A Phase I Trial of Doxorubicin, Paclitaxel, and Valspodar (PSC 833), a Modulator of Multidrug Resistance

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ABSTRACT

Purpose: P-glycoprotein is an efflux pump for many drugs including doxorubicin and paclitaxel. This study evaluated the coadministration of these drugs with the P-glycoprotein inhibitor valsapar (PSC 833) with the aim of determining: (a) maximum tolerated doses (MTDs) of doxorubicin followed by paclitaxel (DP); (b) the MTD of DP combined with PSC 833 (DPV), without and with filgrastim (G-CSF); and (c) the pharmacokinetic interactions of PSC 833 with doxorubicin and paclitaxel.

Experimental Design: For the first cycle, patients received doxorubicin as a 15-min infusion followed by paclitaxel as a 1-h infusion. For the second cycle, patients received reduced doses of DP with PSC 833 at 5 mg/kg p.o., four times a day for 12 doses.

Results: Thirty-three patients with various refractory malignancies were enrolled and assessable. The MTD of DP without PSC 833 was 35 mg/m² doxorubicin and 150 mg/m² paclitaxel. The MTD of DPV with G-CSF was 12.5 mg/m² doxorubicin and 70 mg/m² paclitaxel. The dose-limiting toxicity for both DP and DPV was neutropenia without thrombocytopenia. With G-CSF, the MTD for DPV was 20 mg/m² doxorubicin and 90 mg/m² paclitaxel. No grade 4 nonhematological toxicities were observed. Five partial and two minor tumor remissions were observed. Paired pharmacokinetics with and without PSC 833 revealed substantial drug interactions with both doxorubicin and paclitaxel.

Conclusions: PSC 833 can be administered safely with doxorubicin and paclitaxel. The pharmacokinetic profiles of these drugs are significantly affected by PSC 833, requiring ~60% dose reductions for equivalent degrees of myelosuppression.

INTRODUCTION

P-gp is a transmembrane efflux pump for many drugs including anticancer agents such as vincristine, anthracyclines, taxanes, and epidophyllotoxins. Encoded by the MDR1 gene, this multidrug transporter is highly expressed in many clinically resistant tumors at diagnosis and/or relapse. Expression of MDR1 is an adverse prognostic factor in several cancers (3–10), which suggests that P-gp-mediated drug resistance may contribute to a significant portion of treatment failures (11–13). Several noncytotoxic drugs such as verapamil, phenothiazines, and cyclosporines have been shown to modulate this mechanism of MDR by inhibition of P-gp function (14–22).

PSC 833 is a nonimmunosuppressive, nonnephrotoxic analog of cyclosporine and is ~2- to 10-fold more potent than cyclosporine in its ability to inhibit P-gp (23–24). PSC 833 alters the pharmacokinetics of etoposide causing a decrease in CL, doubling of plasma t½, and an increase in the dose-normalized plasma AUC (25–27). Significant pharmacokinetic effects are also observed when PSC 833 is given in combination with paclitaxel (28) or doxorubicin (29–30). These effects are in part a consequence of inhibition by PSC 833 of endogenous P-gp, which is expressed at high levels in the biliary canaliculi, renal tubules, and intestinal lumen. The drug interactions necessitate dose reductions of the chemotherapeutic agents to avoid excessive toxicity.

PATIENTS AND METHODS

Patient Selection

Patients were eligible if they had a pathological confirmation of incurable malignancy and had measurable disease. No chemotherapy for 3 weeks prior to study entry (6 weeks for nitrosoureas) or no radiation therapy for 4 weeks prior to study entry was permitted. Patients were required to have an Eastern

1 The abbreviations used are: P-gp, P-glycoprotein; MDR, multidrug resistance; PSC 833, valsapar; MTD, maximum tolerated dose; DP, doxorubicin followed by paclitaxel; DPV, DP combined with PSC 833; G-CSF, filgrastim; AUC, area under the curve; StD, stable disease; LVEF, left ventricular ejection fraction; HPLC, high-performance liquid chromatography; MRT, mean residence time.
Cooperative Oncology Group performance status 0–2 and an anticipated survival of >3 months. Adequate renal function (creatinine < 1.5 mg/dL) and hepatic function (serum bilirubin, <1.0 mg/dL, serum aspartate amino transferase <3 × upper limit of normal) was required. Normal bone marrow function was required as indicated by white blood count greater than 3,500 per mm³ and platelet count greater than 100,000 per mm³. Prior doxorubicin treatment was allowed but could not exceed a cumulative dose of 300 mg/m². LVEF was required to be ≥45% at rest by radionuclide ventriculogram. Patients with active central nervous system metastases were excluded. The protocol was approved by the Panel on Medical Human Subjects of Stanford University.

Chemotherapy Administration

To obtain paired pharmacokinetic sampling data, patients were hospitalized in the Clinical Research Center of Stanford University Medical Center. Approximately 30 min prior to chemotherapy, all of the patients received i.v. premedication with dexamethasone 20 mg, famotidine 20 mg, and diphenhydramine 25 mg to prevent hypersensitivity reactions to paclitaxel, as well as an antiemetic (ondansetron 10 mg i.v.) prior to the administration of doxorubicin.

Patients received doxorubicin by i.v. 15-min infusion, followed by paclitaxel 1 h i.v. infusion, for the first cycle. Three weeks later, patients received reduced doses of doxorubicin and paclitaxel (as described below) in combination with PSC 833. PSC 833 (provided by Novartis Pharmaceuticals Corporation, East Hanover, New Jersey) was administered at 5 mg/kg p.o. four times daily for a total of 12 doses using the liquid formulation. If significant toxicity particular to PSC 833 occurred (i.e., grade 3 ataxia), the PSC 833 dose was reduced to 4 mg/kg. Chemotherapy was administered on day 2, after the fifth or sixth dose of PSC 833.

Patients continued to receive this combination of doxorubicin, paclitaxel, and PSC 833 every 3 to 4 weeks, with assessment of toxicity and tumor response. In the absence of grade 3 or 4 neutropenia, or other grade 3 toxicity, the dosage of chemotherapy was increased stepwise, to ensure that the patient was receiving a therapeutic dose. If grade 3 or 4 neutropenia occurred the doses remained fixed for that patient. If grade 3 or 4 toxicity occurred (excluding neutropenia, hyperbilirubinemia, and nausea/vomiting), the patient was removed from the study or treated at a lower dose of chemotherapy, as agreed on by patient and investigator. On documentation of disease progression, or StD after six cycles of the combination treatment, the patient was removed from the study.

Study Design

Separate dose escalation cohorts were planned for cycle 1 (DP) versus cycle 2 (DPV), as further described. If one patient among the first three in a cohort developed grade 3 or 4 toxicity (excluding neutropenia, hyperbilirubinemia, and nausea/vomiting), the cohort was expanded to six. If two of these six patients developed grade 3 or 4 toxicity (exclusions as noted above), this represented dose-limiting toxicity, and the maximal tolerated dose was defined as the next lower dose level.

Because grade 4 neutropenia represents a significant risk to patients, but is no longer usually considered a dose-limiting toxicity, we decided that a >50% incidence of grade 4 neutropenia was unacceptable, even if there were no other dose-limiting toxicities in the cohort. Grade 4 neutropenia was, therefore, handled as follows. If any of the three patients in a cohort of three had grade 4 neutropenia, the cohort was expanded from three to six. If one to three of six patients in a cohort of six had grade 4 neutropenia, without fever, the next cohort was enrolled and treated at the next dose level. If 4 of 6 patients had grade 4 neutropenia, the cohort was expanded to 8 or 10 patients (see cohort I of DP and cohort III of DPV).

Once the maximally tolerated dose of doxorubicin and paclitaxel (DP) with PSC 833 was determined, we proceeded to establish the maximally tolerated dose with G-CSF (Neupogen; Amgen, Thousand Oaks, CA) support. Patients were treated at the maximally tolerated dose of DP for one cycle, and then, on cycle 2, received doxorubicin, paclitaxel, and PSC 833 (DPV) with G-CSF support, at one dose level higher than that previously established as the maximally tolerated dose without G-CSF support. This was based on our finding that the dose-limiting toxicity with DPV is neutropenia. Patients were enrolled in cohorts of three, until the maximally tolerated dose was determined.

Dose levels (all doses are in mg/m²) were as follows. The initial starting doses for DP alone were 40 and 175 mg/m² and for DPV were 10 and 70 mg/m², respectively. Because of dose-limiting neutropenia observed in the first cohort of patients receiving DP (40/175), the second cohort was treated at a lower dose level of 35/150, respectively. For DPV, dose escalation entailed doses of 12.5/70 and then 15/80.

Dose escalation by cohort and determination of the MTD was based on dose-limiting toxicities observed during the first cycle of DP or DPV, respectively. After cycle 2, a patient whose nadir showed only grade 1 or 2 neutropenia received a higher dose of chemotherapy, with the increase according to the scheme presented above to ensure that patients were not undertreated.

Assessment of Toxicities and Antitumor Response

Quantitative hematological and nonhematological toxicities were assessed by the common toxicity criteria of the National Cancer Institute (36). Patients were monitored closely and nadir counts checked on days 11 and 15. Imaging studies were performed after every two cycles of treatment.

Although antitumor efficacy was not a primary objective of the study, tumor response rate and time to progression were recorded. Partial remission was defined as a 50% reduction in the sum of products of the two largest perpendicular diameters of all of the measurable lesions for at least 4 weeks and with no new lesion or progression of assessable disease. Minor response was defined as partial regression of disease of <50%. StD was measured not fulfilling the criteria for response or progression and lasting at least 4 months. Tumor progression was defined as a 25% or greater increase in tumor size measured as the bidimensional product of perpendicular diameters. The time to progression was measured from the time of first study drug administration to documented progressive disease.

Cardiac function was monitored in all of the patients. All of the patients had a baseline radionuclide ventriculogram.
Pharmacokinetic Analyses

Standard, noncompartmental pharmacokinetic calculations of parameters from doxorubicin and paclitaxel concentration-time data were performed using the XLPHARM M-IND program (Dr. V. K. Piotrovskij, VKPharmacokinetics, Turnhout, Belgium) implemented on Microsoft Excel (version 5.0, Microsoft Corporation, Redmond, WA; Ref. 40). The AUC was normalized for differences in the delivered dose by dividing the AUC by the dose administered in mg/m². Volume of distribution at steady state (Vss) and clearance were derived from traditional noncompartmental equations in which:

\[ V_{ss} = \text{Dose} \times \frac{AUMC}{AUC} - \frac{\text{Dose} \times T}{2 \times AUC} \]

where \( T \) is the duration of the infusion, clearance is the dose divided by the plasma AUC, and \( AUMC \) is the area under the first moment curve. The peripheral volume of distribution (\( V_p \)) was calculated by subtracting the central volume of distribution (\( V_c \)) from \( V_{ss} \). The MRT was calculated using the equation:

\[ MRT = \frac{AUMC}{AUC} \times \frac{T}{2} \]

where \( T \) is the duration of drug administration. Time above threshold concentrations for paclitaxel (>0.05 or >0.025 \( \mu \)M) were determined by simulating the decay of a terminal first-order time point concentration (\( C_{p1} - C_{p2} \)), using the derived terminal elimination rate constant (\( K_e \)) value, fitted to the equation:

\[ C_{p2} = C_{p1} \times \exp(-K_e \times T) \]

using Scientist 2.0 (MicroMath Scientific Software, Salt Lake City, UT).

Statistical Considerations

Pharmacokinetic data were tested using the Wilk-Shapiro statistic and were found to be abnormally distributed (41). Differences in pharmacokinetic data and parameters between the doxorubicin or paclitaxel groups in the absence and presence of treatment with PSC 833 were compared using the Wilcoxon signed rank test. The \( a \) priori level of significance was \( P = 0.05 \) (two-sided).

RESULTS

Patient Characteristics

Thirty-nine patients with advanced cancers were entered into the trial between June 1995 and June 1998. Six were excluded from this report because they did not receive DPV because of progressive disease (\( n = 3 \)), toxicity (\( n = 2 \)), or patient’s choice (\( n = 1 \)). The demographic characteristics of the 33 assessable patients are shown in Table 1. Disease categories of these patients were: ovarian cancer (\( n = 15 \)), lung cancer (\( n = 7 \)), sarcoma (\( n = 3 \)), and 1 each for germ cell, fallopian tube, breast, colon, neuroectodermal, peritoneal, vaginal, and

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(MUGA) at rest to determine the LVEF. Patients had a repeat study after every two cycles of treatment once the doxorubicin cumulative dose reached 300 mg/m². Patients were taken off study if there was a >10% decrease in the absolute LVEF or a decline in the resting ejection fraction to ≤45%.

Cerebellar ataxia has previously been shown to be a limiting toxicity for PSC 833 (12). This toxicity was graded as follows: grade 1, slight subjective sense of incoordination, without difficulty in walking and a normal exam; grade 2, definite subjective incoordination on walking but able to walk without assistance and, on physical examination, broad-based gait and/or mild dysdiadochokinesis and difficulty walking heel to toe; grade 3, unable to walk without assistance from another person or a walker and, on examination, a markedly abnormal gait and inability to walk heel to toe; grade 4, unable to walk with assistance.

For grade 3 or 4 ataxia, the PSC 833 dose was reduced to 4 mg/kg, and the patient was observed closely for resolution of toxicity. If grade 3 ataxia still persisted, the PSC 833 dose was held, and the patient restarted at 4 mg/kg. If grade 3 ataxia still persisted, the PSC 833 dose was further reduced to 3 mg/kg.

Pharmacokinetic Studies of Doxorubicin, Paclitaxel, and PSC 833

Pharmacokinetic studies of doxorubicin and paclitaxel were performed on 25 patients during their first cycle of doxorubicin and paclitaxel alone, and subsequently during their first cycle with PSC 833. Plasma and urine levels of these drugs were analyzed. Samples of venous blood (2 ml, one pediatric green top tube) were drawn and patients asked to collect all of the urine for 24 h after the initiation of the chemotherapy infusions, in 24-h aliquots. These samples were analyzed for doxorubicin, paclitaxel, and their respective metabolites by HPLC. For the first doxorubicin and paclitaxel cycle, blood samples were drawn just prior to the chemotherapy infusion, and then 0.5, 1, 2, 3, 4, 6, 8, 12, 18, 24, 36, and 48 h after initiation of the paclitaxel infusion.

Venous sampling during the combined treatment with PSC 833 was similar to that for the chemotherapy alone with the addition of limited sampling for peak and trough PSC 833 values. Whole blood PSC 833 concentrations were determined by simulating the decay of a terminal first-order time point concentration (\( C_{p1} - C_{p2} \)), using the derived terminal elimination rate constant (\( K_e \)) value, fitted to the equation:

\[ C_{p2} = C_{p1} \times \exp(-K_e \times T) \]

using Microsoft Excel (Ref. 40). The AUC was normalized for differences in the delivered dose by dividing the AUC by the dose administered in mg/m². Volume of distribution at steady state (Vss) and clearance were derived from traditional noncompartmental equations in which:

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where \( T \) is the duration of the infusion, clearance is the dose divided by the plasma AUC, and \( AUMC \) is the area under the first moment curve. The peripheral volume of distribution (\( V_p \)) was calculated by subtracting the central volume of distribution (\( V_c \)) from \( V_{ss} \). The MRT was calculated using the equation:

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where \( T \) is the duration of drug administration. Time above threshold concentrations for paclitaxel (>0.05 or >0.025 \( \mu \)M) were determined by simulating the decay of a terminal first-order time point concentration (\( C_{p1} - C_{p2} \)), using the derived terminal elimination rate constant (\( K_e \)) value, fitted to the equation:

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Phase I of Doxorubicin, Paclitaxel, and PSC 833

Table 1  Demographic and clinical characteristics of the 33 patients registered on this trial

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Cohort I</th>
<th>Cohort II</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td>25 female, 8 male</td>
<td>25 female, 8 male</td>
</tr>
<tr>
<td>Median age (range)</td>
<td>53 yr (33–76)</td>
<td>53 yr (33–76)</td>
</tr>
<tr>
<td>Mean prior chemotherapy regimens</td>
<td>3.4</td>
<td>3.4</td>
</tr>
<tr>
<td>Tumor types</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ovarian</td>
<td>15</td>
<td>15</td>
</tr>
<tr>
<td>Lung</td>
<td>7</td>
<td>7</td>
</tr>
<tr>
<td>Sarcoma</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>Other</td>
<td>8</td>
<td>8</td>
</tr>
<tr>
<td>PS, ECOG scale</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PS 0</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>PS 1</td>
<td>12</td>
<td>12</td>
</tr>
<tr>
<td>PS 2</td>
<td>3</td>
<td>3</td>
</tr>
</tbody>
</table>

*PS, performance status; ECOG, Eastern Cooperative Oncology Group.

Table 2  Hematological toxicities of doxorubicin and paclitaxel coadministered without PSC 833

The MTD of doxorubicin and paclitaxel without PSC 833 was established as shown in cohort II based on the experience with the first 10 patients, of whom 5 had grade 4 neutropenia and two had fever with neutropenia. Subsequently, 16 more patients were enrolled at the MTD for a total of 26 patients.

Table 3  Nonhematological toxicities (grade 2 and 3) recorded during the first cycle of treatment with the combination of doxorubicin and paclitaxel, without PSC 833

Table 4  Hematological toxicities recorded during the first cycle of treatment with the combination of doxorubicin, paclitaxel, and PSC 833

Table 5  Hematological toxicities recorded during the first cycle of treatment with the combination of doxorubicin, paclitaxel, and PSC 833

The toxicities for the two dose levels of DP alone are shown in Tables 2 and 3. Seven patients were entered on the starting dose of 40 and 175 mg/m², and six of seven experienced grade 4 neutropenia. Although there were no cases of fever with neutropenia at this dose level, the high rate of grade 4 neutropenia was considered unacceptable without the use of G-CSF. The next dose level was reduced to 35 and 150 mg/m², and this was defined as the MTD because 5 of 10 patients had grade 4 neutropenia with 2 of 10 also experiencing fever. Subsequently, 16 more patients were entered at this dose level. Overall at this dose, 12 of 26 patients had grade 4 neutropenia and 11 of 26 had grade 3 neutropenia. Five of these 26 patients had fever associated with the neutropenia.

Thymic cancers. The median age was 53 years (range, 33–76 years). The patients were pretreated with a mean of 3.4 regimens (excluding hormonal and radiation therapy) prior to study entry. There were 25 women and 8 men, reflecting the relatively large number of patients with ovarian cancers.

Determination of MTD

Doxorubicin and Paclitaxel without PSC 833. The toxicities for the dose levels of DP alone are shown in Tables 2 and 3. Seven patients were entered on the starting dose of 40 and 175 mg/m², and six of seven experienced grade 4 neutropenia. Although there were no cases of fever with neutropenia at this dose level, the high rate of grade 4 neutropenia was considered unacceptable without the use of G-CSF. The next dose level was reduced to 35 and 150 mg/m², and this was defined as the MTD because 5 of 10 patients had grade 4 neutropenia with 2 of 10 also experiencing fever. Subsequently, 16 more patients were entered at this dose level. Overall at this dose, 12 of 26 patients had grade 4 neutropenia and 11 of 26 had grade 3 neutropenia. Five of these 26 patients had fever associated with the neutropenia.

Doxorubicin and Paclitaxel with PSC 833. The toxicities observed for DPV without and with G-CSF are presented in Tables 4 and 5. The MTD of this combination was 12.5 mg/m² doxorubicin and 70 mg/m² paclitaxel with three of six patients experiencing grade 4 neutropenia, and two with both fever and neutropenia (Table 4). No significant thrombocytopenia (platelets <100,000/mm³) was noted in cohorts I through III. It is evident from Tables 2 and 4 that cohort I of DP produced a degree of myelosuppression very similar to that of cohort III of DPV, with mean neutrophil nadir counts of 290 and 350/mm³, respectively. Likewise, cohort II of DP was similar in myelosuppression to cohort II of DPV (Tables 2 and 4), with mean neutrophil nadir counts of 930 and 1,000/mm³, respectively.

The next twelve patients (cohorts IV and V) were treated with escalating dose of DPV with G-CSF support. DPV was well tolerated by six patients at doxorubicin 20 mg/m² and paclitaxel 80 mg/m² with one (of six) grade 3 and one (of six) grade 4 thrombocytopenia without evidence of bleeding. Only one grade 4 neutropenia was observed. The use of G-CSF resulted in higher nadir counts (mean/median 2,000/mm³) compared with the earlier cohorts. Six patients were treated at the next dose level of 20/90 (cohort V) and this dose defined the MTD because two of six patients had grade 4 neutropenia with fever and two of six patients experienced grade 3 or 4 thrombocytopenia, with a total of three of the six patients experiencing dose-limiting toxicities (Table 4).
Table 5  Nonhematological toxicities (grade 2 and 3) recorded during the first cycle of treatment with the combination of doxorubicin, paclitaxel, and PSC 833

<table>
<thead>
<tr>
<th>Doses of doxorubicin and paclitaxel, mg/m²</th>
<th>Cohort I</th>
<th>Cohort II</th>
<th>Cohort III</th>
<th>Cohort IV</th>
<th>Cohort V</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of patients</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>Ataxia</td>
<td>3</td>
<td>4</td>
<td>7</td>
<td>3</td>
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<tr>
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<td>1</td>
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<tr>
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<td>2</td>
<td>3</td>
<td>0</td>
<td>1</td>
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<tr>
<td>Fatigue</td>
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<td>0</td>
<td>2</td>
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<td>Hyperbilirubinemia</td>
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<td>3</td>
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<tr>
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<td>0</td>
<td>2</td>
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<tr>
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<tr>
<td>Neuropathy</td>
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</table>

*No ataxia was noted during treatment cycles without valspodar.

Table 6  Cerebellar ataxia by cohorts of patients treated with doxorubicin, paclitaxel, and PSC 833

<table>
<thead>
<tr>
<th>Grade</th>
<th>Cohort I</th>
<th>Cohort II</th>
<th>Cohort III</th>
<th>Cohort IV</th>
<th>Cohort V</th>
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<tbody>
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<td>4</td>
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</table>

*No ataxia was noted during treatment cycles without valspodar.

Toxicities

The most frequent toxicity for both DP and DPV was hematological as shown in Tables 2 and 4. Grade 4 neutropenia was observed in 18 of 33 patients treated with DP alone and 12 of 20 patients treated with DPV. Fever with neutropenia was observed in 5 of 33 patients on DP and 5 of 20 treated with DPV. Two patients developed fever and neutropenia despite prophylactic G-CSF in cohort V, Table 4. Significant thrombocytopenia (<50,000/mm³) was seen in two of six patients in cohort IV and in three of six patients in cohort V.

Nonhematological toxicities (grades 2 and 3) for DP alone are shown in Table 3 and for DPV in Table 5. No grade 4 toxicities were observed. Transient reversible hyperbilirubinemia was an anticipated side effect of PSC 833 (41) and was seen in 15 of 33 patients. One patient developed a cardiac arrhythmia that required treatment. Other reversible side effects observed were nausea/vomiting (n = 5 for DP; n = 8 for DPV), constipation (n = 1 for DP; n = 8 for DPV), fatigue (n = 6 for DP; n = 7 for DPV), and skin rash (n = 2 for DP and n = 1 for DPV).

Cerebellar neurotoxicity attributed to PSC 833 (42) is shown in Table 6 for each of the cohorts studied. In 9 patients, PSC 833 produced significant (grade 3), but rapidly reversible, ataxia requiring dose reduction of PSC 833 to 4 mg/kg. Two patients required a dose reduction to 3 mg/kg of PSC 833. No grade 4 ataxia was noted.

Antitumor Responses

Five partial responses were noted among 33 assessable patients (non-small cell lung, 12, 6, and 4 months; ovarian cancer, 5 months; and fallopian tube cancer, 11 months), as well as two minor responses (ovarian cancer, 6 months; mediastinal germ cell tumor, 4 months). Six patients had StD (three ovarian cancers and one sarcoma for 4 months, two non-small cell lung cancers for 6 and 9 months, respectively).

Pharmacokinetic Evaluations

The median peak whole blood PSC 833 level in 15 patients evaluated was 3090 ng/ml (range, 1460–7020 ng/ml) with a median trough level of 3400 ng/ml (1460–5500 ng/ml). Paired pharmacokinetic analyses with and without PSC 833 were performed in 18 courses of doxorubicin and in 16 courses of paclitaxel. PSC 833 resulted in substantial drug interaction with both doxorubicin and paclitaxel. In the 18 patients with pharmacokinetic analyses of doxorubicin, an average dose reduction of 65% resulted in an actual doxorubicin AUC decline of 35% during PSC 833 treatment. This was primarily a reflection of a 57% reduction in doxorubicin peak concentration at the end of drug administration. However, PSC 833 treatment resulted in an 86% increase in the dose-normalized AUC of doxorubicin, which correlated with a 44% decrease in CL, a 77% increase in the terminal phase τ_{1/2}, and an 84% increase in MRT (Table 7).

Despite the lower actual AUC values, the longer τ_{1/2} and MRT values indicate that coadministration with PSC 833 results in a prolonged low-level exposure to the drug as illustrated in the concentration-time profile for a patient receiving doxorubicin ± PSC 833 in Fig. 1. PSC 833 had a greater effect on doxorubicin AUC, causing an increase of 259% for this metabolite.

In 16 patients with pharmacokinetic analyses of paclitaxel, coadministration of PSC 833 treatment resulted in a 158% increase in terminal PAU (V_{tu}, a 213% increase in (V_{tu}), a 96% increase in the terminal phase τ_{1/2}, and a 31% increase in time >0.025 mm concentration (Table 8). There was no apparent effect on the AUC of paclitaxel. However, the longer τ_{1/2} and MRT values indicate that PSC 833 prolongs the duration of exposure to paclitaxel as illustrated in the typical concentration-time profile for a patient receiving paclitaxel with and without PSC 833 in Fig. 2. PSC 833 had a greater effect on 6-OH-paclitaxel, causing a 222% increase in the AUC of this metabolite.

DISCUSSION

MDR1 gene expression is seen in a variety of malignancies, both at initial diagnosis and at time of relapse (1, 12), and is one of the mechanisms implicated in clinical drug resistance. Modulation of MDR with inhibitors of P-gp may improve the efficacy of chemotherapy (12, 22). Doxorubicin and paclitaxel are substrates of P-gp (the protein encoded by the MDR1 gene) and are active agents in many solid tumors. Therefore, modulation of resistance to these drugs is an attractive concept. A large number of clinical trials of reversal of MDR have been undertaken with drugs that are relatively weak P-gp inhibitors, often producing limiting toxicities at doses below those necessary to inhibit MDR (11–12). PSC 833 is 2- to 10-fold more potent than...
its parent compound cyclosporine for modulating MDR in vitro and in vivo (23–24). In this study, target levels of PSC 833 capable of MDR1 modulation (>1000 ng/ml) were achieved in all of the 15 patients thus evaluated.

On the basis of our previous experience and those of others using these agents, we anticipated significant pharmacokinetic interactions between PSC 833 and the MDR-related agents, and, therefore, used lower initial doses of doxorubicin and paclitaxel when given with PSC 833 (26, 29, 30, 43). The MTD of DP alone was 35 mg/m² and 150 mg/m² respectively. The addition of PSC 833 to DP resulted in substantial hematological toxicity even at the lowered doses, and, as a result, the MTD of the combination was 12.5 mg/m² doxorubicin and 70 mg/m² paclitaxel. This degree of dose reduction is not surprising. Previous studies with paclitaxel or doxorubicin when administered as single agents combined with cyclosporine or PSC 833 also produced marked alterations in pharmacokinetics, necessitating 50 and 60% dose reductions, respectively (12, 28–30). With the addition of G-CSF, we were able to escalate doses of doxorubicin only marginally because of thrombocytopenia. The MTD of the combination with G-CSF support was 20 mg/m² doxorubicin and 90 mg/m² paclitaxel.

The formulation of paclitaxel contains cremophor EL, which has been shown to cause alterations in the pharmacokinetics of the drug. In this study, the addition of PSC 833 significantly prolonged the plasma t½ and MRT of both doxorubicin and paclitaxel. The observed increase in myelosuppression when PSC 833 was combined with doxorubicin and paclitaxel is most likely related to these pharmacokinetic effects. The pharmacokinetic interactions of these drugs are consistent with clinical, laboratory, and animal model observations (26, 44–45) and corroborate the need for dose reduction of doxorubicin and paclitaxel when they are combined with PSC 833.

Previously, Gianni et al. (46) reported a correlation of the degree of neutropenia to the duration of paclitaxel concentration above 0.05 μM. In paired analyses, we found no difference in time above 0.05 μM of paclitaxel for DP alone versus DPV, despite the fact that the dose of paclitaxel was 2-fold higher in the DP cycle. The prolongation of the elimination phase of paclitaxel by PSC 833 was manifested by a significant increase in time above 0.025 μM for paclitaxel in the DPV cycle. These effects of PSC 833 on the disposition and distribution of paclitaxel are likely to have contributed to the relatively high myelosuppression seen with the low dose of doxorubicin and paclitaxel in DPV.

Table 7  Effect of PSC 833 on pharmacokinetic (PK) parameters of doxorubicin in patients (n = 18) treated sequentially over two cycles with DP and then with DPV

The data are means (±SD).

<table>
<thead>
<tr>
<th>PK parameter</th>
<th>DP</th>
<th>DPV</th>
<th>% change</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dose, μmol/m²</td>
<td>63 (4)</td>
<td>22 (4)</td>
<td>-65</td>
<td>0.0002</td>
</tr>
<tr>
<td>Cmax, μM</td>
<td>0.17 (0.05)</td>
<td>0.07 (0.03)</td>
<td>-57</td>
<td>0.0003</td>
</tr>
<tr>
<td>t½, h</td>
<td>23 (7)</td>
<td>40 (13)</td>
<td>+77</td>
<td>0.0004</td>
</tr>
<tr>
<td>AUC, h·μM</td>
<td>1.6 (0.5)</td>
<td>1.1 (0.4)</td>
<td>-35</td>
<td>0.0006</td>
</tr>
<tr>
<td>AUC per μmol dose</td>
<td>2.6 (0.7)</td>
<td>4.8 (1.5)</td>
<td>+86</td>
<td>0.0003</td>
</tr>
<tr>
<td>CL, liter/h/m²</td>
<td>42 (12)</td>
<td>23 (10)</td>
<td>-44</td>
<td>0.0002</td>
</tr>
<tr>
<td>Vss, liter/m²</td>
<td>1149 (556)</td>
<td>1078 (330)</td>
<td>-6</td>
<td>0.7</td>
</tr>
<tr>
<td>MRT, h</td>
<td>27 (8)</td>
<td>50 (17)</td>
<td>+84</td>
<td>0.0004</td>
</tr>
<tr>
<td>AUC, h·μM</td>
<td>1.3 (0.6)</td>
<td>1.6 (0.6)</td>
<td>+18</td>
<td>0.13</td>
</tr>
<tr>
<td>AUC/μmol dose</td>
<td>0.03 (0.01)</td>
<td>0.05 (0.02)</td>
<td>+259</td>
<td>0.0002</td>
</tr>
</tbody>
</table>

*Pharmacokinetics are based on 48 h of sampling after doxorubicin infusion. Analyses are from 18 patients receiving 18 courses of DP and 18 courses of DPV, without G-CSF.

Ps are Wilcoxon rank-sum test, two-tailed, doxorubicin alone versus doxorubicin plus PSC 833.

CL, clearance; Vss, volume of distribution at steady state.

Fig. 1  Drug concentration and time plots of doxorubicin (D) and doxorubicinol (DOL) in the same patient treated sequentially without and with PSC 833 (PSC), with 96-h sampling. The doses of doxorubicin were 35 mg/m² without and 20 mg/m² with PSC 833.
netics of the doxorubicin (50, 51, 52). In this study, because the paclitaxel dose was lowered with PSC 833, the decreased amount of cremophor EL would have had minimal effects during the PSC 833 cycle.

Recently, Gianni et al. (53), using a schedule of administration similar the one in our study, i.e., a bolus of doxorubicin, followed 15 min later with a 3-h infusion of paclitaxel, found a modest interaction when doxorubicin administration preceded paclitaxel. These effects were observed during the β \( t_{1/2} \) phase with no significant change in the terminal (γ) \( t_{1/2} \), which suggests that the observed changes could be attributable to alterations in pharmacokinetics when the paclitaxel vehicle, cremophor EL, was at its maximal concentrations. In contrast, we found significant increase in dose-normalized AUC (86%) and terminal elimination \( t_{1/2} \) (77%), as well as other parameters (Table 7). Similar findings were observed with the major metabolite doxorubicinol. These data support the contention that cremophor EL did not contribute significantly to the observed effects on doxorubicin pharmacokinetics.

Paclitaxel was well tolerated as a 1-h infusion as has been reported previously (54). The main toxicity of both DP and DPV was hematological (Tables 2 and 4). PSC 833 caused a reversible grade 3 ataxia in nine patients, which required dose reduction to 4 mg/kg in eight patients, and 3 mg/kg in two patients. The mechanism of the cerebellar toxicity of PSC 833 is unknown, but is probably not related directly to inhibition of P-gp, because some other inhibitors of P-gp including cyclosporine do not cause ataxia, and P-gp knockout mice are not ataxic (55). The transient hyperbilirubinemia produced by PSC 833 is similar to that of high-dose cyclosporine (42) and is likely attributable to inhibition of the canalicular multispecific organic anion transporter (cMOAT/MRP2; Refs. 56, 57), and/or other transporters of bilirubin and bilirubin glucuronide in the biliary canaliculi.

Tumor regression was observed in 7 of 33 patients and another 6 patients had stabilization of disease for at least 4 months. The design of this study did not allow us to assess what role MDR modulation may have played in these responding patients. Whether these responses are attributable to reversal of \( MDR1 \) or intrinsic sensitivity to doxorubicin and paclitaxel cannot be determined from the design of this Phase I trial. Moreover, treatment of P-gp-negative tumors with inhibitors of P-gp may result in clinical benefit by preventing the development of MDR. This has been demonstrated in a laboratory model of a human sarcoma cell line, in which PSC 833 reduced the mutation rate for doxorubicin resistance significantly, and suppressed the activation of \( MDR1 \) (58). These findings suggest that treatment of drug-sensitive, P-gp-negative malignancies with effective MDR modulation may suppress the emergence of resistant \( MDR1 \) positive subclones and result in improved outcome (1, 14).

### Table 8  Effect of PSC 833 on pharmacokinetic (PK) parameters of paclitaxel in patients (n = 16) treated sequentially with DP and DPV

<table>
<thead>
<tr>
<th>PK Parameter</th>
<th>DP</th>
<th>DPV</th>
<th>% change</th>
<th>( P^b )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dose, ( \mu \text{mol/m}^2 )</td>
<td>188 (15)</td>
<td>86 (6)</td>
<td>-55</td>
<td>0.0003</td>
</tr>
<tr>
<td>( C_{\text{max}} ), ( \mu \text{M} )</td>
<td>10.6 (6.4)</td>
<td>3.6 (2.8)</td>
<td>-66</td>
<td>0.0008</td>
</tr>
<tr>
<td>( t_{1/2} ), h</td>
<td>8 (4)</td>
<td>16 (9)</td>
<td>+96</td>
<td>0.0003</td>
</tr>
<tr>
<td>AUC, h*( \mu \text{M} )</td>
<td>20 (9)</td>
<td>8 (4)</td>
<td>-58</td>
<td>0.0003</td>
</tr>
<tr>
<td>AUC per ( \mu \text{mol dose} )</td>
<td>0.10 (0.04)</td>
<td>0.10 (0.04)</td>
<td>0</td>
<td>0.5</td>
</tr>
<tr>
<td>CL, liter/h/m(^2)</td>
<td>10.7 (3.4)</td>
<td>11.9 (4.7)</td>
<td>+11</td>
<td>0.5</td>
</tr>
<tr>
<td>( V_e ), liter/m(^2)</td>
<td>45 (26)</td>
<td>116 (66)</td>
<td>+158</td>
<td>0.0005</td>
</tr>
<tr>
<td>( V_v ), liter/m(^2)</td>
<td>30 (19)</td>
<td>97 (64)</td>
<td>+220</td>
<td>0.0003</td>
</tr>
<tr>
<td>( MRT, h )</td>
<td>4 (2)</td>
<td>11 (7)</td>
<td>+156</td>
<td>0.0003</td>
</tr>
<tr>
<td>( h &gt; 0.05 \mu \text{M} )</td>
<td>23 (10)</td>
<td>25 (12)</td>
<td>+9</td>
<td>0.5</td>
</tr>
<tr>
<td>( h &gt; 0.025 \mu \text{M} )</td>
<td>31 (13)</td>
<td>41 (19)</td>
<td>+31</td>
<td>0.014</td>
</tr>
<tr>
<td>6-OH-paclitaxel AUC, h*( \mu \text{M} )</td>
<td>1.7 (1.5)</td>
<td>5.3 (3.3)</td>
<td>+222</td>
<td>0.003</td>
</tr>
<tr>
<td>AUC per ( \mu \text{mol dose} )</td>
<td>0.01 (0.01)</td>
<td>0.06 (0.04)</td>
<td>+589</td>
<td>0.0005</td>
</tr>
</tbody>
</table>

\( ^a \) Analyses are from 16 patients receiving 16 courses of paclitaxel alone and 17 courses of paclitaxel plus PSC 833.  
\( ^b \) \( P \)s are Wilcoxon rank-sum test, two-tailed, paclitaxel without \( v e rsus \) paclitaxel with PSC 833.  
\( ^c \) CL, clearance; \( V_{\text{ss}} \), volume of distribution at steady state; \( V_p \), volume of peripheral compartment.
In conclusion, PSC 833 can be administered safely in combination with doxorubicin and paclitaxel with appropriate dose modification of the cytotoxic drugs. PSC 833 significantly increases the paclitaxel and doxorubicin exposure secondary to decreased clearance, accounting for the need to reduce doses ~2-fold to achieve equivalent myelosuppression. Additional Phase II/III studies are required to determine the clinical role of this intriguing combination.

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REFERENCES


# Clinical Cancer Research

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Ranjana Advani, George A. Fisher, Bert L. Lum, et al.


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