A Phase I Trial of Vorinostat and Alvocidib in Patients with Relapsed, Refractory, or Poor Prognosis Acute Leukemia, or Refractory Anemia with Excess Blasts-2

Beata Holkova1,2, Jeffrey G. Supko7, Matthew M. Ames8, Joel M. Reid6, Geoffrey I. Shapiro7, Edward Brent Perkins1,2, Viswanathan Ramakrishnan1,3, Mary Beth Tomes1, Connie Honeycutt1, Renee M. McGovern9, Maciej Kmieciak1, Ellen Shrader1, Martha D. Wellons1, Heidi Sankala1, Austin Doyle9, John Wright9, John D. Roberts1,2, and Steven Grant1,2,4,5,6

Abstract

Purpose: This phase I study was conducted to identify the maximum-tolerated dose (MTD) of alvocidib when combined with vorinostat in patients with relapsed, refractory, or poor prognosis acute leukemia, or refractory anemia with excess blasts-2. Secondary objectives included investigating the pharmacokinetic and pharmacodynamic effects of the combination.

Experimental Design: Patients received vorinostat (200 mg orally, three times a day, for 14 days) on a 21-day cycle, combined with 2 different alvocidib administration schedules: a 1-hour intravenous infusion, daily \(/C2\); or a 30-minute loading infusion followed by a 4-hour maintenance infusion, weekly \(/C2\). The alvocidib dose was escalated using a standard 3+3 design.

Results: Twenty-eight patients were enrolled and treated. The alvocidib MTD was 20 mg/m2 (30-minute loading infusion) followed by 20 mg/m2 (4-hour maintenance infusion) on days one and eight, in combination with vorinostat. The most frequently encountered toxicities were cytopenias, fatigue, hyperglycemia, hypokalemia, hypophosphatemia, and QT prolongation. Dose-limiting toxicities (DLT) were cardiac arrhythmia-atrial fibrillation and QT prolongation. No objective responses were achieved although 13 of 26 evaluable patients exhibited stable disease. Alvocidib seemed to alter vorinostat pharmacokinetics, whereas alvocidib pharmacokinetics were unaffected by vorinostat. Ex vivo exposure of leukemia cells to plasma obtained from patients after alvocidib treatment blocked vorinostat-mediated p21CIP1 induction and downregulated Mcl-1 and p-RNA Pol II for some specimens, although parallel in vivo bone marrow responses were infrequent.

Conclusions: Alvocidib combined with vorinostat is well tolerated. Although disease stabilization occurred in some heavily pretreated patients, objective responses were not obtained with these schedules.

Clin Cancer Res; 19(7); 1873–83. ©2013 AACR.

Introduction

The outcome for adults with relapsed or refractory acute myelogenous leukemia (AML), acute lymphoblastic leukemia (ALL), high-risk myelodysplastic syndrome (MDS), and chronic myelogenous leukemia in blast crisis (CML-BC) remains poor. Of note, 20% to 30% of patients never achieve complete remission (CR) and fewer than 35% are cured. Furthermore, cures are achieved in fewer than 10% of patients at least 60 years of age. Current treatment strategies include chemotherapy, in which survival beyond 1 year is rare, or allogeneic stem cell transplant, for which there are few eligible patients and long-term survival rates are 20% to 40% (1). Histone deacetylase inhibitors (HDACIs) act by modifying chromatin structure and gene expression (2). They kill transformed cells through diverse mechanisms, including induction of oxidative injury and death receptors, interference with chaperone protein function and DNA repair, and
Translational Relevance

This phase I study defines the maximum-tolerated dose of alvocidib that can be administered in combination with vorinostat in patients with relapsed, refractory, or poor prognosis acute leukemia, or refractory anemia with excess blasts-2. Several heavily pretreated patients exhibited disease stabilization although there were no objective responses. Plasma obtained from a subset of patients the day after alvocidib was administered blocked certain histone deacetylase inhibitor–associated actions (e.g., p21CIP1 induction) in leukemia cells that were exposed to vorinostat in vivo. However, this and other pharmacodynamic effects were detected infrequently in bone marrow blasts obtained from patients who were treated with the 2 agents. These findings highlight the importance of identifying factors that prevent targeted antileukemic agents from recapitulating their in vitro actions in bone marrow blasts following in vivo administration and raise the possibility that circumventing this problem might improve the antileukemic activity of such regimens.

upregulation of proapoptotic proteins (e.g., Bim), among others (3). Multiple HDACis, representing diverse chemical classes, have been investigated as anticancer agents. Vorinostat is a pan-HDACi that received U.S. Food and Drug Administration (FDA) approval for the treatment of patients with progressive, persistent, or recurrent cutaneous T-cell lymphoma (4). Vorinostat has shown some, albeit limited, single-agent activity in AML/MDS (5).

Alvocidib (flavopiridol) is a rohitukine alkaloid that inhibits cyclin-dependent kinases (CDK) 1, 2, 4/6, 7, and 9 (6) and in some cell types downregulates expression of cyclin D1 (7). Alvocidib also acts as a transcriptional repressor by inhibiting the CDK9/cyclin T pTEFb (positive transcription elongation factor b) complex (8). Besides inducing cell-cycle arrest (9), alvocidib potently triggers apoptosis in tumor cells, particularly those of hematopoietic origin (10). Alvocidib downregulates various antiapoptotic proteins including p21CIP1, cyclin D1, McI-1, XIAP, and Bag-1 (11–15). In human leukemia cells, alvocidib-mediated apoptosis has been related to induction of mitochondrial injury, characterized by loss of mitochondrial membrane potential and release of cytochrome c into the cytosol (16). Preclinical studies have also shown synergism between alvocidib and cytotoxic agents (17).

In preclinical studies involving human leukemia cells, coadministration of alvocidib with differentiating agents, such as the HDACis sodium butyrate and vorinostat, potently induces leukemic cell apoptosis (11, 12, 18–21). This may reflect alvocidib-mediated transcriptional repression of several antiapoptotic genes required for leukemic cell differentiation, including p21CIP1 and McI-1 (18). We have reported that in human leukemia cells, alvocidib blocks vorinostat-mediated p21CIP1 induction, diminishes expression of various antiapoptotic proteins and induces pronounced apoptosis (22).

These findings raised the possibility that alvocidib administered either before or concurrently with vorinostat might exert similar actions in vivo, resulting in enhanced antileukemic effects in patients with acute leukemia. The primary goal of this study was to establish the MTD for alvocidib when combined with vorinostat (200 mg orally, 3 times a day, for 14 days) in patients with relapsed, refractory, or poor prognosis acute leukemia, or refractory anemia with excess blasts-2 (RAEB-2). Pharmacokinetic studies were conducted to assess interactions between the drugs. In addition, pharmacodynamic studies were conducted to determine whether patient plasma—after alvocidib treatment—could recapitulate known alvocidib actions (e.g., inhibition of p21CIP1 induction) in human leukemia cells (1937) exposed to vorinostat in vitro and if analogous events occurred in leukemic bone marrow cells following in vivo administration of these agents to patients.

Patients and Methods

Drug supply

Vorinostat (SAHA; NSC 701852) and alvocidib (flavopiridol; NSC 649890) were supplied by Merck and Co., Inc. and Sanofi-Aventis Pharmaceuticals, Inc., respectively, through the U.S. National Cancer Institute (NCI).

Patient eligibility

Eligible patients were at least 18 years of age with an Eastern Cooperative Oncology Group performance status 2 or less and a diagnosis of relapsed or primary refractory AML, ALL, acute leukemia following prior MDS, CML-BC following prior imatinib therapy, RAEB-2, or acute leukemia in patients 60 years or older (no requirement for prior therapy). Additional eligibility criteria included normal to near-normal kidney and liver function; no prior autologous or allogeneic bone marrow/stem cell transplantation; and a baseline white blood cell count (WBC) < 50,000/μL. Hydroxyurea and/or leukapheresis was permitted to reduce the WBC, but hydroxyurea must have been discontinued 24 hours before study initiation.

Treatment plan

This phase I trial was a nonrandomized, dose-escalation study to identify the MTD for alvocidib given in combination with vorinostat. Vorinostat was administered orally, at a dose of 200 mg 3 times a day, on days 1 to 14. Initially, alvocidib was given as a 1-hour intravenous infusion on days 1 to 5 of a 21-day cycle. The starting dose was 10 mg/m² and the dose was escalated by constant increments of 10 mg/m², as permitted by toxicity.

After completing the evaluation of dose level 4 for this daily short infusion (DSI) schedule, the protocol was amended, based on recommendations of CTEP, to evaluate administration of alvocidib with a weekly loading-maintenance infusion (WLMI) schedule based on promising evidence of activity in patients with chronic lymphocytic
leukemia (CLL; ref. 23) and also to improve the safety in patients receiving alvocidib. Specifically, a loading dose given as a 30-minute intravenous infusion was followed immediately by a 4-hour maintenance infusion on days 1 and 8 of a 21-day cycle. The dose levels (loading dose/maintenance dose, in mg/m²) were as follows: dose level 5, 20/20; dose level 6A, 30/20; dose level 6B, 30/30.

In general, patients were treated every 3 weeks. Disease status was assessed by bone marrow aspirate and biopsy before cycle 2 or as clinically indicated. Patients with no evidence of disease progression (reflected by an increase in bone marrow blasts in marrows exhibiting ≥50% cellularity) and no evidence of significant side effects, were eligible to receive multiple treatment courses without interruption.

All patients received full supportive care, including herpes zoster prophylaxis. With the WLMI schedule, QTc interval monitoring was included because of vorinostat safety issues. Clinical issues unique to acute leukemia and alvocidib infusion, such as hyperacute tumor lysis syndrome (TLS) and cytokine release syndrome (characterized by fever, bronchospasm, altered blood pressure, myalgias, arthralgias, tumor pain, and/or urticarial rash; ref. 23), necessitated rigorous supportive care regimens with the WLMI schedule, that is, prophylaxis, monitoring, and treatment of TLS during the first course (doses 1 and 2) of alvocidib and for subsequent infusions if clinically indicated. Patients also received oral dexamethasone, 20 mg before the first and second dose of alvocidib, to prevent cytokine release syndrome.

Study design, definition of DLT, and identification of the MTD

This study used a 3+3 dose-escalation design, with expansion to 6 patients if a dose-limiting toxicity (DLT) was observed in any of the initial 3 patients evaluated. The MTD was defined as the highest dose level at which fewer than 2 of 6 patients experienced a DLT during the first cycle of therapy. DLT was defined as any of the following adverse events that were possibly, probably, or definitely related to study treatment: (i) any grade 4 nonhematologic toxicity, except infection or hyperbilirubinemia; (ii) any grade 3 nonhematologic toxicity lasting longer than 7 days; (iii) a grade 4 absolute neutrophil count or platelet toxicity persisting longer than 6 weeks in the absence of leukemia.

Toxicity evaluation

Adverse events were characterized in terms of attribution, severity, and study treatment relatedness according to the NCI-Common Terminology Criteria for Adverse Events version 3.0.

Response evaluation

Response criteria were as described by Cheson and colleagues (24, 25). Patients who did not experience a CR during the treatment course were allowed to continue on study treatment if they did not experience significant toxicity in the absence of disease progression.

Pharmacokinetic studies

The effect of vorinostat on the plasma pharmacokinetics of alvocidib was assessed during the first cycle of therapy by comparing data for the initial dose of alvocidib with that for a subsequent dose, administered after patients had received alvocidib for either 3 or 7 days, depending upon the alvocidib dosing schedule. For the DSI schedule, blood specimens were obtained before dosing, then at 30, 55, and 70 minutes, and 2, 3, 4, 8, and 24 hours after starting the infusion on days 1 and 4. For the WLMI schedule, blood specimens were obtained before dosing, then at 15 and 25 minutes, and 1, 2, 3, 4, 5, 6, 8, and 24 hours after starting the infusion on days 1 and 8. Blood samples (6 mL) were drawn from a peripheral arm vein into sodium heparin collection tubes and centrifuged (1,100–1,300 × g, 4°C, 10 minutes). The plasma was stored at −70°C until assayed. Plasma samples (100 μL) were prepared for the analysis of alvocidib by adding 7 μL of internal standard working solution [150-ng/mL bis(O-methyl)flavipridol in acetonitrile] and 200 μL of acetonitrile to plasma with vigorous mixing (26). After centrifugation (10,000 × g, 5 minutes), the supernatant (200 μL) was diluted with 25-mmol/L ammonium formate buffer, pH 2.75 (200 μL), whereupon 100 μL was injected onto a 150 mm × 4.6 mm Luna 5-μm C8 high-performance liquid chromatography column (Phenomenex) and eluted with acetonitrile/25-mmol/L ammonium formate buffer, pH 2.75 (30:70 v/v) at 1.0 mL/min. An Agilent Technologies 1100 Series LC/MSD system with an electrospray ionization interface was used for detection. Nitrogen was used as the nebulizing (50 p.s.i.) and drying gas (12 L/min, 350°C). Positive ions corresponding to the [M+H]⁺ ions of alvocidib at m/z 402.1 and the internal standard at 430.1 were measured by selected-ion monitoring with a 2,500 V capillary voltage, 130 V fragmentor voltage, and 289 milliseconds dwell time. Extracted ion chromatograms were integrated to provide peak areas. The analytic method was validated and applied to the routine analysis of study samples according to published guidelines (27). Calibration standards of alvocidib in human donor plasma had concentrations ranging from 1 to 100 ng/mL (2.3–228 nmol/L). Samples were assayed in 28 runs during a period of 24 months. The calibration curves had correlation coefficients 0.998 or more. Interday accuracy was within ±4.4% of the known concentration of the calibration standards and quality control solutions with a precision 7.7% or less.

Blood samples to define the plasma profile for the first dose of vorinostat were collected before dosing and 0.5, 1, 2, 2.5, 3, 4, and 8 hours after dosing. Sampling according to this schedule was also conducted for the morning dose of vorinostat when given together with alvocidib on day 4 for the DSI schedule and day 8 for the WLMI schedule. Serum was harvested from the blood and assayed for vorinostat, vorinostat glucuronide, and 4-anilino-4-oxobutanoic acid (VA) using a validated liquid chromatography–electrospray ionization tandem mass spectrometry method as previously described in refs. (28, 29).
Plasma concentration–time curves for each patient were analyzed by standard noncompartmental methods using WinNonlin Professional 5.0 software (Pharsight Corp.; ref. 30). Mean values of the pharmacokinetic variables were calculated as the geometric mean ± SD of the individual patient values (31, 32). The paired two-tailed t test was used to compare the overall mean total body clearance for the first dose of alvocidib to the doses given on day 4 for the DSI schedule) or day 8 for the WLMI schedule using logarithmically transformed data. \( P < 0.05 \) was considered to be significantly different.

Pharmacodynamics

Collection of plasma and bone marrow samples. Bone marrow aspirate and peripheral blood (3–5 mL) samples were obtained before treatment and on day 2, approximately 24 hours after the first dose of alvocidib. Blood samples were processed to obtain plasma and stored at −80°C before use for ex vivo studies. Bone marrow mononuclear cells were isolated from the aspirates with the Accuspin System-Hisopaque-1077 (Sigma-Aldrich) according to manufacturer’s instructions and stored at −80°C for Western blot analysis as before (33). Only bone marrow samples with a blast count of at least 59% were assayed to avoid problems in interpretation due to cellular heterogeneity.

Vorinostat and alvocidib ex vivo studies. U937 human leukemia (myelomonocytic) cells were obtained from the American Type Culture Collection and cultured, as previously reported (34). Cells (0.2 × 10⁶/mL) were treated with 1-μmol/L vorinostat in the presence of 90% patient plasma obtained before or 24 hours after treatment with alvocidib and stored at −80°C for Western blot analysis. See Supplementary Methods for details.

Human investigation studies

These studies were conducted after Institutional Review Board approval and in accordance with an assurance filed with and approved by the Department of Health and Human Services. Informed consent was obtained from each subject.

Results

Patient characteristics

A total of 14 patients, 8 male and 6 female, were treated with the alvocidib DSI schedule (Table 1). The median age of the patients was 63 years (range, 20–78). Thirteen patients had AML, 1 patient had ALL. The median number of prior regimens was 2.5 (range, 1–5). The patients received a median of 1.5 courses of study treatment (range, 1–4).

A total of 14 patients, 8 male and 6 female, were treated with the alvocidib WLMI schedule (Table 1). The median age of the patients was 66 years (range, 24–76). All 14 patients enrolled to the WLMI schedule had AML. The median number of prior regimens was 3 (range, 1–5). The patients received a median of 1.5 courses of study treatment (range, 1–10).

Safety and tolerability

Overall, the treatment was well tolerated, with toxicities that were transient and/or manageable, and generally as expected for an acute leukemia regimen (Table 2). Fatigue was the most common nonhematologic toxicity. Hyperacute TLS was not seen but aggressive prophylaxis and monitoring were integral to the treatment plan for the WLMI schedule. There were no episodes of cardiac arrhythmias. Ten patients died while enrolled in the study and all deaths were reported as being unrelated to treatment.

DLT and MTD

An MTD was not reached for the alvocidib DSI schedule before the protocol was amended to evaluate the WLMI schedule for all additional patients enrolled into the study. Four patients were initially treated at dose level 5 (20 mg/m² loading infusion; 20 mg/m² maintenance infusion). No DLTs were seen in 3 of the patients, and 1 patient died during cycle 1 due to a fungal infection considered unrelated to treatment. A single patient was treated at dose level 6 (30 mg/m² loading infusion; 20 mg/m² maintenance infusion) without a DLT. This dose level was later noted to have been erroneously defined by the dose-escalation schema.

Table 1. Patient enrollment and characteristics

<table>
<thead>
<tr>
<th></th>
<th>DSI schedule</th>
<th>WLMI schedule</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender (number of patients)</td>
<td>Male</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>Female</td>
<td>10</td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>14</td>
</tr>
<tr>
<td>Age, y</td>
<td>Median</td>
<td>63</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>20–78</td>
</tr>
<tr>
<td>≥60 y old (number of patients)</td>
<td>10</td>
<td>12</td>
</tr>
<tr>
<td>Performance status (number of patients)</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>7</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>14</td>
</tr>
<tr>
<td>Diagnosis (number of patients)</td>
<td>AML</td>
<td>13</td>
</tr>
<tr>
<td></td>
<td>ALL</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>CML-BC</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>RAEB-2</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>14</td>
</tr>
<tr>
<td>Prior treatment (number of regimens)</td>
<td>Median</td>
<td>2.5</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>1–5</td>
</tr>
<tr>
<td></td>
<td>&gt;2 prior regimens (no. of patients)</td>
<td>4</td>
</tr>
<tr>
<td>Study treatment received (number of courses)</td>
<td>Mean</td>
<td>2.07</td>
</tr>
<tr>
<td></td>
<td>Median</td>
<td>1.5</td>
</tr>
<tr>
<td></td>
<td>Range</td>
<td>1–4</td>
</tr>
</tbody>
</table>
This level was redesignated 6A and no further patients enrolled. Dose level 6B was added with the corrected dose combination of 30 mg/m² loading infusion and 30 mg/m² maintenance infusion. Six patients were treated on dose level 6B. Two of these patients experienced a DLT: a grade 3 cardiac arrhythmia-atrial fibrillation in 1 and a grade 3 QT prolongation in the other. The toxicities resolved with discontinuation of vorinostat but recurred when vorinostat was restarted. Dose level 6B was deemed to have exceeded the MTD. Consequently, dose level 5 was expanded. None of the 3 additional patients treated at dose level 5 experienced a DLT, establishing it as the MTD (Table 3).

### Disease response

All but 2 treated patients were evaluable for response. One patient treated with the alvocidib WLMI schedule expired because of fungal infection before the disease assessment time point, and the second patient treated with the DSI schedule was removed from the study due to adverse events and did not complete disease assessment. None of the 26 evaluable patients achieved a CR. Disease stabilization was evident in 6 patients treated with the alvocidib WLMI schedule (median number of treatment cycles: 3; range, 2–5) and 7 patients receiving the WLMI schedule (median number of treatment cycles: 2; range, 2–10). One heavily pretreated patient (7+3, 5+2, FLAG, Mylotarg) with refractory AML remained on treatment for 11 cycles before death due to infection (Supplementary Table S1).

### Pharmacokinetic studies

Pharmacokinetic parameters for both the initial dose of alvocidib and a subsequent dose given to the same patient after receiving vorinostat (200 mg orally, 3 times a day) for either 3 or 7 days were determined for a total of 18 patients. Data were obtained from 9 patients receiving the DSI schedule and 9 patients treated with the WLMI schedule. The more protracted infusion schedule resulted in a lower maximum concentration of the drug in plasma ($C_{\text{max}}$) although the duration of time that drug levels remained near the $C_{\text{max}}$ before decay was prolonged. Mean ($\pm$SD) values of the $C_{\text{max}}$ and clearance of alvocidib at each dose level evaluated in the study are presented in Supplementary Table S2. Drug accumulation upon repeated dosing was insignificant for the daily and weekly infusion schedules as the drug concentration in plasma samples obtained immediately before dosing on days 4 or 8 was less than 1% of the subsequent $C_{\text{max}}$ on average. The overall mean clearance of alvocidib for the initial doses in the DSI ($13.0 \pm 4.8 \text{ L/h/m}^2$) and WLMI ($12.2 \pm 5.3 \text{ L/h/m}^2$) schedules were not significantly different ($P = 0.75$). The mean clearances for the dose given on day 4 of the DSI schedule ($9.9 \pm 5.4 \text{ L/h/m}^2$; $P = 0.15$) and day 8 of the WLMI schedule ($11.0 \pm$)
were not significantly different from the mean clearance for the initial dose of the respective infusion schedule.

Pharmacokinetic data for vorinostat and its major circulating metabolites, vorinostat glucuronide and VA, for the initial dose of the drug were obtained from 28 patients (Supplementary Table S3). When vorinostat was administered with alvocidib using the DSI schedule, there was no change in $T_{\text{max}}$, $C_{\text{max}}$, and AUC0–8 h for vorinostat, VA, or vorinostat glucuronide on day 1 or 4 when alvocidib was escalated over the 10 to 40 mg/m² dose range. Administration of vorinostat with alvocidib using the WLMI schedule resulted in 25% to 50% higher vorinostat $C_{\text{max}}$ and area under curve (AUC) values when compared with the DSI schedule. In contrast, the $C_{\text{max}}$ and AUC of vorinostat glucuronide were 50% lower when given together with the 60 mg/m² alvocidib dose compared with the 40 mg/m² dose using the WLMI schedule.

**Pharmacodynamic studies.**

*In vivo changes in p21$^{\text{CIP1}}$, Mcl-1, and p-RNA Pol II expression.* Protein levels of p21$^{\text{CIP1}}$, Mcl-1, and phosphorylated RNA polymerase II (p-RNA Pol II), which have been shown to be downregulated in leukemia cells exposed to vorinostat and alvocidib in vitro (18, 19), were monitored in bone marrow mononuclear cells from 7 patients before and approximately 24 hours after the first dose of alvocidib (Fig. 1). Changes in the pharmacodynamic markers measured were variable and did not correlate with the DSI versus the WLMI schedules of alvocidib, with p21$^{\text{CIP1}}$ downregulation observed in 1 patient (14.3%), no change in 2 patients, and upregulation of p21$^{\text{CIP1}}$ in 4 patients. For Mcl-1 protein levels, 3 samples (42.9%) exhibited downregulation post-alvocidib, no change was observed in 1 sample, and Mcl-1 upregulation was detected in 3 samples (Fig. 1). Of note, changes in p21$^{\text{CIP1}}$ and Mcl-1 were concordant in some samples (e.g., #32) but discordant in others (e.g., #14). p-RNA Pol II downregulation post-alvocidib treatment was observed in 2 samples (28.6%), no change in 4 samples, and 1 sample displayed p-RNA Pol II upregulation. Of note, concordance was observed between changes in p-RNA Pol II and p21$^{\text{CIP1}}$/Mcl-1 expression in some samples (e.g., #32) but not others (e.g., #2). Downregulation of all 3 assayed proteins occurred in only 1 of 7 patients (14.3%), that is, #32 (WLMI schedule). Conversely, upregulation of all 3 assayed proteins also occurred in 1 of 7 patients (14.3%).

**Table 3. Dose levels and DLTs**

<table>
<thead>
<tr>
<th>DSI schedule</th>
<th>Vorinostat (mg total dose) orally, TID days 1–14</th>
<th>Alvocidib-loading dose</th>
<th>Alvocidib (mg/m²) 1-h intravenous infusion days 1–5</th>
<th>Patients treated/DLT</th>
<th>DLT</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>200</td>
<td>—</td>
<td>10</td>
<td>4/0</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>200</td>
<td>—</td>
<td>20</td>
<td>2/0</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>200</td>
<td>—</td>
<td>30</td>
<td>4/0</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>200</td>
<td>—</td>
<td>40</td>
<td>3/1</td>
<td>Grade 4 infection with neutropenia (sepsis &gt;7d)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>WLMI schedule</th>
<th>Vorinostat (mg total dose) orally, TID days 1–14</th>
<th>Alvocidib-loading dose</th>
<th>Alvocidib (mg/m²) 4-h intravenous infusion days 1, 8</th>
<th>Patients treated/DLT</th>
<th>DLT</th>
</tr>
</thead>
<tbody>
<tr>
<td>5a</td>
<td>200</td>
<td>20</td>
<td>20</td>
<td>7/0</td>
<td>Grade 3 atrial fibrillation grade</td>
</tr>
<tr>
<td>6Aa</td>
<td>200</td>
<td>30</td>
<td>20</td>
<td>1/NEa</td>
<td>3 QTc prolongation (intermittent, precluded vorinostat dosing for &gt;7d)</td>
</tr>
<tr>
<td>6Bb</td>
<td>200</td>
<td>30</td>
<td>30</td>
<td>6/2</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviation: TID, 3 times a day.

aDefined as the MTD.
bExceeded the MTD.
cLevel defined by clerical error in study protocol and was aborted after 1 patient enrolled.
dPatient did not complete cycle 1 of treatment so not evaluable (NE) for DLT.
regulation of p21CIP1, Mcl-1, and p-RNA Pol II as previously reported (18, 19). Interestingly, p21CIP1, Mcl-1, and p-RNA Pol II expression were monitored (Fig. 2). Results for U937 cells treated with 1-μmol/L vorinostat and alvocidib—a 1-hour intravenous loading dose level and in vivo expression of these proteins could be discerned. Collectively, these findings indicate that in vivo administration of vorinostat and alvocidib results in highly variable and in some cases discordant changes in the expression of p21CIP1, Mcl-1, and p-RNA Pol II in leukemic bone marrow cells, and that in vitro perturbations observed in preclinical studies, for example, downregulation of these proteins (18, 19), are recapitulated in only a minority of patients.

Ex vivo studies. Alvocidib is highly bound to plasma proteins (35), raising the possibility that free levels of the compound achieved in the plasma of patients treated at MTDs of the drug may be insufficient to recapitulate the pharmacologic effects observed in vitro (18, 19). Plasma samples from 19 patients were collected before treatment and approximately 24 hours after the first dose of alvocidib. U937 cells were treated with 1-μmol/L vorinostat in the presence of 90% patient pre- or post-alvocidib treatment plasma, after which effects on p21CIP1, Mcl-1, and p-RNA Pol II expression were monitored (Fig. 2). Results for U937 cells treated (24 hours) in vitro with 1-μmol/L vorinostat ± 150-nmol/L alvocidib, showed virtually complete downregulation of p21CIP1, Mcl-1, and p-RNA Pol II as previously reported (18, 19). Interestingly, p21CIP1, Mcl-1, and p-RNA Pol II expression was downregulated by posttreatment plasma (compared with expression in the presence of pre-treatment plasma) in a subset of patients (i.e., 73.7, 68.4, and 84.2%, respectively), regardless of the alvocidib schedule. In some of the samples (e.g., #16), discordance was observed with respect to the ex vivo effects of plasma on U937 cell expression of p21CIP1, Mcl-1, and p-RNA Pol II (i.e., downregulation) and in vivo effects on bone marrow mononuclear cells (i.e., upregulation of each of the proteins; Fig. 1). The discordance between perturbations in expression of p21CIP1, Mcl-1, or p-RNA Pol II were less pronounced than that observed in posttreatment bone marrow samples. As in the case of the in vivo studies, ex vivo effects on these proteins were not clearly associated with dose level. Together, these findings suggest that in a subset of patients, plasma alvocidib concentrations can be achieved that are sufficient to mimic previously described in vitro actions (downregulation of p21CIP1, Mcl-1, and p-RNA Pol II; refs. 18, 19) but raise the possibility that yet to be determined factors limit the ability of alvocidib to recapitulate these actions in bone marrow leukemia cells following in vivo administration of vorinostat and alvocidib to patients.

### Discussion

The results of this study show that vorinostat can be safely administered in combination with alvocidib—administered according to the WLMI schedule—in patients with relapsed/refractory leukemias; whether tolerable doses will be therapeutically relevant remains to be determined. The MTD for this regimen was vorinostat at 200 mg 3 times a day and alvocidib administered as a 1-hour intravenous loading
infusion at a dose of 20 mg/m² followed by a 20 mg/m² 4-hour infusion on days 1 and 8 of a 21-day cycle. Fatigue was the most common nonhematologic toxicity and myelosuppression was the most common hematologic toxicity. Notably, TLS, which has been reported in patients with high-count CLL receiving a WLMI alvocidib schedule (36), was not observed. Nevertheless, given the potentially lethal consequences of this syndrome, aggressive monitoring and prophylaxis seems warranted. In a recently completed phase I study of vorinostat and alvocidib in patients with solid tumors, the MTD was determined to be 800 mg of vorinostat on days 1 to 3 and 15 to 17 with alvocidib given by the WLMI schedule at a dose of 30 mg/m², delivered during the initial 30-minute infusion and another 30 mg/m² given over 4 hours on days 2 and 16, with cycles repeated every 28 days (37). The principal toxicity of this regimen was myelosuppression as observed in the present study.

The concurrent oral administration of vorinostat at a dose of 200 mg 3 times a day did not seem to modify alvocidib pharmacokinetics. The clearance of alvocidib, determined after patients received vorinostat for either 3 or 7 days, was in agreement with the mean values previously reported for

![Figure 2](image-url)
clinical studies of single-agent alvocidib given as a daily 1-hour intravenous infusion or a weekly 30-minute loading and 4-hour maintenance infusion (38-40). As observed previously (37), concomitant administration with alvocidib seemed to modify the pharmacokinetics of vorinostat when alvocidib was administered using the WLM1 schedule, but not with the DSI schedule, and was most apparent when the combined alvocidib dose was increased to 60 mg/m². As the increased vorinostat Cmax and AUC values were accompanied by a concomitant reduction in vorinostat glucuronide, the glucuronide metabolite of vorinostat, this interaction may be attributed to inhibition of one or more glucuronosyl transferases that metabolize both alvocidib and vorinostat (41, 42).

Although for various reasons (e.g., limited patient numbers, treatment with different drug doses), phase I trials are not equipped to assess regimen efficacy, an attempt was made to describe clinical responses. The best response in this heavily pretreated patient population with relapsed or refractory leukemia was stable disease (Supplementary Table S1). Nevertheless, several patients with highly refractory disease were able to continue treatment with stable disease for up to 11 months. Although single-agent vorinostat has some, albeit limited, activity in AML (5), evidence of alvocidib activity in this disease, in contrast to CLL (36), is lacking. Whether alvocidib can enhance vorinostat antileukemic activity in vivo, as observed in vitro (15), remains to be determined, for example, through the conduct of an appropriately powered phase II trial involving uniform drug doses and a larger number of patients. In this context, encouraging results in AML have recently been reported for a regimen in which alvocidib was combined with cytotoxic chemotherapy [mitoxantrone and ara-C (35)]. Additional studies will be necessary to determine, for example, whether HDACIs can increase the activity of genotoxic agents in AML (43), the concept of combining the vorinostat/alvocidib regimen with such agents warrants consideration, if justified by preclinical evidence of efficacy.

Although there were exceptions, the effects of the vorinostat/alvocidib regimen on bone marrow mononuclear cell expression of p21CIP1 (11, 12), Mcl-1 (13, 14), and p-RNA Pol II (44, 45) revealed a lack of downregulation of these proteins in most cases (Fig. 1). Although such findings are in accord with the lack of objective responses, the limited sample size precludes drawing definitive conclusions from these data. The possibility that vorinostat was unable to recapitulate its known in vitro actions seems less likely in view of previous evidence of activity against its biologic targets in vivo (46), and the observation that upregulation of p21CIP1, a major vorinostat target (47), occurred in multiple samples. However, dose-dependent effects of vorinostat cannot be excluded (18, 21). In addition, it has previously been suggested that yet to be defined plasma or microenvironmental factors may limit the impact of alvocidib on bone marrow cells in vivo (35). Additional studies will be required to distinguish between these possibilities.

An alternative explanation for these findings is that sustained plasma alvocidib concentrations were insufficient to exert anticipated biologic effects, or alternatively, plasma protein binding may have substantially reduced effective alvocidib concentrations, as previously observed in the case of other targeted agents (48, 49). To address this issue, U937 cells were exposed to vorinostat in the presence of pretreatment and 24-hour posttreatment plasma, after which p21CIP1, Mcl-1, and p-RNA Pol II expression were monitored. Interestingly, posttreatment plasma downregulated these proteins in vorinostat-treated U937 cells for 68.4% to 84.2% of samples, but analogous changes in bone marrow bone marrow blasts following administration of alvocidib and vorinostat occurred relatively rarely (14.3%-42.9%). Disparate responses were also noted in several instances in which plasma samples and bone marrow mononuclear cells from the same patients were evaluated. The discordance between these ex vivo actions and effects on bone marrow mononuclear cells from the same patients was stable. For example, in vitro actions and effects on bone marrow microenvironmental factors (35), as alluded to earlier. Alternatively, they may reflect disparate responses of continuously cultured (e.g., U937) cells versus primary AML blasts. Finally, the presence of other non-CDK target proteins (40 and 120 kDa) with high affinities to alvocidib in tumor cells cannot be excluded.

In summary, the present results show that a regimen combining alvocidib and vorinostat in patients with refractory AML is tolerable at pharmacologically relevant doses, although clear evidence of activity could not be shown in this heavily pretreated patient population. Moreover, postalvocidib plasma samples recapitulated the in vitro effects of exogenously administered alvocidib in a subset of specimens assayed. On the other hand, in vivo administration of alvocidib failed to downregulate key targets (i.e., p21CIP1, Mcl-1, and p-RNA Pol II) in most patient-derived bone marrow cells, raising the possibility that this phenomenon might contribute to the minimal activity of this regimen observed in the present study. Elucidation of the factors responsible for this discordance may assist in future efforts designed to improve the antileukemic activity of alvocidib and possibly other CDK inhibitors. Finally, while the very limited activity observed, mainly in patients with relapsed/refractory AML, may reduce enthusiasm for successor phase II trials, it is conceivable that the antileukemic activity of this strategy may be improved by the addition of other agents (e.g., ara-C), or the use of newer and potentially more effective CDK inhibitors, for example, SCH727965 (50). Preclinical studies designed to test these possibilities are currently underway. There are currently no plans to proceed with a phase II clinical trial with this particular combination in patients with relapsed or refractory leukemia.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: B. Holkova, J.G. Supko, G.I. Shapiro, J. Wright, J.D. Roberts, S. Grant
Development of methodology: B. Holkova, J.G. Supko, M.M. Ames, J.D. Roberts, S. Grant
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): B. Holkova, J.G. Supko, M.M. Ames, G.I. Shapiro, E.B. Perkins, M.B. Tombes, C. Honeycutt, M. Kniecak, E. Strader, J.D. Roberts

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): B. Hollkova, J.G. Supko, J.M. Reid, V. Ramakrishnan, M.B. Tombres, M. Kmiecik, E. Shrader, S. Grant Writing, review, and/or revision of the manuscript: B. Hollkova, J.G. Supko, J.M. Reid, G.I. Shapiro, M.B. Tombres, M. Kmiecik, E. Shrader, H. Sankala, A. Doyle, J. Wright, S. Grant Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): J.G. Supko, J.M. Reid, G.I. Shapiro, M.B. Tombes, M. Kmieciak, E. Shrader, H. Sankala, A. Doyle, J. Wright, S. Grant

Study supervision: B. Hollkova, J.G. Supko, E.B. Perkins, C. Honeycutt, A. Doyle, J. Wright, J.D. Roberts, S. Grant

Acknowledgments
The authors thank Sarah Kolla for assistance with the pharmacodynamic studies and Kevin T. Hogan for editorial assistance with the article.

References

In Memoriam
The authors would like to dedicate this article to the memory of Dr. Igor Delgado-Espinosa.

Grant Support
This study was supported by NIH grants R21 CA115260, R01 CA93738, P30 CA016059, M01 RR00065, U01 CA9912, 23XS026, P30 CA15083, and P30 CA60516.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received September 14, 2012; revised December 14, 2012; accepted January 3, 2013, published OnlineFirst March 20, 2013.

1882 Clin Cancer Res; 19(7) April 1, 2013 Clinical Cancer Research

Downloaded from clinicanerrse.aacrjournals.org on July 14, 2017. © 2013 American Association for Cancer Research.


A Phase I Trial of Vorinostat and Alvocidib in Patients with Relapsed, Refractory, or Poor Prognosis Acute Leukemia, or Refractory Anemia with Excess Blasts-2

Beata Holkova, Jeffrey G. Supko, Matthew M. Ames, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-12-2926

Supplementary Material
Access the most recent supplemental material at:
http://clincancerres.aacrjournals.org/content/suppl/2013/09/27/1078-0432.CCR-12-2926.DC1

Cited articles
This article cites 49 articles, 35 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/19/7/1873.full#ref-list-1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.