The Orally Administered P-glycoprotein Inhibitor R101933 Does Not Alter the Plasma Pharmacokinetics of Docetaxel

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ABSTRACT

This Phase I study was performed to assess the feasibility of combining docetaxel with the new P-glycoprotein inhibitor R101933 and to determine the dose limiting toxicity of this combination. Fifteen patients received oral R101933 alone at a dose escalated from 200 to 300 mg twice daily (b.i.d.; cycle 0), an escalating i.v. dose of docetaxel (60, 75, and 100 mg/m²) as a 1-h infusion (cycle 1), and the combination (cycle 2 and further). Dose limiting toxicity consisting of mucositis and neutropenic fever was reached at the combination of docetaxel, 100 mg/m², and R101933, 300 mg b.i.d., and the maximum tolerated dose was established at docetaxel, 100 mg/m², and R101933, 200 mg b.i.d. Plasma concentrations of R101933 achieved in patients were in the same range as required in preclinical rodent models to overcome paclitaxel resistance. The plasma pharmacokinetics of docetaxel were not influenced by the R101933 regimen at any dose level tested, as indicated by plasma clearance values of 26.5 ± 7.78 liters/h/m² and 23.4 ± 4.52 liters/h/m² (P = 0.15) in cycles 1 and 2, respectively. These findings indicate that the contribution of a P-glycoprotein inhibitor to the activity of anticancer chemotherapy can now be assessed in patients for the first time independent of its effect on drug pharmacokinetics.

INTRODUCTION

Acquired or intrinsic resistance of malignant cells to taxanes and other naturally occurring drugs has been linked to the so-called “classical” mechanism of MDR resulting in decreased intracellular concentrations of these anticancer drugs. This MDR phenotype is characterized by increased levels of P-glycoprotein, a member of the ATP-binding cassette superfamily of transmembrane transport proteins with a molecular weight of M₉ 170,000 encoded by the MDR1 gene and acting as an energy-dependent drug efflux pump with broad substrate specificity (1, 2).

Since the first observation that verapamil could reverse MDR in vitro, similar properties have been shown for a wide range of drugs (3). These agents are thought to be competitive substrates for P-glycoprotein and thus can increase the intracellular concentration of a coadministered anticancer agent and consequently restore the antitumoral activity (4). Initially, a number of drugs, marketed for other indications than inhibiting P-glycoprotein, have entered clinical trials (5). However, it became evident that pharmacokinetic interactions occurred between these P-glycoprotein inhibitors and the coadministered anticancer drugs due in part to competitive inhibition of cytochrome P-450 enzymes resulting in significantly increased toxicity of the anticancer drug (6). By rational design, new modulators were developed to specifically inhibit P-glycoprotein and to be more suitable candidates for further clinical evaluations (7, 8). The results of most of these clinical studies have been rather disappointing, and the pharmacokinetic interaction between the cytotoxic and the P-glycoprotein inhibiting agent remains a confounding problem (6, 9).

R101933 (Fig. 1) is a new p.o. administered compound that inhibits P-glycoprotein as demonstrated by various in vitro and in vivo models (10, 11). The tolerability, cardiovascular and laboratory safety, and the pharmacokinetics were investigated in healthy subjects. Nausea and vomiting were the dose-limiting adverse events and were reported above the 400-mg single oral dose. Drowsiness was also mentioned as a side effect. No clinically relevant changes in laboratory and cardiovascular safety parameters were observed. In vitro metabolism studies showed that the major metabolic pathway is not cytochrome P450 3A4-dependent. Plasma levels of R101933 at 200 mg b.i.d. are in the range of concentrations that are active in paclitaxel and Adriamycin-resistant human tumor xenograph rodent models.

Docetaxel is a known substrate of P-glycoprotein and has shown to have a higher affinity for the protein than the related compound paclitaxel (12, 13). It also lacks the problems associated with i.v. use of paclitaxel caused by the presence of the...
formulation vehicle Cremophor EL, which is known to (a) alter the pharmacokinetics of the anticancer drug by entrapment in micelles (14) and (b) mask the effects, if any, of endogenously expressed P–glycoprotein on the plasma levels of paclitaxel (15). Therefore, the development of agents that could reverse or prevent the development of resistance to docetaxel is of great interest.

The principal objectives of this Phase I and pharmacokinetic study of R101933 and docetaxel were to determine the clinical utility of the combination and to investigate the potential lack of pharmacokinetic interactions.

PATIENTS AND METHODS

Eligibility. Patients with a histologically confirmed diagnosis of a solid tumor for whom docetaxel as monotherapy was a viable therapeutic option or for whom other treatment options were not available were candidates for this study. Additional eligibility criteria were: age ≥18 and ≤75 years; Eastern Cooperative Oncology Group performance status ≤3; life expectancy of at least 3 months; off previous anticancer therapy for at least 4 weeks; no previous treatment with taxanes or high dose chemotherapy requiring progenitor cell support; adequate bone marrow function (WBC count >3.5 × 10^9/liter, platelet count >100 × 10^9/liter), renal function (serum creatinine ≤2 times the upper limit of normal), and liver function (bilirubin level normal, aspartate/alanine aminotransferase ≤2 times upper limit of normal, and alkaline phosphatase ≤2.5 times upper limit of normal); and symptomatic peripheral neuropathy less than grade 2 (NCI criteria). Written informed consent was obtained from all patients, and the study was approved by the Rotterdam Cancer Institute Ethics Board.

Pretreatment and Follow-up. Pretreatment evaluation consisted of recording the history of the patient, physical examination, laboratory studies, electrocardiography, and chest X-ray. Computer tomographic scans were performed for tumor measurements. Laboratory studies included a complete blood cell count analysis and measurement of WBC differential, electrolytes (including sodium, potassium, chloride, calcium, and inorganic phosphate), creatinine, urea, alkaline phosphatase, aspartate aminotransferase, alanine aminotransferase, lactate dehydrogenase, bilirubin, total plasma proteins, serum albumin, glucose, uric acid, and urinalysis. History, physical examination, and toxicity scoring (according to the NCI-expanded CTC) were repeated once a week. Complete blood cell counts, including WBC differential, were performed twice a week, and the other laboratory tests were done once a week. Electrocardiography was repeated as clinically indicated. A final assessment was to be made after patients went off the study. Formal tumor measurements and chest X-ray were performed at 6-week intervals until documentation of PD. Standard WHO response criteria were used.

Drug Administration. Docetaxel was administered every 3 weeks on day 3 as a 1-h infusion and was started 1 h after intake of R101933. All patients received premedication with dexamethasone, p.o. 8 mg b.i.d., starting 1 day before each infusion of docetaxel for 5 days. R101933 (Janssen Research Foundation, Beerse, Belgium) was supplied as a 10-mg/ml oral solution in 15% hydroxypropyl-β-cyclodextrin. It had to be taken with water at least 1 h after a meal. The drug was administered twice daily from days 1–5. From studies with healthy volunteers, it was known that the terminal half-life of R101933 averaged about 24 h, with peak plasma concentrations attained within 2 h after intake. The MTD after 7-day b.i.d. dosing was 300 mg in healthy volunteers. Seven-day dosing at 200 mg b.i.d. appeared to be safe and well tolerated. Pharmacokinetic data revealed that plasma levels of R101933 at 200 mg b.i.d. achieve concentrations that are in the same range as required in in vivo models to overcome paclitaxel resistance. Hence, the starting dose for our study was set at this dose level. In view of the terminal disposition half-life of docetaxel, a simultaneous exposure to both R101933 and docetaxel for a 3-day period was considered sufficient. This led to the choice of the 5-day R101933 regimen. In the first stage of the study, the dose of docetaxel was escalated and the dose of R101933 was fixed. In the second stage of the study, the dose of docetaxel was fixed and the dose of R101933 was escalated.

First, the patients received five doses of R101933 alone every 12 h (cycle 0) followed by a 48-h wash-out to allow assessment of the terminal half-life of R101933. One week later, cycle 1 was initiated with docetaxel alone. Thereafter, the combination was given triweekly until PD or DLT occurred.

In each cohort, three patients were treated unless DLTc or DLTr was observed. In that case, the accrual of three additional patients was required. DLTc was defined as grade 3 nonhematological toxicity (with the exception of nonhematological toxicity that was still manageable in an out-patient setting, such as nausea/vomiting) or grade 4 neutropenia lasting ≥8 days or grade 4 thrombocytopenia or required delay ≥2 weeks to a subsequent cycle due to toxicity. Febrile neutropenia and neutropenia with severe infection (≥grade 3 infection) was also considered as DLTc. DLTr was defined as any nonhematological toxicity ≥grade 2 in the first 2 days of treatment before chemotherapy was given. For dose-escalation decisions, only DLTs in cycles 0 and 2 were taken into account. The DLT of the combination of R101933 with docetaxel was reached when greater or equal to three of six patients experienced DLTc. The DLT of R101933 alone was reached when greater or equal to one of three (or greater or equal to two of six) patients experienced DLTr. The MTD was defined as the dose level below DLT.

Sample Collection and Processing. Blood specimens were taken in all patients during the first, second, and third courses of treatment. Blood volumes of 6 ml were drawn di-
rectly into Vacutainer tubes containing lyophilized sodium heparin (Becton Dickinson, Meylan, France) from a peripheral venous access device. In each patient, sufficient plasma was obtained before drug administration to evaluate possible interfering peaks in the chromatographic analysis. Samples for docetaxel analysis were collected immediately before infusion and at 0.5, 1, 1.25, 1.5, 2, 3, 7, 11, 23, and 31 h after the start of infusion. For the determination of R101933 concentrations, blood samples were obtained on day 1 (before the first dosing), day 2 (before the second dosing and 12 h thereafter), and day 3 (before the third dosing and 2, 4, 8, 12, 24, 32, and 48 h thereafter). All blood samples were centrifuged immediately for 10 min at 1000 x g to yield plasma, which was stored frozen in polypropylene vials (Eppendorf, Hamburg, Germany) until the time of analysis.

**Analytical Methods.** A pure reference standard of docetaxel (batch, 14PROC9230; purity, 98.0% by reversed-phase high-performance liquid chromatography) and the clinical docetaxel formulation in polysorbate 80 (Taxotere; 40 mg/ml) were kindly supplied by Rhône-Poulenc Rorer (Vitry-sur-Seine, France) and were used as received. Plasma concentrations of docetaxel were determined by a validated liquid chromatographic method described for paclitaxel (16). A stainless steel analytical column (50 x 4.6 mm internal diameter) packed with 3-μm Hypersil BDS C18 material (Alltech, Breda, the Netherlands) was used for chromatographic separation, and gradient elution was performed with a mixture of acetonitrile and 0.02 M ammonium acetate (pH 4.0) at a flow rate of 0.8 ml/min. Paclitaxel (50 μl of 20 μg/ml in acetonitrile) was used as internal standard. Triple quadrupole mass-spectrometric detection was performed with a turboionspray interface used in the positive ion mode according to the square of the model predictions of the concentration-time profiles of docetaxel:internal standard ratio of the log-transformed peak areas of each of the analytes: internal standard were plotted versus nominal concentrations for quantitative computations.

**Pharmacokinetic Data Analysis.** Individual plasma concentration-time profiles of R101933 and its inactive metabolite R102207 were analyzed model independently using a validated macro in the EXCEL software package. The actual times of drug intake and blood sampling were taken into account. Peak plasma concentration (C_max) was determined by visual inspection of the data. The AUC within a 12-h dosing interval was calculated by the trapezoidal rule. In all cases, the AUC was extrapolated to infinity by addition of C_last/A, in which C_last is the last quantifiable concentration in the curve and A is the terminal elimination rate constant determined by linear regression analysis of the terminal points of the ln-linear plasma concentration-time curve. The terminal disposition half-life [1/[β(2)]] was defined as ln2/A. Individual plasma concentration-time curves of docetaxel were analyzed using the software package WinNonlin (Pharsight, Mountain View, CA) by determining the slopes and intercepts of the plotted curves with multieponential functions. All curves were fitted using the actual infusion duration and blood sampling times. In all cases, concentration-time profiles of docetaxel were best fitted to a biexponential equation after zero-order input with weighting according to the square of the model predictions of the concentrations. Final values of the incorporated parameters of the best-fit function were used to calculate the pharmacokinetic parameters using standard equations (17).

**Statistical Considerations.** Pharmacokinetic parameters for docetaxel and R101933 are reported as mean values ± SD. Variability in dose-normalized parameters between the various docetaxel dose levels was evaluated by the Kruskal-Wallis statistic followed, if required, by a Dunn’s test to determine which group differed. To test pharmacodynamic and pharmacokinetic parameter differences for statistical significance among treatment courses, a two-tailed paired Student’s t test was performed. Probability values of <0.05 were regarded as statistically significant. All statistical calculations were performed using the Number Cruncher Statistical System version 5.X (Dr. Jerry Hintze, Kaysville, UT; 1992) or using Statgraphics Plus version 2 (Manugistics Inc., Rockville, MA).

**RESULTS**

Seventeen patients were entered into this study. Patient characteristics are listed in Table 1; all patients were eligible. Two patients were considered not evaluable for toxicity and response. They did not receive the combination therapy, one because of unexpected rapid deterioration of the clinical condition and another because of development of liver enzyme abnormalities due to the malignant disease that would have precluded administration of docetaxel within normal safety limits.
DLTr was not reached at any of the investigated dose levels. Nausea/vomiting and drowsiness, known to be side effects of R101933 in healthy subjects, were not seen in our patients after administration of R101933 alone. Fatigue was often mentioned as a side effect after docetaxel but never exceeded grade 2. Nevertheless, for one patient given docetaxel at 100 mg/m² and R101933, 200 mg b.i.d., it was a reason to refuse further treatment after cycle 3, although an ongoing partial response was noted. Two patients at docetaxel 100 mg/m² and R101933, 300 mg b.i.d., went off the study after cycle 2 because of the observed DLT consisting of mucositis and vomiting, respectively. All other patients went off the study because of PD. Two patients achieved a partial response, and seven had stable disease.

### Plasma Pharmacokinetics.
For the evaluation of docetaxel pharmacokinetics, only the patients who had sampling and complete kinetic data during both treatment courses were included (n = 14 of 15). The results of paired plasma concentration-time profiles of unchanged docetaxel given with and without cotreatment were remarkably similar for all patients studied (Fig. 2). During both treatment courses, disposition phases appeared to be very typical of a biexponential profile, with plasma concentrations of docetaxel decreasing very rapidly immediately after cessation of the infusion, followed by a more prolonged terminal disposition phase of ~11 h, in line with previous observations (18). The mean pharmacokinetic parameters of docetaxel for both treatment courses are summarized as a function of the study cohort in Table 5. The docetaxel total body clearance was normally distributed as judged by the D’Agostino-Pearson omnibus K² test, was independent of the administered dose (Kruskal-Wallis, P = 0.396), and averaged 26.5 ± 7.78 liters/h/m² (mean ± SD) without R101933 and 23.4 ± 4.52 liters/h/m² with R101933 (Kruskal-Wallis, P = 0.608), which is within the same range as described for this compound previously (18). There were no statistically significant differences in any of the studied docetaxel pharmacokinetic parameters, including the clearance (P = 0.15), between the two treatment courses (Table 5), suggesting that R101933 administration did not influence the disposition of the taxane at the dose levels tested. At the final dose level, combining docetaxel at 100 mg/m² and R101933, 300 mg b.i.d, statistical analysis indicated that a 1.3-fold change in docetaxel clearance could have been detected with (1−β) = 0.80 at the observed SD of the mean difference between cycles (δ = 3.13) and a calculated standardized difference of 2δ/σδ (19).

Similarly, docetaxel did not significantly alter the absorption and elimination routes of R101933 (Table 6). In addition, dose-normalized AUC values for R101933 were similar with or without docetaxel cotreatment. Overall, substantial interpatient variability in R101933 kinetic parameters was apparent, with up to a 10-fold variation in peak plasma levels. Over the total dose range studied, the peak plasma levels of R101933 did not increase with values of 133 ± 74 ng/ml (mean ± SD; n = 10) and 136 ± 45 ng/ml (n = 7) respectively, suggesting a dose-dependent kinetic behavior of the compound with saturable absorption characteristics. For this reason, no attempt was made to further increase the dose of R101933. In all patients, there was extensive formation of the pharmacologically inactive compound R102207, the principal circulating metabolite of

### Table 1 Patient characteristics

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients included</td>
<td>17</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>10</td>
</tr>
<tr>
<td>Female</td>
<td>7</td>
</tr>
<tr>
<td>Age, yr</td>
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<tr>
<td>Median</td>
<td>57.5</td>
</tr>
<tr>
<td>Range</td>
<td>42–72</td>
</tr>
<tr>
<td>Performance score (WHO)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>8</td>
</tr>
<tr>
<td>1</td>
<td>9</td>
</tr>
<tr>
<td>Primary tumor</td>
<td></td>
</tr>
<tr>
<td>Urogenital tract</td>
<td>6</td>
</tr>
<tr>
<td>Gastrointestinal tract</td>
<td>5</td>
</tr>
<tr>
<td>Respiratory tract</td>
<td>2</td>
</tr>
<tr>
<td>Melanoma</td>
<td>2</td>
</tr>
<tr>
<td>Sarcoma</td>
<td>1</td>
</tr>
<tr>
<td>Unknown primary</td>
<td>1</td>
</tr>
<tr>
<td>Prior therapy</td>
<td></td>
</tr>
<tr>
<td>Surgery</td>
<td>14</td>
</tr>
<tr>
<td>Radiotherapy</td>
<td>7</td>
</tr>
<tr>
<td>Chemotherapy</td>
<td>11</td>
</tr>
<tr>
<td>None</td>
<td>1</td>
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</table>
R101933, reaching AUC values ~80-fold higher than that of the parent compound. Concentrations of this compound were also not substantially influenced by the administration of docetaxel at any dose level tested (Table 6). Of particular note, plasma levels of R101933 capable of reversal of daunorubicin resistance in A2780 cell cultures with and without P-glycoprotein expression were achieved in all patients (20).

### DISCUSSION

In the present study, we observed that the plasma pharmacokinetic characteristics of docetaxel were not substantially influenced by R101933, a new p.o. administered P-glycoprotein inhibitor. The lack of a pharmacokinetic interaction between docetaxel and R101933 is an important finding that makes it possible to study the contribution of an inhibitor of P-glycoprotein to the toxicity and activity of an anticancer drug independent-
Table 5  Plasma pharmacokinetic parameters of docetaxel in the absence or presence of R101933*

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Docetaxel 60 mg/m² (n = 3)</th>
<th>With R, 200 mg</th>
<th>Docetaxel 75 mg/m² (n = 3)</th>
<th>With R, 200 mg</th>
<th>Docetaxel 100 mg/m² (n = 3)</th>
<th>Without R</th>
<th>With R, 200 mg</th>
<th>Docetaxel 100 mg/m² (n = 5)</th>
<th>Without R</th>
<th>With R, 300 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>AUC (µg·h/ml)</td>
<td>3.18 ± 0.95</td>
<td>2.88 ± 0.52</td>
<td>2.83 ± 1.03</td>
<td>3.59 ± 0.30</td>
<td>3.93 ± 0.67</td>
<td>4.33 ± 1.13</td>
<td>3.62 ± 0.71</td>
<td>4.04 ± 0.84</td>
<td>2.54 ± 0.54</td>
<td>2.86 ± 0.55</td>
</tr>
<tr>
<td>Cₘ₉ (µg/ml)</td>
<td>2.06 ± 0.58</td>
<td>1.77 ± 0.27</td>
<td>1.68 ± 1.01</td>
<td>2.20 ± 0.53</td>
<td>2.47 ± 0.21</td>
<td>2.95 ± 0.26</td>
<td>2.54 ± 0.54</td>
<td>2.86 ± 0.55</td>
<td>0.18 ± 0.03</td>
<td>0.23 ± 0.07</td>
</tr>
<tr>
<td>t₁/₂(α) (h)</td>
<td>0.17 ± 0.02</td>
<td>0.20 ± 0.01</td>
<td>0.37 ± 0.36</td>
<td>0.32 ± 0.12</td>
<td>0.15 ± 0.06</td>
<td>0.24 ± 0.06</td>
<td>0.18 ± 0.03</td>
<td>0.23 ± 0.07</td>
<td>1.10 ± 2.00</td>
<td>10.9 ± 4.80</td>
</tr>
<tr>
<td>t₁/₂(β) (h)</td>
<td>12.5 ± 3.8</td>
<td>11.3 ± 0.4</td>
<td>12.0 ± 2.4</td>
<td>12.1 ± 2.5</td>
<td>11.1 ± 2.3</td>
<td>9.4 ± 4.4</td>
<td>11.0 ± 2.0</td>
<td>10.9 ± 4.80</td>
<td>5.51 ± 1.92</td>
<td>4.59 ± 0.95</td>
</tr>
<tr>
<td>Vₐ (liters/m²)</td>
<td>120 ± 43</td>
<td>119 ± 35</td>
<td>253 ± 227</td>
<td>112 ± 29</td>
<td>137 ± 27</td>
<td>86 ± 41</td>
<td>129 ± 32</td>
<td>104 ± 50</td>
<td>5.31 ± 1.22</td>
<td>4.27 ± 2.02</td>
</tr>
</tbody>
</table>

* Data were obtained from patients after the first (without R101933) and second treatment cycle (with R101933) of a 1-h infusion of docetaxel. The kinetic terms are mean values ± SD.

Table 6  Plasma pharmacokinetic parameters of R101933 and its metabolite R102207 in the absence or presence of docetaxel*

<table>
<thead>
<tr>
<th>Parameter</th>
<th>R101933 200 mg (n = 9)</th>
<th>Without D With D</th>
<th>Without D With D</th>
<th>R102207 300 mg (n = 5)</th>
<th>Without D With D</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cₘ₉ (ng/ml)</td>
<td>120 ± 66</td>
<td>94.3 ± 26.9</td>
<td>127 ± 50</td>
<td>144 ± 42</td>
<td>50.5 ± 14.2</td>
</tr>
<tr>
<td>t₁/₂(α) (h)</td>
<td>19.6 ± 7.4</td>
<td>NA</td>
<td>23.0 ± 9.2</td>
<td>NA</td>
<td>52.4 ± 26.7</td>
</tr>
<tr>
<td>AUC (µg·h/ml)</td>
<td>0.55 ± 0.24</td>
<td>NA</td>
<td>0.91 ± 0.46</td>
<td>0.86 ± 0.49</td>
<td>37.9 ± 20.2</td>
</tr>
<tr>
<td>R102207 AUC (µg·h/ml)</td>
<td>50.5 ± 14.2</td>
<td>52.4 ± 26.7</td>
<td>45.7 ± 11.0</td>
<td>45.7 ± 11.0</td>
<td></td>
</tr>
</tbody>
</table>

* Data were obtained from patients after cycle 0 (without docetaxel) and after cycle 2 (with docetaxel). The kinetic terms are mean values ± SD.

r-verapamil (21), cyclosporin A (22), PSC833 (23), or VX-710 (24), are most likely more related to an overlap in specificity of enzymes responsible for metabolism of the compounds than to modulation of P-glycoprotein activity. Although few clinical data are available, several in vitro studies have shown that docetaxel is extensively metabolized in humans by the cytochrome P450 3A4 system (25, 26). The main pathway of docetaxel metabolism in humans consists of successive oxidations of the tert-buty1 propionate group on the C13 side chain, with spontaneous cyclization occurring for the putative aldehyde and acid derivatives. All metabolites thus far characterized have been found to be >100-fold less cytotoxic than docetaxel itself (27, 28). In this context, it is noteworthy that R101933 did not influence the in vitro metabolism of docetaxel even at concentrations as high as 1 µg/ml and that the major metabolic route to R102207 is cytochrome-P450-unrelated. Clearly, additional experiments are needed to establish the relevance of this principle in humans and to determine for what drugs it will apply. In addition, when given in combination with docetaxel, biologically relevant R101933 concentrations could be achieved and sustained for several hours, simulating optimal pharmacological conditions required for complete reversal of the MDR phenotype in in vitro systems.

Clinically, we observed that single treatment with R101933 given p.o. at the tested dosages was associated with minimal toxicity. The toxicological profile of the combination appeared to be very similar to that reported for docetaxel alone and included neutropenic fever and mucositis as the principal DLTs. Febrile neutropenia requiring hospitalization has been reported in ~15% and severe mucositis in ~10% of cases treated with docetaxel alone (29). In fact, the incidence of neutropenia observed with other inhibitors of P-glycoprotein in studies with anticancer drugs is greater than that observed with the cytotoxic agent alone (24). Fatigue was often mentioned by the patients in this study as a side effect, but never after R101933 alone, and asthenia is also a known side effect of docetaxel.

In conclusion, we have shown that the studied combination of docetaxel with R101933 constitutes a new approach for improving anticancer efficacy.
of oral R101933 and i.v. docetaxel is safe and at the achieved dose levels, lacks the significant kinetic interaction with the anticancer drug as observed previously with other modulators. In the case of a Phase II/III study with the combination of R101933 and docetaxel, 100 mg/m², and in view of the pharmacokinetic data on R101933 presently presented, the recommended dose of R101933 will be 200 mg b.i.d. p.o.

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