

## Potentially Prognostic miRNAs in HPV-Associated Oropharyngeal Carcinoma

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### Abstract

**Purpose:** Deregulation of miRNAs is associated with almost all human malignancies. Human papillomavirus (HPV)-associated oropharyngeal carcinoma (OPC) has a significantly more favorable outcome compared with HPV-negative OPCs; however, the underlying mechanisms are not well understood. Hence, the objectives of this study were to determine whether miRNA expression differed as a function of HPV status and to assess whether such miRNAs provide prognostic value beyond HPV status.

**Methods:** Global miRNA profilings were conducted on 88 formalin-fixed and paraffin-embedded (FFPE) OPC biopsies (p16-positive: 56; p16-negative: 32), wherein the expression levels of 365 miRNAs plus 3 endogenous controls were simultaneously measured using quantitative real-time (qRT)-PCR. Seven FFPE specimens of histologically normal tonsils were used as controls.

**Results:** Overall, 224 miRNAs were expressed in more than 80% of the investigated samples, with 128 (57%) being significantly differentially expressed between tumor versus normal tissues ( $P < 0.05$ ). Upregulated miR-20b, miR-9, and miR-9\* were significantly associated with HPV/p16-status. Three miRNA sets were significantly associated with overall survival (miR-107, miR-151, miR-492;  $P = 0.0002$ ), disease-free survival (miR-20b, miR-107, miR-151, miR-182, miR-361;  $P = 0.0001$ ), and distant metastasis (miR-151, miR-152, miR-324-5p, miR-361, miR-492;  $P = 0.0087$ ), which retained significance even after adjusting for p16 status. The associated biologic functions of these miRNAs include immune surveillance, treatment resistance, invasion, and metastasis.

**Conclusion:** We have identified several miRNAs, which associate with HPV status in OPC; furthermore, three candidate prognostic sets of miRNAs seem to correlate with clinical outcome, independent of p16 status. Furthermore, evaluations will offer biologic insights into the mechanisms underlying the differences between HPV-positive versus HPV-negative OPC. *Clin Cancer Res*; 19(8); 2154–62. ©2013 AACR.

### Introduction

Over the past 2 decades, the incidence of human papilloma virus (HPV)-associated head and neck squamous cell carcinomas (HNSCC) involving the oropharynx has been increasing, nowadays comprising the majority of oropharyngeal carcinoma (OPC) cases seen in North America

(1–3). There are several unique clinical characteristics of HPV-positive OPC, such as younger age at diagnosis, lower likelihood of heavy smoking or alcohol, and greater degree of sexual activity, compared with patients with HPV-negative OPC (1, 2). Intriguingly, HPV-positive patients experience a significantly superior clinical outcome when treated with either radiotherapy alone or combined chemoradiotherapy, despite presenting with higher grade and stage of disease (3). We have previously reported that approximately 60% of patients with OPC seen at the Princess Margaret Cancer Center were HPV-positive, with 3-year overall survival (OS) rates of 88% versus 67% in favor of HPV-positive versus negative patients (4). However, the biologic mechanisms behind this unique clinical entity of HPV-associated OPC remain to be elucidated.

Deregulation of miRNAs is clearly associated with the development and progression of human malignancies. We have conducted global miRNA profiling of HNSCC (5), through which the miR-375~metadherin axis was newly identified as a potentially important pathway that could partially explain the propensity for lung metastases in this

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### Translational Relevance

The incidence of human papilloma virus (HPV)-associated oropharyngeal carcinoma (OPC) has been increasing. HPV-positive OPCs have several unique clinical characteristics and with a significantly more favorable outcome, compared with HPV-negative OPCs. The biologic basis behind this differential outcome is currently unelucidated. Deregulation of miRNAs is associated with oncogenesis of various malignancies, suggesting that miRNA expression profiling have the potential to unravel the complex biology of human tumors. In this study, we evaluated the miRNA profiles of archival formalin-fixed and paraffin-embedded diagnostic biopsy specimens from 88 nonmetastatic OPC samples (p16+ve:58; p16-ve:34). We have identified a panel of p16/HPV-associated miRNAs and three potential miRNA signature sets that are associated with clinical outcome, independent of p16 status. Furthermore, examination of these candidate miRNAs will inform biologic insights into the mechanisms underlying the differences between HPV-positive and HPV-negative OPC.

disease (6). Additional miRNA studies in HNSCC have reported potentially predictive sets of miR-205 and let-7d or miR-210 as a hypoxia marker (7–10). MiRNAs associated with OPC have been reported previously; however, overlap in these candidate prognostic miRNAs has remained limited as well as lack of concordance in HPV-associated miRNAs (11–14). Hence, to better understand the role of miRNAs in HPV-associated OPC, we conducted a comprehensive miRNA profiling focused solely on OPC, using the same samples that have been previously analyzed for HPV status (4).

### Materials and Methods

#### Patient information

A subset of diagnostic formalin-fixed paraffin-embedded (FFPE) blocks from our previously published 111-sample OPC report was evaluated (4). That study showed that HPV status as defined by p16 immunohistochemistry (IHC), HPV16 *in situ* hybridization (ISH), or HPV16 E6 transcript levels using quantitative real-time PCR (qRT-PCR) were strongly concordant (4). Given that these were small diagnostic biopsies, only 88 samples had sufficient remaining tumor tissues to carry out global miRNA profiling.

#### RNA purification from FFPE samples

A representative section from each sample was stained with hematoxylin and eosin stain and reviewed by head and neck cancer pathologists (B. Perez-Ordonez or I. Weinreb) to identify regions containing more than 70% malignant epithelial cells for macroadhesion. Seven normal tonsillar epithelial FFPE tissues from individuals who underwent a tonsillectomy were included as controls. University Health

Network (UHN, Toronto, Canada) Institutional Research Ethics Board approval has been obtained for this study.

All blocks were processed randomly, with clinical outcome unknown, to avoid experimental bias. Total RNA enriched for small RNA species was isolated using the RecoverAll Total Nucleic Acid Isolation Kit for FFPE samples (Ambion), according to the manufacturer's instructions (15).

#### MiRNA profiling using TaqMan low density array

Quality of RNA samples was assessed by qRT-PCR analysis of the endogenous control RNU44 using TaqMan microRNA Assay (Applied Biosystems), as previously described (6, 15). Global miRNA expression on 88 OPC and 7 normal samples was conducted using the TaqMan Low Density Array (TLDA) Human MicroRNA Panel v1.0 (Applied Biosystems), which enabled the simultaneous quantification of 365 human miRNAs plus 3 endogenous controls (RNU6B, RNU44, and RNU48). In brief, total RNA of each sample was first reverse-transcribed with the Multiplex RT pool set, then quantitated with a TLDA array using an Applied Biosystems 7900 HT Real-Time PCR system, with the  $C_t$  values determined by threshold method, according to the manufacturer's protocol (15).

#### Data processing

The TLDA data were processed and analyzed as previously described (15) with some minor modifications. Any miRNAs with undetermined values in more than 80% of the tumors and more than 6 "normals" were eliminated from the analysis. Undetermined  $C_t$  values, or  $C_t$  values more than 36, were imputed to 40. All samples were normalized by the mean of the endogenous controls and converted into a ratio of abundance compared with the geometric mean of the abundance of the 7 normal samples, as we have previously described (15).

Significant differences in miRNA expression between tumor and normal samples were assessed using the Wilcoxon rank sum test, with multiple comparisons adjusted by Benjamini–Hochberg false discovery rate (FDR) correction, conducted in the R statistical environment (v2.6.1; ref. 16). Two-way ANOVA and the Benjamini–Hochberg correction for FDR of multiple testing were conducted on each miRNA to investigate the association with p16 and HPV status. The Spearman correlation coefficient was used to calculate  $P$  values of the consistency of the miRNA signals by using p16 and HPV as outcomes, for each miRNA.

To explore potential prognostic effects, we applied univariate survival analysis to detect association of single miRNA expression level with OS, disease-free survival (DFS), or distant relapse-free survival (DM) as previously described (4). After identifying potentially important miRNAs ( $P < 0.05$ ), we applied multivariate models on these miRNAs. Model selection procedure was conducted using a stepwise selection algorithm, conducted separately for each outcome. Finally, after model selection, additional analyses were conducted to generate a risk score based on the significant miRNAs identified from the multivariate

analysis. The risk score was based on the weighted combination of the miRNAs with the estimated Cox proportional hazard regression model coefficient as the weight (17). For each outcome, patients were dichotomized into 2 categories of low (risk score < median), or high risk (risk score > median); following which, outcomes were compared for each risk group. Test of difference between the risk categories was assessed using the log-rank test or Cox proportional hazard regression model, with *P* values, and HRs (including 95% confidence interval (CI).

To validate the prognostic effect, we carried out internal validation procedures using a bootstrap algorithm (18) and constructed the bootstrap CI. The bootstrap is one of the resampling techniques, and bootstrap datasets were created by sampling with replacement. The bootstrap method has been shown to provide a valid estimation of prediction error and can correct for the bias of the parameter estimate (18). We applied bootstrap based on 500 replications; these results were obtained using the PROC SURVEYSELECT procedure in SAS version 9.3, and presented as the bootstrap HR CIs of the prognostic effect adjusted for p16 status.

## Results

### Differentially expressed miRNAs in OPC

Genome-wide miRNA profiles of 88 OPC patient samples were conducted; the clinical characteristics are provided in Table 1. The median follow-up time for all patients has now extended to 5.9 years, with 5-year OS, and DFS rates at 55% and 52%, respectively (Supplementary Fig. S1). The 5-year OS for HPV-positive versus negative patients were 65% versus 30%; the DFS rates were 70% versus 20% respectively.

Among the 365 interrogated miRNAs, 224 were expressed in more than 80% of the samples, with 128 being significantly differentially expressed between the 88 OPC and 7 normal tonsillar epithelial tissues (*P* < 0.05). Ninety-two of these 128 miRNAs were more than 2-fold differentially

expressed (Supplementary Fig. S2), with the majority (120/128; 94%) being upregulated in OPC (Supplementary Table S1). A detailed comparison of the top 6 most significantly differentially expressed miRNAs (miR-21, let-7g, miR-25, let-7f, miR-130b, and miR-151) are provided in Fig. 1. These dysregulated miRNAs seemed to be randomly distributed among different chromosomal regions, with the exception of 1p34.2 and 7q22.1, within which 6 of the top 40 aberrantly expressed miRNAs are located (bold information in Supplementary Table S1).

### MiRNAs associated with HPV/p16 status

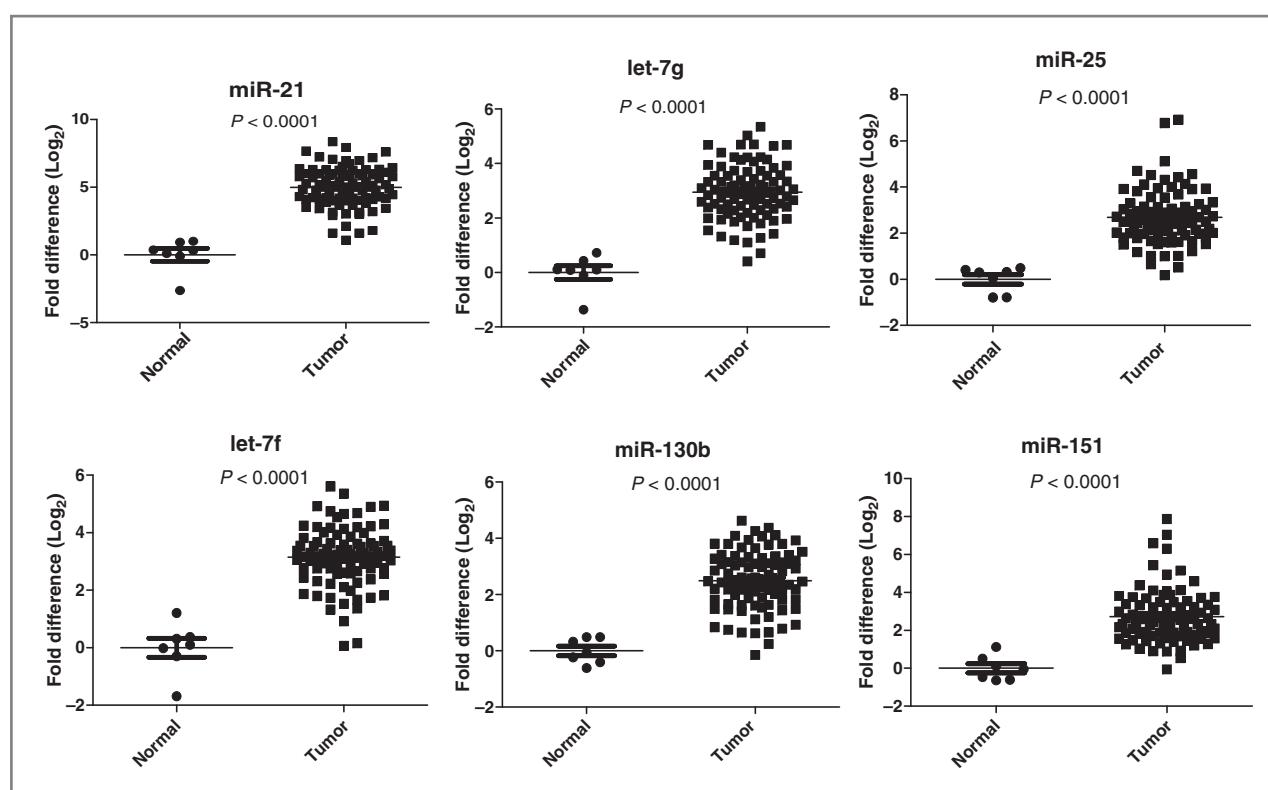
The miRNAs associated with either HPV16 ISH or p16 IHC were next investigated. Comparison of the 2 sets of miRNAs showed that the fold change in miRNA expression defined by either HPV ISH, or p16 IHC, were highly correlated ( $R^2 = 0.78$ ; Supplementary Fig. S3). Figure 2 compared the miRNAs that were significantly associated with p16 IHC or HPV16 ISH as defined by the  $-\log_{10}$  (*P* values), again showing a strong correlation ( $R^2 = 0.8$ ). Using a cutoff of *P* < 0.01, 9 miRNAs were significantly associated with both p16 IHC and HPV16 ISH-positive OPCs, which included upregulated miR-20b, miR-9, miR-9\*, miR-492, miR-545, miR-591, miR-422a, and downregulated miR-193b and miR-107. Because p16 IHC is a broadly accepted surrogate marker for HPV status, shown in multiple studies to correlate with patient outcome (1, 4), subsequent analyses were based on the p16 IHC data for these patients.

Validation of the HPV/p16-associated miRNAs (Supplementary Table S2) was conducted using the samples from an independent HNSCC cohort that we had previously profiled, which contained 11 p16 positive and 8 p16 negative OPC samples (5). Seven of the top 10 most significant HPV/p16-associated miRNAs were also included in the previous list of investigated miRNAs of HNSCC profiles (5). As shown in Fig. 3, miR-9 and miR-9\* retained statistical significance (*P* = 0.04); the remaining 5 miRNAs were also

**Table 1.** Clinical description of the 88 patients with OPC

	Patient characteristics		
	HPV positive ( <i>n</i> = 56; 64%)	HPV negative ( <i>n</i> = 32; 36%)	<i>P</i>
Gender			
Male	41 (73%)	24 (75%)	0.95
Female	15 (27%)	8 (25%)	
Age (median; range)	Mean: 55 ± 11 y (range: 27–80 y)	Mean: 67 ± 11 y (range: 46–93 y)	<0.0001
Stage			
I	0	1 (3%)	
II	2 (5%)	7 (22%)	0.003
III	5 (12%)	7 (22%)	
IV	49 (83%)	17 (53%)	
Treatment			
RT	26 (46%)	26 (81%)	0.003
CRT	30 (54%)	6 (19%)	

Abbreviation: C-RT, chemoradiotherapy; RT, radiotherapy.



**Figure 1.** Expression levels of the top 6 miRNAs associated with OPC (miR-21, let-7g, miR-25, let-7f, miR-130b, and miR-151), comparing 88 tumors with that of the 7 normal tonsillar epithelial tissues. The data are presented as fold change in log<sub>2</sub> space, normalized to that of RNU44, 48, and 6B.

consistently aberrantly expressed, but did not reach statistical significance, likely due to the small sample size of the validation cohort.

#### Potential prognostic miRNA sets

Using the miRNA profiles of these 88 samples, with their median value as the cutoff, and treated as binary predictors, 3 potential miRNA signature sets were identified that were associated with OS, DFS, and DM (Fig. 4). The candidate prognostic panel for OS was defined by upregulated miR-107 and miR-151, and downregulated miR-492 (Fig. 4B;  $P < 0.0001$ ). For the DFS set, this composed of upregulated miR-107, miR-151, miR-182, and miR-361 with downregulated miR-20b (Fig. 4C;  $P \leq 0.0001$ ). The DM set was defined by upregulated miR-151, miR-361 and miR-324-5p, as well as downregulated miR-492 and miR-152 (Fig. 4D;  $P = 0.0088$ ). Even after adjusting for p16 IHC status, these  $P$  values still retained significance (adjusted  $P$  values were 0.0002, 0.0001, and 0.0087 for OS, DFS, and DM, respectively).

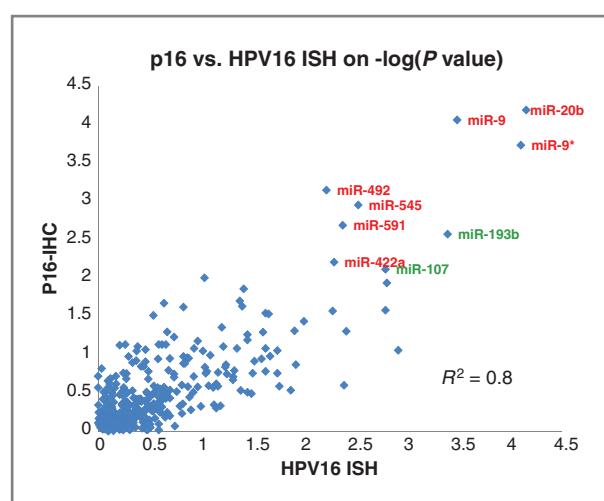
#### In silico analysis

We proceeded to investigate the putative biologic functions of these aforementioned miRNAs using *in silico* analysis (Fig. 5). For miRNAs differentially expressed in OPC (Supplementary Table S2), miR-30c, miR-30e-3p, and miR-30e-5p all resided within the fifth intron of *NFYC* on chromosome 1p34.2 (19). *NFYC* is the nuclear transcrip-

tion factor Y-γ, which binds with CCAAT motifs in the promoter region of several genes, associated with MHC class II determinants in immune response (Fig. 5A; ref. 19). As described in our previous publication (5), overexpression of miR-25, miR-93, and miR-106b and its host gene *MCM7* can be regulated by *E2F1*, interfering with *TGF-β* signaling in HNSCC. Furthermore, the miR-106b-25 cluster has been reported to target *PTEN* (20).

Six of the HPV/p16-associated miRNAs (miR-381, miR-412, miR-380-5p, miR-487b, and miR-382) are all located at a common fragile site (FRA) at 14q32.3 (Fig. 5B; refs. 21, 22). Aberrations of 14q32.31 miRNA cluster have been previously described in human malignancies with an emerging role in immune regulation (23, 24). MiR-9 was found to be induced by lipopolysaccharide-inflammatory stimuli mediated by the proinflammatory cytokines IL-1β and TNF-α (25). Moreover, expression of miR-34a can be induced by p53 under genotoxic stress (26), and miR-20b has been described to target the 3' untranslated region of *hypoxia-inducible factor-1α* and *VEGF* (27).

Overexpression of miR-151 with its host gene protein tyrosine kinase 2 (PTK2/FAK) has been reported to induce tumor invasion and metastasis in hepatocellular carcinoma (HCC; Fig. 5C; ref. 28). It also acts synergistically with FAK to enhance HCC cell motility and spreading (28, 29). Several miR-152 targets have also been identified, including the DNA methyltransferase *DNMT1*, *E2F3*, *MET*, and *Rictor* (30). Moreover, miR-182 was reported to be upregulated in



**Figure 2.** Comparison of the association between miRNAs with p16 IHC, and association between miRNAs with HPV16 ISH, as defined by  $P$  values. The x and y axes refer to  $-\log_{10}$  ( $P$  values) of HPV16 ISH-associated miRNAs, and  $-\log_{10}$  ( $P$  values) of p16 IHC-associated miRNAs, respectively. These 2 sets of  $P$ -values are highly correlated with  $R^2 = 0.8$ . MiRNAs with significant associations ( $P < 0.01$ ) for both HPV16 ISH and p16 IHC are specifically identified; green denotes downregulation; red denotes upregulation.

multidrug-resistant cell lines (31), associated with both metastasis (32) and poor clinical outcome (33). Finally, miR-107 has been associated with mammalian development and cellular metabolism (34).

## Discussion

We have conducted a global miRNA profiling study focused strictly on OPC, with approximately two-third being HPV-positive and one-third being HPV-negative. A panel of miRNAs significantly differentially expressed between tumor versus normal tissues ( $P < 0.05$ ) was identified, with miR-21 being the most significantly upregulated miRNA. MiRNA-21 is one of the most consistently reported aberrant miRNAs in HNSCC, known to target multiple tumour suppressors such as *PTEN* and *TPM1* and *Bcl-2* (5, 7, 10, 12). Upregulated miR-20b, miR-9, and miR-9\* were also significantly associated with HPV/p16-status. Furthermore, there were 3 candidate miRNA sets that were associated significantly with OS, DFS, and DM, even after adjusting for HPV status.

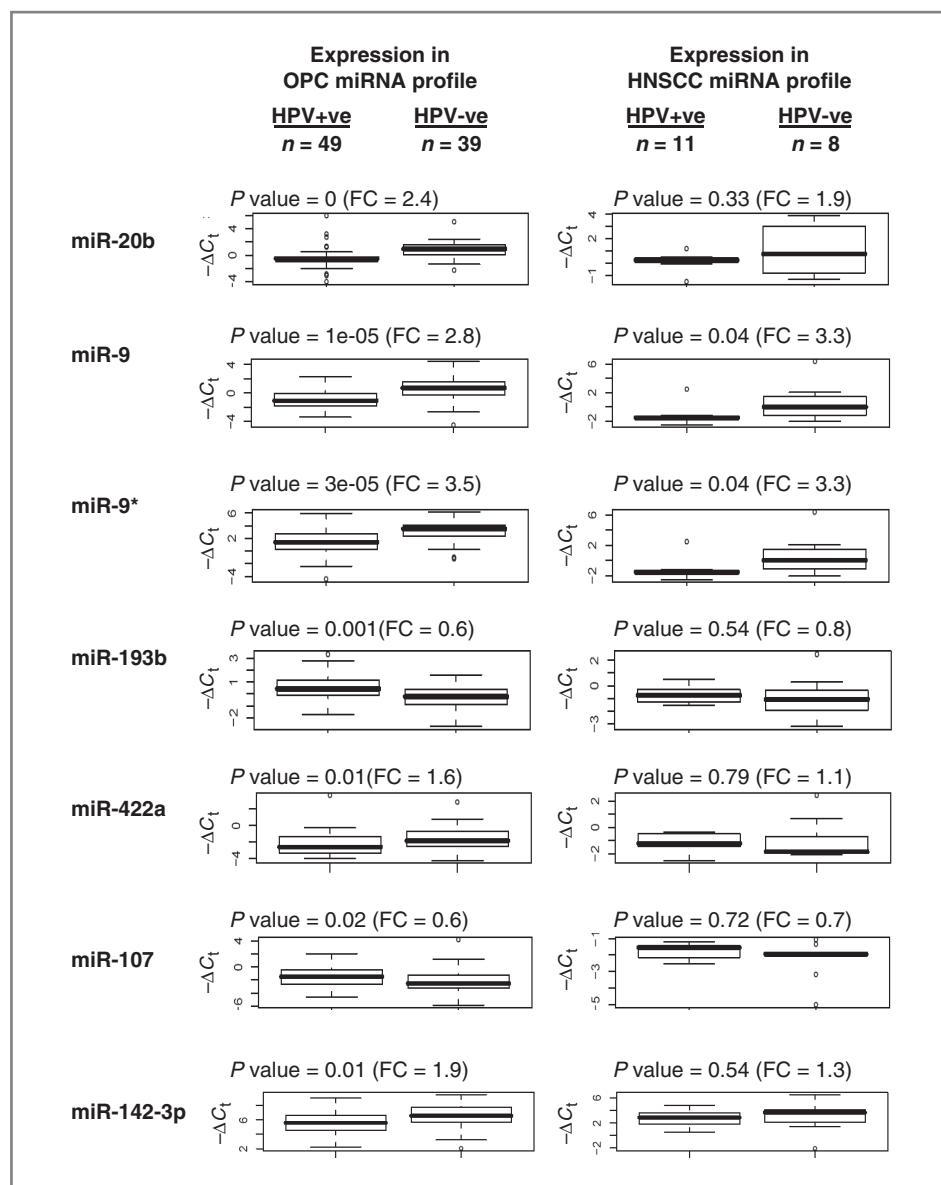
Similar to our previous HNSCC miRNA study (5), upregulation of the miR-106b-25 cluster was also observed in these OPC samples. These included the nonrandom overexpression of miR-25, miR-93, and miR-106b, located on chromosome 7q22.1, within the intronic region of *MCM7*. Overexpression of *MCM7*, as well as regulation of the miR-106b-25 cluster and *MCM7* by E2F1, with subsequent interference of TGF- $\beta$  signaling in HNSCC has been previously noted (5). Interestingly, TGF- $\beta$  polymorphisms have been reported to be a susceptibility marker for HPV16 status amongst patients with OPC (35). Furthermore, the miR-106b-25 cluster can also target *PTEN* (20), which in turn will activate AKT, which correlates with adverse outcome for

patients with OPC (36). Hence, dysregulation of *MCM7* and the miR-106b-25 cluster could lead to OPC development via aberrant TGF- $\beta$  and *PTEN* signaling (Fig. 5A).

Many of the p16-associated miRNAs were located at common FRAs, such as 1p, 1q, 5q, 8p, 14q32.31, 16p13.12, 17p, and Xq (Supplementary Table S2; ref. 21). This is consistent with previous studies reporting that the majority (53%) of deregulated miRNAs were located in cancer-associated genomic regions or FRAs (21). Chromosome 14q32.31 is the only nonrandom region that harbored 5 (miR-381, miR-412, miR-380-5p, miR-487b, and miR-382) of the p16-associated miRNAs, which is also one of the most common HPV integration sites in cervical cancer (22). In fact, a cluster of 46 miRNAs (3.2% of 1,426 human miRNAs; <http://genome.ucsc.edu/cgi-bin/hgTracks>) have been mapped to this small region of approximately 45 kb. Aberrations of the 14q32.31 miRNA cluster has been documented in many human cancers and its role in immune surveillance is an emerging area of evaluation (Fig. 5B; ref. 23, 24). Hence, in OPC, frequent aberrant expression of these p16-associated miRNAs at 14q32.31 might relate to this being a preferential HPV integration site. This postulate would be supported by the recent emerging exome sequencing data from TCGA wherein 14q32.3 amplifications have been noted for HPV-positive HNSCC (<https://wiki.nci.nih.gov/display/TCGAM/03-08-12+HNSCC+AWG>).

Enhanced immune response induced by HPV has been suggested as one possible mechanism for the superior outcome of HPV-associated OPC (37). Among the p16-associated miRNAs, several have been previously described to be associated with immune regulation, such as miR-9, miR-9\*, miR-146a, miR-34a, and miR-155 (Supplementary Table S2 and Fig. 5B; ref. 38). Upregulation of miR-9 and its passenger strand miR-9\* were among the most significantly associated miRNAs with both OPC, and specifically p16-positive OPC. Overexpression of miR-9/9\* has been described in other human cancers (39), with miR-9 potentially involved in immune response during inflammatory stimuli (25). In addition, miR-9 might also have an oncogenic role in mediating angiogenesis and metastasis by inducing Myc to target E-cadherin, priming cancer cells for epithelial–mesenchymal transition (39). MiRNA-20b was the most significant p16-associated miRNA in OPC, and its downregulation was associated with poor DFS (Fig. 4 and 4C). The potential tumor suppressor role of miR-20b has been investigated wherein low circulating levels of miR-20b was reported in lung cancer and mantle cell lymphoma, also associated with poor outcome (40). Inhibition of miR-20b can lead to an increase HIF-1 $\alpha$  and VEGF under normoxia; conversely, an increase of miR-20b in hypoxic tumor cells decreased HIF-1 $\alpha$  and VEGF (27). This negative regulatory loop between HIF-1 $\alpha$  and miR-20b has been suggested as one of multiple mechanisms by which tumor cells can rapidly adapt to varying oxygen concentrations (27).

MiR-34a was found to have higher expression levels in HPV-positive OPC, which is distinctly different from cervix cancer whereby miR-34a is downregulated by E6 via destabilization of p53 (41). In OPC, the relationship between



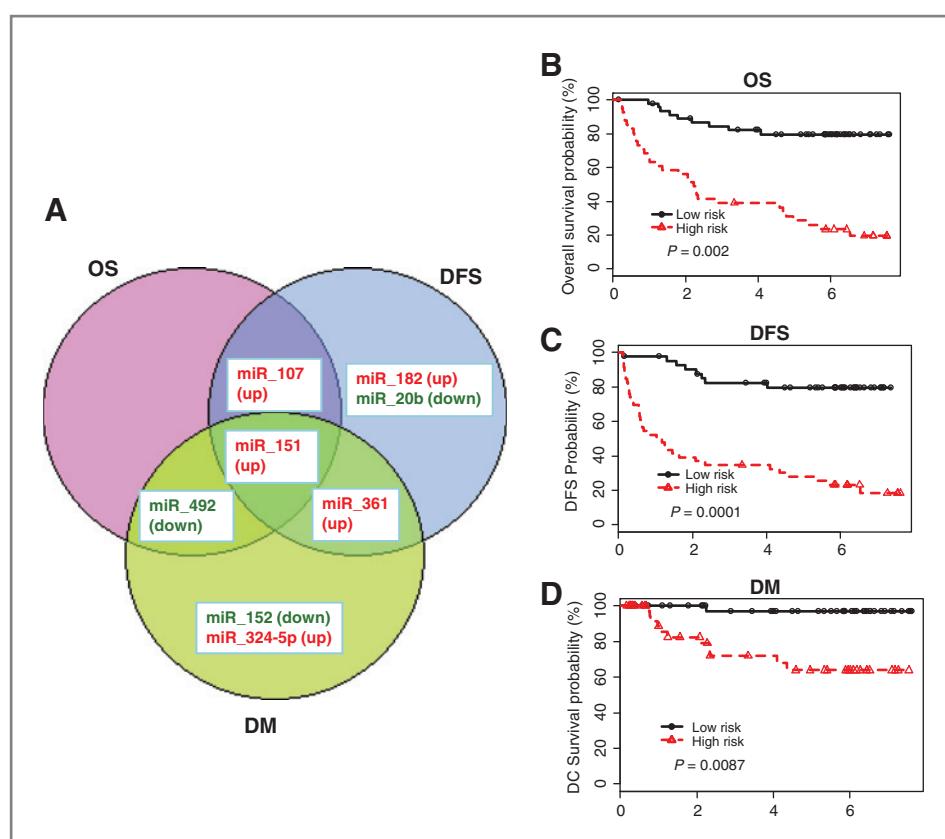
**Figure 3.** Validation of the most significant p16-associated miRNAs was conducted by using the OPC samples from an independent HNSCC cohort ( $N = 19$ ). The top 7 miRNAs were investigated (miR-20b, -9, -9\*, -193b, -422a, -107, and 142-3p); both fold change and  $P$  values are presented. " $-\Delta C_t$ " refers to the differences between the measured miRNAs compared with the average  $C_t$  values of the endogenous controls (RNU44, 48, and 6B).

p53 and HPV remains unclear (4, 42). Our previous study observed p53 to be overexpressed on IHC in 63% of cases and only borderline association with HPV positivity (4). Expression of miR-34a was shown to be induced by p53 under genotoxic stress conditions (Fig. 5B; ref. 26). Furthermore, there is a positive feedback loop wherein p53 induces miR-34a; in turn, miR-34a activates p53 by inhibiting SIRT1, leading to an increase in miR-34a expression in cells with wild-type *p53*, resulting in enhanced apoptosis (43). This might, in part, explain the high incidence (70%) of OPCs harboring p53 immunoexpression, particularly in association with p16 (4).

Finally, in relation to the potential miRNA signature associated with clinical outcome, upregulation of miR-151 was the only miRNA associated with all of OS, DFS, and DM (Fig. 4A), as well as being among the top 10

significantly deregulated miRNAs in OPC (Supplementary Table S1). Its chromosomal location on 8q24.3 is frequently amplified in human cancers, including HNSCCs (44). Overexpression of miR-151 has been reported to promote invasion and metastasis in HCC, by targeting the putative metastasis suppressor *RhoGDIα*, which in turn synergized with FAK to enhance motility (28, 29). Downregulation of miR-152 is another well-characterized tumor suppressor miRNA commonly inactivated by promoter hypermethylation (30, 45), targeting several known important mediators of tumor progression (Fig. 5C; ref. 30). It resides in the intron of the host gene Coatomer protein complex  $\zeta 2$  (COPZ2), which is also a putative tumor suppressor (46).

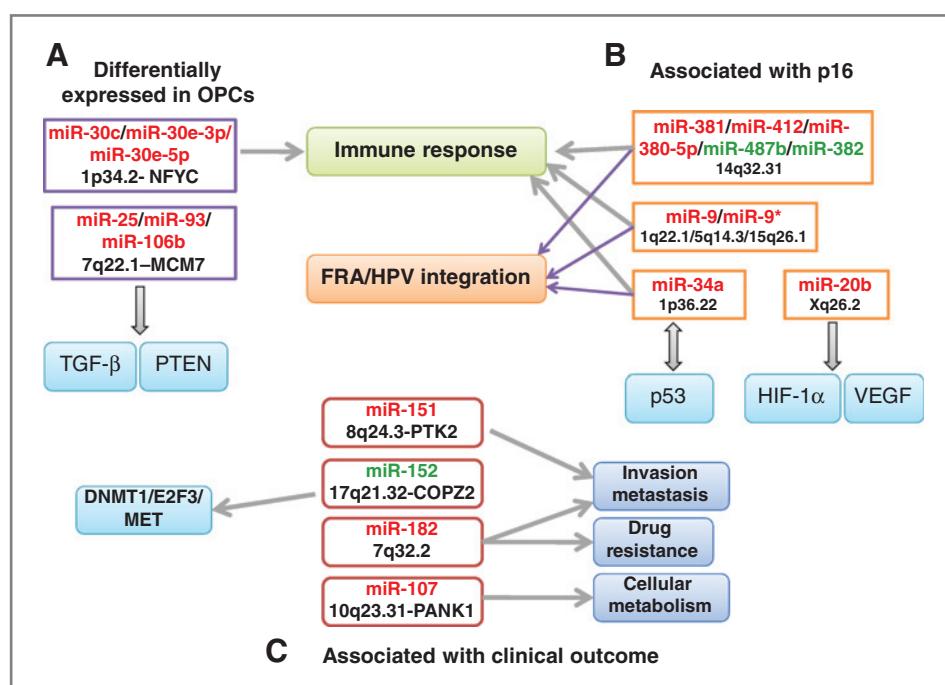
Oncogenic miR-182 (Fig. 4 & 5) belongs to the miR-183-96-182 cluster, which was reported to regulate zinc homeostasis in prostate cancer, and consistently upregulated in



**Figure 4.** Potential miRNA signature sets that are associated with OS, DFS, and DM. A, Venn diagram of the candidate miRNAs as a function of clinical outcome (OS, DFS, and DM). B, Kaplan-Meier survival curves for OS as a function of the median expression level of the 3 miRNAs (miR-107, miR-151, and miR-492). C, DFS actuarial plot as a function of the median expression level of the 5 miRNAs (miR-107, miR-151, miR-182, miR-20b, and miR-361). D, DM actuarial plot as a function of the median expression levels of the 5 miRNAs (miR-151, miR-492, miR-361, miR-152, and miR-324-5p). The P values indicated in all 3 graphs have already been adjusted for p16 status. Green denotes underexpression; red denotes overexpression of each candidate miRNA.

multidrug-resistant cell lines (31). It has been implicated in metastasis and poor survival in cancers (32, 33). Overexpression of miR-107 was observed to be associated with worse outcome (Fig. 4) and inversely related to HPV-pos-

itivity in OPC (Supplementary Table S2). It is a member of the miR-15/107 miRNA gene group associated with mammalian development (34), as well as regulation of the miRNA processing machinery and cellular metabolism



**Figure 5.** *In silico* analysis of selected key OPC-associated miRNAs. A, selected miRNAs significantly differentially expressed between OPC versus normal tissues: miR-30c, miR-30e-5p, miR-25, miR-93, and miR-106b, along with their host genes, and putative mRNA targets. B, selected miRNAs significantly associated with p16 status: miR-381, miR-412, miR-380-5p, miR-487b, miR-382, miR-9/9\*, miR-34a, and miR-20b. C, selected miRNAs significantly associated with clinical outcome: miR-151, miR-152, miR-182, and miR-107. Green denotes underexpression; red denotes overexpression of candidate miRNAs.

(34, 47). It resides within the introns of the host gene *PANK1*, involved in catalyzing the formation of CoA during the Krebs cycle (48). Downregulation of miR-107 has been noted in HNSCC (49), but in contrast, for this study, we report a contradictory observation in that miR-107 overexpression was associated with poor survival. These data indicate that dysregulation of miR-107 in OPC might well be quite complex; although its oncogenic function has been previously reported (34, 47, 50).

A 6 miRNA signature set that is associated with survival in OPC was recently reported (11). None of these 6 miRNAs overlapped with our 3 potential miRNA signature sets. Among the 5 miRNAs that were suggested to be associated with HPV by Gao and colleagues, upregulated miR-9 and miR-155 were also among the significantly HPV associated miRNAs in our current study (Supplementary Table S2). However, no overlap was observed in HPV-associated miRNAs between those 5 miRNAs described by Gao and colleagues (11) with the 21 HPV-associated miRNAs reported by Lajer and colleagues (13). Downregulation of miR-145 was found to be associated with HPV in both our current study and that conducted by Lajer and colleagues (13). MiR-381 and miR-101 were also identified in both studies, but with the fold changes in opposite directions (13). One possible reason for the limited overlap amongst these various studies (Lajer and colleagues vs. current vs. Gao and colleagues) could be due to the different number of miRNAs being included at the outset (847 vs. 365 vs. 96). Moreover, each study included different proportions of HPV-positive patients: 42% versus 64% versus 82% for Lajer and colleagues versus current versus Gao and colleagues, respectively. The methods used to define HPV-positivity were also distinct (combination of 2 positives of p16 IHC, RT-PCR, or HPV ISH vs. p16 IHC vs. RT-PCR). Controversy still remains regarding the best method of HPV detection, with discrepancies ranged from 60% to 92% among these aforementioned methods (4, 13). These variations hopefully will be resolved by the development of miRNA sequencing of FFPE samples, which will enable the interrogation of all known human miRNAs, and the integration of efforts currently being undertaken by the TCGA head and neck cancer group.

## References

- Ang KK, Harris J, Wheeler R, Weber R, Rosenthal DI, Nguyen-Tan PF, et al. Human papillomavirus and survival of patients with oropharyngeal cancer. *N Engl J Med* 2010;363:24–35.
- Marur S, D’Souza G, Westra WH, Forastiere AA. HPV-associated head and neck cancer: a virus-related cancer epidemic. *Lancet Oncol* 2010;11:781–9.
- Sturgis EM, Ang KK. The epidemic of HPV-associated oropharyngeal cancer is here: is it time to change our treatment paradigms? *J Natl Compr Canc Netw* 2011;9:665–73.
- Shi W, Kato H, Perez-Ordóñez B, Pintilie M, Huang S, Hui A, et al. Comparative prognostic value of HPV16 E6 mRNA compared with *in situ* hybridization for human oropharyngeal squamous carcinoma. *J Clin Oncol* 2009;27:6213–21.
- Hui AB, Lenarduzzi M, Krushel T, Waldron L, Pintilie M, Shi W, et al. Comprehensive microRNA profiling for head and neck squamous cell carcinomas. *Clin Cancer Res* 2010;16:1129–39.
- Hui AB, Bruce JP, Alajezi NM, Shi W, Yue S, Perez-Ordóñez B, et al. Significance of dysregulated metadherin and microRNA-375 in head and neck cancer. *Clin Cancer Res* 2011;17:7539–50.
- Chang SS, Jiang WW, Smith I, Poeta LM, Begum S, Glazer C, et al. MicroRNA alterations in head and neck squamous cell carcinoma. *Int J Cancer* 2008;123:2791–7.
- Childs G, Fazzari M, Kung G, Kawachi N, Brandwein-Gensler M, McLemore M, et al. Low-level expression of microRNAs let-7d and miR-205 are prognostic markers of head and neck squamous cell carcinoma. *Am J Pathol* 2009;174:736–45.

## Conclusion

We have conducted a comprehensive miRNA profiling study focused strictly on OPC, which identified 3 potential miRNA signature sets associated with clinical outcome, independent of HPV status. The emerging picture of these dysregulated miRNAs points to a complexity of pathways involved in immune response, and tumor progression, located within FRA and HPV integration sites. Furthermore, examination of these candidate miRNAs will inform biologic insights into the mechanisms underlying the differences between HPV-positive and HPV-negative OPC.

## Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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9. Gee HE, Camps C, Buffa FM, Patiar S, Winter SC, Betts G, et al. hsa-mir-210 is a marker of tumor hypoxia and a prognostic factor in head and neck cancer. *Cancer* 2010;116:2148–58.
10. Ramdas L, Giri U, Ashorn CL, Coombes KR, El-Naggar A, Ang KK, et al. miRNA expression profiles in head and neck squamous cell carcinoma and adjacent normal tissue. *Head Neck* 2009;31:642–54.
11. Gao G, Gay HA, Chernock RD, Zhang TR, Luo J, Thorstad WL, et al. A microRNA expression signature for the prognosis of oropharyngeal squamous cell carcinoma. *Cancer* 2013;119:72–80.
12. Lajer CB, Garnaes E, Friis-Hansen L, Norrild B, Therkildsen MH, Glud M, et al. The role of miRNAs in human papilloma virus (HPV)-associated cancers: bridging between HPV-related head and neck cancer and cervical cancer. *Br J Cancer* 2012;106:1526–34.
13. Lajer CB, Nielsen FC, Friis-Hansen L, Norrild B, Borup R, Garnaes E, et al. Different miRNA signatures of oral and pharyngeal squamous cell carcinomas: a prospective translational study. *Br J Cancer* 2011;104:830–40.
14. Wald AI, Hoskins EE, Wells SI, Ferris RL, Khan SA. Alteration of microRNA profiles in squamous cell carcinoma of the head and neck cell lines by human papillomavirus. *Head Neck* 2011;33:504–12.
15. Hui AB, Shi W, Boutros PC, Miller N, Pintilie M, Fyles T, et al. Robust global micro-RNA profiling with formalin-fixed paraffin-embedded breast cancer tissues. *Lab Invest* 2009;89:597–606.
16. Team RDC, editor. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2008.
17. Reeves GK, Travis RC, Green J, Bull D, Tipper S, Baker K, et al. Incidence of breast cancer and its subtypes in relation to individual and multiple low-penetrance genetic susceptibility loci. *JAMA* 2010;304:426–34.
18. Sauerbrei W, Schumacher M. A bootstrap resampling procedure for model building: application to the Cox regression model. *Stat Med* 1992;11:2093–109.
19. Jabrane-Ferrat N, Nekrep N, Tosi G, Esserman L, Peterlin BM. MHC class II enhanceosome: how is the class II transactivator recruited to DNA-bound activators? *Int Immunol* 2003;15:467–75.
20. Poliseno L, Salmena L, Ricciardi L, Fornari A, Song MS, Hobbs RM, et al. Identification of the miR-106b ~ 25 microRNA cluster as a proto-oncogenic PTEN-targeting intron that cooperates with its host gene MCM7 in transformation. *Sci Signal* 2010;3:ra29.
21. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci U S A* 2004;101:2999–3004.
22. Wentzensen N, Vinokurova S, von Knebel Doeberitz M. Systematic review of genomic integration sites of human papillomavirus genomes in epithelial dysplasia and invasive cancer of the female lower genital tract. *Cancer Res* 2004;64:3878–84.
23. Gattoliat CH, Thomas L, Ciafre SA, Meurice G, Le Teuff G, Job B, et al. Expression of miR-487b and miR-410 encoded by 14q32.31 locus is a prognostic marker in neuroblastoma. *Br J Cancer* 2011;105:1352–61.
24. Vinuesa CG, Rigby RJ, Yu D. Logic and extent of miRNA-mediated control of autoimmune gene expression. *Int Rev Immunol* 2009;28:112–38.
25. Bazzoni F, Rossato M, Fabbri M, Gaudiosi D, Mirolo M, Mori L, et al. Induction and regulatory function of miR-9 in human monocytes and neutrophils exposed to proinflammatory signals. *Proc Natl Acad Sci U S A* 2009;106:5282–7.
26. Mathe E, Nguyen GH, Funamizu N, He P, Moake M, Croce CM, et al. Inflammation regulates microRNA expression in cooperation with p53 and nitric oxide. *Int J Cancer* 2011;131:760–5.
27. Lei Z, Li B, Yang Z, Fang H, Zhang GM, Feng ZH, et al. Regulation of HIF-1alpha and VEGF by miR-20b tunes tumor cells to adapt to the alteration of oxygen concentration. *PLoS ONE* 2009;4:e7629.
28. Luedde T. MicroRNA-151 and its hosting gene FAK (focal adhesion kinase) regulate tumor cell migration and spreading of hepatocellular carcinoma. *Hepatology* 2010;52:1164–6.
29. Ding J, Huang S, Wu S, Zhao Y, Liang L, Yan M, et al. Gain of miR-151 on chromosome 8q24.3 facilitates tumour cell migration and spreading through downregulating RhoGDIα. *Nat Cell Biol* 2010;12:390–9.
30. Tsuruta T, Kozaki K, Uesugi A, Furuta M, Hirasawa A, Imoto I, et al. miR-152 is a tumor suppressor microRNA that is silenced by DNA hypermethylation in endometrial cancer. *Cancer Res* 2011;71:6450–62.
31. Mihelich BL, Khramtsova EA, Arva N, Vaishnav A, Johnson DN, Giangreco AA, et al. miR-183-96-182 cluster is overexpressed in prostate tissue and regulates zinc homeostasis in prostate cells. *J Biol Chem* 2011;286:44503–11.
32. Segura MF, Hanniford D, Menendez S, Reavie L, Zou X, Alvarez-Diaz S, et al. Aberrant miR-182 expression promotes melanoma metastasis by repressing FOXO3 and microphthalmia-associated transcription factor. *Proc Natl Acad Sci U S A* 2009;106:1814–9.
33. Jiang L, Mao P, Song L, Wu J, Huang J, Lin C, et al. miR-182 as a prognostic marker for glioma progression and patient survival. *Am J Pathol* 2010;177:29–38.
34. Finnerty JR, Wang WX, Hebert SS, Wilfred BR, Mao G, Nelson PT. The miR-15/107 group of microRNA genes: evolutionary biology, cellular functions, and roles in human diseases. *J Mol Biol* 2010;402:491–509.
35. Guan X, Sturgis EM, Lei D, Liu Z, Dahlstrom KR, Wei Q, et al. Association of TGF-beta1 genetic variants with HPV16-positive oropharyngeal cancer. *Clin Cancer Res* 2010;16:1416–22.
36. Yu Z, Weinberger PM, Sasaki C, Eggleston BL, Speier WF, Haffty B, et al. Phosphorylation of Akt (Ser473) predicts poor clinical outcome in oropharyngeal squamous cell cancer. *Cancer Epidemiol Biomarkers Prev* 2007;16:553–8.
37. Psyri A, DiMaio D. Human papillomavirus in cervical and head-and-neck cancer. *Nat Clin Pract Oncol* 2008;5:24–31.
38. Tsitsiou E, Lindsay MA. microRNAs and the immune response. *Curr Opin Pharmacol* 2009;9:514–20.
39. Khew-Goodall Y, Goodall GJ. Myc-modulated miR-9 makes more metastases. *Nat Cell Biol* 2010;12:209–11.
40. Di Lisi L, Gomez-Lopez G, Sanchez-Beato M, Gomez-Abad C, Rodriguez ME, Villuendas R, et al. Mantle cell lymphoma: transcriptional regulation by microRNAs. *Leukemia* 2010;24:1335–42.
41. Wang X, Wang HK, McCoy JP, Banerjee NS, Rader JS, Broker TR, et al. Oncogenic HPV infection interrupts the expression of tumor-suppressive miR-34a through viral oncoprotein E6. *RNA* 2009;15:637–47.
42. Kumar B, Cordell KG, Lee JS, Worden FP, Prince ME, Tran HH, et al. EGFR, p16, HPV Titer, Bcl-xL and p53, sex, and smoking as indicators of response to therapy and survival in oropharyngeal cancer. *J Clin Oncol* 2008;26:3128–37.
43. Yamakuchi M, Lowenstein CJ. MiR-34, SIRT1, and p53: the feedback loop. *Cell Cycle* 2009;8:712–5.
44. Gollin SM. Chromosomal alterations in squamous cell carcinomas of the head and neck: window to the biology of disease. *Head Neck* 2001;23:238–53.
45. Das S, Foley N, Bryan K, Watters KM, Bray I, Murphy DM, et al. MicroRNA mediates DNA demethylation events triggered by retinoic acid during neuroblastoma cell differentiation. *Cancer Res* 2010;70:7874–81.
46. Shtutman M, Baig M, Levina E, Hurteau G, Lim CU, Broude E, et al. Tumor-specific silencing of COPZ2 gene encoding coatomer protein complex subunit zeta 2 renders tumor cells dependent on its paralogous gene COPZ1. *Proc Natl Acad Sci U S A* 2011;108:12449–54.
47. Wang WX, Kyriianou N, Wang X, Nelson PT. Dysregulation of the mitogen granulin in human cancer through the miR-15/107 microRNA gene group. *Cancer Res* 2010;70:9137–42.
48. Wilfred BR, Wang WX, Nelson PT. Energizing miRNA research: a review of the role of miRNAs in lipid metabolism, with a prediction that miR-103/107 regulates human metabolic pathways. *Mol Genet Metab* 2007;91:209–17.
49. Liu X, Chen Z, Yu J, Xia J, Zhou X. MicroRNA profiling and head and neck cancer. *Comp Funct Genomics*. 2009 Jun 1. [Epub ahead of print].
50. Trajkovski M, Hausser J, Soutschek J, Bhat B, Akin A, Zavolan M, et al. MicroRNAs 103 and 107 regulate insulin sensitivity. *Nature* 2011;474:649–53.

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