Enhanced Antitumor Activity of an Anti-5T4 Antibody-Drug Conjugate in Combination with PI3K/mTOR inhibitors or Taxanes

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Abstract

Purpose: Targeted treatment of solid or liquid tumors with antibody-drug conjugates (ADCs) can lead to promising clinical benefit. The aim of the study is to investigate combination regimens of auristatin-based ADCs in preclinical models of cancer.

Experimental Design: An auristatin-based anti-5T4 antibody conjugate (5T4-ADC) and auristatin payloads were combined with the dual PI3K/mTOR catalytic site inhibitor PF-05212384 (PF-384) or taxanes in a panel of tumor cell lines. Drug interactions in vitro were evaluated using cell viability assays, apoptosis induction, immunofluorescence, mitotic index, and immunoblotting. Breast cancer cells treated with auristatin analogue or 5T4-ADC were profiled by total- and phospho-proteomics. Antitumor efficacy of selected combinations was evaluated in 5T4-positive human breast or lung tumor xenografts in vivo.

Results: In vitro, auristatin-based agents displayed strong synergistic or additive activity when combined with PF-384 or taxanes, respectively. Further, treatment of 5T4-ADC plus PF-384 resulted in stronger induction of apoptosis and cell line-specific attenuation of pAKT and pGSK. Interestingly, proteomic analysis revealed unique effects of auristatins on multiple components of mRNA translation. Addition of PF-384 further amplified effects of 5T4-ADC on translational components, providing a potential mechanism of synergy between these drugs. In human tumor xenografts, dual targeting with 5T4-ADC/PF-384 or 5T4-ADC/paclitaxel produced substantially greater antitumor effects with longer average survival as compared with monotherapy treatments.

Conclusions: Our results provide a biologic rationale for combining 5T4-ADC with either PI3K/mTOR pathway inhibitors or taxanes and suggest that mechanisms underlying the synergy may be attributed to cellular effects of the auristatin payload. Chin Cancer Res; 22(2); 383–94. ©2015 AACR.
Translational Relevance

Antibody–drug conjugates (ADCs) are emerging as a promising therapeutic modality for the treatment of cancers. However, despite encouraging activity, patients treated with single-agent ADCs show relapse or progression due to the emergence of resistance. Combination of ADCs with other chemotherapeutic or targeted agents is being evaluated in the clinic to improve the overall response. In this study, in preclinical models, we systematically evaluated combination partner(s) for an auristatin-based antibody–conjugate (5T4-ADC) and demonstrate enhanced activity when 5T4-ADC was combined with a PI3K/mTOR inhibitor or with taxanes. Our findings provide a rationale to evaluate 5T4-ADC in combination with PI3K/mTOR pathway inhibitors or taxanes in the clinic. Additionally, our data suggest that the mechanistic basis of the observed synergy may be attributed to the mechanism of action of payload (i.e., auristatin). As several auristatin-based ADCs are in clinical trials, we believe that our findings can be applicable to other ADCs using auristatins.

Materials and Methods

Cell lines and reagents

Human tumor cell lines NCI-H1975, Calu-6, NCI-H358, HCC2429, MDA-MB-468, MDA-MB-231, CAOV-3, TOV-112D, OV-90, OVCAR-3, SKOV-3, HT-29, NCI-N87, Raji, and Ramos were purchased from the ATCC. MDAMB361-DYT2 cells were obtained from Dr. D. Yang (Georgetown University, Washington, DC). Cell lines were authenticated annually by short-tandem repeat analysis (Promega STR profiling service) and routinely tested for mycoplasma contamination (ATCC). MDAMB435/5T4 are cells stably transfected with human 5T4 that were described previously (23). The 37622A1 NSCLC patient-derived xenograft, and the establishment and characterization of primary serum-free culture TUM622 from 37622A1, were described (24). Each cell line was cultured in its standard medium as recommended by the ATCC. For in vitro studies, chemotherapeutic drugs were obtained from Sigma-Aldrich. PF-05212384 (PKI-587), MMAF-Ome, and auristatin 101 were obtained from Pfizer WMC. Preparation of 5T4-ADC (A1mcMMAF) was described previously.

Synergy assays

The effects of drug combinations were evaluated using Chou–Talalay median effect analysis (25). Cells were treated with each drug alone and in combination in two independent 96-well plates in a diagonal matrix format, and proliferation was measured by using a CellTiter Glo kit (Promega). Results were expressed as surviving fractions (fraction affected), based on the measured luminescence counts of treated samples, compared with that of untreated controls. Seven diagonals representing various dose–effect curves with fixed drug ratios were used to measure the combination indexes (CI) for each of the combinations with Calcusyn software (Biosoft). In each experiment, CI indexes at ED50 levels were averaged for the three dose–effect curves that had 7 to 8 data points. The CIs from two to three independent experiments were averaged to generate a single number shown in Figs. 1A and 4A. The Chou–Talalay method was used to calculate CI with the CI values of <0.9 considered as evidence of synergy: 0.9–1.1, additive effects; CI > 1.1, antagonism (25).

Western blotting

Equal amounts of proteins were subjected to immunoblotting analysis using NuPAGE electrophoresis system (Life Technologies). The primary antibodies for P-AKT (S473, S308), AKT, P-4E-BP1 (S632), 4E-BP1, P-4E-BP2, P-4E-BP3, P-eIF4G1, and P-4E-BP1 (S21), P-eIF4B (S125) were obtained from Promega. Secondary antibodies were obtained from Cell Signaling Technology. Antibodies for total 5T4 were from Abcam (EPR5530/ab129058) or described previously (24).

Xenograft efficacy studies

Female athymic nu/nu mice (18–23 g) were obtained from Charles River Laboratories. Mice were injected with tumor cells subcutaneously and animals with staged tumors were administered intravenously with saline (vehicle), 5T4-ADC, PF-384, paclitaxel, or combinations 5T4-ADC plus PF-384, 5T4-ADC plus paclitaxel. ADCs were administered based on mAb protein content at 2 or 3 mg Ab/kg on a q4d (every 4 day) schedule, with 8 to 10 mice per group. 5T4-ADC, PF-384, and paclitaxel were administered at clinically equivalent doses. All procedures using mice were approved by the Pfizer Institutional Animal Care and Use Committee according to established guidelines. Time-to-endpoint (TTE; time to no tumor or rate of tumor tripling) were used for comparison of drug-treated groups. The t test was used to compare individual tumor volumes as indicated in the figure legends.

Additional Materials and Methods are available in the Supplementary Information.

Results

5T4-ADC or auristatins synergize with PI3K/mTOR inhibitor in vitro
We performed a focused combinatorial screen of selected standard of care agents or signaling inhibitors combined with auristatin-based drugs in a panel of breast, lung, or ovarian cancer cells (unpublished data and Supplementary Table S1). Cell lines with previously characterized and clinically relevant ST4 expression levels were also included in the analysis to enable direct comparison of the drug interactions with ST4-ADC conjugate to the unconjugated auristatins (Supplementary Fig. S1, ref. 7). Through this screen, we identified the dual PI3K/mTOR inhibitor PF-384 as showing most consistent interactions with microtubule-destabilizing agents or the ST4-ADC across the panel of 16 cell lines (Fig. 1A). Overall, a similar pattern of synergistic or

![Figure 1](https://example.com/image1.png)

**Figure 1.**
Effects of auristatin-based agents combined with PF-384. A, dot plot summary showing the range of CI index values in a panel of cancer cell lines obtained following analysis of drug combinations with PF-384 or with mTOR-specific inhibitor. CI indexes were determined using the Chou–Talalay method as described in Materials and Methods and are presented at the ED50 level for each of the combinations. Results are the average of at least three independent experiments. The CI has been interpreted as follows: very strong synergy (01.2), strong synergy (0.7–0.1), moderate synergy (0.7–0.85), slight synergism (0.85–0.9), nearly additive (0.9–11), slight antagonism (11–12), and moderate antagonism (1.2–1.45). Dashed lines are at CI values of 1.1 and 0.7. PTX, paclitaxel; VINO, vinorelbine; Aur101, auristatin-101. B, dynamic monitoring of cell growth in MDA-468 cells with the xCELLigence System. B, concentration- and time-dependent cytotoxic effects of ST4-ADC, PF-384, or combination in MDA-468 cells. One representative experiment of the three is shown. B, Right, impedance index values are presented for the 8-day time point and show statistically significant enhancement of cell growth inhibition for the ST4-ADC plus PF-384 combination. (***, P < 0.001; Student t test). The means and SEM are shown. C, MDA-468 cells grown as 3D spheroids Matrigel were treated with increasing doses of ST4-ADC, a fixed dose of PF-384 (10 nmol/L), or a combination of both the drugs for 7 days. Spheroid viability was measured as described in Materials and Methods. Plotted is the percentage of growth relative to the respective untreated control. Values are means ± SEM. Dashed line indicates % viability for the fixed dose of PF-384. B, Right, histogram plots of the selected data points at the indicated drug concentrations. Asterisks represent significant differences from ST4-ADC+PF-384 (**, P < 0.01; Student t test). D, enhanced induction of caspase-3/7 in MDA-468 cells treated with a combination of ST4-ADC and PF-384 (10 µM) plus PF-384 (1 µM) for 24 hours. Fold induction in caspase-3/7 activity was determined as described in Materials and Methods. Means and SEMs of triplicate experiments are shown. *P < 0.05; **P < 0.01; ***P < 0.001. Asterisks show statistically significant differences between each of the single drugs alone and a combination; Student t test. E, H-1975 cells were treated with indicated concentrations of ST4-ADC and PF-384, or concurrently with the combination of both for 24 hours. Cell lysates were subjected to immunoblotting analysis with antibodies to P-AKT, AKT, P-GSK3, GSK, P-H3, cPARP, or with antibodies to GAPDH as a loading control. E (right) densitometric analyses of E was performed using ImageJ, as described in Supplementary Materials and Methods. Densitometric analyses of data shows protein levels of P-AKT (S473, T308), P-GSK3b (S9).
additive effects detected for 5T4-ADC was also observed for the cell permeable version of parental payload MMAF, MMAF-OMe, or for a recently described auristatin analogue PF-06380101 (Aur101; ref. 5). Structurally unrelated microtubule modulators with diverse mechanisms of action (both stabilizer paclitaxel and destabilizer vinorelbine) also showed additive or synergistic relationships when combined with PF-384 in a smaller subset of cell lines, suggesting a shared mechanism of drug interaction that is directly related to the inhibition of microtubules (Fig. 1A).

We also performed real-time monitoring of cell growth using the xCelligence system. Over the course of approximately 11 days, we observed that exposure of MDA-468 cells to ST4-ADC/PF-384 and MMAF-OMe/PF-384 combinations reduced proliferation more than either of the individual agents alone (Fig. 1B; Supplementary Fig. S2A). Furthermore, when tested in 3D spheroid growth assays, significant enhancement of cytotoxicity was detected for the ST4-ADC/PF-384 and MMAF-OMe/PF-384, compared with the single-agent treatments (Fig. 1C; Supplementary Fig. S2B). Similar observations were made for the lung cancer H-1975 spheroid model (data not shown). Thus, these results confirmed our findings above and provided additional insights on the time dependence of synergistic effects in vitro.

Effect of ST4-ADC/PF-384 on apoptosis, cell-cycle, and PI3K/mTOR markers

To evaluate if synergistic growth inhibition induced by the ST4-ADC and PF-384 combinations is due to apoptosis, we determined proapoptotic signal caspase-3/7 activation in H-1975 or MDA-468 cells. ST4-ADC or PF-384 alone led to modest activation of caspase-3/7 in both the cell lines, measured 24 hours after incubation (Fig. 1D). However, the combination treatment showed markedly enhanced induction of caspase-3/7. Similar results were obtained when the same cells were treated with MMAF-OMe/PF-384, suggesting that the induction of apoptosis in ST4-ADC/PF-384 combination is mechanistically linked to the action of parental payload MMAF-OMe (Supplementary Fig. S2C). Immunoblot analysis for cleaved PARP further demonstrated the induction of apoptosis by the combination of ST4-ADC/PF-384 in H-1975 (Fig. 1E). Collectively, these findings demonstrate that anti-proliferative effects observed with combinations could be accounted for, at least in part, by the enhanced apoptotic response mediated by caspase-3/7.

As auristatin-containing drugs have been described to impair cell-cycle progression, we first asked if PF-384 can modify mitotic arrest when combined with ST4-ADC in MDA-468 cells. As expected, ST4-ADC alone markedly induced a significant accumulation of cells in mitosis as evidenced by the levels of phosphorylated histone H3 (P-H3; Supplementary Fig. S3A). Interestingly, the addition of PF-384 substantially reduced ADC-mediated mitotic arrest. The same observations were made in H-1975 cells when PF-384 was combined with ST4-ADC or free auristatins (Supplementary Fig. S3B and data not shown). Analysis of the cell cycle in H-1975 cells showed that ST4-ADC/PF-384 consistently reduced ADC-mediated G2–M arrest, with a modest increase in G1 stage compared with ST4-ADC alone (Supplementary Fig. S3C). These results raised the possibility that combined treatment with ST4-ADC/PF-384 may suppress the spindle assembly checkpoint function, leading to inappropriate transition out of mitosis. Aurora B inhibition has been shown to selectively relax the spindle checkpoint invoked by microtubule inhibitors (26, 27). We observed an increase of Aurora A, B, and C phosphorylation by the ST4-ADC, an effect blocked by the addition of 50 nmol/L PF-384 (Supplementary Fig. S3D). Consequently, these results support the hypothesis that suboptimal Aurora activity may be responsible for accelerated mitotic exit and apoptosis in cells exposed to the combination of both the drugs.

To examine the changes in signaling after treatment with single-agent PF-384 or ST4-ADC, we initially used phosphokinase antibody arrays. In the H-1975 lung tumor model treated with PF-384, we detected expected declines in P-AKT, P-S6k, P-GSK-3β, P-NOS, and P-PRAS40 markers 6 or 16 hours following drug treatment (Supplementary Fig. S4A). Surprisingly, treatment with ST4-ADC has resulted in the inhibition of P-AKT and P-GSK-3β 16 hours after drug exposure (Supplementary Fig. S4B). To confirm these findings, we analyzed by immunoblot the downstream effectors of PI3K/mTOR pathways as well as markers for mitotic arrest and apoptosis, H-1975 (lower ST4-expressor, L558R/T790M EGFR mutations) and MDA-468 (high ST4-expressor, PTEN−/−) are cancer cell lines that are equally sensitive in vitro to the dual inhibitor of PI3K/mTOR or to auristatin derivative MMAF-OMe. We treated H-1975 or MDA-468 cells with the dual PI3K/mTOR inhibitor PF-384, ST4-ADC, or a combination of the two. In agreement with the previous report (16), PF-384 alone effectively reduced phosphorylation of downstream markers reflecting activation status of the PI3K/mTOR pathway: P-AKT S308, P-AKT S473, and P-GSK-3β S21/9 24 hours after drug exposure (Fig. 1E; Supplementary Fig. S4C). In H-1975 cells, treatment with 50 μg/mL ST4-ADC led to modest reduction in P-AKT S308, P-AKT S473, and GSK-3β S9, as quantified by densitometry. In the same 24-hour experiment, ST4-ADC/PF-384 treatment showed stronger decline in phosphorylation levels of PI3K/mTOR downstream effectors than each of the single agents, as confirmed through densitometric analysis (Fig. 1E). Increasing concentration of ST4-ADC alone induced phosphorylation of H3, but the combination appeared nearly equivalent or even somewhat lower than ST4-ADC alone, which is in good agreement with the mitotic index values determined by flow cytometry. Furthermore, we found that ST4-ADC/PF-384 caused a greater induction of cleaved PARP than either ST4-ADC or PF-384 alone. The potentiation of PF-384’s effect on downstream markers by ST4-ADC appears to be cell line specific because experiments on MDA-468 showed no further reduction of P-AKT and P-GSK by the combination treatment as compared with PF-384 alone (Supplementary Fig. S4C). In MDA-468, we detected no suppression of PI3K/mTOR pathway markers with a single-agent ST4-ADC. These findings support the conclusions that enhanced cytotoxicity observed in ST4-ADC/PF-384 combination is linked, at least in part, to the induction of caspase-3/7 and PARP-dependent apoptosis and correlates with the stronger suppression of PI3K/mTOR pathway biomarkers in H-1975 cells.

Auristatin agents cooperate with PF-384 to regulate translation

We hypothesized that previously uncharacterized effects of auristatins on additional intracellular targets may underlie the observed synergy with PF-384. Total proteomics and phosphoproteomics were independently applied to evaluate changes
in protein abundance and phosphorylation in response to auristatin-based agents in MDA-468, a cell line that had no demonstrable changes in phosphorylation of AKT or GSK upon administration of 5T4-ADC.

Analysis of protein interaction networks for all significantly changed proteins in total proteome perturbed with 0.5 nmol/L auristatin showed several clusters of related functional classes of proteins, with a highly distinct group containing components of mRNA translation and mRNA biogenesis (Fig. 2A; Supplementary Fig. S5A). Specifically, we detected remarkable overrepresentation of mRNA translation factors in the downregulated protein set and of ribosomal proteins in the upregulated group after a 24-hour cell exposure to either of the two doses of MMAF-Ome (0.5 and 5 nmol/L; Supplementary Fig. S5B and S5C and Supplementary Table S2A). Label-free phosphoproteomics of cells treated with the MMAF-Ome or 5T4-ADC for 6 hours demonstrated unanticipated enrichment of “translation factors,” “mRNA processing,” and “mRNA splicing” for both MMAF-Ome and 5T4-ADC in the list of downregulated phosphopeptides (Supplementary Fig. S5D and S5E and Supplementary Table S2B). Consistent with the common mechanism of action between the ADC and free unconjugated payload, there was a substantial overlap in differentially expressed phosphopeptides between MMAF-Ome and 5T4-ADC, with the overrepresentation of translation or mRNA processing-related components in the shared datasets (Supplementary Fig. S5D). Thus, our results raise the intriguing possibility that protein synthesis is one of the convergence points for cellular action of MMAF-Ome or 5T4-ADC. This notion, together with the well-known role for the PI3K/mTOR signaling

Figure 2.

Cooperative suppression of protein translation by ADC/PF-384 combination. A, STRING network analysis of all significantly changed proteins in a total proteomics experiment for MDA-48 cells treated with 0.5 nmol/L MMAF-Ome. Major cluster of interacting proteins includes the translation factors, ribosomal proteins, and ribonucleoproteins that form a densely connected module, which is denoted by a red rectangle. Only connected nodes are shown for simplicity. STRING network analysis and visualization was performed using the online STRING database 9.1 (45). B, total protein abundance and phosphorylation status of selected proteins involved in mRNA translation. MDA-468 (left) or H-1975 (right) cells were treated with the 5T4-ADC, PF-384 or combinations of both drugs for 24 hours at the indicated concentrations. Total protein abundance or phosphorylation levels were measured with antibodies indicated and as described in Materials and Methods. C, effect of auristatin-based agents alone or in combination with PF-384 on cap-dependent synthesis of luciferase reporter. MDA-468 cells stably transduced with monocistronic luciferase reporter were used to assay inhibition of cap-dependent translation as described in Materials and Methods. Cells were treated with individual drugs or with combinations for 24 hours. The concentrations of drugs were: PF-384 50 nmol/L, 5T4-ADC 10 mg/mL, PTX (paclitaxel) 10 mg/mL, CHX (cycloheximide) 30 mg/mL, 5T4-ADC/PF-384: 5T4-ADC 10 mg/mL and PF-384 50 nmol/L. Results are mean ± SE of biologic triplicates from a single experiment representative of the two. 5T4-ADC and PF-384 were statistically significantly different compared with vehicle-treated control by the two-tailed Student t test. 5T4-ADC/PF-384 combination was significantly different compared with each of the single drug controls by the two-tailed Student t test.
To substantiate the above findings, we focused on the effects of 5T4-ADC/PF-384 combination on steady-state level and phosphorylation status of key translation factors involved in the PI3K/mTOR pathway. Single-agent 5T4-ADC showed modest cell type-specific effects, each modulating expression and/or phosphorylation of a select subset of proteins to a different extent (Fig 2B). In both the cell lines, treatment with 5T4-ADC decreased levels of eIF4G1, eIF4G2, eIF4B, and eIF3A, but upregulated P-eIF2α and P-eEF2. More importantly, combination treatments caused cooperative changes in a distinct set of translational regulators. For example, in MDA-468, a cell model that was used for proteome-wide analysis, 5T4-ADC/PF-384 combination led to a stronger decline in eIF4G2, eIF4B, eIF2α, and eIF3A levels, which coincided with greater induction of P-eIF2α and P-eEF2 than for each agent alone (Fig 2B). In the H-1975 model, the same drug combination caused decreases in the expression levels of P-eIF4G1, P-eIF4B, eIF4B, P-EF-BP1, and 4E-BP1 with a concomitant increase in P-eIF2α and P-eEF2. To test whether these observations also translate into functional impairment of general protein synthesis, we monitored the activity of firefly luciferase in MDA-468 cells stably transduced with a cap-dependent monocistronic reporter. Addition of PF-384 or 5T4-ADC significantly suppressed production of luciferase by approximately 40% to 50% after 16- or 24-hour incubation with drugs, whereas treatment with a positive control, protein synthesis inhibitor cycloheximide, fully reduced luciferase production in this system. Treatment with 5T4-ADC/PF-384 led to a stronger decline in luciferase activity when compared with either single agent alone (Fig 2C). No reduction of luciferase transcription was observed in this experiment in response to drug exposure as measured by RT-PCR (data not shown). Overall, our results suggest that combining auristatin-based drugs with inhibitors of the PI3K/mTOR pathway can lead to a specific reprogramming of translational factor repertoire at the level of expression and/or phosphorylation, which causes suppression of protein synthesis.

**In vivo combination therapy with 5T4-ADC and PF-384**

To investigate whether cooperative action of 5T4-ADC/PF-384 in vitro could be observed in the *in vivo* setting, we tested the efficacy of the respective single agents and of the combination in two previously characterized tumor xenograft models with broad range of ST4 expression levels as shown by flow cytometry and IHC staining (Supplementary Fig. S1B; ref. 7). Treatment of animals bearing MDA-468 breast cancer xenografts with 2 mg/kg 5T4-ADC caused initial robust tumor suppression followed by stasis, whereas PF-384 at 7.5 mg/kg showed a very minor inhibition of tumor growth over the vehicle-treated arm (Fig 3A). In contrast, concurrent administration of both the drugs led to a more complete tumor regressions clearly observed in all tumors treated by the end of the study. Subsequent TTE analysis of time to no tumor (tumor regression) showed that a much shorter time was needed to achieve complete tumor regressions in the combination arm versus 5T4-ADC alone (*P* < 0.0001 by the log-rank test), with all animals in the 5T4-ADC/PF-384 group becoming tumor-free by the day 36 (Fig 3B). In the H-1975 lung cancer model, treatment with 3 mg/kg 5T4-ADC resulted in tumor stasis followed by regrowth of tumors, whereas 7.5 mg/kg PF-384 elicited only nominal antitumor activity (Fig. 3C). The 5T4-ADC/PF-384 combination resulted in a more complete, but still unsustainable suppression of tumor growth. The percentage of animals with less than three-fold increase in tumor volume was used as survival endpoint for the analysis of H-1975 model. TTE analysis indicated a statistically significant delay in tumor tripling rate for the combination group compared with 5T4-ADC (3 mg/kg; *P* = 0.0356, log-rank test) or PF-384 alone (7.5 mg/kg, *P* < 0.0001, log-rank test, Fig. 3D).

**Combination of 5T4-ADC with taxanes**

The initial drug interaction screen also identified paclitaxel as an agent that potentiated MMAF-OMe- or 5T4-ADC growth inhibitory effects. The combination results varied from synergism to additivit in most of the cell models tested as measured by the CI index (Fig. 4A). This effect was not unique to paclitaxel, as docetaxel, a structurally similar taxane, also showed favorable interactions with MMAF-OMe or 5T4-ADC. Moreover, when MMAF-OMe or 5T4-ADC was substituted for another auristatin analogue Aur101, or an unrelated microtubule-depolymerizing agent vinorelbine, we also observed potentiation of their cytotoxicity by paclitaxel. According to a high-resolution structural data obtained for dolastatin-10 (30) or for the new auristatin analogue bound to tubulin (5), auristatins bind at a site adjacent to the vinca binding site at the interface of two tubulin molecules and in close proximity to the β-tubulin nucleotide exchange (Fig. 4A, Right). We examined additional cellular changes after cotreatment with paclitaxel. Modest enhancement of cytotoxicity in a 3D spheroid assay and stronger induction of caspase-3/7 was observed in the MDA-468 cells treated with 5T4-ADC plus paclitaxel for 48 hours compared with the single drug controls (Fig. 4B and C). A potential explanation for the cooperative action between 5T4-ADC and paclitaxel includes modulation of cell-cycle progression and altered microtubule dynamics. As expected, an M-phase-specific marker P-H3 was markedly induced in the MDA-468 cells treated with paclitaxel or 5T4-ADC. The cotreatment with both drugs slightly enhanced the mean mitotic increase relative to single agents, but this trend did not reach statistical significance (Supplementary Fig. S6A). Furthermore, we observed enhanced PARP cleavage and increase in levels of P-H3 in MDA-468 or H-1975 cells treated with 5T4-ADC/paclitaxel combination versus either single drug (Fig. 4D). Interestingly, fluorescent microscopy with an anti-tubulin antibody demonstrated a significant collapse of microtubule network around the nucleus and formation of the microtubule aggregates with 5T4-ADC/paclitaxel combination in MDA-468 cells (Supplementary Fig. S6B). Unlike the combination's effect, the 5T4-ADC alone treated cells showed disintegration of microtubule bundles with more intense staining at cell periphery and lesser cytoplasmic volume. Collectively, these findings demonstrate that antiproliferative effects observed with the 5T4-ADC/paclitaxel could be accounted for, at least in part, by the enhanced apoptotic response mediated by caspase-3/7 and PARP with parallel induction of P-H3 and more pronounced loss of microtubule integrity.
Given the favorable interactions observed between 5T4-ADC and taxanes in vitro, we also evaluated the potential antitumor activity of this combination in vivo. MDA-468 xenografts were tested with vehicle, 5T4-ADC (i.v. 2 mg/kg, q4d), PF-384 (i.v. 7.5 mg/kg, q4d), or a combination. A, tumor growth curves. Tumor volume was determined at the indicated times after the onset of treatment. Points, mean of values from 10 mice/group; bars, SE. Time to endpoint (TTE) plots for the treatment groups in A show change in percentage of animals with tumors over the time. Endpoint is defined as the time elapsed for animal to become tumor-free. TTE analysis demonstrates significantly enhanced rate of tumor regressions with combination of 5T4-ADC plus PF-384 compared with the single-agent activity of 5T4-ADC ($P < 0.0001$, log-rank Mantel-Cox test). PF-384 did not elicit regressions in this experiment. C and D, mice bearing subcutaneous H-1975 human lung tumor xenografts were treated with 5T4-ADC (i.v. 3 mg/kg, q4d), PF-384 (i.v. 7.5 mg/kg, q4d) or a combination. C, tumor growth curves. Tumor volume was determined at the indicated times after the onset of treatment. Points, mean of values from 10 mice/group; bars, SE. TTE plots for the treatment groups in C show the percentage of animals with less than three-fold increase in tumor volume over time. Endpoint is defined as the time at which tumor volume has tripled. TTE analysis of data demonstrates significant delay at rate of tumor tripling for the 5T4-ADC plus PF-384 combination compared with the single-agent activity of 5T4-ADC ($P = 0.0356$, log-rank test) or PF-384 ($P < 0.0001$, log-rank test) alone. To minimize fluctuations in the tumor growth curve plots and facilitate interpretation of the data, the mean tumor volume for each group was plotted until $>10\%$ of the mice in the group were sacrificed.

Combination of 5T4-ADC with PI3K/mTOR Inhibitors or Taxanes
Figure 4.
Effects of auristatin-based agents combined with taxanes. A, summary of CI values in a panel of cancer cell lines. Dot plot showing the range of CI index values obtained following analysis of drug combinations with microtubule inhibitors. CI indexes were determined using the Chou-Talalay method as described in Materials and Methods and are presented at the ED50 level for each of the combinations. Results are the average of at least three independent experiments. The CI has been interpreted as in Fig. 1A. Dashed lines are at CI values of 1.1 and 0.7. B, combination of 5T4-ADC or MMAF-OMe with paclitaxel (PTX) leads to stronger suppression of cell growth in 3D culture. MDA-468 cells were treated with increasing doses of 5T4-ADC, fixed dose of paclitaxel (1 nmol/L), or a combination of both drugs for 7 days. Spheroid viability was measured as described in Materials and Methods. Plotted is the percentage of growth relative to the respective untreated control. Dashed line indicates% viability for the fixed dose of MMAF-OMe. Right, histogram plots of the selected data points at the indicated drug concentrations. C, induction of caspase-3/7 activity by the combination of 5T4-ADC with paclitaxel (PTX) leads to stronger suppression of cell growth in 3D culture. MDA-468 cells were treated with increasing doses of 5T4-ADC, fixed dose of paclitaxel (1 nmol/L), or a combination of both drugs for 7 days. Spheroid viability was measured as described in Materials and Methods. Plotted is the percentage of growth relative to the respective untreated control. Dashed line indicates% viability for the fixed dose of MMAF-OMe. Right, histogram plots of the selected data points at the indicated drug concentrations. C, induction of caspase-3/7 activity by the combination of 5T4-ADC with paclitaxel (PTX) leads to stronger suppression of cell growth in 3D culture. MDA-468 cells were treated with increasing doses of 5T4-ADC, fixed dose of paclitaxel (1 nmol/L), or a combination of both drugs for 7 days. Spheroid viability was measured as described in Materials and Methods. Plotted is the percentage of growth relative to the respective untreated control. Dashed line indicates% viability for the fixed dose of MMAF-OMe. Right, histogram plots of the selected data points at the indicated drug concentrations.

Discussion
Optimizing the efficacy of ADCs by systematic nonclinical assessment of combinations remains an important objective for ADC development. We hypothesized that major synergistic effects for the antibody–auristatin conjugates may be mediated by the pharmacologic action of payload itself and therefore searched for the common chemotherapeutic agents or signaling inhibitors that could potentiate either free or conjugated auristatins, such as 5T4-ADC. Here, we describe novel and previously uncharacterized potentiation of auristatin-based agents by PF-384 or taxanes in vitro that translates to enhanced antitumor efficacy in tumor xenograft models. The combination of MMAF-OMe or 5T4-ADC and PF-384 has resulted in consistently synergistic drug effect in tumor cell lines of lung, breast, and ovarian cancer origin, the three putative tumor types that show broad 5T4 expression (7). Analysis of the in vitro cytotoxicity data for the cell line panel used in this study revealed that neither common mutations in these cell lines nor their tissue lineage or sensitivity of the individual drugs alone could be used to predict synergistic responses to the combinations involving auristatins and other drugs. The favorable pharmacologic interactions were also observed when different microtubule-targeting agents paclitaxel (polymerizing) or
Vinorelbine (depolymerizing, unpublished observation) were combined with PF-384, suggesting that microtubule damage is a global signal that can be potentiated by the suppression of PI3K/mTOR signaling. Surprisingly, we also found reproducible but cell- and drug-dependent inhibition of AKT and/or GSK3 phosphorylation in response to single-agent 5T4-ADC.

Figure 5.
The 5T4-ADC and paclitaxel (PTX) combination treatment leads to enhanced therapeutic effects in breast and lung cancer models in vivo. A and C, mice bearing subcutaneous MDA-468 human breast tumor xenografts were treated with 5T4-ADC (i.v. 2 mg/kg, q4d), paclitaxel (p.o. 10 mg/kg, q4d), or a combination. A, tumor growth curves. Tumor volume was determined at the indicated times after the onset of treatment. Points, mean of values from 10 mice/group; bars, SE. B, time to endpoint (TTE) plots for the treatment groups in A show change in the percentage of animals with tumors over the time. Endpoint is defined as the time elapsed for animal to become tumor-free. TTE analysis of data demonstrates significantly faster rate of complete tumor regressions achieved with the combination of 5T4-ADC and paclitaxel compared with the single-agent activity of 5T4-ADC (P = 0.0071, log-rank test) or paclitaxel (P = 0.01, log-rank test). C, similar to A, but paclitaxel was used at a dose of 22.5 mg/kg (p.o. q4d). D, TTE analysis of data from C shows significantly faster rate of complete tumor regressions achieved with the combination of 5T4-ADC plus paclitaxel compared with the single-agent activity of 5T4-ADC (P = 0.0082, log-rank test) or paclitaxel, which has not produced any tumor regressions at this dose. E, mice bearing subcutaneous H-1975 tumors were treated with 5T4-ADC (i.v. 3 mg/kg, q4d), paclitaxel (p.o. 10 mg/kg, q4d), or a combination. 5T4-ADC combined with paclitaxel is more efficacious than treatment with single agents. F, TTE analysis of data performed similarly to Fig. 3B, demonstrates significant delay of rate of tumor tripling for the 5T4-ADC plus paclitaxel combination compared with the single-agent activity of 5T4-ADC (P < 0.0001, log-rank test) or paclitaxel (P = 0.0001, log-rank test) alone.
This finding is unexpected but in agreement with the report by Asnaghi and colleagues who demonstrated inhibitory effects of nocodazole on phosphorylation of mTOR at Ser 2448 (31). We, however, could not consistently detect synergistic suppression of PI3K/mTOR pathway markers in MDA-468 cells. Unlike H-17975, an MDA-468, a PTEN-/- breast cancer model is more sensitive to each single agent alone, which may make the detection of cooperative effects on downstream pathways technically difficult.

We also applied a system-wide approach to explore the additional cellular action of auristatins that could help explain molecular mechanisms behind the synergistic interactions with the PF-384. Both total proteomics with MMAF-OMe and phosphoproteomics performed with 5T4-ADC and MMAF-OMe uncovered mRNA translation as one of the predominantly affected processes. These findings were further corroborated by Western blot analysis of selected translation initiation or elongation components. More proteins were modulated by MMAF-OMe, 5T4-ADC, or PF-384 alone, with only some showing cooperative effects by combined treatment with 5T4-ADC/PF-384 or MMAF-OMe/PF-384. This comports with other large-scale phosphoproteomic studies that identified a number of initiation factors differentially phosphorylated in response to the nocodazole treatment (32, 33). Whereas targeting of translational components by auristatin-based agents is a novel finding, the effects of PF-384 are quite expected, given the known role of PI3K and mTOR kinases in the control of protein synthesis. One anticipated consequence of ADC-mediated modulation of translation machinery is a decline in global translation rates. Stronger reduction of cap-dependent translation of luciferase reporter by the 5T4-ADC/PF-384 combination relative to single-agent drugs is consistent with the upregulation of P-eIF2α (S51) and P-eEF2 (S56), changes that are suggestive of a slowdown in translation. Thus, cooperative suppression of protein synthesis can, at least in part, be linked to the observed cellular synergy with PF-384. Notably, structurally unrelated and highly specific mTOR inhibitors WYE-132, which is known to disrupt the cap-dependent mRNA translation and inhibit global protein synthesis, also showed synergy when combined with auristatin derivative in vitro (Supplementary Table S3). This implies that reducing cap-dependent translation by targeting mTOR kinase activity alone may be sufficient to enhance therapeutic effects of the auristatin-based agents.

Given the highly specific effects of MMAF-OMe and 5T4-ADC on microtubules, how these agents perturb mRNA translation and how PF-384 can potentiate this mechanism? One potential explanation is based on the substantial evidence of interactions between tubulin cytoskeletal components and ribosomal proteins, translation initiation factors and various mRNAs. In addition, many mRNAs encoding mitotic regulators or translational components are known to localize to mitotic spindles (34–40). This and the evidence that ongoing translation is maintained throughout the cell cycle, without substantial decline during mitosis (35) is consistent with the notion that tubulin-localized protein synthesis maybe especially important for the efficient progression through mitosis. Hence, we propose that the entire class of auristatin-containing agents, including ADCs, can disrupt the tubulin-bound pools of translational components, thereby modulating their abundance. We speculate that the combined action of ADC and PF-384 disrupts protein synthesis during mitotic transition, or collectively impacts translation of the key mRNAs required for the survival during mitotically arrested state. Furthermore, it is also possible that both the drug classes could affect mRNA translation in different stages of cell cycle. Interestingly, cotreatment with PF-384 impairs Aurora kinase phosphorylation in our experiments, a phenotype that is generally consistent with compromised spindle checkpoint and decreased mitotic index in these cells. These data are also in agreement with what has been previously described for combination of docetaxel and another PI3K inhibitor GDC-0941, where decreased time of mitotic arrest was mechanistically linked to the induction of apoptosis in synchronized cells (41). Our results provide one potential scenario to explain the synergistic activity and it is likely that multiple mechanisms may determine favorable pharmacologic outcomes in tumor cells.

Remarkably, the combination therapy with 5T4-ADC/PF-384 in vivo significantly improved antitumor activity and reduced tumor volumes in models of breast and lung cancer as compared with the effects of single drug treatment alone. Other important preclinical work demonstrated that T-DM1 plus the pan-PI3K inhibitor GDC-0941 or plus dual PI3K/mTOR combinations resulted in the enhanced antitumor activity both in vitro and in trastuzumab-resistant or in PIK3CA mutant breast cancer xenograft models in vivo (13). The T-DM1 plus GDC-0941 was tested in a 3 × 3 design dose-escalation phase I study in patients with advanced HER2-positive metastatic breast cancer. Results have been reported only on 13 patients, with dose-limiting toxicities of a grade 4 thrombocytopenia and grade 3 fatigue observed in two initial cohorts. The combination regimen was better tolerated in a third cohort that enrolled at a reduced dose of T-DM1 (3.0 mg/kg) and GDC-0941 (100 mg; ref. 42). It remains to be seen if combining T-DM1 with PI3K inhibitors generates meaningful clinical activity in these patients.

Several lines of evidence illustrate an important therapeutic potential for another promising combination between the 5T4-ADC and a taxane, paclitaxel. Enhanced antitumor activity of 5T4-ADC plus paclitaxel is intriguing but not counterintuitive. Whereas both agents act on the microtubules, MMAF-OMe binds to a distinct site than the taxane binding site, in a manner similar to the vinca alkaloids and thus may affect additional tubulin-dependent functions in a paclitaxel-independent manner. This may lead to therapeutic synergy when combined with paclitaxel. There is considerable interest in the ADC field in testing the clinical activity and safety of the conjugates with other cytotoxic antiangiogenic agents, such as taxanes. This hypothesis is currently under investigation in clinical trials evaluating combination therapy of T-DM1 plus docetaxel in early-stage HER2 breast cancer (43) and in a phase III study comparing brentuximab vedotin plus AVD versus ABVD (doxorubicin, bleomycin, vinblastine, and dacarbazine) alone (44).

In conclusion, our report provides strong preclinical framework and the rationale for combination therapy of 5T4-ADC with taxanes or 5T4-ADC with PF-384 in clinical trials for the treatment of lung, breast, or ovarian cancer. The dual-targeting approach presented here, with both an auristatin-based agent and PI3K/mTOR pathway inhibitor or taxanes, could serve as an important model for enhancing antitumor activity of other auristatin-based ADCs and overcoming potential drug resistance in the clinic.
Disclosure of Potential Conflicts of Interest

P. Sapra and B. Shor have ownership interest in Pfizer and are listed as co-inventors on a provisional patent, which is owned by Pfizer, directed to subject matter including auristatin combinations. No potential conflicts of interest were disclosed by the other authors.

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References


Enhanced Antitumor Activity of an Anti-5T4 Antibody–Drug Conjugate in Combination with PI3K/mTOR inhibitors or Taxanes

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