Angiogenesis, Thymidine Phosphorylase, and Resistance of Squamous Cell Head and Neck Cancer to Cytotoxic and Radiation Therapy

Michael I. Koukourakis,2 Alexandra Giatromanolaki, George Fountzilas, Efthimios Sivridis, Kevin C. Gatter, and Adrian L. Harris

Tumour and Angiogenesis Research Group, Department of Radiotherapy and Oncology, Medical School, University of Thessalia, Larissa 41222, Greece [M. I. K.]; Department of Pathology, Democritus University of Thrace, Alexandroupolis 68100, Greece [A. G., E. S.]; Department of Medical Oncology, Hellenic Cooperative Oncology Group, AHEPA Hospital, Aristotle University of Thessaloniki, Thessaloniki 54006, Greece [G. F.]; Department of Cellular Science and Imperial Cancer Research Fund Medical Oncology Unit, Oxford Radcliffe Hospital, Headington, Oxford OX3 7LI, United Kingdom [K. C. G., A. L. H.]

ABSTRACT

Thymidine phosphorylase (TP), an enzyme involved in the thymidine synthesis and degradation, has been shown to promote tumor angiogenesis. Both TP expression and tumor vascularization are putative postoperative prognostic markers of cancer. Because of its bifunctional role, TP may have interactions with cytotoxic drugs or radiation via pathways requiring thymidine or prodrug activation. The microvessel score and TP expression were examined immunohistochemically on paraffin-embedded biopsial material from 94 locally advanced squamous cell head and neck carcinomas. All patients were treated with conventionally fractionated radiotherapy combined with induction (platinum- and 5-fluorouracil-based) or concurrent platinum chemotherapy. The follow-up of patients ranged from 6 to 108 months (median, 48 months). Nuclear TP expression was significantly associated with increased microvessel score ($P < 0.0001$, $r = 0.45$). A low percentage of cancer cells with nuclear TP expression in pretreatment biopsies was associated with a high rate of CR after combined chemoradiotherapy ($P = 0.006$) and induction chemotherapy (0.01). A better local relapse-free and overall survival was also observed in these patients ($P = 0.001$ and $P = 0.0005$, respectively). Biopsies on the day after the delivery of 20 Gy of conventionally fractionated radiotherapy showed residual cancer cell nests, frequently of high vascularization and of intense nuclear TP reactivity. It is concluded that thymidine phosphorylase is associated with angiogenesis, with resistance to radiotherapy and cytotoxic therapy, and with poorer survival in squamous cell head and neck cancer. A strong rationale is provided for subsequent clinical trials of concurrent radiotherapy and chemotherapy with antiangiogenic agents or with specific TP inhibitors.

INTRODUCTION

TP is an enzyme involved in DNA synthesis (1). It catalyzes the reversible synthesis of thymidine from thymine but also transforms deoxyribose to a pyrimidine base, resulting in the formation of a deoxynucleoside. A putative role of TP in human malignancies has been suggested since the early 1960s (2, 3).

The importance of TP and TS in cancer chemotherapy with antimetabolites has focused our attention recently. In vitro data suggest that high levels of TP correlate with an enhanced response to fluoropyrimidines, because increased 5-fluorouracil transformation to fluorodeoxyuridine results in higher amounts of FdUMP production, increased DNA FdUTP incorporation, and strong TS inactivation by ternary complex formation. TP may also prevent thymidine salvage, and therefore, TP can potentiate 5-fluorouracil or methotrexate from a different mechanism besides activation (4). On the other hand, high levels of TS correlate with decreased sensitivity to fluoropyrimidines. If high levels of TS remain free after fluoropyrimidine chemotherapy, DNA synthesis and cancer cell proliferation will efficiently go on. Although several clinicopathological studies have been reported recently, providing evidence for an important role of TS in resistance to antimetabolites (5, 6), the role of TP has not been studied extensively (7).

TP is identical to platelet-derived endothelial cell growth factor, which is a protein involved in the stimulation of endothelial cell migration and proliferation (8–10). The association of TP expression with increased intratumoral angiogenesis has been verified in several studies in breast (11), lung (12), and colorectal cancer (13). Experimental data support the idea of an important synergistic role of antiangiogenic agents with radiotherapy (14). Indeed, in a recent study, we observed an impor-

Received 9/7/99; revised 11/16/99; accepted 11/22/99.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

1 Designed, analyzed, interpreted, and financially supported by the Tumor and Angiogenesis Research Group and the Imperial Cancer Research Fund.

2 To whom requests for reprints should be addressed, at Tumour and Angiogenesis Research Group, 18 Dimokratias Avenue, Iraklion 71306, Crete, Greece. Phone: 30-932-480808; Fax: 30-81-392848; E-mail: targ@her.forthnet.gr.

3 The abbreviations used are: TP, thymidine phosphorylase; TS, thymidine synthase; MS, microvessel score; CR, complete response; PR, partial response; NR, no response; APAAP, alkaline phosphatase/anti-alkaline phosphatase; MAb, monoclonal antibody; LVG, low vessel grade; MVG, medium vessel grade; HVG, high vessel grade; CI, confidence interval; LRFS, local relapse-free survival; OS, overall survival.
Thymidine Phosphorylase and Chemoradiotherapy

and localization in a subset of patients undergoing radiotherapy. We therefore measured changes of TP expression in less vascularized non-small cell lung cancer cases and found that TP expression in these patients was used to assess the changes induced by radiotherapy and chemotherapy. These patients were not included in the analysis of TP expression in squamous cell carcinomas. Patients and disease characteristics are shown in Table 1. All cases were locally advanced cancers and were treated with conventionally fractionated radiotherapy combined with chemotherapy. None of the patients underwent surgery. The follow-up of patients ranged from 1 to 108 months (median, 12 months). For patients alive at the follow-up, the follow-up ranges from 6 to 108 months (median, 48 months).

In an additional cohort of 14 patients treated prospectively with radiotherapy alone, biopsies of the primary tumor were performed, either endoscopically or directly, 1 day before and immediately after the delivery of 20 Gy of conventionally fractionated radiotherapy (15 days after the beginning of fractionated radiotherapy). These patients were not included in the response and survival analysis. Six of these patients suffered from nasopharyngeal, 3 hypopharyngeal, 1 laryngeal, and 4 oral cavity squamous cell carcinomas. The biopsy material from these patients was used to assess the changes induced by radiotherapy in the patterns of vascularization and of TP cancer cell expression.

**MATERIALS AND METHODS**

The MS and TP expression were examined immunohistochemically in 2-μm tissue sections of paraffin-embedded biopsy material from 94 primary head and neck cancers, histologically shown to be squamous cell carcinomas. Patients and disease characteristics are shown in Table 1. All cases were locally advanced cancers and were treated with conventionally fractionated radiotherapy combined with chemotherapy. None of the patients underwent surgery. The follow-up of patients ranged from 1 to 108 months (median, 12 months). For patients alive at the follow-up, the follow-up ranges from 6 to 108 months (median, 48 months).

In an additional cohort of 14 patients treated prospectively with radiotherapy alone, biopsies of the primary tumor were performed, either endoscopically or directly, 1 day before and immediately after the delivery of 20 Gy of conventionally fractionated radiotherapy (15 days after the beginning of fractionated radiotherapy). These patients were not included in the response and survival analysis. Six of these patients suffered from nasopharyngeal, 3 hypopharyngeal, 1 laryngeal, and 4 oral cavity squamous cell carcinomas. The biopsy material from these patients was used to assess the changes induced by radiotherapy in the patterns of vascularization and of TP cancer cell expression.

**TREATMENT AND RESPONSE ASSESSMENT.** Baseline studies included physical examination, chest X-rays, whole blood count with differential and platelet count, complete biochemical profile, bone scan, and computed tomography of the cranium, neck, and chest. Treatment characteristics are shown in Table 2. Although the four treatment cohorts included in this study were not randomized, these were sequential in time, and there was no patient selection according to the regimen. Thirty-eight patients received three cycles of induction chemotherapy, followed by radical radiotherapy (LINAC 6 MV or Cobalt 60 Unit; 2 Gy/fraction; five fractions/week; total tumor dose, 70 Gy). Fifty-six patients were treated with concurrent chemoradiotherapy (same radiotherapy regimen).

The response to treatment was assessed with a computed tomography scan of the head and neck area immediately after the completion of induction chemotherapy and 45–60 days after
radiation treatment completion. The duration of response was measured from the time the criteria for the objective response were first met, with the computed tomography scan done every 2 months for the first 6 months and every 3–4 months (or earlier if necessary) thereafter. Complete response was defined as the disappearance of all measurable lesions within 2 months after treatment completion that lasted for at least 2 months after response documentation. A remnant scar on computed tomography scan measuring <5% of the initial tumor volume and with no signs of progression within 2 months after response documentation was considered as complete response. Similarly, partial and minimal response refers to a 50–95% and 25–49% reduction in tumor size, respectively. Small reductions in tumor size (0–24%) were considered stable disease. All other cases were considered progressive disease. In the present study, minimal response, stable, and progressive disease were considered in one group of NR.

**Assessment of TP Expression.** TP expression was assessed with the P-GF.44C monoclonal antibody (18) using the APAAP method. Two-μm paraffin-embedded sections were dewaxed and rehydrated. P-GF.44C monoclonal antibody as undiluted supernatant was applied at room temperature for 30 min and washed in TBS. Rabbit antimouse antibody 1:50 was applied for 30 min, followed by application of mouse APAAP

*Fig. 1* A, a squamous cell head and neck carcinoma with predominantly nuclear TP reactivity (×200). B, a squamous cell head and neck carcinoma with intense nuclear and cytoplasmic TP reactivity (×200).
complex 1:1 for 30 min. After washing in TBS, the last two steps were repeated for 10 min each. The color was developed by 15-min incubation with New Fuchsin solution. Normal rabbit IgG was substituted for primary antibody as the negative control (same concentration as the test antibody). Tumor-infiltrating macrophages were used as a positive internal control.

In a previous study, we observed that TP staining in squamous lung and head and neck cancer could be either cytoplasmic or nuclear (16, 17). Tissue samples were scanned at low (×40 and ×100) power, and the percentage of nuclear (TPnuc) and strong cytoplasmic (TPcyt) expression was separately assessed in six chosen optical ×250 fields of the highest reactivity. The percentage of staining was the mean value of the scores obtained. We also assessed the overall TP staining (TPall), which is the percentage of positive cells no matter whether the reactivity was in the nucleus or in the cytoplasm or in both subcellular localizations. The results were analyzed as continuous variables. We also considered the mean percentage of positive cells as a cutoff point to group cases in two categories of low and high TPnuc or TPcyt reactivity. The specimens were scanned at low optical power (×40 and ×100), and microvessel counting was performed on ×250 fields. Three areas (per case) of high vascularization were chosen for microvessel counting. The overall MS was the mean value of all appraised fields of the case. Vessels with a clearly defined lumen or well-defined linear vessel shape but not single endothelial cells were taken into account for microvessel counting. Overall MS was used for statistical analysis, both as a continuous and a categorical variable. Microvessel analysis was performed by three independent observers, and results were assessed for interobserver variability. Disagreement was resolved from a conference microscope. The vascular grade grouping of our cases was based on a previous study (15). Briefly, patients with an intermediate microvessel score (10–50 vessels per ×250 field) had a statistically significant better CR rate, local relapse, and OS as compared with patients with poor (<10 vessels per ×250 field) or high (>50 vessels per ×250 field) vascularization. Cases with poor and high vascularization...
had a similarly poor prognosis. Therefore, in the present study, we adopted three groups of vascular grade: LVG, MVG, and HVG.

**Statistical Analysis.** Statistical analysis and graphs were performed using the GraphPad Prism 2.01 and the Instat 3.0 packages (San Diego, CA). Nonparametric unpaired two tailed t test with Welch’s correction or Fisher’s exact t test was used for testing relationships between categorical tumor variables, as appropriate. Linear regression analysis was used to assess interobserver variability or correlation between continuous variables. Multiple regression analysis was also used as appropriate. Survival curves were plotted using the method of Kaplan and Meier, and the log-rank test was used to determine statistical differences between life tables (20, 21). The end points for analysis were the response rate, the local progression-free survival, and the OS starting from the last day of radiotherapy. CR rate was separately assessed from PR rate because head and neck cancer is a curable disease, with the PR always indicative of treatment failure. A Cox proportional hazard model was used to assess which of the tumors variables were independently correlated with response, local relapse, and death events (22).

**RESULTS**

**TP Reactivity Assessment.** The mean cytoplasmic TP-cyt reactivity was 38.2 ± 24%, ranging from 0% to 90% (95% CI, 33–43%). The mean nuclear TPnuc reactivity was 32.2 ± 25%, ranging from 0% to 90% (95% CI, 24–37%). Overall, the mean TPall reactivity was 47.8 ± 28% (95% CI, 42–53%). Interobserver variability was minimal (P < 0.0001, r > 0.86). Linear regression analysis of TPnuc versus TPcyt showed a significant association between variables (P < 0.0001, r = 0.58). As cutoff points for identifying high and low TP reactivity, the mean percentage of positive cells assessed is used. Using the cutoff point of 32% for TPnuc reactivity, 56 of 94 cases had low TPnuc reactivity, and 38 of 94 cases had high reactivity. Nine of 38 cases with high TPnuc reactivity had low cytoplasmic reactivity, and 19 of 48 cases with high TPcyt had low nuclear reactivity.

None of the TP reactivity patterns showed a statistical significant correlation with T stage, N stage, histological grade, or localization of the primary tumor (data not shown).

**TP Reactivity and MS.** The overall MS ranged from 6 to 63 microvessels per ×250 optical field (mean, 34.2 ± 21; 95% CI, 29–38). Interobserver variability was minimal (P < 0.0001, r > 0.91). Linear regression analysis of TP score and MS showed a significant association of TPnuc with increasing MS (P < 0.0001, r = 0.45). TPcyt was marginally associated with MS, and a poor goodness of fit was noted (P = 0.01, r = 0.24). Multiple regression analysis of TPnuc, TPall, and TPcyt showed that only TPnuc was significantly associated with the MS (P = 0.02; t ratio, 2.35).

**TP Reactivity and Response.** In a bivariate model, we analyzed the association of TPnuc and TPcyt reactivity with radius to chemotherapy (all cases). Only the nuclear patterns of staining showed a significant association with response (P = 0.04; t ratio, 2.0).

We analyzed the percentage of TPnuc-reactive cells in tumors with CR, PR, or less than PR (NR) to radiotherapy and chemotherapy. Cases with CR after combined therapy (all cases together) had a significantly lower percentage of TPnuc reactivity (27.1 ± 23) as compared with cases with PR (42.0 ± 30; P = 0.03) and NR (35.9 ± 28; P = 0.17; Fig. 2a).

Cases that showed CR to induction chemotherapy also had a low percentage of TPnuc reactivity (20.6 ± 20) as compared with cases with PR (43.6 ± 29; P = 0.03) or with NR (41.3 ± 23; P = 0.05; Fig. 2b). Similarly, a lower percentage of TPnuc cells was present in cases that had a CR after concurrent chemoradiotherapy (P = 0.05). Analysis of cases that showed PR/NR to induction chemotherapy but CR after radiotherapy that followed the induction chemotherapy showed no significant difference in the percentage of TPnuc reactivity as compared with cases that had PR or NR after radiotherapy. The CR rate of this latter category of patients was lower as compared with the CR rate observed after concurrent chemoradiotherapy [12 of 30 (40%) versus 33 of 56 (59%); P = 0.11].

Table 3 shows the responses (CR, PR, and NR) obtained after radiotherapy, induction chemotherapy, radiotherapy after induction chemotherapy, and concurrent chemoradiotherapy stratified for TP nuclear reactivity.

**Nuclear TP Reactivity and Prognosis.** In a bivariate model, we examined the impact of TPnuc and TPcyt reactivity (high versus low) on the local relapse and death events. A significant association was found only for TPnuc reactivity (P = 0.003, t ratio is 2.9 for local relapse; P = 0.001, t ratio is 3.2 for OS).

Fig. 3 shows the Kaplan-Meier survival curves (all cases) for LRFS and OS, stratifying for TPnuc reactivity and using as a cutoff point of the mean percentage of cells with nuclear staining (32%). The median LRFS for patients with high TPnuc reactivity was 5 months as compared with 32 months of median LRFS obtained in patients with low TPnuc reactivity (P = 0.001). The median OS in patients with high and low TPnuc reactivity was 10 and 36 months, respectively (P = 0.0005).

![Table 3 Nuclear TP reactivity and response to radiotherapy and chemotherapy](attachment:table3.png)

<table>
<thead>
<tr>
<th>TP nuclear reactivity</th>
<th>Low (≤32% of cells)</th>
<th>High (&gt;32% of cells)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>All cases</td>
<td>CR 38</td>
<td>15</td>
<td>0.006</td>
</tr>
<tr>
<td></td>
<td>PR 9</td>
<td>11</td>
<td></td>
</tr>
<tr>
<td></td>
<td>NR 9</td>
<td>12</td>
<td></td>
</tr>
<tr>
<td>Induction chemotherapy</td>
<td>CR 7</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td></td>
<td>PR 8</td>
<td>11</td>
<td>0.01</td>
</tr>
<tr>
<td></td>
<td>NR 4</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>Radiotherapy after</td>
<td>induction chemotherapy</td>
<td>CR 6</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>induction chemotherapy</td>
<td>PR 4</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>induction chemotherapy</td>
<td>NR 2</td>
<td>6</td>
</tr>
<tr>
<td>Concurrent chemoradiotherapy</td>
<td>CR 25</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td></td>
<td>PR 5</td>
<td>5</td>
<td>0.06</td>
</tr>
<tr>
<td></td>
<td>NR 7</td>
<td>6</td>
<td></td>
</tr>
</tbody>
</table>

*Patients with incomplete response to chemotherapy.*
Further analysis in the group of patients treated with induction chemotherapy revealed similar results. Patients with high TPnuc reactivity had a worse LRFS and OS (\(P = 0.01\) and \(P = 0.01\), respectively) as compared with patients bearing tumor with low TPnuc reactivity. Patients with high TPnuc reactivity treated with concurrent chemoradiotherapy had similarly worse prognosis (\(P = 0.10\) and \(P = 0.04\), respectively).

**Nuclear TP Reactivity, MS, and Prognosis.** T stage, N stage, histological grade, or primary site localization was not associated with response or prognosis both in terms of LRFS and OS. A higher CR rate, a better LRFS, and OS have been observed in cases with MVG (MS 11–50) as compared with cases with LVG (MS <11) and HVG (MS >50). This finding has been reported and discussed in a previous study (17). In a bivariate model taking into account the VG (MVG versus LVG, HVG) and TPnuc reactivity (low versus high), we observed that, despite the previously noted association of TPnuc with high angiogenesis, TPnuc reactivity was an independent factor defining response to radiotherapy (\(P = 0.01\); \(t\) ratio, 2.5), whereas VG was not (\(P = 0.22\); \(t\) ratio, 1.20). As far as the local relapse events is concerned, both TPnuc and VG were independent prognostic variables (\(P = 0.0007\); \(t\) ratio 3.5 for TPnuc; \(P = 0.0001\); \(t\) ratio 5.0 for VG). Similarly, both parameters had an independent prognostic meaning as for death events (\(P = 0.003\); \(t\) ratio 2.99 for TPnuc; \(P = 0.0001\); \(t\) ratio 4.0 for VG).

**Comparative Analysis of MS and TPnuc Changing Patterns before and after Radiotherapy.** Sections from head and neck carcinomas treated with 20 Gy of radiotherapy showed bands and islands of cancer cells immersed within avascular degenerated tissue areas that were frequently of high vascularization and of intense nuclear TP reactivity (Fig. 4). Degenerated cancer cells were negative for nuclear TP expression, whereas cytoplasmic expression was occasionally present. In patients with CR, the mean MS in these areas of viable cells was 35.3 ± 19 before radiotherapy and decreased to 19.3 ± 15 after radiotherapy (\(P = 0.11\)). In patients with PR/NR, the initial score was 39.5 ± 23 and was maintained at 42.5 ± 32 (\(P = 0.81\)). Similarly, the mean TPnuc reactivity in patients with CR was decreased from 40.2 ± 32 before radiotherapy to 28.1 ± 27 after radiotherapy (\(P = 0.05\)). In patients with PR/NR, this remained stable (50.1 ± 1 versus 50.1 ± 34; \(P = 1\)).

**DISCUSSION**

The role of TP as an enzyme responsible for antimetabolite drug activation as well as a factor responsible for endothelial cell migration and proliferation has been well established in experimental studies (1–10). Several clinicopathological studies have shown a direct association of cancer cell or stromal cell TP overexpression with increased tumor vascularization and poor prognosis (11–13). Moreover, Fox et al. (7) found a positive association of TP overexpression with better outcome in breast cancer cases treated with cyclophosphamide-methotrexate-5-fluorouracil chemotherapy, providing an *in vivo* evidence of TP-mediated antimetabolite activation.

The ominous prognostic value of high intratumoral vascu-
larization has been reported by a large number of studies in different tumors (23). Because TP is associated with high vascularization, its overexpression should be associated with worse prognosis. However, the role of TP in the outcome of chemotherapy and radiotherapy is balanced by at least three different TP-related features: (a) antimetabolite activation; (b) increased intrinsic cancer cell aggressiveness; and (c) increased tumor vascular density and angiogenicity.

In the present study, we examined the response to platinum and 5-fluorouracil induction chemotherapy in a cohort of 38 squamous cell head and neck cancers. Cases that showed a CR after induction chemotherapy had a very low percentage of TP nuclear and cytoplasmic reactivity as compared with cases with PR or NR. It seems, therefore, that cancer cells with very low levels of TP are sensitive to DNA-damaging agents. Impaired DNA repair may underlie the phenomenon. The observation that induction chemotherapy with 5-fluorouracil did not result in a higher response rate in TP-overexpressing cases may be a consequence of the relative importance of platinum activity in TP-deficient cells. In a recent study by Fujieda et al. (24), susceptibility to 5-fluorouracil cytotoxicity in oropharyngeal cancer was observed only in extremely high TP expression. Our study shows that TP expression is not a major factor defining favorable response to 5-fluorouracil.

The predictive role of nuclear TP reactivity was also observed in a cohort of 56 patients treated with concurrent che-

![Fig. 4 Postirradiation (20 Gy) tissue sample of a squamous cell head and neck carcinoma immunostained for CD31 endothelial membrane antigen (anti-CD31 MAb) and for TP (PGF-44c MAb). Note a cancer cell nest with intense neovascularization (A; ×250) and intense nuclear TP expression within the same area (B; ×250).](clincancerres.aacrjournals.org)
Thymidine phosphorylase and chemoradiotherapy. LRFS and OS were significantly affected by TP overexpression. The median LRFS was 42 months in cases with reduced TP expression versus 5 months in cases with TP overexpression. Similar results have been reported by Metzger et al. (25), where increased TP levels in colorectal cancer were associated with poorer response to chemotherapy. Because radiotherapy and both platinum and 5-fluorouracil exert their cytotoxic activity through DNA damage, either cross-linking or single and double strand breaks, it may be that DNA-damaging agents are more effective in TP-deficient cancers. Although Fujieda et al. (24) did not find TP levels associated with resistance to cisplatin, in our study we used combined modality therapy with radiation and also 5-fluorouracil with cisplatin.

A possible alternative explanation for why tumors with high TP had a poorer response to therapy may be suggested from the comparative study of angiogenesis and TP expression, in tumors before and after 20 Gy of radiotherapy. The most frequent patterns of tissue structure in irradiated tumors was the presence of viable cancer cell foci immersed within degenerate tumor areas. The MS and the percentage of TP-positive cells in viable tumor areas were significantly decreasing after radiotherapy in tumors that reached CR, whereas these were stable or even increased in cases that did not respond completely to radiotherapy. This observation may show that the ability of cancer cells to maintain active angiogenesis and TP pathways may be important for the results of radiotherapy. The concept of “rapid cancer cell repopulation” during radiotherapy has been suggested to explain the reduced control rate after overall treatment time prolongation (26–28). However, the nature of the phenomenon is unclear because the cancer cell potential doubling time and the cell cycle time are unlikely to decrease during radiotherapy (29–31). Whether the so called “rapid tumor repopulation” phenomenon is a result of a more complicated process involving angiogenic pathways is an issue raised that should be investigated further.

Recently, novel antiangiogenic therapies have emerged (14, 32). Several studies suggest that antiangiogenesis approaches may significantly improve the efficacy of radiotherapy. After administration of angiotatin concurrently with radiotherapy a substantial increase of tumor control is achieved, although no cancer cell radiosensitization is mediated (14). In a recent study involving angiogenic pathways is an issue raised that should be investigated further.

We conclude that TP is an important molecule associated with angiogenesis, with resistance to radiotherapy and chemotherapy, and with poorer survival of locally advanced squamous cell head and neck cancer treated with chemoradiotherapy. Although TP is one pathway for metabolic activation of 5-fluorouracil, it seems that TP-related angiogenicity and a possible sensitivity of TP-deficient tumors to DNA-damaging agents may also confer two distinct mechanisms that should be further studied to clarify the role of the protein in tumor behavior. The cancer cell ability to maintain high angiogenesis and TP expression during radiotherapy may also prove of predictive value. The present study provides a strong rational for subsequent clinical trials of concurrent use of radiotherapy and chemotherapy with antiangiogenic agents and with specific TP inhibitors.

ACKNOWLEDGMENTS

We express special thanks to all medical doctors and scientists that participated in the diagnosis and treatment of the patients analyzed in this study.

REFERENCES


Angiogenesis, Thymidine Phosphorylase, and Resistance of Squamous Cell Head and Neck Cancer to Cytotoxic and Radiation Therapy

Michael I. Koukourakis, Alexandra Giatromanolaki, George Fountzilas, et al.


Updated version

Access the most recent version of this article at:

http://clincancerres.aacrjournals.org/content/6/2/381

Cited articles

This article cites 31 articles, 10 of which you can access for free at:

http://clincancerres.aacrjournals.org/content/6/2/381.full#ref-list-1

Citing articles

This article has been cited by 3 HighWire-hosted articles. Access the articles at:

http://clincancerres.aacrjournals.org/content/6/2/381.full#related-urls

E-mail alerts

Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions

To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions

To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.