Targeting Src and Tubulin in Mucinous Ovarian Carcinoma

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Abstract

Purpose: To investigate the antitumor effects of targeting Src and tubulin in mucinous ovarian carcinoma.

Experimental Design: The in vitro and in vivo effects and molecular mechanisms of KX-01, which inhibits Src pathway and tubulin polymerization, were examined in mucinous ovarian cancer models.

Results: In vitro studies using RMUG-S and RMUG-L cell lines showed that KX-01 inhibited cell proliferation, induced apoptosis, arrested the cell cycle at the G2–M phase, and enhanced the cytotoxicity of oxaliplatin in the KX-01–sensitive cell line, RMUG-S. In vivo studies showed that KX-01 significantly decreased tumor burden in RMUG-S and RMUG-L mouse models relative to untreated controls, and the effects were greater when KX-01 was combined with oxaliplatin. KX-01 alone and in combination with oxaliplatin significantly inhibited tumor growth by reducing cell proliferation and inducing apoptosis in vivo. PTEN knock-in experiments in RMUG-L cells showed improved response to KX-01. Reverse phase protein array analysis showed that in addition to blocking downstream molecules of Src family kinases, KX-01 also activated acute stress-inducing molecules.

Conclusion: Our results showed that targeting both the Src pathway and tubulin with KX-01 significantly inhibited tumor growth in preclinical mucinous ovarian cancer models, suggesting that this may be a promising therapeutic approach for patients with mucinous ovarian carcinoma. Clin Cancer Res; 19(23); 6532–43. © 2013 AACR.

Introduction

Mucinous ovarian carcinoma is a relatively rare subset of ovarian cancer, accounting for 2% to 10% of all subtypes of epithelial ovarian cancer (1, 2). Although standard therapy for ovarian cancer includes taxane and platinum-based chemotherapy (3, 4), such therapy is largely ineffective in patients with mucinous ovarian carcinoma. Consequently, the outcome of women with mucinous ovarian carcinoma is poorer than patients with other epithelial ovarian cancers (5–8). Novel therapeutic approaches are urgently needed to improve outcomes of patients with mucinous ovarian carcinoma.

The Src family of nonreceptor protein tyrosine kinases (SFK) regulates various aspects of tumor progression via multiple signaling pathways, including cell survival (PI3K/Akt), growth (Ras/MEK/ERK), metastasis (FAK/paxillin), and angiogenesis (STAT3/VEGF; ref. 9). Increased Src activity has been demonstrated in colorectal, pancreatic, lung, breast, ovarian, and prostate carcinomas, highlighting Src as a potential therapeutic target (10, 11). Preclinical data have demonstrated the antitumor activity of Src inhibitors in prostate, colon, and breast cancers (12–14). Recently, we reported that Src kinase is overexpressed in mucinous ovarian carcinoma (15). Several agents targeting Src, including dasatinib (BMS-354825; Bristol-Meyers Squibb), saracatinib (AZD0530; AstraZeneca), and bosutinib (SKI-606; Wyeth/Pfizer) are currently in clinical trials for the treatment of various solid tumors. However, monotherapy with an Src inhibitor shows little activity in unselected solid tumor patients (16), emphasizing the need for multitargeted inhibitors and biomarker-driven studies.

KX-01 (Kinex Pharmaceuticals) is a novel Src signaling inhibitor that, unlike other Src inhibitors, does not bind to the ATP site (17). In addition, KX-01 binds to a novel site on the tubulin heterodimer, resulting in inhibition of...

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Src Pathway and Tubulin in Mucinous Ovarian Carcinoma

Translational Relevance
Mucinous ovarian carcinoma responds poorly to currently available treatments. Recently, we reported that Src kinase is overexpressed in mucinous ovarian carcinoma, and an Src inhibitor significantly inhibited tumor growth and enhanced the antitumor effects of oxaliplatin. In this study, we show that KX-01 (Src and tubulin inhibitor) significantly inhibits cell proliferation, induces apoptosis, arrests the cell cycle at the G2-M phase in mucinous ovarian cancer cell lines, and suppresses tumor growth in preclinical mucinous ovarian cancer models. Moreover, KX-01 showed synergistic antitumor activity when combined with oxaliplatin. These findings suggest that targeting both Src and tubulin may be a promising therapeutic approach for patients with mucinous ovarian carcinoma.

Microtubule polymerization and broader antitumor activity (18). In this study, we evaluated the preclinical activity of KX-01 in mucinous ovarian carcinoma.

Materials and Methods
Cell lines and culture
The derivation and source of RMUG-S and RMUG-L cell lines has been previously described (15). YDOV-151 (a kind gift from Dr. Kwong K. Wong, Department of Gynecologic Oncology and Reproductive Medicine, The University of Texas MD Anderson Cancer Center) and EFO-27 (a kind gift from Dr. Gordon Mills, Department of Systems Biology, MD Anderson Cancer Center) cell lines were originally isolated from women with mucinous ovarian carcinoma (19, 20). Cell lines were maintained and propagated in RPMI-1640 medium supplemented with 10% FBS and 0.1% gentamicin sulfate (Gemini Bioproducts). Clear cell ovarian cancer cell lines (ES-2 and TOV21G) were purchased from American Type Culture Collection, and were maintained in specific medium as described previously (15, 21). Cell lines were characterized by the Cell Line Core Facility at MD Anderson, which provides authenticated cell lines. The derivation and source of RMUG-S and RMUG-L cells used in this study has been described previously (15). RMUG-S-ip2 or RMUG-L-ip2 cells were inoculated into the peritoneal cavity of 40 orthotopic nude mice (4 × 10^6 cells per mouse). Mice were randomized into 4 treatment groups of 10 mice each: control, oxaliplatin, KX-01, and oxaliplatin plus KX-01. Treatments were initiated 4 weeks after inoculation. Oxaliplatin was dissolved in 5% dextrose and diluted with Hank’s Balanced Salt Solution (HBSS) and administered intraperitoneally twice weekly (5 mg/kg per mouse; ref. 22). KX-01 was solubilized in distilled water and administered orally every day (15 mg/kg per mouse, according to the dose-finding experiment; see Supplementary Fig. S1A). Control mice received HBSS intraperitoneally twice weekly and oral distilled water daily. Mice were monitored on a daily basis and weighted weekly. After 8 weeks of treatment, the mice were sacrificed and total mouse body weight, tumor location and weight, and number of tumor nodules were recorded. Tumor specimens were preserved in either optimum cutting temperature medium (OCT; Miles Inc.; for frozen slides) or fixed in formalin (for paraffin slides) for further analysis.

Reagents and antibodies
KX-01 was provided by Kinex Pharmaceuticals in water-soluble powder form (KX-01: methanesulfonic acid salt). Oxaliplatin (Wyeth/Pfizer) and paclitaxel (Bristol-Myers Squibb) were purchased from the institutional pharmacy. PP2 (Src inhibitor) and Akt 1/2 kinase inhibitor were obtained from Sigma-Aldrich. Antibodies used in this study were phospho-Src (Tyr419), total-Src, phospho-Akt (ser473), total-Akt, phospho-P130cas (Tyr410), phospho-paxillin (Tyr118), total-paxillin, phospho-FAK (Tyr576/577), phospho-FAK (Tyr925), CDC2, α/β-tubulin, and PARP (Cell Signaling Technology); total-P130cas (Thermo Scientific); total-FAK (BD Biosciences); phospho-FAK (pY397), phospho-FAK (pY861; Invitrogen); glu-Tubulin, lamin B1 (Abcam); CD31 (BD Pharmingen); Ki67 (Neomarkers Inc.); cleaved caspase-3 (Biocare Medical LLC); β-actin (Sigma-Aldrich); horseradish peroxidase–conjugated rat anti-mouse IgG2a (Serotec Harlan Bioproducts for Science Inc.); horseradish peroxidase–conjugated goat anti-rat immunoglobulin G (IgG), and Alexa Fluor 488–conjugated anti-rabbit antibody (Jackson ImmunoResearch Laboratories).

In vivo orthotopic model of mucinous ovarian carcinoma
Female athymic nude mice were purchased from the National Cancer Institute-Frederick Cancer Research and Development Center (Frederick, MD), housed in specific pathogen-free conditions, and cared for in accordance with the guidelines set forth by the American Association for Accreditation for Laboratory Animal Care and the US Public Health Service Policy on Human Care and Use of Laboratory Animals. All animal experiments were approved and supervised by the MD Anderson Institutional Animal Care and Use Committee.

The in vivo model of mucinous ovarian carcinoma (RMUG-S-ip2 and RMUG-L-ip2) used in this study has been described previously (15). RMUG-S-ip2 or RMUG-L-ip2 cells were inoculated into the peritoneal cavity of 40 orthotopic nude mice (4 × 10^6 cells per mouse). Mice were randomized into 4 treatment groups of 10 mice each: control, oxaliplatin, KX-01, and oxaliplatin plus KX-01. Treatments were initiated 4 weeks after inoculation. Oxaliplatin was dissolved in 5% dextrose and diluted with Hank's Balanced Salt Solution (HBSS) and administered intraperitoneally twice weekly (5 mg/kg per mouse; ref. 22). KX-01 was solubilized in distilled water and administered orally every day (15 mg/kg per mouse, according to the dose-finding experiment; see Supplementary Fig. S1A). Control mice received HBSS intraperitoneally twice weekly and oral distilled water daily. Mice were monitored on a daily basis and weighed weekly. After 8 weeks of treatment, the mice were sacrificed and total mouse body weight, tumor location and weight, and number of tumor nodules were recorded. Tumor specimens were preserved in either optimum cutting temperature medium (OCT; Miles Inc.; for frozen slides) or fixed in formalin (for paraffin slides) for further analysis.

Reverse phase protein arrays
RMUG-S and RMUG-L cells were treated with KX-01 at a concentration of 100 nmol/L for 24 hours. Cells were homogenized using a digital homogenizer in the following lysate buffer: 1% Triton X-100, 50 mmol/L HEPES (pH 7.4), 150 mmol/L MgCl₂, 1 mmol/L EGTA, 100 mmol/L NaF, 10 mmol/L Na-pyrophosphate, 1 mmol/L Na₃VO₄, 10% glycerol, and freshly added protease and phosphatase inhibitors. Cellular proteins were denatured using 1% SDS, and five 2 × serial dilutions were performed in lysis buffer containing 1% SDS (dilution buffer). These diluted lysates...
were arrayed on nitrocellulose-coated FAST slides (Whatman Inc.) using an Aushon 2470 Arrayer (Aushon BioSystems). Slides were probed with 152 validated primary antibodies and a biotin-conjugated secondary antibody. The Dako CytoTmage-catalyzed system (Dako) was used for signal amplification. DAB colorimetric reaction was used for visualization. Slides were then scanned, analyzed, and quantified using customized Microvige software (VigeneTech.), and spot intensity was generated.

A logistic model ("Supercurve Fitting," developed by the Department of Bioinformatics and Computational Biology at the MD Anderson Cancer Center; http://bioinformatics.mdanderson.org/OOMPA) was used to generate a fitted curve for each dilution. For both observed and fitted data, the fitted curve was then plotted with the signal intensities on the y-axis and the log2 concentration of proteins on the x-axis. From each slide, the protein concentrations were normalized using median polish. Positive fold-change was calculated by dividing each linear value (>1.0) by the average control linear value for each antibody tested, and negative fold-change (for linear values <1.0) was calculated using the formula (−1/linear fold-change) and plotted in a bar graph.

**Western blot analysis**

Cell lysates were obtained with RIPA lysis buffer [50 mmol/L Tris-HCl (pH 7.4), 150 mmol/L NaCl, 1% Triton, 0.5% deoxycholate, 25 μg/mL leupeptin, 10 μg/mL aprotinin, 2 mmol/L EDTA, and 1 mmol/L sodium orthovanadate] and centrifuged for 15 minutes at 4°C. Protein concentration was then determined using the Bio-Rad Protein Assay Kit (Bio-Rad). Following protein loading (25 μg/well), bands were separated on 8% gel using SDS-PAGE and transferred to nitrocellulose paper, blocked with 5% milk for 1 hour at room temperature, and incubated with primary antibodies overnight at 4°C. The bands were then incubated with horseradish peroxidase-conjugated anti-mouse or anti-rabbit (GE Healthcare) for 1 hour at room temperature. Blots were developed using an enhanced chemiluminescence detection kit (Pierce Biotechnology). β-Actin or vinculin was used for loading control, and all experiments were repeated in duplicate. Densitometry (Image J; National Institutes of Health) was used to assess the differences in results.

**Cytotoxicity assays**

Cells were plated in each well of a 96-well plate and maintained overnight. Cells were then exposed to KX-01 at concentrations of 1, 10, 100, or 1,000 μmol/L for 48 or 72 hours, or to Akt1/2 inhibitor at concentrations of 1, 5, 10, 50, or 100 μmol/L for 72 hours. Controls were treated with an equal volume of vehicle. To assess cell survival, 50 μL of 0.15% MTT (Sigma-Aldrich) was added to each well and incubated for 2 hours at 37°C. The medium containing MTT was then removed and 100 μL of dimethyl sulfoxide (Sigma-Aldrich) was added; cells were then incubated at room temperature for 10 minutes. Absorbance was then read at 570 nmol/L by a 96-well Synergy HT-microplate reader (Ceres UV 900C; Bio-Tek Instrument Inc.). Cell survival was defined as the percentage of cells surviving relative to the control group (number of viable cells in the treatment group/number of viable cells in the control group). The experiments were repeated separately at least 3 times. The median inhibitory concentrations (IC50) were calculated.

Isobologram analysis was performed to evaluate the cytotoxicity of KX-01 and oxaliplatin in RMUG-S cells using dose–response cell-survival curves (23). The interaction index was calculated as described previously (24). Additive effects (equal to 1), synergistic effects (less than 1), and antagonistic effects (greater than 1) of the combination of oxaliplatin plus KX-01 were determined by the interaction index. Results were analyzed using R (version 2.14.2).

**Cell-proliferation assays**

To determine cell proliferation, we used the Click-IT EdU Flow Cytometry Assay Kit (Invitrogen) according to the manufacturer's protocol. Briefly, cells were incubated with EdU for 2 hours before being harvested, then fixed with Click-IT fixative for 15 minutes at room temperature. After washing was completed, cells were permeabilized with Click-IT Saponin-based permeabilization and wash reagent for 15 minutes, and Click-IT reaction cocktail was added for 30 minutes. Cells were then analyzed using a Beckman Coulter XL 4-color flow cytometer (FACS; Beckman-Coulter).

**Apoptosis and cell-cycle assays**

Apoptosis was evaluated using the Annexin V-phycocyanin (PE) apoptosis detection kit (BD Biosciences). Cells were incubated in trypsin-EDTA, and cell pellets were suspended in 1 mL of 1× Annexin V binding buffer. One hundred microliters of each cell suspension was incubated with 5 μL of Annexin V-PE and 5 μL of 7-aminoactinomycin D (7-AAD) at room temperature (25°C) in the dark for 30 minutes. Following this incubation, 400 μL of 1× binding buffer was added to each tube, and samples were analyzed using FACS. Each experiment was repeated in triplicate.

For cell-cycle analysis, cells were maintained in serum-free media for 24 hours and then lifted with trypsin, washed with phosphate-buffered saline (PBS), fixed in 70% cold ethanol, and stored overnight at 4°C. Cells were then centrifuged (pelleted) at 1,200 rotations per minute for 10 minutes at 4°C. After 1 wash with PBS, cells were suspended in propidium iodide (Roche) at 50 μg/mL and RNAseA (Qiagen) at 100 μg/mL and incubated in the dark at room temperature. Cells were then assessed for cell-cycle status using FACS.

**Immunohistochemical and immunofluorescence staining**

Immunohistochemical staining was used to detect angiogenesis (CD31), cell proliferation (Ki67), and apoptosis (cleaved caspase-3) in frozen and paraffin-embedded tumor specimens. Paraffin sections were deparaffinized and rehydrated, then antigen retrieval was performed using citrate buffer (pH 6.0). Sections were then blocked with 3% hydrogen peroxide in methanol and 4% fish gelatin at room temperature, then incubated with Ki67 antibody (1:400) overnight at 4°C. After washing with PBS, the
sections were incubated with horseradish peroxidase–conjugated rat anti-mouse IgG2a (1:100) for 1 hour. For cleaved caspase-3 staining, sections were treated similarly but incubated with the monoclonal mouse antibody against cleaved caspase-3 (1:100). For CD31 staining, frozen sections were fixed in cold acetone for 15 minutes, washed with PBS, endogenous proteins blocked (4% fish gelatin), and then incubated with rat monoclonal anti-mouse CD31 (1:800) overnight at 4°C. Sections were washed with PBS and incubated with horseradish peroxidase–conjugated goat anti-rat IgG (1:200) for 1 hour. Reactive tissues were visualized by staining with 3, 3-diaminobenzidine (Research Genetics) and counterstaining with Gill’s hematoxylin (Biogenex Laboratories).

For quantification, 5 sections from each group were examined. To quantify microvessel density for each sample, the microvessels within 5 randomly selected 0.159-mm² fields at ×200 magnification were counted. A single microvessel was defined as a discrete cluster or single-cell stained positive for CD31 (CD31⁻). To quantify Ki67 and cleaved caspase-3, the percentage of positive cells was determined in 5 random 0.159-mm² fields at ×200 magnification.

Tubulin and lamin B1 were analyzed by indirect immunofluorescence staining. Briefly, sections were fixed in cold acetone for 10 minutes, endogenous proteins blocked for 20 minutes at room temperature, incubated with tubulin antibody (α/β-tubulin; 1:50, glu-tubulin; 1:500) or lamin B1 antibody (1:1000) overnight at 4°C, and then incubated with Alexa Fluor 488–conjugated anti-rabbit antibody (1:1000) for 1 hour at room temperature. After washing with PBS, the samples were incubated with the DNA stain Hoechst 33342 (1:10000; Invitrogen) or DAPI (1:10000; Roche) for 10 minutes at room temperature. Mounted samples were visualized using a fluorescent microscope with the appropriate filter.

**PTEN transfection and Src knockdown**

For PTEN knock-in experiments, PTEN wild-type cDNA was subcloned into the Not I site of a PLNCX expression vector (Clontech Laboratories). PTEN mutants (G129R and PTIZB) were generated using a standard mutagenesis method. For retrovirus production, 239T cells were transfected with vector using the calcium phosphate transfection method. Twenty-four hours after transfection, the standard media was replaced with media containing G418 sulfate (Mediatech Inc.). After 1 to 2 weeks, cell lines stably infected with viral particles were established. The cells were treated with various concentrations of KX-01 for 72 hours, and cell survival was assessed using the MTT assay. Src siRNA (Sense: CUGAAGCGGGCGGCUUGUUGGUAIdTdT; Antisense: UACCCACCGGCCUCUCAGCAGdTdT) was from Sigma-Aldrich. Transient knockdown experiments with siRNA performed as described previously (25).

**Statistical analysis**

Continuous variables were assessed for normal distribution (Kolmogorov–Smirnov test) and expressed as approximate (mean with SD or median with range). Normally distributed variables were compared using the Student t test (2 groups) or analysis of variance (multiple groups). For variables with nonparametric distribution, the Mann–Whitney U test or Kruskal–Wallis test (multiple groups) was used. A P value <0.05 in 2-tailed testing was deemed statistically significant. The Statistical Package for Social Scientists software (SPSS, version 18.0; SPSS, Inc) was used for all analyses.

For the reverse phase protein array (RPPA) data analysis, to limit the false discovery rate to 5%, each cell line was examined individually. We first completed a 2 × 2 factorial analysis of variance to detect a treatment effect on expression and then used the β-uniform mixture distribution method (26) to determine a P-value cutoff for statistical significance that would limit our false discovery rate to 5%. Finally, if a treatment effect was evident, we tested for differences between KX-01 and control. After examining each cell line, we used t-tests to determine whether RMUG-S controls differed from RMUG-L controls. We then used the β-uniform mixture method again to limit our false discovery rate to 5%. Linear fold-change was calculated by dividing each linear value by the average control linear value. Positive fold-change (>).0.1 was the same as the linear fold-change value; negative fold-change (for linear values <1.0) was calculated as −1/ linear fold-change.

**Results**

**In vivo effects of KX-01 in tumor xenografts**

To recapitulate the unique features of mucinous ovarian carcinoma, we have recently established an orthotopic mouse model of metastatic mucinous ovarian carcinoma (15). The RMUG-S and RMUG-L models were utilized to evaluate the biological effects of KX-01 in vivo. In the RMUG-S model, tumor weight was reduced by 75% (P < 0.001) and tumor nodules decreased (P < 0.01) following treatment with KX-01 compared with the control group (Fig. 1A). In the RMUG-L model, tumor weight was reduced by 50% and 48% fewer tumor nodules were observed in the KX-01 treatment group compared with the control group (P < 0.05; Fig. 1B).

Because oxaliplatin shows potential as a chemotherapeutic drug for mucinous ovarian carcinoma, we also evaluated the effects of KX-01 in combination with oxaliplatin in the RMUG-S and RMUG-L models. In the RMUG-S model, tumor weight was reduced by 90% (combination vs. KX-01, oxaliplatin, 90% vs. 75%, 72%, P < 0.0001) and 85% fewer tumor nodules (P < 0.001) were observed in the combination group compared with the control group (Fig. 1A). Combination therapy led to significantly greater tumor suppression effects than did either monotherapy (combination vs. KX-01, oxaliplatin, 90% vs. 75%, 72%, P < 0.05). In the RMUG-L model, although a significant reduction in tumor weight and number of nodules was observed in the combination group compared with the control group, the antitumor effects of the combination therapy did not differ from those of either monotherapy (P > 0.05; Fig. 1B), signifying that the antitumor effects of
the combination therapy were greater in the RMUG-S model than in the RMUG-L model. The body weight of the mice after treatment did not differ between the treatment and control groups in either the RMUG-S or RMUG-L model (Supplementary Fig. S1B and S1C).

To investigate potential mechanisms of KX-01’s antitumor activity, we tested markers of cell proliferation (Ki67), angiogenesis (CD31), and apoptosis (cleaved caspase-3) in the tumor tissues. In the RMUG-S model, tumor cell proliferation was reduced in both the KX-01 group (33% reduction, \( P < 0.05 \)) and the oxaliplatin group (29% reduction, \( P < 0.05 \)) compared with the control group. Tumor cell proliferation was most significantly decreased in the combination group compared with the control group (47% reduction, \( P < 0.01 \); Fig. 1C). Tumor angiogenesis was significantly reduced in all KX-01–based treatment groups compared with the control group (KX-01 vs. control, 43% reduction, \( P < 0.01 \); combination vs. control, 69% reduction, \( P < 0.0001 \); Fig. 1D). This difference was not observed in the oxaliplatin group compared with the control group (\( P > 0.05 \); Fig. 1D).

Tumor cell apoptosis was increased in the oxaliplatin and KX-01 groups compared with the control group (\( P < 0.05 \); Fig. 1E), with the greatest increase in the combination group (\( P < 0.01 \); Fig. 1E). In the RMUG-L model, similar biological effects of KX-01 on angiogenesis and cell apoptosis were observed, but the effects of KX-01 on cell proliferation were less pronounced than in the RMUG-S model (21% reduction in RMUG-L vs. 33% reduction in RMUG-S, \( P < 0.01 \); Supplementary Fig. S2).
In vitro effects of KX-01 in mucinous ovarian carcinoma cells

To identify potential mechanisms by which KX-01 exerts its antitumor activity, we next tested its effects on cell survival, proliferation, apoptosis, and migration in mucinous ovarian cancer cells. Treatment with KX-01 consistently resulted in decreased cell survival in RMUG-S, RMUG-L, YDOV-151, and EFO-27 mucinous ovarian carcinoma cells with IC₅₀ levels from 72 to 203 nmol/L (Fig. 2A). Similar effects on survival were also noted in clear cell ovarian cancer cell lines (Supplementary Fig. S3A). In the RMUG-L cells, the inhibition rate did not reach 50%, indicating the relative resistance of RMUG-L cells to KX-01 treatment. KX-01 treatment reduced proliferation by 73% (vs. control, P < 0.001) in RMUG-S cells and 46% (vs. control, P < 0.01) in RMUG-L cells (Fig. 2B). KX-01 induced comparable late apoptosis in both cell lines (Fig. 2C). Immunoblotting demonstrated that cleaved PARP expression was significantly increased after KX-01 treatment in both cell lines (Supplementary Fig. S3B). A shift toward the G₂–M phase was
seen in cells treated with KX-01 (Fig. 2D). Furthermore, KX-01 inhibited the migration of both cell lines (Supplementary Fig. S3C and S3D).

We also tested the effects of various concentrations of combined oxaliplatin and KX-01 in RMUG-S cells. The IC50 of oxaliplatin alone was 14 µg/mL; the effects of oxaliplatin, based on increased apoptosis, were significantly enhanced with KX-01 (Fig. 2E and F). In isobologram analysis, the interaction index was < 1 at all examined points, suggesting synergistic effects of oxaliplatin and KX-01 (Fig. 2G).

Because KX-01 has activity against Src and tubulin, we investigated the effects of a pure Src inhibitor (PP2) or paclitaxel in RMUG-S cells. PP2 had minimal effects on cell survival (Supplementary Fig. S4A), and paclitaxel had effects on cell survival at higher concentrations (IC50 = 147 ng/mL; Supplementary Fig. S4B). The greater effect observed with KX-01 (Fig. 2A) may be secondary to dual targeting of Src and tubulin. The effects of Src siRNA, PP2, or paclitaxel with oxaliplatin were also tested, demonstrating that Src siRNA and PP2, but not paclitaxel, resulted in lower IC50 level of oxaliplatin (Supplementary Fig. S4C–S4E).

**Pathways affected by KX-01 treatment in mucinous ovarian carcinoma cells**

Because KX-01 was constructed to inhibit phosphorylation of Src substrates, we also tested the effects of KX-01 on Src and FAK, and various Src substrates (Akt, paxillin, and P130cas). In the sensitive RMUG-S cells, expression of p-Akt (ser473), p-paxillin (Tyr118), and p-P130cas (Tyr410) was significantly lower in cells treated with various concentrations of KX-01 for 24 hours than in control cells (Fig. 3A). In RMUG-L cells, expression of p-paxillin (Tyr118) and p-P130cas (Tyr410) was significantly lower whereas expression of p-Akt (ser473) was higher in cells treated with KX-01 than in control cells (Fig. 3B). Expression of p-Src (Tyr416), total-Src, p-FAK (pY397, Tyr576/577, pY861, Tyr925), and total-FAK was not significantly different between the treatment and control cells (data not shown).

The effects of treatment with oxaliplatin alone and the combination of oxaliplatin plus KX-01 on the expression and activation of Src substrates were analyzed in the RMUG-S cell line. Induction of p-Akt, p-paxillin, and p-P130cas activity was seen in the cells in response to oxaliplatin monotherapy, whereas KX-01 blocked oxaliplatin induced p-Akt, p-paxillin, and p-P130cas levels in the cells treated with the combination therapy (Fig. 3C).

To explore other potential pathways disturbed by KX-01, we utilized RPPAs. This analysis demonstrated that KX-01 inhibited p-Akt (Ser473) expression in RMUG-S cells, but not in RMUG-L cells, consistent with our Western blot results (Supplementary Table S1 and S2; Supplementary Fig. S5A and S5B). Expression of 72 molecules in RMUG-S cells and 60 molecules in RMUG-L cells significantly differed between cells treated with KX-01 and control cells, and expression of 35 of these molecules differed between treatment and control cells in both RMUG-S and RMUG-L cells (P < 0.05; Fig. 3D, Supplementary S5A and S5B). The majority of these proteins were direct or indirect downstream SFK members, acute stress-inducing proteins (MAPK pathway), and those associated with apoptosis and cell cycle regulation (Fig. 3E).

In both RMUG-S and RMUG-L cells treated with KX-01, expression of most SFK downstream proteins, such as p-p70s6k, p-S6, and mTOR, were inhibited, and expression of acute stress-inducing and apoptosis-related proteins was increased. However, changes in expression of proteins associated with cell-cycle regulation were completely opposite in the 2 cell lines, except for p21 (Fig. 3E); cyclin B1/CDC2 protein expression with Western blot confirmed these results (Supplementary Fig. S5C). Because one of the functions of cyclin B1 in combination with CDC2 (CDK1) is nuclear membrane breakdown, we stained RMUG-S and L cells with lamin B1 after treatment with KX-01. Lamin B1 in the nuclear membrane was disrupted to a greater extent in the RMUG-S cells than RMUG-L cells (Supplementary Fig. S5D and S5E). This suggests the relative sensitivity of RMUG-S and resistance of RMUG-L cells to KX-01 observed may be related to cell-cycle proteins.

**KX-01 inhibited tubule polymerization and disrupted the microtubule network**

To test whether KX-01 could directly affect the organization of the microtubule network, we treated RMUG-S and RMUG-L cells with KX-01 or vehicle for 24 or 48 hours and subsequently fixed and stained the cells with FITC-conjugated anti-α/β-tubulin antibody and Hoechst 33342 or DAPI for in situ observation of the microtubule network and nuclei with fluorescence microscopy. The microtubule network in control cells exhibited normal arrangement, with microtubules traversing intracellularly throughout the cell, and nuclei were intact and normal in appearance (Fig. 4A and B and Supplementary Fig. S6A and S6B). In contrast, cells treated with KX-01 had a smaller microtubule network, with a diffuse stain visible throughout the cytoplasm. Compared with RMUG-L cells, microtubules in RMUG-S cells were affected to a greater extent after treatment with KX-01 (Fig. 4A and B and Supplementary Fig. S6A and S6B). Condensation and fragmentation of nuclei were apparent in cells treated with KX-01 (Fig. 4A and B). Moreover, endoreduplication was also observed in cells treated with KX-01 (Fig. 4B and Supplementary Fig. S6A).

In tumor tissue from orthotopic mouse models, the staining patterns were similar to those of the in vitro tumor cells, and fluorescence intensity was markedly reduced in the KX-01 and combination groups compared with the control group (Fig. 4C and D). In the oxaliplatin group, the staining pattern of tubulin was unchanged from the control group (Fig. 4C and D).

**PTEN status affects mucinous ovarian carcinoma cell sensitivity to KX-01**

As shown, p-Akt (Ser473) was reduced by KX-01 in the sensitive RMUG-S cells, but not in RMUG-L cells. Expression of p-Akt (Ser473) was significantly higher in RMUG-L cells compared with RMUG-S cells (Fig. 3A and B). Given that PTEN regulates Akt activity via inhibition of Akt
phosphorylation, we examined PTEN status in both cell lines to explore potential underlying mechanisms involved in sensitivity to KX-01. PTEN is wild type in RMUG-S cells, whereas RMUG-L cells have absent PTEN expression (Supplementary Fig. S7A). As previous studies have shown that PTEN deficiency is associated with de novo resistance to epidermal growth factor receptor inhibitors (27, 28), we examined the relationship between PTEN deficiency and response to KX-01 in RMUG-L cells. RMUG-L cells were transfected with wild type or domain mutant (G129R, PTZB) PTEN and then exposed to various concentrations of KX-01 for 72 hours. Sensitivity to KX-01 was significantly enhanced in PTEN-restored cells; IC_{50} was undetectable in the control cells, but was 464 nmol/L in the PTEN-restored cells. In RMUG-L cells transfected with mutant PTEN, the sensitivity to KX-01 did not differ from that of control cells (Fig. 5A).

Next, we assessed cell proliferation, apoptosis, and cell-cycle status in control, wild type, and PTEN domain mutant RMUG-L cells treated with KX-01. In PTEN wild-type cells, compared with control and domain mutant cells, cell proliferation was significantly inhibited, apoptosis was significantly increased, and the proportion of S-phase cells was significantly decreased (Fig. 5B–D). In addition, Western blot showed that expression of p-Akt (ser473) and CDC2 was significantly inhibited in PTEN wild-type cells, whereas no change in expression was observed in the control and PTEN domain mutant cells (Fig. 5E).
Because *PTEN* is associated with lower levels of p-Akt (29), we also tested both RMUG-S and RMUG-L cells for sensitivity to an Akt1/2 inhibitor. Both RMUG-S and RMUG-L cells were sensitive to the Akt1/2 inhibitor (Supplementary Fig. S7B and S7D), and p-Akt expression was significantly decreased in both RMUG-S and RMUG-L cells after treatment with the Akt1/2 inhibitor (Supplementary Fig. S7C).

**Discussion**

This study establishes the ability of KX-01 to suppress cell growth effectively through targeting of both the Src pathway and tubulin in mucinous ovarian carcinoma cells *in vitro* and *in vivo*, and highlights *PTEN* deletion as a predictor of poor response to KX-01 treatment. Furthermore, KX-01 demonstrates synergistic antitumor activity in combination with oxaliplatin. These data suggest targeting both the Src pathway and tubulin with KX-01 alone, or in combination with oxaliplatin, may represent a more promising treatment for mucinous ovarian carcinoma than those currently available.

Preclinical studies in estrogen receptor–positive and estrogen receptor/progesterone receptor/HER2-negative breast cancers have demonstrated that KX-01 reduced proliferation and angiogenesis and increased apoptosis in xenograft tumors, and led to synergistic growth inhibition when combined with chemotherapeutic drugs (30, 31). KX-01 is currently in phase I/II clinical trials for the treatment of malignancies, including prostate cancer and acute myeloid leukemia (30).

Our RPPA data demonstrate that KX-01 significantly increased expression of MAPK pathway proteins, including p38, p-JNK, and p-Jun. Because microtubule disruption has been shown to activate MAPK pathway molecules (32), alterations of p38, p-JNK, and p-Jun observed after treatment with KX-01 suggest that it leads to microtubule disruption.
disruption. Immunofluorescence staining confirms that the microtubule network in both cell lines was disrupted by KX-01, and the appearance of the microtubules after treatment with KX-01 was similar to the appearance of microtubules after treatment with another tubulin polymerization inhibitory reagent (33). RPPA data did not demonstrate alterations in tubulin levels in RMUG-S and RMUG-L cells, suggesting that KX-01 may disrupt microtubule structure and organization, but not affect tubulin expression levels. As seen with other microtubule inhibitors, endoreduplication was observed in cells following treatment with KX-01 (34).

Our PTEN knock-in experiments reveal that PTEN mutation may contribute to relative resistance to KX-01. Cell-proliferation and cell-cycle data showed that the inhibition of cell proliferation and the proportion of S-phase cells differed between RMUG-S and RMUG-L cells, suggesting that KX-01 may disrupt microtubule structure and organization, but not affect tubulin expression levels. As seen with other microtubule inhibitors, endoreduplication was observed in cells following treatment with KX-01 (34).

Figure 5. PTEN deletion is involved in resistance to KX-01. A, MTT assay results showing cytotoxicity of KX-01 in RMUG-L cells transfected with wild-type (WT) and domain mutation (G129R, PTZB) PTEN and exposed to various concentrations of KX-01 for 72 hours. Cell survival was calculated as the number of cells surviving relative to the number of cells surviving in the control group. IC50 indicates median inhibitory rate. B, EdU assay results showing inhibition of cell proliferation in cells treated with 100 nM KX-01 for 72 hours. C, Annexin V and 7-aminoactinomycin D assay results showing cell apoptosis in cells treated with 100 nM KX-01 for 72 hours. D, flow cytometry results showing induction of mitotic arrest in cells treated with 100 nM KX-01 for 72 hours. E, Western blot results showing that p-Akt and CDC2 (cdk1) were inhibited in PTEN WT cells. Cells were treated with 100 nM KX-01 for 24 hours. β-Actin was used as the loading control. All experiments were repeated at least three times, and bars represent means with standard errors. *P < 0.05 and **P < 0.01 compared with control treatment (no KX-01).
Chemotherapeutic agent considered standard treatment for advanced-stage colorectal carcinoma, is a logical prospect in the treatment of mucinous ovarian carcinoma (15, 40). Sato and colleagues showed that 2 of the 5 mucinous ovarian carcinoma cell lines examined were sensitive to oxaliplatin, and the combination of oxaliplatin and fluorouracil was significantly inhibitory in 4 of the 5 cell lines (41). On the basis of these studies, we combined KX-01 with oxaliplatin in our experiments. Both our in vitro and in vivo experiments show the combination to be synergistic, resulting in enhanced antitumor activity compared with either monotherapy alone. These results validate the potential of the combination treatment strategy for translation to the clinical setting.

The mechanism underlying the effect of combination therapy is not completely clear. Our previous data showed that treatment with oxaliplatin alone induced Src kinase activity, which has been proposed as a potential mechanism of resistance to oxaliplatin in mucinous ovarian carcinoma, and that adding the Src/Abi inhibitor dasatinib to the treatment with oxaliplatin could attenuate the oxaliplatin-induced Src kinase activity, leading to synergistic effects (15). Because induction of Src kinase activity is thought to be associated with drug resistance via the Akt and Ras pathways (22, 42), our results suggest that KX-01 may inhibit oxaliplatin-induced expression of p-Akt, leading to the synergistic effects of the combination therapy.

In summary, our results demonstrate that KX-01 significantly suppresses mucinous ovarian carcinoma by targeting both Src substrates and tubulin. These dual effects may make KX-01 a particularly effective treatment for mucinous ovarian carcinoma. In addition, our results suggest that PTEN mutation may be a predictive biomarker of response to KX-01 treatment. Clinical studies using KX-01 in the treatment of mucinous ovarian carcinoma and investigating PTEN mutation as a potential biomarker are warranted.

Disclosure of Potential Conflicts of Interest

D. Hangauer has ownership interest (including patents) in Kinex Pharmaceuticals. No potential conflicts of interest were disclosed by the other authors.

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