Molecular Pathways

Linking Tumor Microenvironment to Epithelial–Mesenchymal Transition in Metastasis

Hae-Yun Jung1, Laurent Fattet1, and Jing Yang1,2

Abstract

During tumor development, tumor cells constantly communicate with the surrounding microenvironment through both biochemical and biophysical cues. In particular, the tumor microenvironment can instruct carcinoma cells to undergo a morphogenesis program termed epithelial-to-mesenchymal transition (EMT) to facilitate local invasion and metastatic dissemination. Growing evidence uncovered a plethora of microenvironmental factors in promoting EMT, including proinflammatory cytokines secreted by locally activated stromal cells, hypoxia conditions, extracellular matrix components, and mechanical properties. Here, we review various biochemical and biophysical factors in the tumor microenvironment that directly impact upon the EMT program. Specifically, cytokines such as TGFβ, TNFα, and IL6 and hypoxia are capable of inducing EMT in various tumors. Several extracellular matrix (ECM) proteins, including collagen-I, fibronectin, and hyaluronan, and ECM remodeling via extracellular lysyl oxidase are also implicated in regulating EMT. In preclinical studies and ongoing clinical trials, targeting these tumor microenvironmental signals has shown promise in halting tumor progression in various human cancers. Clin Cancer Res; 21(5): 962–8. ©2014 American Association for Cancer Research.

Background

During tumor metastasis, the epithelial-to-mesenchymal transition (EMT) program has been indicated in giving rise to the dissemination of single tumor cells from primary epithelial tumors (1). EMT refers to a global cellular and molecular transition by which polarized epithelial cells gain mesenchymal properties to migrate. During EMT, epithelial cells reorganize cytoskeleton and resolve cell–cell junctions, which are accompanied with switching off the expression of epithelial markers and turning on mesenchymal genes. Although changes in epithelial and mesenchymal markers during EMT can vary significantly in different biologic contexts, a network of transcription factors, including TWIST1/2, SNAIL1/2, ZEB1/2, and FOXC2, are consistently required to orchestrate the EMT program (2). Numerous studies have shown that the expression of these transcription factors is associated with poor prognosis and distant metastasis in various human cancers (3). Besides its role in promoting tumor cell invasion, EMT is shown to confer tumor cells with resistance to apoptosis (4) and anoikis (5), thus allowing cell survival in the blood stream after intravasation. EMT could also facilitate tumor cells’ escape from the senescence program, especially through TWIST1 and ZEB1 (6, 7). Furthermore, EMT has been shown to endow cancer cells with cancer stem cell (CSC)–like features, which further aid tumor dormancy and chemoresistance (8, 9).

Studies with tumor samples or experimental tumor xenograft models have provided convincing evidence for the activation of EMT in various primary epithelial tumors. Interestingly, recent studies reveal a dynamic requirement of EMT in tumor metastasis: activation of EMT promotes local tumor invasion, intravasation, and extravasation of the systemic circulation, whereas reversion of EMT is essential to establish macrometastases in distant organs (1, 10). The “reversible” EMT model implies that EMT is unlikely to be regulated by permanent genetic and epigenetic changes in tumor cells; instead, EMT is dynamically controlled by various proinvasion signals from the tumor microenvironment (TME).

The TME is defined as the cellular and physical environment surrounding the primary tumor—including endothelial, inflammatory and immune cells, fibroblasts, extracellular matrix (ECM) components, and soluble factors. In this review, we discuss the most relevant and direct connections between TME signals and EMT-inducing transcription factors in cancer. On the basis of the properties of the TME signals, we divide our discussion into four major categories: inflammatory signals, hypoxia, ECM components, and ECM mechanical properties (Fig. 1).

Inflammatory cytokines

An association between cancer development and inflammation has long been observed. During tumor progression, tumor cells recruit activated fibroblasts and immune cells that in turn secrete many cytokines to affect tumor development and metastasis (11). Interestingly, such cytokines have been shown to directly regulate the EMT program. Transforming growth factor-β (TGFβ), abundantly secreted by cancer-associated fibroblasts, platelets, and tumor cells, is the best-characterized EMT inducer. TGFβ has been...
Figure 1. Regulation of EMT transcription factors by tumor microenvironmental signals. TGFβ regulates upregulation of TWIST1, SNAIL1, and SNAIL2 via the SMAD signaling pathway. Drugs that inhibit TGFβ are AP 12009, GC1008, and LY573636, which are in clinical trials for advanced solid tumors. TNFα activates NF-κB to induce TWIST1, SNAIL2, and ZEB1/2 expression and TNFα/NF-κB activation also increases SNAIL1 protein stability. Therapeutic approaches to inhibit TGFβ signaling include TNFα antagonist (infliximab and etanercept) and NF-κB inhibitor (bortezomib), all of which have been assessed in phase II clinical trials for several cancer types. IL6 induces TWIST1 and SNAIL1 expression via JAK/STAT3 signaling and increases TWIST1 stability through CK2-dependent phosphorylation. An IL6 ligand-blocking antibody, CNTO 328, has been tested in phase I/II clinical trials with metastatic renal cell carcinoma. HIF1α induces TWIST1 and SNAIL1 expression and HIF1α either alone or in cooperation with TGFβ promotes SNAIL1 nuclear localization to stabilize SNAIL. Agents to inhibit HIF1α include EZN-2698, PX-478, and topotecan. Topotecan has been tested in phase I/II clinical trials in combination with conventional chemotherapy, and EZN-2698 and PX-478 are currently being tested in phase I clinical trials. Collagen I can promote SNAIL1 stability through binding to its receptor DDR2 and activating SRC/ERK2 pathway. HA binding to CD44 induces nuclear translocation of CD44 to directly induce lysyl-oxidase (LOX) expression, which in turn increases TWIST1 expression.
shown to induce TWIST1 and SNAIL2 expression in prostate and non–small cell lung cancer (12, 13). TGFβ can also induce SNAIL1 and SNAIL2 via IKKα and SMAD signaling in pancreatic cancer cells (14). Furthermore, Vincent and colleagues (15) showed that SNAIL-SMAD3/4 transcriptional repressor complex could promote TGFβ-mediated EMT in breast cancer. Tumor necrosis factor-α (TNFα) is a crucial activator of the NF-κB signaling pathway, and activated NF-κB has been shown to induce multiple EMT transcription factors expression, including TWIST1, SNAIL2, and ZEB1/2 (16–18). Furthermore, Wu and colleagues (19) found that NF-κB activation could stabilize SNAIL1 to further promote cell migration and invasion. The release of interleukins by immune cells, endothelial cells, and fibroblasts can also contribute to EMT. IL6 promotes EMT in head and neck cancer cells and correlates with increased TWIST1 and SNAIL1 expressions (20). Sullivan and colleagues (21) showed that an IL6-TWIST1 positive feedback loop induces EMT in breast cancer cells. Taken together, various inflammatory cytokines from TME can regulate the expression and/or protein stability of EMT transcription factors to activate EMT and tumor invasion.

**Hypoxia**

Hypoxia condition has been shown to select tumor cells to become more invasive and metastatic. Specifically, hypoxia can promote EMT via hypoxia-inducible factor-1α (HIF1α; ref. 22). HIF1α is found to increase SNAIL1 protein stability, leading to suppression of E-cadherin in ovarian carcinoma (23). Yang and colleagues (24) found that HIF1α could induce TWIST1 expression by binding directly to the TWIST1 promoter. In addition, HIF1α cooperates with inflammatory cytokines to promote EMT. For example, HIF1α, together with TGFβ, promotes SNAIL1 nuclear translocation to induce EMT through the suppression of estrogen receptor β in prostate carcinoma (25). Also, HIF1α could enhance the expression of TWIST1 by upregulating TNFα, IL6, and TGFβ in prostate cancer (26). Hypoxia, together with the Wnt/β-catenin signaling, can also promote SNAIL1 stability by inhibiting GSK3β (27). Taken together, HIF1α, often in cooperation with additional TME factors, can induce EMT, suggesting a promising strategy to target hypoxic signaling for cancer therapeutics.

**ECM components**

ECM includes structural and nonstructural components that can activate cellular signaling through membrane-bonded receptors such as integrins. The critical role of ECM in promoting EMT was already evident in the original experiments conducted by Greenburg and Hay (28). They showed that epithelial cells from embryonic and adult anterior lens cultured in three-dimensional collagen gels can elongate and migrate as individual cells. Indeed, Greenburg and Hay (28) concluded that "interactions with ECM may be a major factor in the ability of a cell to become mesenchymal." Recently, Zhang and colleagues (29) unraveled a direct connection between ECM structural protein collagen-I and SNAIL1. They found that collagen-I binds to its receptor DDR2 and activates downstream SRC/ERK2 to stabilize SNAIL1 in breast tumor cells. SNAIL1 further upregulates MT1-MMP and collagen-I to promote tumor cell invasion. Another ECM structural component, fibronectin, partly through binding to integrin receptors, induces SNAIL1 expression in tumor cells. This study demonstrated that cooperation of fibronectin and TGFβ was required to activate the downstream SRC and ERK/MAPK kinases and induce EMT (30). Hyaluronan (HA) is a major component of ECM and signals through its membrane receptor CD44, which is overexpressed in many human cancers. HA binding to tumor cells was found to induce CD44 nuclear translocation and activate LOX expression, which in turn upregulates TWIST1 expression to promote breast cancer metastasis (31). Periostin, a nonstructural ECM component highly expressed in human tumors, could signal through integrins to increase cell survival and promote metastatic progression of colon cancer in vivo (32). Kim and colleagues (33) identified differential roles of periostin in EMT: it induces SNAIL1 expression in prostate cancer cells, whereas it inhibits TWIST1 expression in bladder cancer cells. These studies show that many ECM components are key regulators of EMT and tumor invasion.

**ECM mechanical properties**

During tumor progression, ECM is constantly remodeled by various cell types in the TME. Specifically, increasing matrix stiffness through LOX-mediated collagen cross-linking plays a critical role in tumor invasion and metastasis. Pioneer study by Paszek and colleagues (34) showed that increasing ECM stiffness induced a malignant phenotype, associated with activated FAK and ERK signaling. LOX-mediated ECM stiffening promoted tumor progression in vivo partially via an activated FAK signaling (35). Conversely, treatment with a LOX inhibitor reduced focal adhesions and PI3K signaling, demonstrating that LOX modulates tumor progression through ECM stiffening to drive focal adhesion assembly. Furthermore, ECM stiffening was required to corporate with TGFβ to induce EMT in human breast tumor cells (36), further strengthening the notion that mechanical properties of the tumor microenvironment are key factors regulating EMT and promoting tumor progression.

**Clinical-Translational Advances**

Accumulating evidence supports a critical role of EMT in many aspects of tumor development, including resistance to apoptosis and senescence, CSCs, and invasion and metastasis, thus suggesting that targeting this process could be a promising therapeutic approach. However, the core EMT transcription factors remain technically challenging to target. Instead, a number of preclinical studies suggest that inhibiting EMT-inducing TME signals could serve as alternative approaches to impinge upon the EMT program. Here, we summarize therapeutics in preclinical and clinical studies that target TME to prevent tumor progression (Table 1).

**Inflammatory cytokines**

Preclinical studies support the importance of inflammatory cytokines, including TNFα and IL6, in promoting EMT and tumor invasion. Several TNFα inhibitors have been tested in clinical trial in different types of cancers. For example, infliximab, a TNFα monoclonal blocking antibody, has been tested in phase II clinical trials in renal cell carcinoma and advanced cancers (37, 38). These studies suggested that TNFα inhibitor was effective to suppress the levels of IL6 and CCL2 in patients and improved progress-free survival. Two clinical studies examined the therapeutic effects of etanercept, a TNFα antagonist, in recurrent ovarian cancer and metastatic breast cancer. Etanercept is well tolerated in patients and significantly improved progress-free survival with consistent decrease in CCL and IL levels (39, 40). Because NF-κB is the essential downstream activator of the TNFα signaling, several clinical trials tested whether inhibition of NF-κB signaling could
<table>
<thead>
<tr>
<th>Target</th>
<th>Drug</th>
<th>Types of drug</th>
<th>Cancer types</th>
<th>Clinical studies</th>
<th>Response</th>
<th>PFS Median/mo</th>
<th>OS Median/mo</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNFα</td>
<td>Infliximab</td>
<td>Monoclonal antibody</td>
<td>Renal cell carcinoma</td>
<td>Phase II</td>
<td>Study 1: 32% PR + SD (7.7)</td>
<td>Study 1: 3.1</td>
<td>Study 1: 10</td>
<td>37</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Ovarian, renal, cervical cancer, endometrial stromal cell sarcoma, metastatic melanoma, and metastatic colorectal cancer</td>
<td>Phase I</td>
<td>62% SD (3.9)</td>
<td>Study 2: 61% SD (6.2)</td>
<td>Study 2: 4.1</td>
<td>Study 2: 13.1</td>
</tr>
<tr>
<td></td>
<td>Etanercept (Enbrel)</td>
<td>Monoclonal antibody</td>
<td>Metastatic breast cancer</td>
<td>Phase II</td>
<td>62.5% SD (4.1)</td>
<td>Study 2: 4.1</td>
<td>Study 2: 13.1</td>
<td>39</td>
</tr>
<tr>
<td></td>
<td>Bortezomib</td>
<td>Proteasome inhibitor</td>
<td>Unresectable/metastatic gastric and gastroesophageal junction adenocarcinoma</td>
<td>Phase II</td>
<td>62.5% SD (3.3)</td>
<td>1.28</td>
<td>5.08</td>
<td>41</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Recurrent ovarian cancer</td>
<td>Phase I</td>
<td>55.3% SD (6.2)</td>
<td>Study 2: 4.1</td>
<td>Study 2: 13.1</td>
<td>40</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Recurrent and/or metastatic head and neck squamous cell carcinoma</td>
<td>Phase II</td>
<td>53% PR + SD (3.0)</td>
<td>3.0</td>
<td>9.4</td>
<td>42</td>
</tr>
<tr>
<td></td>
<td>Siltuximab (CNTO-328)</td>
<td>Monoclonal antibody</td>
<td>Metastatic renal cell cancer</td>
<td>Phase I/II</td>
<td>Part 2: 54% PR + SD (7.6)</td>
<td>3.4</td>
<td>5</td>
<td>43</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Unresectable/metastatic non-small cell lung cancer</td>
<td>Phase II</td>
<td>43.8% SD (4.2)</td>
<td>2.69</td>
<td>8.48</td>
<td>46</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Unresectable/metastatic soft tissue sarcoma</td>
<td>Phase II</td>
<td>46.0% PR + SD (4.4)</td>
<td>2.64</td>
<td>8.71</td>
<td>47</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Unresectable/metastatic melanoma</td>
<td>Phase II</td>
<td>47.1% CR + PR + SD (6.6)</td>
<td>2.6</td>
<td>9.6</td>
<td>48</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Advanced solid tumors and metastatic renal cell carcinoma</td>
<td>Phase I (ongoing)</td>
<td>49</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Advanced metastatic cancer</td>
<td>Phase I (ongoing)</td>
<td>49</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Advanced refractory non-small cell lung cancer</td>
<td>Phase I</td>
<td>69.1% PR + SD (5.1)</td>
<td>5.2</td>
<td>11.5</td>
<td>50</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>GBM, anaplastic astrocytoma (AA), anaplastic oligodendroglioma (AO)</td>
<td>Pilot</td>
<td>GBM: SD (13.5)</td>
<td>AA: SD (13)</td>
<td>AO: SD (29)</td>
<td>53</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Non-GBM: 60</td>
<td>53</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Advanced solid tumors</td>
<td>Phase I</td>
<td>34% SD (15)</td>
<td>17% SD (9)</td>
<td>59</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: CR, complete response; OS, overall survival; PFS, progress-free survival; PR, partial response; SD, stable disease.

*Median/mo: median duration/months.
suppress tumor progression and metastasis. Bortezomib, a proteasome inhibitor that suppresses NF-kB activation, was tested in phase II clinical studies with metastatic gastric adenocarcinoma, and recurrent and metastatic head and neck squamous cell carcinoma (41, 42). Although bortezomib alone showed poor response in patients, combination therapy with docetaxel or targeted inhibition of other oncogetic pathways are currently under way in solid tumors. Finally, various blocking antibodies against cytokines have been used in various clinical studies. CNTO-328, an IL6 ligand-blocking antibody, was tested in phase I/II clinical trials for the treatment of metastatic renal cell carcinoma. This study showed that CNTO-328 could increase patient survival and more than 50% of progressive metastatic renal cell carcinoma patients presented stable diseases upon treatment (43). Together, these clinical trials in progress could bring a number of promising anti-inflammatory cytokine agents to the forefront of antimetastasis therapeutics.

The TGFβ signaling is extensively targeted to block tumor progression and metastasis, and various approaches have been taken to inhibit the TGFβ signaling. AP-12009, an antisense oligonucleotide against TGFβII, was tested in patients with high-grade glioma and significantly improved survival compared with standard chemotherapy treatment (44). Furthermore, TGFβ-neutralizing antibody GC-1008 showed promises in phase I trial for metastatic melanoma and renal cell carcinoma (45). Small-molecule inhibitor, LY-573636, used in phase II clinical studies in patients with metastatic NSCLC, soft tissue sarcoma, and melanoma, has also shown modest activity as a second/third-line therapy (46–48). These studies showed that inhibiting TGFβ signaling pathway is safe, well tolerated in patients and could provide promising new therapeutics against tumor invasion.

Hypoxia

Several HIF1α inhibitors have also shown remarkable antitumor activities in a variety of preclinical and clinical trials. EZN-2698, an antisense oligonucleotide against of HIF1α, is being tested in a phase I clinical trial with advanced solid tumors (49). Another HIF1α inhibitor, PX-478, which inhibit HIF1α expression, is currently tested in phase I clinical trials in patients with advanced metastatic cancer (49). Several novel compounds have also been identified in a high-throughput screen using a cell-based reporter of HIF1α transcriptional activity. One such compound, topotecan has been tested in phase I/II clinical trials with conventional chemotherapies such as cisplatin or bevacizumab in patients with advanced lung cancer. Clinical results indicate that combination treatment is well tolerated and worthy of further clinical investigation (50), thus making them promising agents against tumor metastasis.

ECM components

Disruption of tumor ECM integrity has shown promising results in halting tumor metastasis in preclinical studies. Methyllumbrelliforone, a HA synthesis inhibitor, was effective in preventing bone metastasis of lung cancer in vivo (51). Neutralizing antibody directed against periostin resulted in 40% inhibition of tumor growth (P < 0.001), 80% inhibition of lung metastasis (P < 0.001), and significant increase in survival (P < 0.05) using mouse breast tumor xenografts (52). Because cells that have undergone EMT secrete many unique ECM components, these ECM molecules have also been used for targeting drug delivery to tumors. For example, a promising approach has been used in clinical trials for patients with glioblastoma multiforme (GBM), linking anti-Tenascin C antibody to radioactive particles to specifically target tumor cells. Result showed minimal toxicity associated with a promising antitumor benefit and encouraging overall outcomes (53). Recently, engineered HA-based conjugates have emerged as a promising strategy to efficiently target tumors with drugs exerting poor solubility and strong side effects, such as paclitaxel (54). These strategies take advantage of unique EMT-associated TME components to achieve targeted delivery of traditional chemotherapeutics, thus presenting a new anticaner therapeutic strategy.

Conclusion and Discussion

As discussed, a number of inhibitors targeting TME are being tested in preclinical and clinical trials and well-tolerated in patients and several showed promising results. Because these TME signals regulate various signaling pathways, the impacts of these inhibitors on tumor progression are likely beyond the EMT program. Given the critical role of EMT in multiple steps of tumor progression, targeting the EMT–inducing TME signals is indeed worth pursuing to combat metastatic cancers. However, there are also a number of issues to be resolved to better decide how to effectively affect tumor progression by targeting the EMT program. First, current clinical trials largely aim to shrink established metastases, in which the EMT program may not be involved. Instead, metastasis prevention trials in patients with cancer with high metastasis risk would be the appropriate setting to test the effect of EMT inhibition on metastasis occurrence. Second, recent studies demonstrated the dynamic involvement of EMT in tumor metastasis: activation of EMT promotes tumor dissemination and reversion of EMT is essential for outgrowth of macrometastases. Therefore, EMT inhibitor alone could be counter-productive in preventing distant metastases if patients already have disseminated tumor cells in distant organs. In these cases, combining therapies targeting TME signals with traditional chemotherapy and targeted therapies to simultaneously inhibit EMT and cell proliferation could be a more powerful approach to eradicate both migrating as well as proliferating tumor cells, thus halting tumor progression.
Disclosures of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors’ Contributions

Conception and design: H.-Y. Jung, L. Fattet, J. Yang

Writing, review, and/or revision of the manuscript: H.-Y. Jung, L. Fattet, J. Yang

Acknowledgments

The authors apologize to the many researchers in this field whose work they were unable to cite due to space restrictions.

References


Grant Support

L. Fattet is supported by a postdoctoral fellowship from the Fondation pour la Recherche Medicale (SFRPM20130362457). J. Yang is supported by the NCI of the NIH under award number 1RO1CA168689, American Cancer Society grant RSG-09-282-01-CM3, the Hartwell Foundation, and the U.S. Department of Defense Breast Cancer Program under award number W81XWH-13-1-0132.

Received May 15, 2014; revised July 10, 2014; accepted July 18, 2014; published OnlineFirst August 8, 2014.
Clinical Cancer Research

Molecular Pathways: Linking Tumor Microenvironment to Epithelial–Mesenchymal Transition in Metastasis

Hae-Yun Jung, Laurent Fattet and Jing Yang


Updated version

Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-13-3173

Cited articles

This article cites 59 articles, 15 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/21/5/962.full.html#ref-list-1

Citing articles

This article has been cited by 2 HighWire-hosted articles. Access the articles at:
/content/21/5/962.full.html#related-urls

E-mail alerts

Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions

To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions

To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.