Integrative and Comparative Genomic Analysis of HPV-Positive and HPV-Negative Head and Neck Squamous Cell Carcinomas

Tanguy Y. Seiwert1,2,3, Zhixiang Zuo1, Michaela K. Keck1, Arun Khattri1, Chandra S. Pedamallu4,5, Thomas Stricker6, Christopher Brown7, Trevor J. Pugh4,5, Petar Stojanov4, Juok Cho4, Michael S. Lawrence8, Gad Getz8, Johannes Brägelmann1, Rebecca DeBoer1, Ralph R. Weichselbaum2,8, Alexander Langerman2,9, Louis Portugal2,9, Elizabeth Blai2,9, Kerstin Stenson2,9, Mark W. Lingen2,10, Ezra E.W. Cohen1,2, Everett E. Vokes1,2, Kevin P. White2,3, and Peter S. Hammerman4,5

Abstract

Purpose: The genetic differences between human papilloma virus (HPV)–positive and –negative head and neck squamous cell carcinomas (HNSCC) remain largely unknown. To identify differential biology and novel therapeutic targets for both entities, we determined mutations and copy-number aberrations in a large cohort of locoregionally advanced HNSCC.

Experimental Design: We performed massively parallel sequencing of 617 cancer-associated genes in 120 matched tumor/normal samples (42.5% HPV-positive). Mutations and copy-number aberrations were determined and results validated with a secondary method.

Results: The overall mutational burden in HPV-negative and HPV-positive HNSCC was similar with an average of 15.2 versus 14.4 somatic exonic mutations in the targeted cancer-associated genes. HPV-negative tumors showed a mutational spectrum concordant with published lung squamous cell carcinoma analyses with enrichment for mutations in TP53, CDKN2A, MLL2, CCND1, NSE1, PIK3CA, and NOTCH genes. HPV-positive tumors showed unique mutations in DDX3X, FGFR2/3 and aberrations in PIK3CA, KRAS, MLL2/3, and NOTCH1 were enriched in HPV-positive tumors. Currently targetable genomic alterations were identified in FGFR1, DDR2, EGFR, FGFR2/3, EPHA2, and PIK3CA. EGFR, CCND1, and FGFR1 amplifications occurred in HPV-negative tumors, whereas 17.6% of HPV-positive tumors harbored mutations in fibroblast growth factor receptor genes (FGFR2/3), including six recurrent FGFR3 S249C mutations. HPV-positive tumors showed a 5.8% incidence of KRAS mutations, and DNA-repair gene aberrations, including 7.8% BRCA1/2 mutations, were identified.

Conclusions: The mutational makeup of HPV-positive and HPV-negative HNSCC differs significantly, including targetable genes. HNSCC harbors multiple therapeutically important genetic aberrations, including frequent aberrations in the FGFR and PI3K pathway genes. Clin Cancer Res 21(3): 632–41. ©2014 AACR.

See related commentary by Krigsfeld and Chung, p. 495

Introduction

Head and neck squamous cell carcinoma (HNSCC) is the fifth most common non-skin cancer worldwide with an annual incidence of 600,000 cases and a mortality rate of 40% to 50% despite aggressive treatment (1, 2). The major known risk factors are environmental exposure to tobacco products, alcohol, and infection with high-risk human papilloma viruses (HPV). The incidence of HPV-positive tumors is rising rapidly in Western countries and HPV status is the strongest clinically applicable prognostic marker, portending a favorable prognosis (3, 4).

Although HNSCC is widely viewed as comprised of two distinct clinical entities, HPV-positive and HPV-negative tumors, a comprehensive list of differential molecular abnormalities, in particular therapeutically relevant genetic aberrations, has not been reported. A particular problem is the lack of study of HPV-positive HNSCC: Currently no large series of HPV-positive tumors exist reported. A particular problem is the lack of study of HPV-positive HNSCC: Currently no large series of HPV-positive tumors exist.
Head and neck squamous cell carcinoma (HNSCC) is comprised of two distinct clinical entities, HPV-positive and HPV-negative HNSCC. A comprehensive list of differential molecular abnormalities, in particular therapeutically relevant genetic aberrations, has not been reported. Furthermore, although the incidence of HPV-positive tumors is rising rapidly, there is a lack of HPV-positive cohorts. In our study, we investigate 120 locoregionally advanced HNSCC (including 42.5% HPV-positive tumors). Tumors show a differential genetic profile based on HPV status. We discover multiple, novel therapeutic targets with predicted driver character that are of high translational relevance: EGFR, CCND1, and FGFR1 amplifications occurred in HPV-negative tumors, whereas 17.6% of HPV-positive tumors harbored mutations in fibroblast growth factor receptor genes (FGFR2/3). HPV-positive tumors showed a 5.8% incidence of KRAS mutations, and 7.8% incidence of BRCA1/2 mutations. Our study provides strong support for further clinical investigation of novel therapeutic targets and candidate biomarkers in patients with HNSCC.

Translational Relevance
Head and neck squamous cell carcinoma (HNSCC) is comprised of two distinct clinical entities, HPV-positive and HPV-negative HNSCC. A comprehensive list of differential molecular abnormalities, in particular therapeutically relevant genetic aberrations, has not been reported. Furthermore, although the incidence of HPV-positive tumors is rising rapidly, there is a lack of HPV-positive cohorts. In our study, we investigate 120 locoregionally advanced HNSCC (including 42.5% HPV-positive tumors). Tumors show a differential genetic profile based on HPV status. We discover multiple, novel therapeutic targets with predicted driver character that are of high translational relevance: EGFR, CCND1, and FGFR1 amplifications occurred in HPV-negative tumors, whereas 17.6% of HPV-positive tumors harbored mutations in fibroblast growth factor receptor genes (FGFR2/3). HPV-positive tumors showed a 5.8% incidence of KRAS mutations, and 7.8% incidence of BRCA1/2 mutations. Our study provides strong support for further clinical investigation of novel therapeutic targets and candidate biomarkers in patients with HNSCC.

Unlike lung or breast adenocarcinomas, there are currently no defined targetable genetic aberrations for HNSCC, and no approved therapies are tied to genetic alterations as predictive biomarkers. All patients with HNSCC are treated with a largely uniform approach based on stage and anatomic location, typically using surgery, radio-, and chemotherapy alone or in combination (9). Cetuximab, an anti-EGFR antibody, is the only approved targeted therapy for HNSCC with a single-agent response rate of 10% to 13%. Despite the modest response rate, there are no validated predictive biomarkers for benefit from cetuximab (10, 11).

Previous studies have demonstrated frequent mutations of several genes in cohorts of largely HPV-negative HNSCC, most notably TP53, PIK3CA, CDKN2A, the TERT promoter, and NOTCH pathway gene alterations (12–16). HPV-positive HNSCC remains unclear (15).

In this study, we investigated a fully annotated patient cohort of 120 locoregionally advanced HNSCC (including 42.5% HPV-positive tumors), treated uniformly with organ-preserving chemoradiotherapy using massively parallel sequencing, copy-number profiling, and validation.

We discover distinct mutational and copy-number profiles in HPV-positive and HPV-negative tumors and identify for the first time potentially targetable mutations and copy-number aberrations that are of high translational relevance.

Materials and Methods
Chicago head and neck cancer genomics cohort
Pretreatment tumor tissues (n = 120) and matched normal DNA for patients with locoregionally advanced HNSCC treated at the University of Chicago (Chicago, IL) were obtained from the HNSCC tissue bank (UCCCC#8980).

Sample preparation
An overview of the tissue processing is provided in Supplementary Fig. S1 and described in detail in the Supplementary Methods.

HPV consensus testing
HPV16/18 status was determined by E6/E7–specific qRT-PCR. Results were corroborated by additional tests to increase accuracy, including an E6/E7 DNA–based multiplex PCR for five high-risk HPV types (17) as well as p16/CDKN2A expression, and TP53 mutations (18).

Sequencing data generation and analysis
DNA sequencing libraries were prepared following published protocols (19) and enriched using custom capture reagents (Agilent; Nimblegen (validation)). Of note, 2 × 100 bp paired-end sequencing occurred using Illumina HiSeq 2000/2500 sequencers. Of note, 617 cancer-associated genes (Supplementary Table S1A) were targeted and sequenced at high-depth in tumor/normal pairs. Median coverage at targeted bases was 231 × in the tumors and 254 × in the matched normal. Mutation and indel calling was done using bioinformatic pipelines as described previously using MuTect (20–22). Furthermore, we used Var-Walker to determine potentially relevant genetic aberrations (mutations and copy-number aberrations) using prioritization via a protein–protein interaction network focusing on frequently mutated and cancer gene census genes (23).

We used an established machine learning–based approach—Cancer-Specific High-Throughput Annotation of Somatic Mutations (CHASM)—to predict and prioritize missense mutations leading to functional changes and, thus, likely driving tumorigenesis (24). This approach was previously validated to show high

Table 1. Overview of clinical and histopathologic characteristics of samples

<table>
<thead>
<tr>
<th>Total</th>
<th>HPV+</th>
<th>HPV−</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients</td>
<td>120 (100%)</td>
<td>51 (42.5%)</td>
</tr>
<tr>
<td>Age, y</td>
<td>56.72</td>
<td>56.69</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>24 (20.0%)</td>
<td>4 (3.3%)</td>
</tr>
<tr>
<td>Male</td>
<td>96 (80.0%)</td>
<td>47 (39.2%)</td>
</tr>
<tr>
<td>Anatomic site</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oropharynx</td>
<td>67 (55.8%)</td>
<td>47 (39.2%)</td>
</tr>
<tr>
<td>Oral cavity</td>
<td>25 (20.2%)</td>
<td>2 (1.7%)</td>
</tr>
<tr>
<td>Larynx</td>
<td>19 (15.8%)</td>
<td>1 (0.8%)</td>
</tr>
<tr>
<td>Hypopharynx</td>
<td>8 (6.7%)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Others</td>
<td>3 (2.5%)</td>
<td>1 (0.8%)</td>
</tr>
<tr>
<td>Tobacco use</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Never/light smoker (&lt;10 pack years)</td>
<td>54 (45.0%)</td>
<td>33 (27.5%)</td>
</tr>
<tr>
<td>Heavy smoker (≥10 pack years)</td>
<td>66 (55.0%)</td>
<td>18 (15.0%)</td>
</tr>
<tr>
<td>Alcohol use</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Never/light drinker</td>
<td>49 (40.8%)</td>
<td>26 (21.7%)</td>
</tr>
<tr>
<td>Heavy drinker</td>
<td>71 (59.2%)</td>
<td>25 (20.8%)</td>
</tr>
<tr>
<td>T stage</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Small primary (T1-2)</td>
<td>39 (32.5%)</td>
<td>23 (19.2%)</td>
</tr>
<tr>
<td>Large primary (T3-4)</td>
<td>81 (67.5%)</td>
<td>28 (23.3%)</td>
</tr>
<tr>
<td>N stage</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0-1</td>
<td>20 (16.7%)</td>
<td>5 (4.2%)</td>
</tr>
<tr>
<td>2-3</td>
<td>100 (83.3%)</td>
<td>46 (38.3%)</td>
</tr>
<tr>
<td>Clinical stage</td>
<td></td>
<td></td>
</tr>
<tr>
<td>I-II</td>
<td>5 (4.2%)</td>
<td>3 (2.5%)</td>
</tr>
<tr>
<td>IV</td>
<td>115 (95.8%)</td>
<td>48 (40.0%)</td>
</tr>
</tbody>
</table>
specificity, and to a lesser degree sensitivity to identify "drivers" of oncogenesis (24).

Copy-number analysis was performed using sequencing data and the CONTRA algorithm (25). Results were validated on the Nanostring nCounter using predefined/custom cancer gene panels (Nanostring).

Detailed methods are available in the Supplementary Methods.

Results

Patient cohort description and sequencing metrics

The median age of patients was 56 years with a median follow-up of 48 months (Table 1). Fifty-one patients (42.5%) were HPV-positive. Of note, 45% of patients had no or light smoking histories whereas 55% had significant smoking histories, including 35% of HPV-positive tumors (N = 18). The cohort consisted of locoregionally advanced tumors with 67 (55.8%) oropharynx and 115 (95.8%) stage IV tumors (Table 1).

Detection of somatic aberrations

Analysis of targeted hybrid capture sequencing data identified a total of 5,476 point mutations and 4,562 insertion/deletion (indel) events across 120 tumor/normal pairs in targeted 617 genes (Supplementary Table S1A). Of note, 1,536 of the point mutations and 200 of the indel events were in coding regions. Of the 1,536 point mutations in coding regions, 1,209 resulted in an amino acid change in the corresponding proteins. Individuals displayed a mean point mutation rate of 2.33 mutations per Megabase (Mb) in the targeted regions and 0.22/Mb in the coding territory alone with a range of 0 to 96 mutations in the coding regions of the targeted genes. Among nonsynonymous substitutions, transitions and transversions at CpG sites, which are typically associated with tobacco exposure (26), were the most commonly observed mutational context with a rate of 21/Mb. Mutations at TpCp(A/T/G) sites, a substitution type commonly associated with virally induced cancers (26), were the second most commonly observed context at 13/Mb and enriched in the HPV-positive patients. We did not detect a significant difference in overall mutation rates by HPV status at all targeted bases (HPV-negative 2.22/Mb, HPV-positive 2.46/Mb, P = 0.64) or in the coding territory (HPV-negative 0.23/Mb, HPV-positive 0.21/Mb, P = 0.79). Coding mutations were validated at a rate of 87% (338/390) in a second independent hybrid capture experiment when evaluating statistically enriched mutations (Supplementary Table S1B and S1C).

We applied the MutSig method to identify cancer-relevant genes demonstrating evidence of statistical selection for...
Figure 2.
GISTIC-like copy-number analysis derived from next-generation sequencing data. Shown are the aberrant regions of the chromosome and the respective negative log2 of the q value of the alteration in each panel, respectively. A, copy-number gains in HPV(−) samples. B, copy-number gains in HPV(+) samples. C, copy-number losses in HPV(−) samples. D, copy-number losses in HPV(+) samples.
mutations in our cohort (20–22). (27) MutSig identified 11 genes displaying significant enrichment for mutations as defined by a q value of <0.1 in the overall cohort (Supplementary Fig. S2): TP53, CDKN2A, PIK3CA, MLL2, TPRX1, CUL3, FLG, NSD1, DDX3X, RIPK4, and HRAS. FGFR3 and FBXW7 demonstrated q values between 0.1 and 1. A second MutSig analysis, in which only genes annotated in the COSMIC database were considered, rediscovered TP53, CDKN2A, PIK3CA, HRAS, FGFR3, and FBXW7 and additionally nominated KRAS, MLL3, FGFR2, ZNF217, and RIMS2 as statistically significant with q < 0.1. We performed additional significance analyses on the separate HPV-negative and HPV-positive cohorts using the same MutSig algorithm. Analysis of mutated genes displaying enrichment in the HPV-negative cohorts demonstrated statistical enrichment for mutations of TP53, CDKN2A, MLL2/3, NOTCH1, PIK3CA, NSD1, FBXW7, DDR2, and CUL3, in the HPV-negative samples (Fig. 1A). Genes displaying statistical enrichment in HPV-positive tumors were PIK3CA, MLL3, DDX3X, FGFR2/3, NOTCH1, NF1, KRAS, and FBXW7 (Fig. 1B).

Somatic copy-number alterations were inferred from sequencing data by read-depth analysis using the CONTRA methodology (Supplementary Methods). We applied a modification of GISTIC 2.0 analysis (28) compatible with the focal sequencing data to identify recurrent peaks of amplification and deletion and identified 192 genes with false discovery rate of less than 0.25 (Supplementary Table S1D–S1G). Copy-number alterations were validated at a rate of 83% (45/54) using the Nanostring nCounter, focusing on the genes found to be significantly altered in this dataset (Supplementary Table S1H). Copy-number analysis demonstrated many previously demonstrated regions of amplification and deletion, including focal gains of EGRF, REL, BCL6, PIK3CA, TP53, CCND1, and MDM2 and losses of ATM, CDKN2A, RB1, NOTCH1, and NF1 (Fig. 1C and D; refs. 12–15, 29). Significantly amplified regions that occurred primarily in HPV-negative tumors were amplifications of 11q13 likely targeting CCND1, 7p11 (EGFR; Fig. 2A). Amplification of 3q26–28, a region containing SOX2/TP63/PIK3CA (Fig. 2A and B) occurred in both HPV-positive and HPV-negative tumors. 3p loss and CDKN2A deletions (Fig. 2C) occurred primarily in HPV-negative tumors whereas ATM deletions occurred primarily in HPV-positive tumors (Fig. 2D).

Comparison of altered networks for HPV-positive tumors versus HPV-negative tumors

To identify additional biologic differences between HPV-positive and HPV-negative tumors, we used an unbiased objective protein–protein interaction network-based analysis of genetic aberrations (VarWalker; Fig. 3). The altered networks for HPV-negative tumors contained 84 proteins connected by 76 interactions (Fig. 3A). For HPV-positive tumors, the altered networks contained 88 proteins connected by 83 interactions (Fig. 3B). The two altered networks were substantially different: Alteration of p53 signaling and cell-cycle pathway genes occurred almost exclusively in HPV-negative tumors. The alteration of oxidative stress pathway genes (CUL3, NFE2L2, and KEAP1) occurred more often in HPV-negative tumors (Fig. 3A). The alteration of the DNA damage pathway (BRCA1, BRCA2, FANC, FANCA, FANCD2, and ATM), FGFR signaling (FGFR2, FGFR3, and FGFR4), JAK/STAT signaling (STAT1, JAK1, and JAK2), and immunology related genes (HLA-A and HLA-B) favored HPV-positive tumors (Fig. 3B). HPV-positive and HPV-negative tumors shared network alterations such as PI3K signaling, Notch aberrations, and SMAD signaling (Fig. 3A and B).

Targetable copy-number aberrations and mutations

Somatic mutations in potentially targetable kinase genes occurred in FGFR2 and FGFR3 uniquely in HPV-positive...
Figure 4.
Genomic alterations with translational or clinical relevance by HPV status. A, shown is the incidence of selected potentially targetable genes and their association with key pathways in HPV (−) and HPV (+) samples. B, gene diagrams for a selection of key mutations in potentially targetable genes, or presumed driver genes. Shown are the gene domains that are affected by somatic mutations; the number of cases that harbor the respective mutation is indicated on the ordinate axis, the CHASM scores, and P values for each mutation (red, <0.5 CHASM score, which indicates predicted oncogenic driver character), as well as prior reports of mutations based on occurrence in COSMIC, or prior publications.
tumors with an incidence of 17.6% among HPV-positive tumors. FGFR2 mutations included N569D and N569K (both in HPV-positive), and FGFR3 mutations included six S249C mutations, which were validated by Sanger Sequencing (all in HPV-positive), as well as a K413N somatic mutation (Fig. 4A and B). We used a previously validated machine learning-based approach named CHASM to predict and prioritize mis-sense mutations with a high likelihood of being drivers of oncogenesis (24). Both FGFR2 N569K and FGFR3 S249C have been described in several cancer types (30, 31) and showed low CHASM scores suggestive of oncogenic driver character (Fig. 4A and B). Importantly, the FGFR3 S249C mutation was identified recurrently in six HPV-positive tumors. S249C has been reported in one HPV-positive tumor in the TCGA HNSCC cohort and in the TCGA lung squamous cell and bladder carcinoma cohorts. A fraction of DDR2 and EPHA2 mutations also showed low CHASM scores, but were not recurrent, and did not cluster in any domain (Fig. 4A and B).

**PIK3CA** was the most commonly altered oncogene, including a number of established canonical mutations (E542K, E545K, and H1047R; Fig. 4A and B). Other PI3K pathway mutations included somatic events in PIK3R1, PTEN, TSC1, and TSC2, all of which have been reported to harbor mutations in other tumor types (Fig. 4A). PIK3CA mutations were more commonly observed in HPV-positive individuals, though the difference was not statistically significant in our cohort ($P = 0.20$).

Mutations in the MAPK pathway genes included seven canonical HRAS (G13V, Q61L, and K117N), KRAS (G12D, G13D, and L19F), and NRAS (Q61R) events (incidence 5.8%), as well as multiple mutations of uncertain significance in the tumor suppressor NF1 (Fig. 4A and B).

**Clinical correlation**

We first examined the mutation difference in smokers versus nonsmokers (Supplementary Table S2). In the overall cohort, tumors from smokers displayed a higher overall mutation burden as compared with nonsmokers 11.5 mutations per patient ($\geq 10$ pack years) versus 5.6 mutations per patient (nonsmokers; $P = 0.0007$, t-test). We noted a significant association among smoking and mutations in TP53 ($P = 0.0009$), CSMD3 ($P = 0.0062$), RB1CC1 ($P = 0.0161$), THSD7A ($P = 0.0319$). Mutations in ZFHX4 ($P = 0.0167$, Fisher exact test) and TRRAP ($P = 0.0392$, Fisher exact test) were significantly enriched in smokers compared with nonsmokers. Additional subgroup analyses, including drinkers versus nondrinkers, large primary versus small primary, and node negative versus node positive are provided in Supplementary Table S2, including subgroup analyses for oropharyngeal tumors from smokers displayed a higher overall mutation burden 6.6 mutations per patient (nonsmokers; $P = 0.004$, Supplementary Table S2). In the overall cohort ($\geq 10$ pack-year smoking history) versus 5.6 mutations per patient (nonsmokers; $P = 0.0167$, Fisher exact test) and enrichment for PI3K pathway alterations and rarer KRAS mutations. Somatic aberrations in DNA-repair genes (BRCA1/2, Fanconi anemia genes, and ATM) may contribute to chemotherapeutic and/or radiosensitivity of HPV-positive tumors, and interestingly occurred in HPV-positive tumors in non-/light smokers. Both tobacco smoke and defects in DNA repair are known to induce a large number of genetic aberrations, and may be distinct ways to accumulate genetic aberrations required for the emergence of cancer. Of note, a recent RAD51B was reported as an integration target for HPV16 leading to loss of its DNA-repair function (5).

In contrast, HPV-positive tumors show a distinct genetic profile with unique mutations in DDX3X, CYLD, and FGFR and enrichment for PI3K pathway alterations and rarer KRAS mutations. Somatic aberrations in DNA-repair genes (BRCA1/2, Fanconi anemia genes, and ATM) may contribute to chemotherapeutic and/or radiosensitivity of HPV-positive tumors, and interestingly occurred in HPV-positive tumors in non-/light smokers. Both tobacco smoke and defects in DNA repair are known to induce a large number of genetic aberrations, and may be distinct ways to accumulate genetic aberrations required for the emergence of cancer. Of note, a recent RAD51B was reported as an integration target for HPV16 leading to loss of its DNA-repair function (5).

FGFR and PI3K pathway aberrations are potential therapeutic targets in this population. FGFR2/3 mutations are of particular interest as they occurred in 17.6% of HPV-positive tumors, most commonly the S249C mutation, which has been shown to be an oncogenic driver in bladder cancer.

One HPV-negative tumor harbored amplification of FGFR1, which is being explored clinically for lung SCCAs (33, 34). Furthermore, oncogenic FGFR3–TACC3 fusions were recently reported in HPV-negative HNC, supporting a prominent role for...
oncogenic FGFR signaling for HPV-positive HNC whereas other kinases are genetically unaltered (33–37).

In contrast with prior smaller reports, the overall mutational burden in HPV-negative and HPV-positive HNSCC was comparable (15.2 vs. 14.4 somatic exonic mutations in the targeted cancer-associated genes), a finding corroborated when analyzing publically available data from the TCGA HNSCC cohort (5).

HPV-positive tumors in patients with significant tobacco use had a significantly higher mutational burden. Although the number of KRAS (N = 3) and TP53 (N = 2) mutations in HPV-positive oropharyngeal tumors was small, they occurred in patients with significant tobacco history, and showed 4- to 6-fold higher average mutational burden (not significant). Interestingly, the two TP53 mutations (H179R and G361S) cooccurred with KRAS (G12D and L19F) and one PIK3CA mutation (F872L), suggesting multiple oncogenic drivers in highly mutated altered HPV-positive tumors. The KRAS L19F mutation is thought to be modestly transforming (38).

Low-frequency canonical HRAS, KRAS, and NRAS mutations were identified in both HPV-positive and HPV-negative tumors. RAS mutations have been associated with poor outcome in other cancer types (e.g., lung adenocarcinomas), as well as resistance to cetuximab in colon cancer (39). Further study in HNC will be important, especially about the implications of KRAS mutations in otherwise good-prognosis HPV-positive tumors, which were present in 5.8% of HPV-positive tumors. One additional case of a canonical KRAS mutation in an HPV-positive tumor was previously reported (29).

The specific genetic events in HPV-positive or HPV-negative HNSCC likely contribute to the distinct biologic behavior and may affect therapy in the future. The lack of targeted therapies for squamous cell tumors remains challenging. In addition to FGFRs, we identify multiple potentially targetable genomic alterations, including mutations in the DDR2, EPHA2 kinases, PIK3CA, and PI3K pathway genes (e.g., TSC1/2 and NFI2) that are predicted to be oncogenic drivers. Therapeutic implications for these particular changes are being investigated clinically and preclinically (33, 40, 41).

The lack of EGFR aberrations in HPV-positive tumors is consistent with prior reports (5, 42), and clinical data (10, 11). Other potential therapeutic targets include alterations of genes, which regulate the cell cycle (e.g., CCND1 amplification as a target for cyclin-dependent kinase 4/6 (CDK4/6) inhibitors).

We identify a number of additional novel mutated pathways, providing further insight into the molecular pathogenesis of HNSCC: (i) Similar to lung SCC, MLL2 and MLL3 mutations were observed in both HPV-positive and HPV-negative tumors. MLL genes encode histone lysine methyltransferases that are involved in chromatin remodeling and recurrently mutated in several cancer types (43–46). Other studies have demonstrated an association with poor clinical outcomes (47); (ii) CUL3, NFE2L2, and KEAP1 mediate cellular responses to oxidative stress and are a frequent target for mutations in lung SCCs as well as the HPV-negative HNSCC cohort. KEAP1 binds to NFE2L2 and targets NFE2L2 for ubiquitination and subsequent degradation by the CUL3/RBX1 complex (48). Mutations are typically associated with heavy tobacco exposure, and distributed across the gene, which is suggestive of loss of function mutations; (iii) The DDXX gene was exclusively mutated in HPV-positive tumors. DDXX encodes an ATP-dependent RNA helicase and is involved in RNA processing (Supplementary Fig. S4). It has been described to be significantly mutated in medulloblastomas, and can potentiate β-catenin activity by promoter transactivation (43, 44); (iv) In addition, HPV-positive tumors harbor mutations in CYLD, a gene that has not been implicated as a hotspot for mutations, but is well known for its role in HPV-positive cervical cancer: CYLD is involved in differentiation and NFkB signaling and in response to hypoxia activates prooncogenic processes (Supplementary Fig. S4; ref. 49); (v) Finally, UBR5 mutations are involved in DNA damage and apoptosis signaling, and in addition to our discovery in HPV-positive and HPV-negative tumors, UBR5 is recurrently mutated in mantle cell lymphomas (Supplementary Fig. S4). Investigations into its role in radio- or chemosensitivity are promising (50).

In conclusion, the mutational landscape of HPV-positive HNSCC differs from HPV-negative HNSCC and may help explain the distinct clinical behavior and prognosis. Further study of clinical implications of the observed mutations will be vital with respect to targeted therapies, radio-, and chemotherapy. For the first time, we identify targetable genetic aberrations in both HPV-positive and HPV-negative HNSCC. In particular, frequent aberrations in the FGFR family are promising as well as PI3K pathway and cell-cycle abnormalities. The significant number of potential targets, including rare aberrations (DDR2, TSC1/2, and EPHA2), or candidate biomarkers such as HRAS/KRAS, indicates a need for tumor profiling of HNSCC in the future—including on clinical trials, as well as functional validation in preclinical studies.

Disclosure of Potential Conflicts of Interest
P.S. Hammerman is a consultant/advisory board member for ARIAD, Janssen, and ImmClone. No potential conflicts of interest were disclosed.

Authors’ Contributions
Development of methodology: T.Y. Seiwert, Z. Zuo, M.K. Keck, M.S. Lawrence
Writing, review, and/or revision of the manuscript (i.e., drafting or revising sections of the manuscript): T.Y. Seiwert, Z. Zuo, M.K. Keck, A. Khatatri, T. Stricker, A. Langerman, E. Blair, K. Stenson, M.W. Lingen, E.E.W. Cohen, E.E. Vokes, K.P. White, P.S. Hammerman
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): T.Y. Seiwert, Z. Zuo, R. Deboer, E.E. Vokes, P.S. Hammerman
Study supervision: T.Y. Seiwert
Other funding (i.e., reporting or organizing data, constructing databases): T.Y. Seiwert, Z. Zuo, R. Deboer, E.E. Vokes, P.S. Hammerman
Grant Support
This work was supported by an ASCO Translational Professorship award (to E.E. Vokes), the Grunt Achatz and Nick Kokonas/Alinea Head and Neck Cancer Research Fund (to E.E. Vokes and T.Y. Seiwert), a Flight Attending Medical Research Institute Young Clinical Scientist Award (FAMRI, YCSA; to T.Y. Seiwert), and a NCi K08 Award 1K08CA163677 (to P.S. Hammerman).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received December 10, 2013; revised May 9, 2014; accepted June 24, 2014; published OnlineFirst July 23, 2014.

www.aacrjournals.org
Clin Cancer Res; 21(3) February 1, 2015
639

Head and Neck Squamous Cell Carcinoma Genomic Analysis

Published OnlineFirst July 23, 2014; DOI: 10.1158/1078-0432.CCR-13-3310

Downloaded from clinicancerres.aacrjournals.org on July 13, 2017. © 2015 American Association for Cancer Research.
References


Integrative and Comparative Genomic Analysis of HPV-Positive and HPV-Negative Head and Neck Squamous Cell Carcinomas


Updated version
Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-13-3310

Supplementary Material
Access the most recent supplemental material at:
http://clincancerres.aacrjournals.org/content/suppl/2014/07/24/1078-0432.CCR-13-3310.DC1

Cited articles
This article cites 45 articles, 20 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/21/3/632.full#ref-list-1

Citing articles
This article has been cited by 26 HighWire-hosted articles. Access the articles at:
http://clincancerres.aacrjournals.org/content/21/3/632.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.