Phase I/II Study of G17-DT, an Anti-Gastrin Immunogen, in Advanced Colorectal Cancer

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ABSTRACT

Gastrin is a growth factor for colorectal cancer, and therefore, anti-gastrin hormone therapy has a potential role in treatment of this disease. The gastrin immunogen gastrin-17-diphtheria toxoid (G17-DT; Gastrimmune) produces anti-G17 antibodies that have been shown to be effective in the treatment of colorectal carcinoma in preclinical models. Fifty patients with advanced colorectal cancer were treated with G17-DT in a multicenter, sequential group, open label Phase I/II study. Primary injections with two booster doses were given by i.m. injection. The main aim of the study was to assess the safety and efficacy of the production of anti-gastrin antibodies. Locally developed and standard WHO toxicity measurements with RIA and Scatchard analysis for antibody assessment were used. One center measured tumor response radiologically. Eighty % of patients produced a measurable antibody response. Antibodies of high affinity (median $K_d$, 0.295 nM; interquartile range, 0.16–0.41 nM) were detected between 4 and 12 weeks after primary injection. The antigen binding capacity was high at 2.8 × 10$^{-10}$ to 7.25 × 10$^{-9}$ M. The treatment was well tolerated with no systemic side effects seen. Myalgia at the injection site was seen in 46% of patients with severe pain caused by the formation of a sterile abscess seen in 14% of patients. The abscesses were all drained under ultrasound guidance, and the patients recovered fully within 6 weeks. No radiological responses were seen, but two patients had stable disease. G17-DT immunization produces anti-G17 antibodies in patients with advanced colorectal cancer. The antibodies were of an affinity high enough to compete with the cholecystokinin B/gastrin receptor for G17 binding with adequate capacity to neutralize postprandial gastrin surges. Additional dose-ranging studies have been performed in patients with gastric cancer using 100- and 200-µg doses of G17-DT formulated without adjuvant and the emulsifier aluminum monostearate. In addition, the effect of immunizing at different time intervals has been determined.

INTRODUCTION

Colorectal cancer is the second most common cause of cancer deaths in the United Kingdom, accounting for ~16,000 deaths/year. Despite advances in surgery, chemotherapy, and radiotherapy, only modest improvements in overall mortality have been achieved in the past 30 years. Cancer vaccines offer a new potential treatment option.

Gastrin is a peptide hormone that is important in the regulation of acid secretion and the growth of both normal and malignant gastrointestinal epithelium (1), and it exists in two amidated forms composed of 17 and 34 amino acids, denoted G17$^3$ and G34, respectively. Colorectal tumor cell lines and human cancers may express a number of isoforms of the gastrin/CCKB receptors (2–8) and produce their own gastrin (9–13). Therefore, the tumor cells can potentially respond to circulating endocrine gastrin (14–17) and the locally produced gastrin, which acts in an autocrine or paracrine manner (18, 19). The tumor-associated gastrin is composed mainly of precursor forms, particularly glycine-extended G17, which have also been demonstrated to have a proliferative effect (20, 21).

Elevation of the circulating gastrin level has been shown in animal models and humans to enhance the proliferation of the normal colon mucosa (20–23). As a result, there is an increase in incidence of colorectal cancer in hypergastrinemic patients (24). In addition, studies in the APC$^{Min}$ model of polyposis coli have shown hypergastrinemia induced by omeprazole administration and after infection with Helicobacter pylori promotes the proliferation and progression of colorectal adenomas (25). Effective inhibition of gastrin may, therefore, be beneficial as a therapy in colorectal cancer, potentially in prevention, beneficial as an adjuvant, or even beneficial in advanced disease.

Gastrin/CCKB receptor antagonists have been assessed in patients with gastrointestinal tumors. The antagonists appeared to be ineffectual (26, 27), and it was initially assumed to be because of their lack of potency, with relatively high concentrations required to displace amidated G17 (28, 29). The results of studies assessing tumor receptor profiles indicate that the lack of efficacy is probably also a result of the fact that several receptor subtypes mediate the action of gastrin (2, 8).

A potential method to circumvent the problem created by the presence of numerous gastrin/CCKB receptor isoforms is to...
neutralize gastrin before it interacts with its receptor. Gastrimmune, an anti-gastrin immunogen, produces in situ neutralizing antibodies directed against the forms of gastrin that have a proliferative action before they interact with the receptor.

Gastrimmune is an immunoconjugate consisting of the nine NH₂-terminal amino acids of human G17 linked via a peptide spacer to DT. DT acts as the immunogenic carrier, and the G17 sequence acts as a B-cell epitope. The peptide spacer allows the gastrin moiety to be orientated spatially in such a way that B cells recognize the whole sequence and thus raise high affinity, neutralizing antibodies (30, 31).

Because the immunogen is composed of the NH₂-terminal amino acids, the raised antibodies neutralize both amidated G17 and glycine-extended G17. In an animal model, after priming injections, antibody titers were raised that were higher than those required to neutralize all serum G17, and thus excess unbound antibody was available for neutralization of tumor-associated gastrin molecules (32).

By using active immunization with species-specific Gastrimmune in a pig model, immunoneutralization of G17 was shown to inhibit acid secretion without having any antiproliferative effects on the gastrointestinal mucosa, indicating that normal trophic effects of other gastrin species, such as G34, are preserved (33). Antibodies raised by Gastrimmune have been evaluated in a number of colorectal tumor models and have been shown to exert strong therapeutic effects in primary tumors (34) and in both hepatic and lung metastasis models (35, 36).

After it was demonstrated that Gastrimmune has an antitumor effect in animal models, a Phase I/II study was performed in patients with advanced colorectal cancer. The primary aim of this study was to assess the safety and efficacy, in terms of antibody response, of Gastrimmune in patients with colorectal adenocarcinoma. Secondary aims were to assess the effect of formulation (the dose of Gastrimmune; the addition of an adjuvant, nor-MDP, in the primary injection; and the presence of the emulsifier, AMS) on local tolerance and antibody production. The effect on tumor parameters was also determined.

PATIENTS AND METHODS

Methods. The trial was designed as a multicenter, sequential group, open label study. It was planned that 54 patients with advanced colorectal cancer would be admitted to the study in sequential blocks of six. The first patient entered in June 1995, and the last patient enrolled in October 1997.

Patient Selection. Patients, ages 18 years or older, with histologically verified adenocarcinoma of the colon or rectum that was either locally advanced, inoperable, recurrent, or metastatic and that was not amenable to curative surgery and/or radiotherapy, were candidates for this study. Other eligibility criteria included: (a) life expectancy >3 months; (b) Karnofsky index ≥50%; (c) no other neoplasms apart from colorectal cancer (except treated basal cell carcinoma of the skin or cancer of the uterine cervix, stages 0–1); (d) no acute intercurrent illness; (e) no patients considered to be a poor medical risk because of nonmalignant systemic disease or grossly abnormal laboratory results; (f) no patients who had received any other anticancer therapy within 3 weeks; (g) no patients immunologically compromised, including those on systemic corticosteroid therapy; (h) no women of childbearing age; (i) no positive immediate hypersensitivity reaction to skin testing with Gastrimmune; and (j) adequate hemopoiesis (hemoglobin >10 g/dl; WBC count >4.0 × 10⁹/liter; platelets >100 × 10⁹/liter). Written informed consent was obtained according to institutional guidelines.

Gastrimmune Formulation and Administration. Gastrimmune was supplied by Apton Corporation (Woods-lands, CA). Gastrimmune is a sterile, milky white, semisemous water-in-oil emulsion. The G17 immunogen was dissolved in a PBS aqueous phase that was mixed with a surfactant and oil phase to form a water-in-oil emulsion. Three formulations of Gastrimmune were used to determine whether they would affect local tolerance and antibody production. The basic formulation included an emulsifier, AMS. The second formulation contained an adjuvant in the priming dose, nor-MDP, with the emulsifier AMS. The third formulation was a simple water-in-oil emulsion without adjuvant or emulsifier.

Gastrimmune was administered as an i.m. injection. The primary injection and subsequent boosters were given at different sites. The first immunization was injected into the left thigh, the second immunization was injected into the right thigh, and the third immunization was injected into the left buttock. Each dose of Gastrimmune was drawn up into a syringe immediately before administration to minimize contact with plastic and possible adsorption of the peptide conjugate.

Drug Dosage and Treatment Schedule. Each patient was to receive three injections of Gastrimmune. After a 1-week assessment period, a primary injection was to be given, followed by two additional immunizations spaced 4 weeks apart. The dose level for each group of six patients was to escalate from 100 µg of immunogen for the first group, to 250 µg for the second group, to 10 µg for the third group. The fourth group was to receive 10 µg of immunogen with 3 µg of nor-MDP adjuvant in the first injection only, and the fifth group was to receive 100 µg of immunogen with 3 µg of nor-MDP adjuvant in the first injection only. An unadjuvanted formulation without the emulsifier AMS was then administered in an escalating dose, from 165 µg to the sixth group, 330 µg to the seventh group, 495 µg to the eighth group, and 990 µg to the ninth group (Table 1).

Patient Monitoring. The patients were seen in specially designated clinics. Before the treatment commenced, histories and physical examinations were performed, and the following evaluations were obtained: (a) Karnofsky score; (b) full blood count; (c) urea and electrolytes; (d) liver function tests; (e) 24-h creatinine clearance; (f) CEA; (g) immunoglobulins and autoantibodies; (h) fasting G17 and anti-gastrin antibody; (i) chest radiograph; and (k) CT scan (Nottingham patients only). After each immunization, the patient was given a daily diary to assess local tolerance of the injection and interference with the activities of daily life. The patient was required to record the pain at the injection site and how the discomfort from the injection interfered with the patient’s normal daily activities on an ordinal scale from 0 to 10. The diaries were checked at each visit. The investigator recorded an assessment of the degree of discomfort from the injection site and the degree of functional impairment attributable to the injection site that the patient
Patients’ sera were diluted in Veronal buffer initially at a 1:10 dilution and in a 3-fold dilution series thereafter. A positive control antiserum (Rabbit antihuman Gastrimmune) was also prepared in buffer at the same dilutions.

Sera dilutions were then aliquoted into glass tubes that had been coated previously with 2.5% polyethylene glycol (M_w 10,000; Sigma, Poole Dorset, United Kingdom). Stripped sera were added to the control tubes. This was prepared from pooled human serum that was added to charcoal that had been previously soaked overnight in 100 ml of 0.02 M Veronal buffer. This was mixed and incubated at 4°C for 30 min, after which it was centrifuged at 3000 × g for 10 min. The supernatant was collected and passed through a C18 Sep-pak filter that had been coated previously with 2.5% polyethylene glycol (M_w 8000; Sigma) and 250 µl of 25% polyethylene glycol (M_w 10,000; Sigma) were added to each tube, which were then vortexed. The tubes were then centrifuged at 3000 × g for 30 min at 4°C. The supernatant was aspirated, and the pellet from each tube was counted on a gamma counter (83% counting efficiency). The mean of each triplicate was used, and the products of the largest perpendicular diameters of all measurable disease sites. Progressive disease was indicated by a >25% increase in the size of at least one measurable lesion or the appearance of a new lesion. Stable disease was defined as an increase in disease measurements of ≤25% or a decrease of ≥50%.

Anti-Gastrin Antibody Assay. Serum was obtained by centrifugation at 4°C after the formation of a clot and stored at −70°C until analysis. 125I-labeled G17 (New England Nuclear-DuPont; specific activity, 2200 mCi/mmol) was reconstituted in distilled water to a concentration of 500 Ci/ml. The radiolabeled G17 was diluted in Veronal buffer [0.02 M (pH 8.4)].

### RESULTS

Fifty patients entered into the study. Thirty-three (66%) of the patients were male, and 17 (34%) were female. The mean age at entry was 65.1 years, with a minimum age of 40 and a maximum age of 84. Karnofsky scores ranged from 60 to 100%, with a mean of 84% and a median of 100%. Mean weight was 66.3 kg, with a minimum weight of 41 kg and a maximum weight of 106 kg.

Six patients died during the study from advanced disease. Thirty-four patients received all three immunizations, 15 patients received two injections, and 1 patient received only a
single immunization. Failure to complete a course of immunizations was because of either advancing disease (7 patients) or as a result of a reaction at the local injection site (9 patients).

**Assessment of Safety**

**Systemic Effects.** The immunogen was well tolerated. Only two patients complained of nausea and vomiting that were potentially related to treatment. No other systemic symptoms were noted. Apart from hematological changes that would be expected from advancing colorectal cancer, there were no significant changes in the blood tests throughout the study. There was no effect on renal function, as measured by serial urea and electrolytes and by pretreatment and posttreatment 24-h creatinine clearance. Importantly, there was no change in immunoglobulin levels and no development of autoantibodies (parietal cells, smooth muscle, thyroid, intrinsic factor, reticulin, antinuclear and mitochondrial antibodies, and rheumatoid factor) after administration of G17-DT.

**Local Tolerance.** Twenty-four patients (46%) experienced a mild myalgia at the injection site after at least one of the injections. In most cases, the patient experienced mild discomfort that lasted between 2 and 5 days and did not interfere with any activities of daily life. The mean pain score for all groups was 0.54 (maximum, 10). There was no significant difference in the pain scores among the groups or the formulations (Table 3). The onset of the myalgia was variable and did not occur more frequently after a particular immunization. When patients experienced injection site pain, no abnormality could be found on examination in most cases.

However, a group of seven patients (14%) developed sterile abscesses at an immunization site. Before the detection of the abscess, there was severe pain and limitation of movement of the affected limb in every patient. The diagnosis was made after clinical examination and confirmed by ultrasound. The collection was effectively drained percutaneously and contained pus in all cases. Microscopy of the sample revealed the presence of pus cells; however, no growth was seen on the culture, and a sterile abscess was made. After drainage, the patients made a complete recovery, which took up to 6 weeks.

After the development of abscesses in the group receiving adjuvant, no additional patients were recruited to the adjuvant formulation groups. The abscesses in group 8 (495 µg) did not occur until relatively late in the study, and four patients had been recruited to group 9 (990 µg). No additional patients were entered into the study.

**Assessment of Efficacy**

**Anti-Gastrin Antibody Response.** Using the criteria that 10% of 125I-labeled G17 binding was the threshold for a measurable response, 40 of the 50 patients produced antibodies after immunization with Gastrimmune. The response rate varied among the groups, as seen in Table 4. The lowest response rate was seen in group 1 (10 µg).

All patients achieved a ≥10% 125I-labeled G17 binding between weeks 4 and 12. Differences in the formulation of Gastrimmune did not alter the time to antibody formation.

At week 12, the $K_d$ was 0.295 nM (interquartile range, 0.16–0.41 nM). The median antigen binding capacity of the antibodies was $2.8 \times 10^{-9}$ mol/liter (interquartile range, $5.1 \times 10^{-10}$ to $7.25 \times 10^{-9}$ M). There was no significant difference in the characteristics of the antibodies produced by different dose groups. Several patients have been followed-up after the completion of the study. Antibody titers are maintained, up to a maximum of 2 years.

We have been able to measure the presence of excess anti-G17 antibodies and excess amounts of bound G17. However, measurement of free G17 in the presence of antibodies and gastrin/antibody complexes is technically difficult. Initially, several serum manipulations are required to try to separate the free gastrin fraction, which appears to be small. The numerous

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**Table 3** Number of immunizations received and the pain scores of each group

<table>
<thead>
<tr>
<th>Dose (µg) emulsion</th>
<th>10</th>
<th>100</th>
<th>250</th>
<th>10 + 3</th>
<th>100 + 3</th>
<th>165</th>
<th>330</th>
<th>495</th>
<th>990</th>
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</thead>
<tbody>
<tr>
<td>Volume (ml)</td>
<td>0.1</td>
<td>0.1</td>
<td>0.25</td>
<td>0.1</td>
<td>0.1</td>
<td>0.05</td>
<td>0.1</td>
<td>0.15</td>
<td>0.3</td>
</tr>
<tr>
<td>Group</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
</tr>
<tr>
<td>No. of patients receiving immunization</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>One</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
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<td>0</td>
<td>0</td>
<td>0</td>
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<tr>
<td>Two</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
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<td>Three</td>
<td>6</td>
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<td>6</td>
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<td>6</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>6</td>
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<tr>
<td>Local pain or reaction reported</td>
<td>0</td>
<td>5</td>
<td>2</td>
<td>2</td>
<td>4</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Maximum mean pain score</td>
<td>0.01</td>
<td>0.03</td>
<td>0.61</td>
<td>0.08</td>
<td>0.33</td>
<td>1.07</td>
<td>0.58</td>
<td>2.08</td>
<td>0.8</td>
</tr>
<tr>
<td>Maximum mean interference with daily activities score</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>No. abscesses/fluid collections</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>0</td>
<td>1</td>
<td>4</td>
<td>0</td>
</tr>
</tbody>
</table>

* Pain score: 0, no pain; 10, worst pain. Interference score: 0, no interference; 10, completely inactive or bedbound.

**Table 4** Positive antibody responders by group

<table>
<thead>
<tr>
<th>Group</th>
<th>Gastrimmune (µg)</th>
<th>Adjuvant (µg)</th>
<th>Responder</th>
<th>% responder</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>10</td>
<td></td>
<td>2/6</td>
<td>34</td>
</tr>
<tr>
<td>2</td>
<td>100</td>
<td></td>
<td>5/6</td>
<td>83</td>
</tr>
<tr>
<td>3</td>
<td>250</td>
<td></td>
<td>5/6</td>
<td>83</td>
</tr>
<tr>
<td>4</td>
<td>10</td>
<td>Yes</td>
<td>3/4</td>
<td>75</td>
</tr>
<tr>
<td>5</td>
<td>100</td>
<td></td>
<td>5/6</td>
<td>83</td>
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<tr>
<td>6</td>
<td>165</td>
<td></td>
<td>5/6</td>
<td>83</td>
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<td>7</td>
<td>330</td>
<td></td>
<td>6/6</td>
<td>100</td>
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<tr>
<td>8</td>
<td>495</td>
<td></td>
<td>5/6</td>
<td>83</td>
</tr>
<tr>
<td>9</td>
<td>990</td>
<td></td>
<td>4/4</td>
<td>100</td>
</tr>
</tbody>
</table>
manipulations result in an assay that is not reproducible. Advice was sought from Professor John Walsh, who believes that measurement of the free fraction in vitro is technically difficult at this time. He suggested that an in vivo bioassay would be more effective. This has been performed in a rat gastric fistula model. The induction of anti-gastrin antibodies resulted in significant inhibition of G17-stimulated acid output. This bioassay confirms the effectiveness of the antibodies to bind G17 (25).

**Disease Response.** Although this was a Phase I/II study, response data were collected. Four patients had lung metastases evident on their original screening chest radiograph. These had all progressed by the time of the poststudy examination. Two patients developed lung metastases during the study period.

Metastatic burden was assessed by a CT scan. Sixteen patients had pretreatment CT scans, and 14 patients had the investigation repeated at the end of the study. There were no complete or partial responses, with 12 of 14 patients having progressive disease reflected in elevated CEA levels. Two patients had stable disease, both having had progressive disease before study entry. One patient had a local rectal recurrence; the other patient had liver metastases (data not shown).

Overall, the CEA levels rose during treatment in all study groups. However, in five patients the CEA remained stable, and one patient showed a modest reduction (data not shown).

**REFERENCES**


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