Analysis of Chemokines and Chemokine Receptor Expression in Ovarian Cancer Ascites

INTRODUCTION

Chemokines are a large subfamily of chemoattractant cytokines, which are classified into four highly conserved groups, CXC, CC, C, and CX3C, based on the position of the first two cysteines adjacent to the NH2 terminus. Chemokines selectively regulate the recruitment and trafficking of leukocyte subsets to inflammatory sites. Chemotactic and growth signals initiated by chemokines are mediated by the activation of G-protein-coupled receptors.

Early growth of epithelial ovarian cancer is usually confined to the ovaries, whereas in advanced stages of ovarian cancer, the peritoneal wall, diaphragm, and omental structures are seeded with micro- and macrometastases of tumor cells. Growth of ovarian cancer (stages I–IV) is also associated with the development of ascites (1). Ascitic fluid arises as a plasma exudate; formation results from an imbalance between the influx and efflux of fluid from the peritoneal compartment (2). This is attributable to a combination of increased microvascular permeability mediated by the angiogenic development of immature blood vessels (3); the production of vascular permeability agents, including vascular endothelial growth factor (4–6); and the blockade of lymphatic drainage by cells present in the ascites (7). Ascites consists of a proteinaceous fluid with variable numbers of suspended cells and debris. The cellular population consists of differing proportions of tumor cells, mesothelial cells, fibroblasts, macrophages, leukocytes, and RBCs (8, 9). Peritoneal dissemination of ovarian carcinoma occurs by the growth of malignant cells in the protein-rich ascitic fluid (10).

Ovarian cancer is one epithelial malignancy in which there is evidence for a complex cytokine network. Published information on individual cytokines has suggested a number of autocrine and paracrine cytokine loops in the biology of this tumor (11). Ascitic fluid is rich in cytokines and growth factors that are secreted by the tumor cells and by mesothelial cells lining the peritoneal cavity, some of which can act to directly stimulate tumor cell growth (12–15). It is possible that ascitic fluid also contains chemokines that may influence its cellular composition, but this has not been investigated in detail. Previously our group has detected mRNA for CCL2 (MCP-1), CCL3 (MIP-1α), CCL4 (MIP-1β), and CCL5 (RANTES) in solid ovarian tumors by in situ hybridization. CCL2 was the predominant chemokine detected in both epithelial and stromal areas, and this chemokine was also found in ascites (16).

With the aim of further understanding chemokine action in human ovarian cancer, we investigated CC chemokine receptors and their ligands in ascitic fluid samples. This, to our knowledge, is the first detailed study of CC chemokines and their receptors in human ovarian cancer ascites.

MATERIALS AND METHODS

Samples. Samples of ascitic fluid were collected from patients with ovarian carcinoma at the time of surgery or by paracentesis for palliative/diagnostic purposes. Volumes of...
1–1.5 liters were taken. All 66 ascites samples were from patients with epithelial ovarian cancer. Each sample was spun down, and the ascitic fluid was removed for ELISA analysis. The cell isolate was treated with ACK lysing buffer to remove RBCs before RNA extraction or flow cytometry. In some patients, a peripheral blood sample was also collected, plasma was removed and stored at −80°C, and PBMCs were isolated by Ficoll-Hypaque density centrifugation (Lymphoprep; Nycomed, Birmingham, United Kingdom). Control PBMCs were isolated from heparinized venous blood or cytophoresis buffy coats from normal volunteer donors. In five samples, the pO2 and pCO2 were measured.

RNA Extraction and RT-PCR. Total RNA was prepared from all samples with Tri Reagent (Sigma Chemical Co., Poole, United Kingdom) according to the manufacturer’s instructions. For RT-PCR and RNase protection analysis, total RNA was treated with DNase to remove contaminating genomic DNA, using RNase-free DNase I (Pharmacia Biotech, St. Albans, United Kingdom). cDNA was synthesized from DNase-treated total RNA using the Ready-to-Go T-primed First Strand kit (Pharmacia Biotech). The primers for CCL2 and CCL22 (MCP-2) were designed from sequences submitted to GenBank, using Primer 3.0. CCL3 and CCL4 sequences were from Hosaka et al. (17), and CCL5 and CCL8 sequences were from Mattei et al. (18) and Van Coillie et al. (19), respectively. The primer sequences and product sizes for CCL2 and CCL22 (MCP-2) were as follows:

CCL2: forward, 5′-CAAACTGAAGCTCGCACTCTCGCC-3′; reverse, 5′-ATTCTTGGGTTGTGGAGTGAGTGTTCA-3′ (product, 354 bp)

CCL22: forward, 5′-CCCTACCTCCCTGCCATTAT-3′; reverse, 5′-CAGGGAGCTAGAACCCAACA-3′ (product, 338 bp)

For each sample, a 25-μl volume was used, containing 200 ng of cDNA, 1 unit of AmpliTaq DNA polymerase, GeneAmp PCR buffer, GeneAmp deoxynucleotide triphosphates (all from Perkin-Elmer, Beaconsfield, United Kingdom), and 4 mM each primer. The following protocol was used in a GeneAmp PCR System 9700 thermal cycler: 94°C for 5 min; 35 cycles of 94°C for 30 s, 60°C for 30 s, and 72°C for 30 s; with a final step at 72°C for 7 min. PCR products were electrophoresed through 1.2% agarose gels and visualized by ethidium bromide. Markers (123 bp; Life Technologies, Inc., Paisley, United Kingdom) were used to estimate band sizes. PCR products were extracted from the gels and sequenced to confirm their identity.

RPA. The hCR5 template set from PharMingen (Becton Dickinson, Oxford, United Kingdom) contained DNA templates for CCR1, CCR2, CCR2a, CCR2b, CCR3, CCR4, CCR5, CCR8, GAPDH, and L32. RPA was carried out using [α-35S]UTP (Amersham International plc, Aylesbury, United Kingdom) rather than [α-32P]UTP. The RNase-protected fragments were separated on an acrylamide-urea sequencing gel (Bio-Rad Laboratories Ltd., Hemel Hempstead, United Kingdom), which was then adsorbed to filter paper and dried under vacuum. Autoradiography was subsequently carried out using Kodak Biomax MS film with a Transcreen LE.
Intensifying screen (Sigma). Densitometry was performed using NIH Image 1.61.

**ELISA of Chemokines in Ascitic Fluid and Plasma Samples.** Concentrations of the chemokines CCL2, CCL3, CCL4, and CCL5 and the cytokine TNFα were measured in 66 samples of ascitic fluid and 12 patient plasma samples, using Quantikine ELISA kits (R&D Systems, Abingdon, United Kingdom) according to the manufacturer’s protocol. The sensitivities of the assays were as follows: 5 pg/ml for CCL2, 7 pg/ml for CCL3, 4 pg/ml for CCL4, 8 pg/ml for CCL5, and 4 pg/ml for TNFα.

**Monoclonal Antibodies and Flow Cytometry.** FITC- and phycoerythrin-labeled monoclonal antibodies against CD3 (UCHT1), CD4 (MT310), CD8 (DK25; all from DAKO, Ely, United Kingdom), CD14 (UCHM-1; Sigma), CCR1 (a kind gift from Shixin Qin, Millennium Pharmaceuticals, Cambridge, MA), CCR2 (48607.211), and CCR5 (45531.111; R&D Systems) and isotype-matched labeled controls were obtained and used to characterize cell surface phenotypes by flow cytometry. For staining, cells were washed and resuspended in PBS supplemented with 1% heat-inactivated fetal bovine serum and 0.01% NaN₃. Antibodies were diluted in this buffer and used at a final concentration of 2–20 μg/ml. Incubations with antibodies were carried out for 30 min on ice. After the final washing step, labeled cells were fixed with 1% formaldehyde solution (Sigma). Appropriate gates were drawn around ascites cell populations after propidium iodide staining was used to eliminate dead cells and debris; 10,000 gated cells were analyzed by flow cytometry on a FACScan flow cytometer using Cellquest software (Becton Dickinson). For determination of CD4⁺/CD8⁺...
T-cell populations, only those cells within a gate drawn around the distinct CD3⁺ population were counted to minimize the inclusion of any CD4⁺ macrophages.

**Statistical Analysis.** To determine correlation coefficients for the ascitic fluid chemokine concentrations, the non-parametric Spearman’s rank correlation was used to calculate P because the data were not normally distributed (20). For all other statistical analysis, we used the Welch’s t test (a more stringent form of Student’s t test).

**RESULTS**

**Cell Populations within Ascitic Fluid.** Fluorescence-activated cell-sorting analysis of 20 ascites samples from patients with ovarian cancer showed considerable variation within the cell populations. The numbers of CD14⁺ cells were variable (median, 27%; range, 0–66%). There were usually high numbers of CD3⁺ T lymphocytes (median, 59%; range, 1–89%) forming a discrete population of cells. The CD3⁺ population contained predominantly CD4⁺ T cells (median, 51%; range, 1–65%), rather than CD8⁺ T cells (median, 10%; range, 0–31%). The T-cell subsets in ascitic fluid differed significantly from those in solid ovarian tumors, which contain predominantly CD8⁺ T cells (with few or no CD4⁺ T cells) and macrophages (16). Because T lymphocytes and macrophages typically respond to CC chemokines, we studied a range of these and their related receptors in ascites.

**Chemokine Protein Levels in Ovarian Cancer Ascites.** CCL2, CCL3, CCL4, and CCL5 protein levels were measured by ELISA in 66 samples of ascitic fluid from ovarian cancer patients (Fig. 1A). The total protein concentration in ascites was reasonably constant (range, 13–75 mg/ml; median, 44 mg/ml); therefore, chemokine concentrations are expressed as pg/ml. All 66 samples contained the CC chemokine CCL2 at a range of 103–19,195 pg/ml (median, 1778 pg/ml); this was the most abundant CC chemokine. The median concentrations of CCL3, CCL4, and CCL5 were approximately one log lower, at 60, 182, and 44 pg/ml, respectively. CCL3 was found in 62 of 66 samples (range, 6–4840 pg/ml), CCL4 was found in all 66 samples (range, 33–11,040 pg/ml), and CCL5 was found in 63 of 66 samples (range, 3–1903 pg/ml). TNFα levels were also assayed in 37 of the ascites samples. Only 16 of the 37 samples had detectable levels of TNFα (range, 20–41 pg/ml).

**Chemokine Protein Levels in Ovarian Cancer Patient Plasma.** In addition to ascitic fluid, plasma was obtained from a few patients. Protein concentrations for CCL2, CCL3, CCL4, and CCL5 were determined by ELISA in these plasma samples. CCL2, CCL3, and CCL4 were present in low concentrations, with medians of 341, 12, and 54 pg/ml, respectively (Fig. 1B). CCL2 was present in 12 of 12 samples (range, 110–1888 pg/ml), CCL3 was present in 4 of 10 samples (range, 3–19 pg/ml), and CCL4 was present in 12 of 13 samples (range, 5–120 pg/ml). CCL5 was detected at extremely high levels, with a median of 28,950 pg/ml (data not shown); however, this was likely attributable to the release of stored CCL5 from platelets (21). These results suggest that a chemokine gradient may exist between plasma and ascites, which may partially explain the leukocyte infiltration seen in ascites.

**CC Chemokine mRNA Expression in Cell Isolates from Ovarian Cancer Ascites.** RT-PCR was used to screen for six CC chemokines (CCL2, CCL3, CCL4, CCL5, CCL8, and CCL22), which bind to the receptors studied below, in cell isolates from ascites samples. Each of these chemokines was expressed in 85–100% of the samples studied (data not shown).

**Chemokine Receptor Expression in Cell Isolates from Ascitic Disease.** We found that CC chemokine receptor expression in seven samples of RNA isolated from ascites was similar to that of the normal PBMC control (Fig. 2, A and B), with a majority of samples positive for each receptor. This contrasts with our findings on RNA from solid ovarian tumor biopsies (22), where CCR1 was the only CC chemokine detected in the majority of cases. In solid tumors, CCR1 localized to infiltrating leukocytes and macrophages. The level of chemokine receptor expression in five of the ascites samples was compared densitometrically with the expression in PBMCs from the same patients. When normalized for the “housekeeping” gene, L32, the levels of expression for CCR2, CCR3, and CCR4 were similar in PBMCs and ascites, but CCR1 and CCR5 appeared to be more strongly expressed by the ascites cells (P < 0.05, Fig. 2C).

**CC Chemokine Receptor Protein on Ascites Cells.** Two-color flow cytometric analysis of cells freshly isolated from three samples of ovarian cancer ascites detected cell surface expression of CCR1, CCR2, and CCR5 on the majority of CD14⁺ cells (Fig. 3 and Table 1). CCR1 was expressed by the majority of CD4⁺ T cells. CCR2 expression was detected on 25–80% of cells, and CCR5 was detected on 23–70% of the cells (Fig. 3 and Table 1). CCR1 was expressed by the majority of CD8⁺ T cells, and CCR2 and CCR5 were expressed by ~5–45% of cells (Fig. 3 and Table 1). Thus, more CD4⁺ than CD8⁺ T cells expressed CCR2 and CCR5, suggesting a difference in the chemokine receptor profile of these cell types.

![Fig. 3](image-url) Chemokine receptor expression on cells isolated from ovarian ascites as determined by flow cytometry. CCR1, CCR2, and CCR5 expression on CD4⁺ (left panels), CD8⁺ (middle panels), and CD14⁺ (right panels) cells.
Table 1  Percentage of each leukocyte type expressing CCR1, CCR2, and CCR5, in three samples of ascites.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Cell type</th>
<th>CCR1 (%)</th>
<th>CCR2 (%)</th>
<th>CCR5 (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>CD4⁺</td>
<td>82</td>
<td>82</td>
<td>70</td>
</tr>
<tr>
<td></td>
<td>CD8⁺</td>
<td>84</td>
<td>44</td>
<td>45</td>
</tr>
<tr>
<td></td>
<td>CD14⁺</td>
<td>45</td>
<td>94</td>
<td>93</td>
</tr>
<tr>
<td></td>
<td>CD14⁻</td>
<td>88</td>
<td>75</td>
<td>42</td>
</tr>
<tr>
<td>B</td>
<td>CD8⁺</td>
<td>87</td>
<td>48</td>
<td>27</td>
</tr>
<tr>
<td></td>
<td>CD14⁺</td>
<td>80</td>
<td>96</td>
<td>68</td>
</tr>
<tr>
<td></td>
<td>CD4⁺</td>
<td>57</td>
<td>26</td>
<td>19</td>
</tr>
<tr>
<td>C</td>
<td>CD8⁺</td>
<td>91</td>
<td>6</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>CD14⁺</td>
<td>87</td>
<td>88</td>
<td>60</td>
</tr>
</tbody>
</table>

Table 2  Correlations between the chemokines and cell populations within ovarian ascites, Spearman correlation coefficients and (P) between the chemokines.
P < 0.05 is statistically significant (bold).

<table>
<thead>
<tr>
<th>Chemokine</th>
<th>CCL2</th>
<th>CCL3</th>
<th>CCL4</th>
<th>CCL5</th>
</tr>
</thead>
<tbody>
<tr>
<td>CCL2</td>
<td>0.04 (0.78)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CCL3</td>
<td></td>
<td>0.11 (0.11)</td>
<td>0.56 (&lt;0.0001)</td>
<td></td>
</tr>
<tr>
<td>CCL4</td>
<td></td>
<td></td>
<td>0.29 (0.02)</td>
<td>0.28 (0.02)</td>
</tr>
<tr>
<td>CCL5</td>
<td></td>
<td></td>
<td></td>
<td>0.32 (0.008)</td>
</tr>
</tbody>
</table>

Correlations. The nonparametric Spearman’s rank correlation was used to examine the relationship between the protein concentrations of the four CC chemokines studied. Significant correlations were found between the levels of CCL3 and CCL4 (r = 0.56; P = 0.0001; Table 2) and between the levels of CCL2 and CCL5 (r = 0.32; P = 0.008). Weaker associations were observed between CCL3 and CCL5 (r = 0.29; P = 0.02) and between CCL4 and CCL5 (r = 0.28; P = 0.02). No other chemokine pair gave a significant correlation. These correlations may suggest that CCL3 and CCL4 are produced by the same cell type or are induced by the same stimulus in ovarian cancer ascites. This could also be true for CCL2 and CCL5. Despite high levels of chemokine protein, there were few correlations found between the total cell counts in the ascites and the concentration of any chemokine (Table 3). CCL5 was expressed at higher levels when CD3⁺ cells were prevalent (r = 0.54; P = 0.01), and there was a negative correlation between CCL5 and the presence CD14⁺ cells (r = −0.47; P = 0.03). There appeared to be no correlations between the stage of disease and the presence of any chemokine.

DISCUSSION

Previous studies by our group and others have addressed the relationship between leukocyte infiltration into solid tumors and chemokine expression (14, 16, 23). To our knowledge, this work is the first comprehensive study of CC chemokines and chemokine receptor expression in human ovarian cancer ascites.

We found that ascitic fluid is rich in CC chemokines and that the CD14-expressing cells and T cells present in ovarian cancer ascites express CC chemokine receptor mRNA and protein.

Is the extent and phenotype of the leukocyte infiltration in ovarian ascites related to chemokines and chemokine receptor expression? Gradients of chemokines usually cause tissue recruitment of leukocytes through effects on adhesion and endothelial transmigration (24). Our data suggest that CC chemokine protein levels are significantly higher in ascitic fluid than in patient plasma samples. Therefore, chemokines present in ascites could form a gradient for leukocyte migration into the peritoneal cavity. However, we did not find many associations between chemokine concentrations and leukocyte numbers; this could be because multiple chemokines cooperate to attract a particular leukocyte. Apical production of chemokines during infection or peritoneal inflammation is well documented (25); both CC and CXC chemokines are produced by mesothelial cells, and the resulting chemokine gradient chemoattracts an infiltrate of polymorphonuclear leukocytes.

There was a correlation between CCL5 expression in ascites and the presence of CD3⁺ T cells, suggesting that this chemokine could be important for recruiting T cells. This chemokine also correlated with the presence of CD8⁺ T cells in solid ovarian tumors (16). The fact that the CD4:CD8 ratio differs between solid tumor (16) and ascites may be related to CC chemokine receptor expression on the T cells. There were differing proportions of CD4⁺ and CD8⁺ T cells expressing CCR2 and CCR5 (the receptor for CCL5). CCR2 was expressed more by CD4⁺ T cells than by CD8⁺ T cells, and the same was true of CCR5. The lower number of CD8⁺ T cells expressing CCR5 could account for the defective recruitment of these cells to the ascites compared with the solid tumor. We cannot rule out that a small proportion of the CD4⁺ cells may be CD14⁺ macrophages rather than T cells; however, the CD3⁺ T-cell population was clearly distinct from the CD14⁺ population by flow cytometry.

Overall, the profile of CC chemokine receptor expression in ascites was similar to that of leukocytes in peripheral blood and contrasted with the restricted expression of CC chemokine receptors seen in solid ovarian tumors (22). This may be attributable to differences in the solid tumor and ascitic microenvironment. TNFα levels in ascites are low; therefore, this cytokine is unlikely to be responsible for the stimulation of chemokine production or down-regulation of CC chemokine expression (26). Another important stimulus regulating chemokine production is hypoxia (27, 28). Gas analysis of four ascites samples revealed that pH (range, 59–95 mm Hg) and pCO₂ (range, 41–52 mm Hg) were similar to that of arterial blood (~95 and 40 mm Hg, respectively). Solid tumors are likely to be hypoxic and have high TNF levels, whereas ascites is normoxic with low TNF levels. These factors may contribute to the differences in CC chemokine receptor expression between the two tumor states.
There was a negative correlation between CCL5 and CD14+ cells. A majority of CD14+ expressing cells within ascites consistently expressed CCR1, CCR2, and CCR5. CCR5 and CCR1 are both receptors for CCL5, suggesting that CCL5 may have an antagonistic effect under certain circumstances. Alternatively, we found a correlation between the concentrations of CCL2 and CCL5, although CCL2 levels were consistently one log higher. Thus, at high CCL5 concentrations, CCL2 levels may become antagonistic, as has been shown for CXCL12 (SF1-1; Ref. 29).

CCR1 was the predominant receptor found on leukocytes within the solid tumor (22); this receptor was also found on the majority of leukocytes within the ascites. The expression of CCR2 on CD14+ cells within the ascites contrasts with the results obtained by Sica et al. (26), who suggested CCR2 down-regulation on ovarian tumor infiltrating macrophages. However, this may reflect the fact that the cells used in this study are freshly isolated from ascitic fluid and not purified or cultured. Recently, we have demonstrated that ovarian tumor cells express functional CXCR4 (30), and high concentrations of the ligand CXCL12 have been detected in ovarian cancer ascites. Similar observations in breast and pancreatic carcinoma (31, 32) lead us to suggest that ovarian cancer cells may respond to chemokine gradients in the process of metastasis.

We conclude that there is a complex chemokine/chemokine receptor network in ovarian cancer ascites. We found few associations between chemokine receptor expression, chemokine levels, and cell composition; this is in contrast to the associations found by us and others in solid tumors. It is possible that high levels of chemokine protein in ascites cause receptor desensitization.

ACKNOWLEDGMENTS

We would like to thank the surgeons at the Department of Obstetrics and Gynaecology, Guy’s and St. Thomas’ Hospital for providing patient material.

REFERENCES


Downloaded from clincancerres.aacrjournals.org on October 15, 2017. © 2002 American Association for Cancer Research.


Analysis of Chemokines and Chemokine Receptor Expression in Ovarian Cancer Ascites

David Milliken, Chris Scotton, Shanti Raju, et al.

*Clin Cancer Res* 2002;8:1108-1114.

Updated version  Access the most recent version of this article at:
http://clincancerres.aacrjournals.org/content/8/4/1108

Cited articles  This article cites 30 articles, 13 of which you can access for free at:
http://clincancerres.aacrjournals.org/content/8/4/1108.full#ref-list-1

Citing articles  This article has been cited by 12 HighWire-hosted articles. Access the articles at:
http://clincancerres.aacrjournals.org/content/8/4/1108.full#related-urls

E-mail alerts  Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions  To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions  To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.