Tumor-Suppressive Effects of Neutral Endopeptidase in Androgen-independent Prostate Cancer Cells

Jie Dai, Ruoqian Shen, Makoto Sumitomo, Jonathan S. Goldberg, Yiping Geng, Daniel Navarro, Su Xu, Jason A. Koutcher, Mark Garzotto, C. Thomas Powell, and David M. Nanus

Department of Urology, Urological Oncology Research Laboratory [J. D., R. S., M. S., J. S. G., Y. P., D. N., D. M. N.], and Department of Medicine, Division of Hematology and Medical Oncology [D. M. N.], Joan and Stanford I. Weill Medical College of Cornell University, and Departments of Medical Physics and Radiology [S. X., J. A. K.], and the Department of Urology [M. G., C. T. P.], Memorial Sloan-Kettering Cancer Center, New York, New York 10021

ABSTRACT

Expression of neutral endopeptidase (NEP) 24.11 is diminished in metastatic, androgen-independent prostate cancers (PCs; C. N. Papandreou et al., Nat. Med., 4: 50–57, 1998). To determine the effects on androgen-independent PC cells of overexpressing cell-surface NEP, an inducible tetracycline-regulatory gene expression system was used to stably introduce and express the NEP gene in androgen-independent TSU-Pr1 cells generating WT-5 cells, which expressed high levels of enzymatically active NEP protein when cultured in the absence of tetracycline. TN12 cells, which contain the identical vectors without the NEP gene and do not express NEP, were used as control. Expression of NEP in WT-5 cells after removal of tetracycline from the media resulted in a >80% inhibition in cell proliferation over a 1-week period (P < 0.005) compared with control cells. Tumor formation occurred in the prostate glands of orthotopically injected athymic mice killed at 30 days in 4 of 5 mice that were given injections of 2 × 10⁶ WT-5 cells and were fed doxycycline (NEP suppressed), and in all mice that were given injections of TN12 cells and were fed with or without doxycycline. In contrast, only 1 of 5 mouse prostates developed a tumor in mice that were given injections of WT-5 cells and that did not receive doxycycline. Analysis of the mechanisms of NEP-induced growth suppression revealed that NEP expression in WT-5 cells induced a 4-fold increase in the number of PC cells undergoing apoptosis, and increased the expression of p21 tumor suppressor gene protein and the level of unphosphorylated retinoblastoma protein as determined by Western blot. Flow cytometric analysis show that induced NEP expression in WT-5 cells resulted in a G₁ cell cycle arrest. These data show that NEP can inhibit PC cell growth and tumorigenicity and suggest that NEP has potential as therapy for androgen-independent PC.

INTRODUCTION

NEP³ 24.11 (neprilysin, enkephalinase, CD10, EC 3.4.24.11) is a M₉ 90,000–110,000 cell-surface metalloproteinase that is normally expressed by numerous tissues, including prostate, kidney, intestine, endometrium, adrenal glands, and lung. This enzyme cleaves peptide bonds on the amino side of hydrophobic amino acids and inactivates a variety of physiologically active peptides, including atrial natriuretic factor, substance P, bradykinin, oxytocin, Leu- and Met-enkephalins, neurotensin, bombesin, endothelin-1, and bombesin-like peptides (1–3). NEP reduces the local concentration of peptide available for receptor binding and signal transduction. The biological function of NEP appears to be organ-specific. In the central nervous system, NEP regulates enkephalin-mediated analgesia (4); in the kidney and vascular epithelium, the enzyme is involved in regulating levels of circulating atrial natriuretic factor (5); in the lung, NEP modulates tachykinins, such as substance P, that mediate inflammation (6); in the endometrium, NEP regulates endothelin-1, which causes vasoconstriction of endometrial arterioles during specific phases of the ovulatory cycle (7, 8). NEP has also been implicated in controlling cellular proliferation by hydrolyzing bombesin-like peptides, which are potent mitogens for fibroblasts and bronchial epithelial cells (9).

Loss or decreases in NEP expression have been reported in a variety of malignancies, including renal cancer (10), invasive bladder cancer (11), poorly differentiated stomach cancer (12), small cell and non-small cell lung cancer (13), endometrial cancer (14), and PC (15). Reduced NEP may promote peptide-mediated proliferation by allowing an accumulation of higher peptide concentrations at the cell-surface and may facilitate the development or progression of neoplasia (16, 17).

In PCs, NEP protein is expressed in androgen-sensitive LNCaP cells but not in androgen-independent PC cell lines (15). Furthermore, expression of NEP is transcriptionally activated by androgen in LNCaP cells and decreases with androgen-with-

³ The abbreviations used are: NEP, neutral endopeptidase; PC, prostate cancer; FBS, fetal bovine serum; Ab, antibody; mAb, monoclonal Ab; FAK, focal adhesion kinase.
MATERIALS AND METHODS

Cell Culture. PC cell lines were maintained in RPMI 1640 supplemented with 2 mM glutamine, 1% nonessential amino acids, 100 units/ml streptomycin and penicillin, and 10% FBS. TSUGK27 is a TSU-Prl cell line stably transfected with pGK hygro and PUHD 15–1 (4.4 kb) containing the coding sequence for the COOH-terminal domain of VP16 (named the tetracycline-responsive transactivator or (TA), downstream of the hCMV promoter (19). This cell line was cultured in media containing 150 μg/ml of hygromycin and 1 μg/ml of tetracycline.

Plasmid Construction and Gene Transfer. To construct the tetracycline-repressible NEP-expression vector, 1.7 kb SacII-XhoI and 1.6 kb XhoI-BamHI DNA fragments containing the entire wild-type NEP coding sequence, were isolated from the pCIShENK vector (provided by Arris Pharmaceutical Corp., San Francisco, CA) and ligated into the pTRE vector (Clontech Laboratories, Inc., Palo Alto, CA) at the SacII and XhoI sites to generate pwtNEP. The pTRE empty vector was used as control.

Enzyme Assays. Enzyme assays were performed to determine NEP-specific enzyme activity as described previously (20) using Suc-Ala-Ala-Phe-pNA (Bachem Bioscience, Inc., Philadelphia, PA) as substrate. Briefly, cells were rinsed in cold lysis buffer (50 mM Tris/150 mM NaCl) and lysed in lysis buffer containing 0.5% 3-(3-cholamidopropyl)dimethylammonio)-1-propanesulfonate (CHAPS), which did not affect NEP enzyme specific activity. Protein concentrations were measured using the Bio-Rad DC protein assay kit (Bio-Rad Laboratories, Hercules, CA). Thirty μl of cell membrane suspension were added to a mixture of 200 μl of 100 mM Tris-HCl (pH 7.6), 10 μl of 20 mM substrate (dissolved in DMSO), and 10 μl of aminopeptidase N enzyme solution (EC 3.4.11.2; Boehringer Mannheim, Indianapolis, IN) and were incubated at 37°C for 10 min. The reaction was stopped by adding 10% trichloroacetic acid, and the mixture was centrifuged at 2500 rpm × 5 min; 250 μl of supernatant was then removed for colorimetric analysis. The absorbance of the chromogen was immediately read at 540 nm against a reaction mixture without cell membrane as blank. Specific activities were expressed as pmol/μg protein/min and represent an average of two separate measurements performed in duplicate. The SE of measurement of independent duplicate experiments was ~10–20% of the mean value.

Cell Cycle Analysis. Flow cytometry analysis was performed as described previously (21). Briefly, cells were rinsed twice in cold PBS, trypsinized, washed twice with PBS, and fixed in cold ethanol overnight. The following day, cells were resuspended in 500 μl of PBS, digested with 20 μg/ml RNase at 37°C for 1 h, chilled on ice for 10 min, and stained with propidium iodide (50 μg/ml) by incubation for 1 h at room temperature in the dark. Cell cycle distribution was analyzed by flow cytometry using a Becton Dickinson FACS system.

Apoptosis Assay. Apoptotic cells stained with acridine orange and ethidium bromide were assessed by fluorescence microscopy. Briefly, 2 μl of stock solution containing 100 μg/ml acridine orange and 100 μg/ml ethidium bromide were added to 25 μl of cell suspension. Total number of cells, as well as apoptotic cells that showed shrinkage, blebbing, and/or apoptotic bodies, were counted. DNA fragmentation analysis was performed as described previously (22).

Protein Extraction, Immunoprecipitation, and Western Blot Analysis. Protein was extracted from exponentially growing cells and was analyzed by Western blotting as described previously (23) using primary antibodies 5B5 (anti-NEP; Arris Pharmaceutical Corp.), p21, retinoblastoma, Bcl-2, p53, and cyclins A and D (Oncogene Research Products, Cambridge, MA). Blots were incubated with enhanced chemiluminescent (ECL) detection reagents (Amersham Pharmacia BioTech, Arlington Heights, IL), and proteins were detected by autoradiography by exposure of blots to Kodak XAR film for 2–15 min. Membranes were stained with 0.2% Ponceau red to assure equal protein loading and transfer. Immunoprecipitation using mAb J5, which recognizes NEP (CD10; Beckman-Coulter Pharmaceutical, Inc., Hialeah, FL), was performed as described previously (24).

Orthotopic Injection. Male nude mice (25–30 g) were anesthetized with 300 μl of 5 mg/ml phentobarbital (10 μg body weight) administered i.p. The abdomen was steriley prepped with Betadine, and a 1.5-cm vertical incision was made in the middle line of the abdomen through the skin and peritoneum to expose the bladder and seminal vesicles. A 26-gauge
needle was inserted into the parenchyma of the dorsal lobe of the prostate and advanced just below the capsule, into which 2 × 10^6 tumor cells diluted in 0.1 ml of sterile PRMI1640–10% FBS were injected. The lack of significant extraprostatic leakage and formation of a visible bulla between prostate parenchyma and capsule were criteria for a successful injection. The incision was closed using metal clips. Dox-Diet was obtained from Bio Serve (Frenchtown, NJ).

**NMR Imaging Assay.** Mice were imaged on a 4.7T 33-cm bore Bruker Omega NMR system. Images were obtained using a homebuilt 4-turn foil solenoid radiofrequency coil tuned to 200 MHz. Image acquisition parameters included eight slices, field of view of 25 mm, slice thickness of 2 mm, 128 × 128, matrix (200 units in plane resolution), repetition interval (TR) of 500 ms, and echo time (TE) of 16 ms.

**RESULTS**

**Expression of NEP in TSU-Pr1 Cells.** TSUGK27 cells, a derivative of TSU-Pr1 cells that contain the tetracycline regulator and a hygromycin resistance plasmid (19), do not express NEP protein nor exhibit NEP enzyme activity. TSUGK27 cells were transfected with pwtNEP (or pTRE control vector) together with a neomycin resistance vector and antibiotic-resistant stable clones, screened by measuring NEP catalytic activity in cells cultured in the presence or absence of 1 μg/ml tetracycline (Fig. 1A). Removal of tetracycline from the medium resulted in a significant increase in NEP enzyme activity in at least four clones (WT5, WT6, WT12, and WT24). TSUGK27 cells, which contained the pTRE empty vector without the NEP gene (TN cells), did not exhibit enzyme activity. Integration of pTRE into genomic DNA of TN cell lines was verified by Southern blotting with a probe specific for pTRE vector sequence (not shown). Western blot of NEP protein immunoprecipitated with mAb J5, which recognizes NEP, showed the presence of NEP protein in WT5 and WT24 cells after the removal of tetracycline from the medium, similar to what was observed in LNCaP cells, which constitutively express NEP, but which was not observed in control TN12 or TN15 cells (Fig. 1B). Flow cytometry confirmed the expression of cell-surface NEP in WT cell lines (data not shown). WT5 cells were further characterized for NEP enzyme inducibility. NEP enzyme activity was suppressed when WT-5 cells were cultured in 1 μg/ml tetracycline, and enzyme activity increased with decreasing concentrations of tetracycline ranging from 1000 to 0 ng/ml (Fig. 1C). Enzyme activity reached maximum levels at 72–96 h after the removal of tetracycline from the medium (Fig. 1D). Thus, induction of NEP

---

**Fig. 1** Analysis of TSUGK27 cells stably transfected with pwtNEP. **A.** TSUGK27 cells stably transfected with pwtNEP were cultured in the presence or absence of 1 μg/ml tetracycline (Tet) for 48–72 h, and NEP-specific enzyme activity was determined. Data shown represent the average of one to three independent experiments for each clone. **Error bars,** SE. **B.** cells cultured in RPMI medium containing FCS and no tetracycline were assessed for NEP expression. Identical numbers of cells were lysed and immunoprecipitated with mAb J5, separated by SDS-PAGE, transferred to nitrocellulose, and probed with rabbit polyclonal Ab 5B5, which recognizes NEP. LNCaP cells, which constitutively express NEP, were used as control. **C.** enzyme activities of WT5 cells were determined after incubation in media containing various concentrations of tetracycline and (D) at various time points after removal of 1 μg/ml tetracycline from the cells. Representative data are shown from one experiment performed in duplicate on at least two separate occasions. Data are expressed as relative NEP activity with 100% equal to the average enzyme activity in cells cultured without tetracycline (C), or enzyme activity in cells 96 h after removal of tetracycline (D). **Error bars,** SE.
enzyme activity was both time course-dependent and dose-dependent.

**NEP Expression Induces Growth Inhibition in Vitro and Inhibits Tumorigenicity in Athymic Mice.** WT5 and control TN12 cells were cultured for 7 days with and without tetracycline (Tet), and the cell number was determined. Data are representative of one experiment performed in triplicate. Error bars, SE. 

Fig. 2  A. NEP expression induces growth inhibition in vitro. WT5 and TN12 were cultured for 7 days with and without tetracycline (Tet), and the cell number was determined. Data are representative of one experiment performed in triplicate. Error bars, SE. 

**B.** NMR imaging of mouse prostates into which PC cells were injected. High spatial resolution 1H NMR images of the pelvis were obtained of mice in which $2 \times 10^6$ cells of TN12 or WT5 cells had been injected orthotopically into the prostate. Mice were fed with regular feed or Dox-Diet (feed containing doxycycline), beginning 3 days prior to orthotopic injection. All of the mice were killed 30 days after implantation. Imaging was performed prior to sacrifice in randomly selected animals. Top two panels, images of prostate glands into which TN12 control cells were injected in mice that were fed with Dox-Diet (top left) or regular feed without doxycycline (top right). Note tumor formation in both animals. Bottom two panels, images of prostate glands into which WT-5 cells were injected in mice that were fed with Dox-Diet (bottom left) or regular feed without doxycycline (bottom right). Tumor formation is absent in mice not receiving doxycycline (NEP expressed). T, tumor; B, bladder; P, prostate.

As illustrated in Fig. 2B, tumors were detected in the prostate of two animals that received injections of TN12 cells regardless of whether they received tetracycline, and in the prostate of one animal that was fed with tetracycline (NEP expression suppressed) and that was given injections with WT5 cells. How-
ever, no tumor was detected in the animal that was given injections of WT5 cells and did not receive tetracycline (NEP expressed). Autopsies of all of the animals revealed 100% tumor formation in animals receiving TN12 cells, and in four (80%) of five animals given injections of WT5 cells and fed with tetracycline. Only one of five animals that were given injections of WT5 cells and did not receive tetracycline developed a tumor, which was appreciably smaller than other tumors formed (Table 1).

### Table 1 NEP inhibits xenograft tumor formation in the prostates of athymic mice

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Dox</th>
<th>Tumor formation</th>
<th>Tumor sizea (mm³)</th>
<th>Mouse weightb (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT-5</td>
<td>+</td>
<td>4/5 (80%)</td>
<td>0.43 (0.11)</td>
<td>6.1 (2.4)</td>
</tr>
<tr>
<td>WT-5</td>
<td>−</td>
<td>1/5 (20%)</td>
<td>0.07 (0.07)</td>
<td>5.7 (2.8)</td>
</tr>
<tr>
<td>TN-12</td>
<td>+</td>
<td>4/4 (100.0%)</td>
<td>0.91 (0.35)</td>
<td>5.5 (2.3)</td>
</tr>
<tr>
<td>TN-12</td>
<td>−</td>
<td>4/4 (100.0%)</td>
<td>1.04 (0.42)</td>
<td>6.2 (1.9)</td>
</tr>
</tbody>
</table>

a Tumor size measured in mm³ and calculated as length × width × depth/2. Tumor measurements were performed by two researchers independently; in parentheses, SE.
b Average change in animal weight over 30 days of experiment; in parentheses, SE.

**DISCUSSION**

NEP inhibits xenograft tumor formation in the prostate of athymic mice, which suggests that NEP can function as a tumor suppressor of PC. The mechanism of NEP-induced growth inhibition presumably involves the inactivation of its neuropeptide substrates, such as bombesin, endothelin-1, and neurotensin, each of which has been implicated in androgen-independent PC growth (28). We observed that overexpression of NEP in these cells (20). However, the growth inhibition observed was modest (~20–30%) and the level of NEP specific enzyme activity achieved after androgen-stimulation was relatively low. Recombinant NEP also inhibits androgen-independent PC cell growth, but it is difficult to obtain sustained serum levels of NEP in mice using recombinant NEP (13), and we did not observe significant inhibition of tumorigenicity of PC xenografts in athymic mice receiving i.p. recombinant NEP daily for 30 days. Therefore, we constructed an inducible system of NEP expression to more fully examine the effects of expressing cell-surface NEP in androgen-independent PC cells.

Our data show that NEP can regulate androgen-independent PC cell proliferation. Moreover, expression of NEP inhibits xenograft tumor formation in the prostate gland of athymic mice, which suggests that NEP can function as a tumor suppressor of PC. The mechanism of NEP-induced growth inhibition presumably involves the inactivation of its neuropeptide substrates, such as bombesin, endothelin-1, and neurotensin, each of which has been implicated in androgen-independent PC growth (28). We observed that overexpression of NEP in these cells (20). However, the growth inhibition observed was modest (~20–30%) and the level of NEP specific enzyme activity achieved after androgen-stimulation was relatively low. Recombinant NEP also inhibits androgen-independent PC cell growth, but it is difficult to obtain sustained serum levels of NEP in mice using recombinant NEP (13), and we did not observe significant inhibition of tumorigenicity of PC xenografts in athymic mice receiving i.p. recombinant NEP daily for 30 days. Therefore, we constructed an inducible system of NEP expression to more fully examine the effects of expressing cell-surface NEP in androgen-independent PC cells.

Thus, expression of NEP, which leads to inactivation of these neuropeptides, would allow for greater numbers of cells to undergo cell death. In addition, overexpression of cell-surface NEP led to G1 cell cycle arrest, presumably mediated in part by the induction of p21 protein expression and dephosphorylation of Rb. We recently have shown that overexpression of NEP in WT-5 cells inhibits neurotensin-mediated phosphorylation of FAK (p125FAK). Zhao et al. (32) reported that overexpression of a dominant-negative FAK mutant inhibited cell cycle progression at G1 and induced p21 expression, which leads to the possibility that the induction of growth arrest by NEP results in part from the inhibition of FAK phosphorylation.

LNCaP cells constitutively express NEP and exhibit some in vitro, and in tumor cells of metastatic biopsy specimens in vivo from patients with androgen-independent PC (15). Expression of NEP is transcriptionally activated by androgen in androgen-dependent PC cells and decreases with androgen withdrawal (15). Consequently, PC cells that survive androgen withdrawal can emerge with reduced levels of NEP. This decrease in NEP expression can contribute to the development of androgen-independent PC by allowing PC cells to use neuropeptides as an alternative source to androgen to stimulate cell proliferation.

The current study was aimed at further delineating the antitumor effects of NEP and its potential use for inhibiting androgen-independent PC cell growth. In this regard, we had previously shown that androgen-induced growth repression of androgen-independent PC3 cells expressing androgen receptor (PC3/AR) and of an androgen-independent subline of LNCaP cells results in part from androgen-induced expression of NEP in these cells (20). However, the growth inhibition observed was modest (~20–30%) and the level of NEP specific enzyme activity achieved after androgen-stimulation was relatively low. Recombinant NEP also inhibits androgen-independent PC cell growth, but it is difficult to obtain sustained serum levels of NEP in mice using recombinant NEP (13), and we did not observe significant inhibition of tumorigenicity of PC xenografts in athymic mice receiving i.p. recombinant NEP daily for 30 days. Therefore, we constructed an inducible system of NEP expression to more fully examine the effects of expressing cell-surface NEP in androgen-independent PC cells.
of the characteristics observed in WT-5 cells expressing NEP, including a longer doubling time (33), diminished ability to migrate in extracellular matrix (34), and decreased tumorigenicity in athymic mice (34, 35), compared with androgen-independent PC cells that lack NEP expression. In addition, the inhibition of NEP enzyme activity in LNCaP cells results in an increase in FAK phosphorylation. However, the complete phenotype observed in WT-5 cells that are induced to express NEP is not present in LNCaP cells. This may result from the lower levels of NEP protein and enzyme activity in LNCaP cells.
compared with WT-5 cells, or from a lack of downstream mediators of NEP action that are present in TSU-Pr1 cells but not in LNCaP cells.

In summary, the current study emphasizes the consequences of decreased NEP expression in PC cells by studying the effects of reexpressing cell-surface NEP. These experiments suggest overexpression of NEP in PC cells results in multiple effects, including growth inhibition, induction of apoptosis, cell cycle arrest, and the inhibition of tumor formation. Augmentation of NEP expression by delivery of exogenous cell-surface NEP using gene constructs is a potential approach to the treatment of hormone-refractory PC.

ACKNOWLEDGMENTS

We thank Dr. Hui-Kang Lai for technical assistance, Dr. Christos Papandreou for useful discussions, and Catherine Kearney and Lana Winter for secretarial assistance.

REFERENCES


Tumor-Suppressive Effects of Neutral Endopeptidase in Androgen-independent Prostate Cancer Cells

Jie Dai, Ruoqian Shen, Makoto Sumitomo, et al.


Updated version
Access the most recent version of this article at:
http://clincancerres.aacrjournals.org/content/7/5/1370

Cited articles
This article cites 29 articles, 7 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/7/5/1370.full.html#ref-list-1

Citing articles
This article has been cited by 7 HighWire-hosted articles. Access the articles at:
/content/7/5/1370.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.