Imaging, Diagnosis, Prognosis

Potentially Prognostic miRNAs in HPV-Associated Oropharyngeal Carcinoma

Angela B.Y. Hui1, Alice Lin1, Wei Xu2, Levi Waldron6, Bayardo Perez-Ordonez7, Ilan Weinreb7, Wei Shi1, Jeff Bruce1, Shao Hui Huang5,8, Brian O’Sullivan5,8, John Waldron5,8, Patrick Gullane3, Jonathan C. Irish3, Kelvin Chan4, and Fei-Fei Liu1,5,8

Abstract

Purpose: Deregulation of miRNAs is associated with almost all human malignancies. Human papillomavirus (HPV)-associated oropharyngeal carcinoma (OPC) has a significantly more favorable outcome compared with HPV-negative OPCs; however, the underlying mechanisms are not well understood. Hence, the objectives of this study were to determine whether miRNA expression differed as a function of HPV status and to assess whether such miRNAs provide prognostic value beyond HPV status.

Methods: Global miRNA profilings were conducted on 88 formalin-fixed and paraffin-embedded (FFPE) OPC biopsies (p16-positive: 56; p16-negative: 32), wherein the expression levels of 365 miRNAs plus 3 endogenous controls were simultaneously measured using quantitative real-time (qRT)-PCR. Seven FFPE specimens of histologically normal tonsils were used as controls.

Results: Overall, 224 miRNAs were expressed in more than 80% of the investigated samples, with 128 (57%) being significantly differentially expressed between tumor versus normal tissues ($P < 0.05$). Upregulated miR-20b, miR-9, and miR-9/* were significantly associated with HPV/p16-status. Three miRNA sets were significantly associated with overall survival (miR-107, miR-151, miR-492; $P = 0.0002$), disease-free survival (miR-20b, miR-107, miR-151, miR-182, miR-361; $P = 0.0001$), and distant metastasis (miR-151, miR-152, miR-324-5p, miR-361, miR492; $P = 0.0087$), which retained significance even after adjusting for p16 status. The associated biologic functions of these miRNAs include immune surveillance, treatment resistance, invasion, and metastasis.

Conclusion: We have identified several miRNAs, which associate with HPV status in OPC; furthermore, three candidate prognostic sets of miRNAs seem to correlate with clinical outcome, independent of p16 status. Furthermore, evaluations will offer biologic insights into the mechanisms underlying the differences between HPV-positive versus HPV-negative OPC. Clin Cancer Res; 19(8); 2154–62. ©2013 AACR.

Introduction

Over the past 2 decades, the incidence of human papillomavirus (HPV)-associated head and neck squamous cell carcinomas (HNSCC) involving the oropharynx has been increasing, nowadays comprising the majority of oropharyngeal carcinoma (OPC) cases seen in North America (1–3). There are several unique clinical characteristics of HPV-positive OPC, such as younger age at diagnosis, lower likelihood of heavy smoking or alcohol, and greater degree of sexual activity, compared with patients with HPV-negative OPC (1, 2). Intriguingly, HPV-positive patients experience a significantly superior clinical outcome when treated with either radiotherapy alone or combined chemoradiotherapy, despite presenting with higher grade and stage of disease (3). We have previously reported that approximately 60% of patients with OPC seen at the Princess Margaret Cancer Center were HPV-positive, with 3-year overall survival (OS) rates of 88% versus 67% in favor of HPV-positive versus negative patients (4). However, the biologic mechanisms behind this unique clinical entity of HPV-associated OPC remain to be elucidated.

Deregulation of miRNAs is clearly associated with the development and progression of human malignancies. We have conducted global miRNA profiling of HNSCC (5), through which the miR-375/*metadherin axis was newly identified as a potentially important pathway that could partially explain the propensity for lung metastases in this...
Translational Relevance

The incidence of human papilloma virus (HPV)-associated oropharyngeal carcinoma (OPC) has been increasing. HPV-positive OPCs have several unique clinical characteristics and with a significantly more favorable outcome, compared with HPV-negative OPCs. The biologic basis behind this differential outcome is currently unelucidated. Derepression of miRNAs is associated with oncogenesis of various malignancies, suggesting that miRNA expression profiling have the potential to unravel the complex biology of human tumors. In this study, we evaluated the miRNA profiles of archival formalin-fixed and paraffin-embedded diagnostic biopsy specimens from 88 nonmetastatic OPC samples (p16+; p16−). We have identified a panel of p16/HPV-associated miRNAs and three potential miRNA signature sets that are associated with clinical outcome, independent of p16 status. Furthermore, examination of these candidate miRNAs will inform biologic insights into the mechanisms underlying the differences between HPV-positive and HPV-negative OPC.

Materials and Methods

Patient information

A subset of diagnostic formalin-fixed paraffin-embedded (FFPE) blocks from our previously published 111-sample OPC report was evaluated (4). That study showed that HPV status as defined by p16 immunohistochemistry (IHC), HPV16 in situ hybridization (ISH), or HPV16 E6 transcript levels using quantitative real-time PCR (qRT-PCR) were strongly concordant (4). Given that these were small diagnostic biopsies, only 88 samples had sufficient remaining tumor tissues to carry out global miRNA profiling.

RNA purification from FFPE samples

A representative section from each sample was stained with hematoxylin and eosin stain and reviewed by head and neck cancer pathologists (B. Perez-Ordonez or I. Weinreb) to identify regions containing more than 70% malignant epithelial cells for macrodissection. Seven normal tonsillar epithelial FFPE tissues from individuals who underwent a tonsillectomy were included as controls. University Health Network (UHN, Toronto, Canada) Institutional Research Ethics Board approval has been obtained for this study. All blocks were processed randomly, with clinical outcome unknown, to avoid experimental bias. Total RNA enriched for small RNA species was isolated using the RecoverAll Total Nucleic Acid Isolation Kit for FFPE samples (Ambion), according to the manufacturer’s instructions (15).

MiRNA profiling using TaqMan low density array

Quality of RNA samples was assessed by qRT-PCR analysis of the endogenous control RNU44 using TaqMan microRNA Assay (Applied Biosystems), as previously described (6, 15). Global miRNA expression on 88 OPC and 7 normal samples was conducted using the TaqMan Low Density Array (TLDA) Human MicroRNA Panel v1.0 (Applied Biosystems), which enabled the simultaneous quantification of 365 human miRNAs plus 3 endogenous controls (RNU6B, RNU44, and RNU48). In brief, total RNA of each sample was first reverse-transcribed with the Multiplex RT pool set, then quantitated with a TLDA array using an Applied Biosystems 7900 HT Real-Time PCR system, with the C<sub>t</sub> values determined by threshold method, according to the manufacturer’s protocol (15).

Data processing

The TLDA data were processed and analyzed as previously described (15) with some minor modifications. Any miRNAs with undetermined values in more than 80% of the tumors and more than 6 “normals” were eliminated from the analysis. Undetermined C<sub>t</sub> values, or C<sub>t</sub> values more than 36, were imputed to 40. All samples were normalized by the mean of the endogenous controls and converted into a ratio of abundance compared with the geometric mean of the abundance of the 7 normal samples, as we have previously described (15).

Significant differences in miRNA expression between tumor and normal samples were assessed using the Wilcoxon rank sum test, with multiple comparisons adjusted by Benjamini–Hochberg false discovery rate (FDR) correction, conducted in the R statistical environment (v2.6.1; ref. 16). Two-way ANOVA and the Benjamini–Hochberg correction for FDR of multiple testing were conducted on each miRNA to investigate the association with p16 and HPV status. The Spearman correlation coefficient was used to calculate P values of the consistency of the miRNA signals by using p16 and HPV as outcomes, for each miRNA.

To explore potential prognostic effects, we applied univariate survival analysis to detect association of single miRNA expression level with OS, disease-free survival (DFS), or distant relapse-free survival (DM) as previously described (4). After identifying potentially important miRNAs (P < 0.05), we applied multivariate models on these miRNAs. Model selection procedure was conducted using a stepwise selection algorithm, conducted separately for each outcome. Finally, after model selection, additional analyses were conducted to generate a risk score based on the significant miRNAs identified from the multivariate...
The risk score was based on the weighted combination of the miRNAs with the estimated Cox proportional hazard regression model coefficient as the weight (17). For each outcome, patients were dichotomized into 2 categories of low (risk score < median), or high risk (risk score > median); following which, outcomes were compared for each risk group. Test of difference between the risk categories was assessed using the log-rank test or Cox proportional hazard regression model, with P values, and HRs (including 95% confidence interval (CI)).

To validate the prognostic effect, we carried out internal validation procedures using a bootstrap algorithm (18) and constructed the bootstrap CI. The bootstrap is one of the resampling techniques, and bootstrap datasets were created by sampling with replacement. The bootstrap method has been shown to provide a valid estimation of prediction error and can correct for the bias of the parameter estimate (18). We applied bootstrap based on 500 replications; these results were obtained using the PROC SURVEYSELECT procedure in SAS version 9.3, and presented as the bootstrap HR CIs of the prognostic effect adjusted for p16 status.

Results

Differentially expressed miRNAs in OPC

Genome-wide miRNA profiles of 88 OPC patient samples were conducted; the clinical characteristics are provided in Table 1. The median follow-up time for all patients has now extended to 5.9 years, with 5-year OS, and DFS rates at 55% and 52%, respectively (Supplementary Fig. S1). The 5-year OS for HPV-positive versus negative patients were 65% versus 30%; the DFS rates were 70% versus 20% respectively.

Among the 365 interrogated miRNAs, 224 were expressed in more than 80% of the samples, with 128 being significantly differentially expressed between the 88 OPC and 7 normal tonsillar epithelial tissues (P < 0.05). Ninety-two of these 128 miRNAs were more than 2-fold differentially expressed (Supplementary Fig. S2), with the majority (120/128; 94%) being upregulated in OPC (Supplementary Table S1). A detailed comparison of the top 6 most significantly differentially expressed miRNAs (miR-21, let-7g, miR-25, let-7f, miR-130b, and miR-151) are provided in Fig. 1. These dysregulated miRNAs seemed to be randomly distributed among different chromosomal regions, with the exception of 1p34.2 and 7q22.1, within which 6 of the top 40 aberrantly expressed miRNAs are located (bold information in Supplementary Table S1).

Table 1. Clinical description of the 88 patients with OPC

<table>
<thead>
<tr>
<th>Patient characteristics</th>
<th>HPV positive (n = 56; 64%)</th>
<th>HPV negative (n = 32; 36%)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>41 (73%)</td>
<td>24 (75%)</td>
<td>0.95</td>
</tr>
<tr>
<td>Female</td>
<td>15 (27%)</td>
<td>8 (25%)</td>
<td></td>
</tr>
<tr>
<td>Age (median; range)</td>
<td>Mean: 55 ± 11 y (range: 27–80 y)</td>
<td>Mean: 67 ± 11 y (range: 46–93 y)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Stage</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>0</td>
<td>1 (3%)</td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>2 (5%)</td>
<td>7 (22%)</td>
<td>0.003</td>
</tr>
<tr>
<td>III</td>
<td>5 (12%)</td>
<td>7 (22%)</td>
<td></td>
</tr>
<tr>
<td>IV</td>
<td>49 (83%)</td>
<td>17 (53%)</td>
<td></td>
</tr>
<tr>
<td>Treatment</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RT</td>
<td>26 (46%)</td>
<td>26 (81%)</td>
<td>0.003</td>
</tr>
<tr>
<td>CRT</td>
<td>30 (54%)</td>
<td>6 (19%)</td>
<td></td>
</tr>
</tbody>
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Abbreviation: C-RT, chemoradiotherapy; RT, radiotherapy.
consistently aberrantly expressed, but did not reach statistical significance, likely due to the small sample size of the validation cohort.

Potential prognostic miRNA sets

Using the miRNA profiles of these 88 samples, with their median value as the cutoff, and treated as binary predictors, 3 potential miRNA signature sets were identified that were associated with OS, DFS, and DM (Fig. 4). The candidate prognostic panel for OS was defined by upregulated miR-107 and miR-151, and downregulated miR-492 (Fig. 4B; \( P < 0.0001 \)). For the DFS set, this composed of upregulated miR-107, miR-151, miR-182, and miR-361 with downregulated miR-20b (Fig. 4C; \( P \leq 0.0001 \)). The DM set was defined by upregulated miR-151, miR-361 and miR-324-5p, as well as downregulated miR-492 and miR-152 (Fig. 4D; \( P = 0.0088 \)). Even after adjusting for p16 IHC status, these \( P \) values still retained significance (adjusted \( P \) values were 0.0002, 0.0001, and 0.0087 for OS, DFS, and DM, respectively).

In silico analysis

We proceeded to investigate the putative biologic functions of these aforementioned miRNAs using in silico analysis (Fig. 5). For miRNAs differentially expressed in OPC (Supplementary Table S2), miR-30c, miR-30e-3p, and miR-30e-5p all resided within the fifth intron of NFYC on chromosome 1p34.2 (19). NFYC is the nuclear transcription factor Y-\( \gamma \), which binds with CCAAT motifs in the promoter region of several genes, associated with MHC class II determinants in immune response (Fig. 5A; ref. 19).

As described in our previous publication (5), overexpression of miR-25, miR-93, and miR-106b and its host gene MCM7 can be regulated by E2F1, interfering with TGF-\( \beta \) signaling in HNSCC. Furthermore, the miR-106b-25 cluster has been reported to target PTEN (20). Six of the HPV/p16-associated miRNAs (miR-381, miR-412, miR-380-5p, miR-487b, and miR-382) are all located at a common fragile site (FRA) at 14q32.3 (Fig. 5B; refs. 21, 22). Aberrations of 14q32.31 miRNA cluster have been previously described in human malignancies with an emerging role in immune regulation (23, 24). MiR-9 was found to be induced by lipopolysaccharide-inflammatory stimuli mediated by the proinflammatory cytokines IL-1 and TNF-\( \alpha \) (25). Moreover, expression of miR-34a can be induced by p53 under genotoxic stress (26), and miR-20b has been described to target the 3\(^{\prime} \) untranslated region of hypoxia-inducible factor-1a and VEGF (27).

Overexpression of miR-151 with its host gene protein tyrosine kinase 2 (PTK2/FAK) has been reported to induce tumor invasion and metastasis in hepatocellular carcinoma (HCC; Fig. 5C; ref. 28). It also acts synergistically with FAK to enhance HCC cell motility and spreading (28, 29). Several miR-152 targets have also been identified, including the DNA methyltransferase DNMT1, E2F3, MET, and Rictor (30). Moreover, miR-182 was reported to be upregulated in

Figure 1. Expression levels of the top 6 miRNAs associated with OPC (miR-21, let-7g, miR-25, let-7f, miR-130b, and miR-151), comparing 88 tumors with that of the 7 normal tonsillar epithelial tissues. The data are presented as fold change in log2 space, normalized to that of RNU44, 48, and 6B.
upregulated miR-20b, miR-9, and miR-9*

R tumour suppressors such as
tified, with miR-21 being the most significantly upregulated
miRNAs, respectively. These 2 sets of
ISH-associated miRNAs, and
ment and cellular metabolism (34).
miR-107 has been associated with mammalian develop-
metastasis (32) and poor clinical outcome (33). Finally,
multidrug-resistant cell lines (31), associated with both
metastasis (32) and poor clinical outcome (33). Finally,
miR-107 has been associated with mammalian develop-
and cellular metabolism (34).

Discussion
We have conducted a global miRNA profiling study
focused strictly on OPC, with approximately two-third
being HPV-positive and one-third being HPV-negative. A
panel of miRNAs significantly differentially expressed
between tumor versus normal tissues (P < 0.05) was identi-
fied, with miR-21 being the most significantly upregu-
lated miRNA. MiRNA-21 is one of the most consistently reported
aberrant miRNAs in HNSCC, known to target multiple
tumour suppressors such as PTEN and TPM1 and Bcl-2
(5, 7, 10, 12). Upregulated miR-20b, miR-9, and miR-9∗
were also significantly associated with HPV/p16-status.
Furthermore, there were 3 candidate miRNA sets that
were associated significantly with OS, DFS, and DM, even after
adjusting for HPV status.

Similar to our previous HNSCC miRNA study (5), upregu-
lation of the miR-106b-25 cluster was also observed in
these OPC samples. These included the nonrandom over-
expression of miR-25, miR-93, and miR-106b, located on
chromosome 7q22.1, within the intronic region of MCM7.
Overexpression of MCM7, as well as regulation of the miR-
106b-25 cluster and MCM7 by E2F1, with subsequent
interference of TGF-β signaling in HNSCC has been previ-
ously noted (5). Interestingly, TGF-β polymorphisms have been
reported to be a susceptibility marker for HPV16 status
amongst patients with OPC (35). Furthermore, the
miR-106b-25 cluster can also target PTEN (20), which in turn
will activate AKT, which correlates with adverse outcome for

Figure 2. Comparison of the association between miRNAs with p16
IHC, and association between miRNAs with HPV16 ISH, as defined
by P values. The x and y axes refer to –log10 (P values) of HPV16
ISH-associated miRNAs, and –log10 (P values) of p16 HPV-associated
miRNAs, respectively. These 2 sets of P-values are highly correlated
with R² = 0.8. MiRNAs with significant associations (P < 0.01) for both HPV16
ISH and p16 IHC are specifically identified; green denotes
downregulation; red denotes upregulation.

multidrug-resistant cell lines (31), associated with both
metastasis (32) and poor clinical outcome (33). Finally,
miR-107 has been associated with mammalian develop-
and cellular metabolism (34).

Many of the p16-associated miRNAs were located at
common FRAs, such as 1p, 1q, 5q, 8p, 14q32.31, 16p13.12, 17p, and Xq (Supplementary Table S2; ref. 21).
This is consistent with previous studies reporting that the
majority (53%) of deregulated miRNAs were located in
cancer-associated genomic regions or FRAs (21). Chromo-
some 14q32.31 is the only nonrandom region that har-
tored 5 (miR-381, miR-412, miR-380-5p, miR-487b, and
miR-382) of the p16-associated miRNAs, which is also one
of the most common HPV integration sites in cervical cancer
(22). In fact, a cluster of 46 miRNAs (3.2% of 1,426 human
miRNAs; http://genome.ucsc.edu/cgi-bin/hgTracks) have
been mapped to this small region of approximately 45 kb.
Aberrations of the 14q32.31 miRNA cluster has been docu-
mented in many human cancers and its role in immune
surveillance is an emerging area of evaluation (Fig. 5B;
ref. 23, 24). Hence, in OPC, frequent aberrant expression
of these p16-associated miRNAs at 14q32.31 might relate to
this being a preferential HPV integration site. This postulate
would be supported by the recent emerging exome sequenc-
ing data from TCGA wherein 14q32.3 amplifications have
been noted for HPV-positive HNSCC (https://wiki.nci.nih.gov/display/TCGAM/03-08-12+HNSCC+AWG).

Enhanced immune response induced by HPV has been
suggested as one possible mechanism for the superior
outcome of HPV-associated OPC (37). Among the p16-
associated miRNAs, several have been previously described
to be associated with immune regulation, such as miR-9,
miR-9∗, miR-146a, miR-34a, and miR-155 (Supplementary
Table S2 and Fig 5B; ref. 38). Upregulation of miR-9 and its
passenger strand miR-9∗ were among the most significantly
associated miRNAs with both OPC, and specifically p16-
positive OPC. Overexpression of miR-9-9∗ has been
described in other human cancers (39), with miR-9 poten-
tially involved in immune response during inflammatory
stimuli (25). In addition, miR-9 might also have an onco-
genic role in mediating angiogenesis and metastasis by
inducing Myc to target E-cadherin, priming cancer cells for
epithelial–mesenchymal transition (39). MiRNA-20b was
the most significant p16-associated miRNA in OPC, and its
downregulation was associated with poor DFS (Fig. 4 and
4C). The potential tumor suppressor role of miR-20b has
been investigated wherein low circulating levels of miR-20b
was reported in lung cancer and mantle cell lymphoma, also
associated with poor outcome (40). Inhibition of miR-20b
can lead to an increase HIF-1α and VEGF under normoxia;
conversely, an increase of miR-20b in hypoxic tumor cells
decreased HIF-1α and VEGF (27). This negative regulatory
loop between HIF-1α and miR-20b has been suggested as
one of multiple mechanisms by which tumor cells can
rapidly adapt to varying oxygen concentrations (27).
MiR-34a was found to have higher expression levels in
HPV-positive OPC, which is distinctly different from cervix
cancer whereby miR-34a is downregulated by E6 via desta-
bilization of p53 (41). In OPC, the relationship between
p53 and HPV remains unclear (4, 42). Our previous study observed p53 to be overexpressed on IHC in 63% of cases and only borderline association with HPV positivity (4). Expression of miR-34a was shown to be induced by p53 under genotoxic stress conditions (Fig. 5B; ref. 26). Furthermore, there is a positive feedback loop wherein p53 induces miR-34a; in turn, miR-34a activates p53 by inhibiting SIRT1, leading to an increase in miR-34a expression in cells with wild-type p53, resulting in enhanced apoptosis (43). This might, in part, explain the high incidence (70%) of OPCs harboring p53 immunoexpression, particularly in association with p16 (4).

Finally, in relation to the potential miRNA signature associated with clinical outcome, upregulation of miR-151 was the only miRNA associated with all of OS, DFS, and DM (Fig. 4A), as well as being among the top 10 significantly deregulated miRNAs in OPC (Supplementary Table S1). Its chromosomal location on 8q24.3 is frequently amplified in human cancers, including HNSCCs (44). Overexpression of miR-151 has been reported to promote invasion and metastasis in HCC, by targeting the putative metastasis suppressor RhoGDIA, which in turn synergized with FAK to enhance motility (28, 29). Downregulation of miR-152 is another well-characterized tumor suppressor miRNA commonly inactivated by promoter hypermethylation (30, 45), targeting several known important mediators of tumor progression (Fig. 5C; ref. 30). It resides in the intron of the host gene Coatomer protein complex v2 (COPZ2), which is also a putative tumor suppressor (46).

Oncogenic miR-182 (Fig. 4 & 5) belongs to the miR-183-96-182 cluster, which was reported to regulate zinc homeostasis in prostate cancer, and consistently upregulated in
multidrug-resistant cell lines (31). It has been implicated in metastasis and poor survival in cancers (32, 33). Overexpression of miR-107 was observed to be associated with worse outcome (Fig. 4) and inversely related to HPV-positivity in OPC (Supplementary Table S2). It is a member of the miR-15/107 miRNA gene group associated with mammalian development (34), as well as regulation of the miRNA processing machinery and cellular metabolism.

Figure 4. Potential miRNA signature sets that are associated with OS, DFS, and DM. A, Venn diagram of the candidate miRNAs as a function of clinical outcome (OS, DFS, and DM). B, Kaplan–Meier survival curves for OS as a function of the median expression level of the 3 miRNAs (miR-107, miR-151, and miR-492). C, DFS actuarial plot as a function of the median expression level of the 5 miRNAs (miR-107, miR-151, miR-182, miR-20b, and miR-361). D, DM actuarial plot as a function of the median expression levels of the 5 miRNAs (miR-151, miR-492, miR-361, miR-152, and miR-324-5p). The P values indicated in all 3 graphs have already been adjusted for p16 status. Green denotes underexpression; red denotes overexpression of each candidate miRNA.

Figure 5. In silico analysis of selected key OPC-associated miRNAs. A, selected miRNAs significantly differentially expressed between OPC versus normal tissues: miR-30c, miR-30e-5p, miR-25, miR-93, and miR-106b, along with their host genes, and putative mRNA targets. B, selected miRNAs significantly associated with p16 status: miR-381, miR-412, miR-380-5p, miR-487b, miR-382, miR-9/9*, miR-34a, and miR-20b. C, selected miRNAs significantly associated with clinical outcome: miR-151, miR-152, miR-182, and miR-107. Green denotes underexpression; red denotes overexpression of candidate miRNAs.
(34, 47). It resides within the introns of the host gene PANK1, involved in catalyzing the formation of CoA during the Krebs cycle (48). Downregulation of miR-107 has been noted in HNSCC (49), but in contrast, for this study, we report a contradictory observation in that miR-107 overexpression was associated with poor survival. These data indicate that dysregulation of miR-107 in OPC might well be quite complex; although its oncogenic function has been previously reported (34, 47, 50).

A 6 miRNA signature set that is associated with survival in OPC was recently reported (11). None of these 6 miRNAs overlapped with our 3 potential miRNA signature sets. Among the 5 miRNAs that were suggested to be associated with HPV by Gao and colleagues, upregulated miR-9 and miR-155 were also among the significantly HPV associated miRNAs in our current study (Supplementary Table S2). However, no overlap was observed in HPV-associated miRNAs between those 5 miRNAs described by Gao and colleagues (11) with the 21 HPV-associated miRNAs reported by Lajer and colleagues (13). Downregulation of miR-145 was found to be associated with HPV in both our current study and that conducted by Lajer and colleagues (13). MiR-381 and miR-101 were also identified in both studies, but with the fold changes in opposite directions (13). One possible reason for the limited overlap amongst these various studies (Lajer and colleagues vs. current vs. Gao and colleagues) could be due to the different number of miRNAs being included at the outset (847 vs. 365 vs. 96).

Moreover, each study included different proportions of HPV-positive cases: 42% versus 64% versus 82% for Lajer and colleagues versus current versus Gao and colleagues, respectively. The methods used to define HPV-positivity were also distinct (combination of 2 positives of p16 IHC, RT-PCR, or HPV ISH vs. p16 IHC vs. RT-PCR). Controversy still remains regarding the best method of HPV detection, with discrepancies ranged from 60% to 92% among these aforementioned methods (4, 13). These variations hopefully will be resolved by the development of miRNA sequencing of FFPE samples, which will enable the interrogation of all known human miRNAs, and the integration of efforts currently being undertaken by the TCGA head and neck cancer group.

Conclusion

We have conducted a comprehensive miRNA profiling study focused strictly on OPC, which identified 3 potential miRNA signature sets associated with clinical outcome, independent of HPV status. The emerging picture of these dysregulated miRNAs points to a complexity of pathways involved in immune response, and tumor progression, located within FRA and HPV integration sites. Furthermore, examination of these candidate miRNAs will inform biologic insights into the mechanisms underlying the differences between HPV-positive and HPV-negative OPC.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors' Contributions

Conception and design: A.B.Y. Hui, W. Xu, J.C. Irish, F.-F. Liu


Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): A.B.Y. Hui, A. Lin, B. Perez-Ordonez, J. Weinreb, S. Hui Huang, B. O’ Sullivan, J. Waldron, P. Guillaume

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): A.B.Y. Hui, W. Xu, L. Waldron, J. Bruce, S. Hui Huang, B. O’Sullivan, J. Waldron, K. Chan

Writing, review, and/or revision of the manuscript: A.B.Y. Hui, W. Xu, J. Weinreb, S. Hui Huang, B. O’Sullivan, P. Guillaume, J.C. Irish, K. Chan, F.-F. Liu

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): A.B.Y. Hui, W. Shi, S. Hui Huang, B. O’Sullivan, J. C. Irish, F.-F. Liu

Study supervision: A.B.Y. Hui, J. Waldron, J.C. Irish, F.-F. Liu

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