-RNAs in 2 different ciOKC cell clones and 3 HNSCC cell lines (VU-SCC-OE, UM-SCC-6, and VU-SCC-120). We could not confirm the tumor-specific lethal effect of thirteen mi-RNAs. The
remaining 6 mi-RNAs showed the expected phenotype (Supplementary Fig. S1): a decreased proliferation in HNSCC cells but not in ciOKC cells and these mi-RNAs were further investigated. Visual inspection indicated that the tumor cells had disappeared from the wells indicating that they likely ceased proliferation and died. As yet, we did not elucidate the precise molecular mechanism behind the cell death.

For the initial large scale discovery screens, we had to rely on the ciOKC cell model to study the effect of the mi-RNAs. In the subsequent small-scale validation experiments we included primary keratinocytes. The mi-RNAs miR-181a, miR-323, miR-326, miR-342, miR-345, and miR-371 all showed a significant decrease in cell proliferation in 3 HNSCC cell lines and not or only to a limited extent in primary keratinocytes (Fig. 1C). The sequence identity of these 6 mi-RNAs was confirmed by Sanger sequencing (data not shown).

mi-RNA expression in HNSCCs

As the ectopic expression of the 6 tumor-selective lethal mi-RNAs caused cell death in tumor cell lines but not or less in mucosal keratinocytes, we were interested in the expression of these mi-RNAs in HNSCC tumors and normal oral mucosa. Hence, we determined expression levels for the 6 mi-RNAs in RNA extracted from both microdissected tumor and mucosal epithelium. miR-371 was neither expressed in normal mucosa nor in the 5 tumors analyzed (data not shown). For the other 5 mi-RNAs, expression was observed in all tumor and mucosal epithelium samples but apparently at a low level (Fig. 2). The expression levels of the mi-RNAs were in all cases lower than the expression of the RNU44 reference gene. In addition, only small differences in expression levels were observed between mucosa and tumor samples. The expression of miR-181a was slightly increased in tumors but not significant (Fig. 2A). For miR-326, miR-342, and miR-345, the expression level in tumor tissue was decreased when compared to normal mucosal epithelium but not significantly. Only the expression level of miR-323 was significantly lower in tumor cells (Fig. 2B).

Tumor-specific lethal phenotype in cell lines of different cancer types

We next questioned whether the effect observed in the HNSCC cell lines was specific for HNSCCs. Therefore, we tested the 6 mi-RNAs with HNSCC-specific lethal effects in other cancer cell lines. The introduction of the various mi-RNAs in cervical carcinoma cell line (SiHa) and breast carcinoma cell line (MCF7) had no effect on proliferation except for mi-RNA 181a (Supplementary Fig. S2A and S2B). However, when the mi-RNAs were introduced in colon adenocarcinoma cell line (HT29) or glioblastoma cell line (U87), a decrease in cell proliferation was observed, although with a less severe phenotype than the tested...
HNSCC cell lines (Supplementary Fig. S2C and S2D). The effects observed vary with the specific microRNA. This strongly suggests that genes are targeted that show synthetic lethal interactions in relation to the mutational status of specific cancer genes or deregulated signaling pathways in the various cell lines of different tissue origin, and it might be worthwhile to repeat these functional screens for other tumor types as it might reveal other candidate microRNAs. The lethal interaction is likely not related to a mutation in TP53 as all mi-RNAs showed the tumor-selective lethal effect in all 3 HNSCC cell lines, whereas one cell line was TP53 wild-type, one showed two missense mutations, and one a large deletion (see Materials and Methods for details).

**Target gene analysis**

mi-RNAs regulate gene expression at the posttranscriptional level, so we were interested in the downstream gene targets of these 6 tumor-specific lethal mi-RNAs that might explain the cell growth inhibitory phenotype. First, we conducted an in silico analysis. There is a multitude of software tools available for target identification (reviewed in ref. 24), and we chose TargetScan and DIANA-microT. We also defined the overlap between genes detected by the 2 programs (Supplementary Table S4). There was a wide variety of potential genes identified with more than 700 for miR-181a. This analysis did not give a direct clue.

Therefore, we conducted microarray-based gene expression analysis to identify candidate target genes. As ectopic expression of these mi-RNAs caused a decrease in cell proliferation and cell death as phenotype in HNSCC cell lines, we were unable to analyze cells stably transduced with the mi-RNAs. We therefore decided to transiently transfect VU-SCC-120 cells with the retroviral vector plasmids instead of transduction with retroviral particles. Transient transfection is efficient, and overexpression can be observed almost immediately. RNA was isolated 72 hours posttransfection, the time point with high mi-RNA expression, but before cell death was observed (Supplementary Fig. S3). The expression level of the transfected mi-RNAs was also determined. Depending on the endogenous expression of the mi-RNAs in VU-SCC-120 cells, expression increased from 23- to 4,000-fold after transient transfection (Fig. 3A and B).

![Figure 3](https://example.com/figure3.png)

**Figure 3.** Ectopic expression of mi-RNAs and their effect on gene expression. VU-SCC-120 cells were transiently transfected with the six different mi-RNA plasmids and overexpression of the indicated mi-RNAs was compared with miR-Vec-Ctrl–transfected cells for (A) miR-181a, miR-326, and miR-345 as well as (B) miR-323, miR-326, and miR-371. Significant differences are indicated by an * (P < 0.05, Student t test). Two separate groups became apparent by hierarchical cluster analysis (C).
Next, total RNA of the transfected cells was labeled and hybridized for gene expression profiling by microarray hybridization. We assumed that some of these 6 mi-RNAs might in fact target the same genes, and we therefore focused on a groupwise comparison. In the expression profiles, we indeed observed significant correlations between miRNA-associated mRNA expression profiles (Table 1). Highly significant correlations were observed for miR-181a and miR-326 ($r = 0.572$), miR-326 and miR-345 ($r = 0.573$), miR-323 and miR-371 ($r = 0.602$), and miR-342 and miR-323 ($r = 0.577$). Cluster analysis revealed 2 groups of each 3 miRNAs with downstream target effects that showed significant correlations (Fig. 3C; Supplementary Tables S2 and S3). miR-181a, miR-326, and miR-345 clustered together in group A, and group B was composed of miR-323, miR-371, and miR-323. The profiles of the mi-RNAs per group were combined and analyzed against the empty vector control (in triplicate hybridized on the arrays) to detect significantly differentially expressed genes.

In total, we observed 187 and 15 genes (FDR-corrected $P$ value < 0.1) that were significantly differently expressed between groups A and B, respectively, as compared to the empty vector control. Subsequently, we applied several rankings on the differentially expressed genes to distinguish primary effects from secondary effects. First, we ranked the genes for a decreased level of expression, as mi-RNAs are considered to cause downregulation of expression of their target genes. Second, as for many genes multiple probes were present on the array, we subsequently ranked the genes for the number of probes per gene with an FDR-corrected $P$ value < 0.1 (Supplementary Tables S2 and S3). One of the most striking target genes that is apparently regulated by the mi-RNAs from group A is the ATM gene. First, the differences of many probes were highly significant given the limited sample size (4 controls vs. 3 mi-RNAs of group A analyzed in duplicate) and the conservative $P$ value adjustment. Second, in total 9 of 12 ATM probes were significantly regulated. Unfortunately, we did not find such an apparent lead target for the mi-RNAs in group B.

### Knockdown of ATM causes tumor-specific lethality

ATM is a nuclear protein kinase that senses DNA damage and activates downstream pathways (24). To validate the observed decrease in expression, we carried out qRT-PCR for ATM in the same samples that were transiently transfected with the mi-RNAs and that were analyzed by microarray hybridization. Indeed we confirmed that ATM expression is inhibited in the RNA samples from group A (Fig. 4A).

As ectopic expression of miR-181a, miR-326, and miR-345 results in tumor-specific lethality and downregulation of ATM expression, we hypothesized that knockdown of ATM may also be accompanied by tumor-specific lethality in HNSCC cells. Therefore, we introduced 5 lentiviral shRNA constructs designed to specifically knockdown ATM expression in ciOKC cells and VU-SCC-120. Each shRNA sequence was complementary to a unique part of the ATM mRNA sequence. Introduction of the ATM shRNAs resulted in a minor inhibition of cell proliferation in ciOKC cells compared with cells transduced with a control construct (ctrl), but proliferation was significantly inhibited in HNSCC cell line VU-SCC-120, ranging from 21% to 90% (Fig. 4B). The maximum window was observed with shRNA ATM1. It was at least 8 times more active in the tumor cell line as compared to the ciOKC cells. To check the knockdown, ATM expression was analyzed by qRT-PCR and expression levels were compared with the cells transduced with the control construct. Introduction of 4 of 5 ATM shRNAs resulted in more than 70% downregulation of ATM expression levels. Only transduction with ATM shRNA number 5 did not result in significant downregulation of ATM expression (Fig. 4C).

ATM is a kinase and druggable by kinase inhibitors. To confirm the tumor-specific decrease in cell viability by ATM inhibition, we also subjected ciOKC and HNSCC cells to different concentrations of the commercially available specific ATM inhibitor CP466722. Analysis of cell viability showed that HNSCC cells were more sensitive to the inhibitor compared to the ciOKC cells (Fig. 4D). The $IC_{50}$ of 8.2 $\mu$mol/L in ciOKC cells shifts to 2.6 $\mu$mol/L in VU-SCC-120, a significant change of 3-fold ($P < 0.05$ by t testing).

Inhibition of ATM by microRNA overexpression, specific ATM shRNAs, or kinase inhibitors results in a decrease in cell proliferation in HNSCC cells. When ATM is the direct effector, ectopic expression of ATM should rescue the HNSCC cells from cell death. To investigate this, VU-SCC-120 cells were transfected with either wild-type ATM (ATMwt) or a kinase-dead mutant ATM (ATMkd) in an expression cassette that lacks the 3'UTR of ATM. Overexpression of ATMwt or ATMkd was confirmed by qRT-PCR (Supplementary Fig. S4). Next mi-RNAs, miR-181a, miR-326, and miR-345 were introduced in these two cell lines and compared with

| Table 1. Correlations of gene expression profiles between different microRNAs |
|-----------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|
| miR-181a     | miR-326 | miR-371 | miR-345 | miR-323 | miR-342 |
| miR-181a | 1.000  |          |          |          |          |
| miR-326     | 0.572  | 1.000   |          |          |          |
| miR-371     | 0.113  | 0.119   | 1.000    |          |          |
| miR-345     | 0.373  | 0.573   | 0.158    | 1.000    |          |
| miR-323     | 0.099  | 0.087   | 0.602    | 0.170    | 1.000    |
| miR-342     | 0.074  | 0.065   | 0.388    | 0.123    | 0.577    | 1.000    |
untransduced VU-SCC-120 (ctrl). The mi-RNAs all showed a decrease in cell proliferation in the untransduced VU-SCC-120 (Fig. 4F). However, in the cells with the ATMwt expression construct, cell proliferation was rescued up to about 80% (miR-181a) and about 50% (miR-326 and miR-345). Rescue was not observed when the kinase-dead mutant ATM was ectopically expressed in VU-SCC-120 cells. These data indicate that the miRNAs inhibit ATM expression, which elicits

Figure 4. ATM expression is regulated by miR-181a, miR-326, and miR-345 from group A. A, endogenous ATM expression in ciOKC cells was analyzed by qRT-PCR after transient transfection with miR-Vec-Ctrl or the miRNA constructs indicated. In (B), cell proliferation of ciOKC (dark gray bars) and VU-SCC-120 (medium gray bars) is depicted after ATM knockdown with 5 different shRNA constructs and control vector (ctrl). In (C), the knockdown of endogenous ATM expression in ciOKC cells is depicted after lentiviral transduction with either a control construct (ctrl) or ATM shRNA constructs 1 to 5. In (D), sensitivity of HNSCC cell lines VU-SCC-120 and ciOKC cells to the ATM drug CP466722 is determined. In (E) the effect of the miRNAs of group A on the ATM 3'UTR luciferase reporter construct is shown. The 3'UTR of ATM was cloned behind the firefly luciferase. Firefly luciferase activity was normalized using Renilla luciferase activity to correct for transfection efficiency. In (F) rescue experiments with ATM encoding expression constructs are depicted. Wild-type ATM (ATM; medium gray bars) or kinase-dead ATM (ATMkd; light gray bars) were transfected and the effect on the tumor-selective phenotype of miR-181a, miR-326, miR-345, or miR-Vec-Ctrl (miCtrl) compared with untransfected cells (ctrl; dark gray bars). The average of triplicate experiments is shown with standard deviations as error bars. Significant differences are indicated with an * (P < 0.05, Student t test).
the tumor-selective lethal phenotype and it depends on the kinase activity of ATM.

Targeting of ATM by miR-181a, miR-326-, and miR-345

mi-RNAs are known to regulate gene expression posttranscriptionally via binding to the 3′ UTR or coding sequences of an mRNA sequence. To show an effect of miR-181a, miR-326, and miR-345 on the 3′UTR of the ATM gene mi-RNAs were cotransfected with a luciferase reporter construct cloned to the ATM 3′UTR sequence. Transfection of ciOKCs with miR-181a and miR-326 suppressed the activity of a luciferase reporter gene cloned to the 3′UTR of ATM. miR-345 did show an effect but not significant (Fig. 4E).

The knockdown of ATM by the respective microRNAs seems to inversely correlate with the effect of ectopic ATM expression. In addition, ectopic ATM expression does not rescue the lethal phenotype to 100%. This suggests that some mi-RNAs might not target only the 3′UTR but also the 5′UTR or coding sequences. These sequences are still present in the expression construct, only the 3′UTR of ATM has been substituted. We further hypothesized that another gene target might be involved as well. First, we re-inspected our expression data and found another gene significantly down-regulated in 9 of 10 available probes: CD40. We next checked the presence of microRNA target sites using RNA22 and TargetScan on both ATM and CD40. Multiple binding sites were found for these microRNAs in these genes, albeit mostly not in the 3′UTR region (Supplementary Fig. S5).

The relevance of the various putative mi-RNA target sites and the role of CD40 and its interaction with ATM requires further elucidation.

Discussion

In the present study, we identified 6 mi-RNAs that, when ectopically expressed, lead to a tumor-specific inhibition of proliferation in HNSCC cells and not in primary keratinocytes. mi-RNAs are generally classified into different families based on their sequence homology, most particularly of the seed sequence, and it is hypothesized that different mi-RNAs within the same family may have similar effects on gene expression. Although all mi-RNAs highlighted in this study have a tumor-specific lethal effect, some even recognize at least one identical and apparently critical target gene, they do not belong to the same family.

Many mi-RNAs have already been implicated in cancer development and progression. For miR-345, a role in head and neck cancer development has been suggested previously by profiling studies. In these studies, mi-RNA expression profiles in tumor samples were compared with normal mucosal epithelium, and it was shown that miR-345 showed an increased expression level in malignant cells (10). miR-181a has been shown by functional studies to have tumor-suppressive properties in HNSCCs (15). Here, we show a tumor-specific lethal effect for ectopic expression of these 2 miRs as well as miR-326, miR-342, miR-371, and miR-323 in head and neck cancer cells. We also show that a similar lethal effect was observed when those 6 mi-RNAs were introduced in colon cancer and glioblastoma cell lines, although there was a considerable variation in the lethal phenotype between cell lines of different origin. It might therefore be of interest to repeat similar screens in other tumor types. Unfortunately, it is not always possible to have access to matched normal (or near normal) cells, which are required to allow a tumor-specific lethal screen and subsequent validation. Assuming that the identified hits are synthetic lethal with cancer-associated alterations in conserved cellular pathways such as cell-cycle control or DNA repair, the precise source and histotype of the normal reference cells might be less critical, and normal fibroblasts may be sufficient.

mi-RNAs regulate gene expression at the posttranscriptional level. One gene may be regulated by multiple mi-RNAs but also one mi-RNA may regulate multiple genes. Consequently, it is difficult to study downstream targets of a certain mi-RNA, particularly as the target genes may also be involved in the regulation of expression of other genes. Therefore, it is not surprising that many genes were shown to be significantly differentially expressed when the effect of the 6 tumor-specific lethal mi-RNAs was analyzed. Although the expression profiles of the 6 mi-RNAs were not similar, it was interesting to observe that 2 groups could be discriminated. Unfortunately, we could not identify an apparent candidate target gene in group B. Likely multiple repeats of the array analysis in multiple cell lines will be required to define more convincing candidate target genes. The limited sample size combined with the strict FDR correction of the P values may have skewed the analyses. Moreover, mi-RNAs might also influence genes at the translational level, which would necessitate alternative approaches to identify the candidate target genes.

However, in group A, several probes for ATM showed significant differences. Using ATM 3′UTR luciferase reporter experiments, we showed that both miR-181a and miR-326 seem to directly target the 3′UTR sequence of ATM. We could not confirm this for miR-345. We also showed that specific downregulation of ATM via shRNA knockdown elicits a similar tumor-specific lethal phenotype. Although there was a slight toxic effect in the normal squamous epithelial cells, the lethal phenotype observed in HNSCC cell lines was much more severe, and HNSCC cells are more sensitive to the specific ATM inhibitor CP466722. Finally, the tumor-selective phenotype of the mi-RNAs could be rescued by introduction of an ATM expression construct that lacks the 3′UTR of ATM itself, whereas the phenotype could not be rescued by expression of a kinase-dead mutant.

Intriguingly, there seemed an inverse correlation between reduction of luciferase activity with the ATM 3′UTR and the rescue level when expressing an ATM cDNA sequence without the 3′UTR, strongly suggesting that target sites in the coding cDNA sequence are highly relevant as well. When analyzing the potential target sites by RNA22 and TargetScan, a binding site for miR-181a was found in the 3′UTR of ATM, but many more in the coding cDNA sequence. Mutation of predicted target sites will be required to proof the relevance of all of them, which will be a major future task. A
worrying observation is that the various prediction programs come up with different potential binding sites, hampering a focused approach. Nonetheless, such experiments will be required to establish the ultimate proof of ATM targeting by the microRNAs of group A.

We also observed that the rescue level never reaches 100% cell survival. There might be many explanations for this observation, but it suggests that also another target gene might be involved, and we could pinpoint CD40 as a first hint. Additional experiments will be required to prove this and that combined targeted treatment might be of benefit.

Our data thus far suggest that HNSCC cells are depending on ATM signaling for their survival. When ATM expression is lost, either by shRNA knockdown or when targeted by an miRNA, the cell is unable to survive, at least under culture conditions. ATM senses DNA damage and phosphorylates CHEK2 which in turn activates p53 by phosphorylation. This route plays a major role in maintaining genome integrity and cell-cycle control (25). Intriguingly, TP53 is mutated in 2 of the 3 HNSCC cell lines tested and ATM may have become more critical to organize DNA repair or induce a cell-cycle block during S–G2 phase after DNA damage independent of p53. The third cell line UM-SCC-6 is TP53 wild-type but it is unclear whether p53 is still functionally active. Detailed analyses of the ATM/ATR-CHEK1/CHEK2 pathway should reveal which critical downstream signaling route causes the lethal phenotype and what the mechanism is. Obviously ATM has many downstream targets, and the tumor-selective phenotype might be related to alterations in other signaling routes. Nonetheless, our data suggest that ATM is an interesting drug target for HNSCCs.

Previously, it has been shown that miR-181a targets KRAS, and this was put forward as explanation that this particular microRNA inhibits proliferation of squamous cancer cells (15). This seemed a little remarkable as KRAS mutations are hardly found in HNSCCs, suggesting that this is not the most critical driving gene or even pathway in squamous oncogenesis. On the basis of our data, we assume that the targeting of ATM by miR-181a is the likely event that causes a proliferation stop and cell death in squamous cancer cells.

In summary, we show here that functional screens by microRNA expression libraries are an effective way to identify novel druggable targets. We showed that the ectopic expression of miRNAs such as miR-323, miR-345, miR-371, miR-181a, miR-342, and miR-326 may serve as a new treatment in HNSCCs. Particularly in head and neck cancer where access to the tumor is relatively easy, introduction of miRNAs, for instance by intratumoral injection combined with electroporation, may be a therapeutic possibility in the future (26). Also, the application of ATM inhibiting drugs might be a very interesting approach to study in clinical trials specifically focusing on HNSCCs.

Disclosure of Potential Conflicts of Interest

R.H. Brakenhoff has a commercial research grant from InterRNA Technologies BV. InterRNA Technologies BV has filed a patent application regarding the inventions described in this manuscript. No potential conflicts of interest were disclosed by the other authors.

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References


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