Enhanced Safety Profiles of the Telomerase-Specific Replication-Competent Adenovirus by Incorporation of Normal Cell-Specific microRNA-Targeted Sequences

Kumiko Sugio1,3, Fuminori Sakurai1,3, Kazufumi Katayama1, Katsuhisa Tashiro3, Hayato Matsui1,3, Kenji Kawabata2,3, Atsushi Kawase4, Masahiro Iwaki4, Takao Hayakawa5, Toshiyoshi Fujiwara6, and Hiroyuki Mizuguchi1,3

Abstract

Purpose: Oncolytic adenoviruses (Ad) have been actively pursued as potential agents for cancer treatment. Among the various types of oncolytic Ads, the telomerase-specific replication-competent Ad (TRAD), which possesses an E1 gene expression cassette driven by the human telomerase reverse transcriptase promoter, has shown promising results in human clinical trials; however, the E1 gene is also slightly expressed in normal cells, leading to replication of TRAD and cellular toxicity in normal cells.

Experimental Design: To overcome this problem, we utilized a microRNA (miRNA)-regulated gene expression system. Four copies of complementary sequences for miR-143, -145, -199a, or let-7a, which have been reported to be exclusively downregulated in tumor cells, were incorporated into the 3’-untranslated region of the E1 gene expression cassette.

Results: Among the TRAD variants (herein called TRADs) constructed, TRADs containing the sequences complementary to miR-143, -145, or -199a showed efficient oncolytic activity comparable to the parental TRAD in the tumor cells. On the other hand, replication of the TRADs containing the miRNA complementary sequences was at most 1,000-fold suppressed in the normal cells, including primary normal cells. In addition, to suppress the replication of the TRADs in hepatocytes as well as other normal cells, we constructed a TRAD containing 2 distinct complementary sequences for miR-199a and liver-specific miR-122a (TRAD-122a/199aT). TRAD-122a/199aT exhibited more than 10-fold reduction in viral replication in all the normal cells examined, including primary hepatocytes.

Conclusions: This study showed that oncolytic Ads containing the sequences complementary to normal cell-specific miRNAs showed significantly improved safety profiles without altering tumor cell lysis activity. Clin Cancer Res; 17(9); 1–12. ©2011 AACR.

Introduction

Oncolytic adenoviruses (Ad) are genetically engineered Ads which can kill tumor cells by tumor cell-specific replication (1, 2). Several clinical trials using oncolytic Ads have been carried out, and promising results have been reported (3-5). Various types of oncolytic Ads have been developed, and can be mainly classified into 2 groups. One type of oncolytic Ads show tumor-selective replication via deletion of certain genes, such as the E1B-55K gene, which are dispensable for the replication of Ads in tumor cells. The other type of oncolytic Ads possess an E1 gene expression cassette driven by tumor-specific promoters. Various types of tumor-specific promoters are used in oncolytic Ads, including the α-fetoprotein promoter (6), prostate-specific antigen promoter (7), osteocalcin promoters (8), and cyclooxygenase-2 promoter (9).

Among these oncolytic Ads possessing tumor-specific promoters, the telomerase-specific replication-competent
Oncolytic adenoviruses (Ad) are promising anticancer agents and have been used in human clinical trials. However, though a lesser extent than in tumor cells, some oncolytic Ads also replicate in normal human cells, resulting in unexpected toxicity. In this study, we included a microRNA (miRNA)-regulated posttranscriptional detargeting system into a telomerase-specific replication-competent Ad (TRAD), which has been used in clinical trials. Complementary sequences for miR-143, -145, and -199a, which have been shown to be exclusively downregulated in tumor cells, were inserted into the E1 gene expression cassette. The TRAD containing these miRNA complementary sequences exhibited significantly reduced replication in normal cells (up to 1,000-fold reductions), including human primary cells, and comparable tumor cell lysis activity to the conventional TRAD. These results indicate that an miRNA-regulated posttranscriptional detargeting system offers a potential strategy to reduce the replication of TRAD in normal cells without altering tumor cell lysis activity, and makes it possible to increase the injected doses, leading to enhanced antitumor effects.

**Materials and Methods**

**Cells**

A549 (a human non–small cell lung cancer cell line), HepG2 (a human hepatocellular carcinoma cell line), and 293 cells (a transformed embryonic kidney cell line) were cultured in Dulbecco’s modified Eagle’s Medium containing 10% fetal bovine serum (FBS) and antibiotics. HT29 (a human colorectal cancer cell line) and WI38 cells (a normal human lung diploid fibroblast) were cultured in Minimum Essential Medium containing 10% FBS and antibiotics. H1299 cells (a human non–small cell lung cancer cell line) were cultured in RPMI1640 containing 10% FBS and antibiotics. These cell lines were obtained from the cell banks, including the Japanese Collection of Research Resources (JCRB) cell bank. The normal human lung fibroblasts (NHFL), normal human prostate stromal cells (PrSC), normal human small airway epithelial cells (SAEC), and normal human hepatocytes (Nhep; Lonza) were cultured in the medium recommended by the manufacturer.

**Construction of TRADs**

All TRADs were prepared by means of an improved in vitro ligation method described previously (21–23). hTERT
miRNA-Regulated Replication of Oncolytic Adenovirus

promoter-driving E1 gene-expressing shuttle plasmids having multiple tandem copies of sequences perfectly complementary to miRNAs in the 3′-UTR of the E1 gene expression cassette were constructed as described below. A KpnI/AfIII fragment of pHMCMV5 (22) was ligated with oligonucleotides miR-143T-S1 and miR-143T-AS1, which contain miR-143 complementary sequences, resulting in pHMCMV5-143T-1. The sequences of the oligonucleotides are shown in Supplementary Table S1. Next, a PacI/AfIII fragment of pHMCMV5-143T-1 was ligated with oligonucleotides miR-143T-S2 and miR-143T-AS2. The resulting plasmid, pHMCMV5-143T, was digested with I-CeuI after digestion with Nhel followed by Klenow treatment, and then ligated with the I-CeuI/Pmel fragment of pSh-hAIB (10), in which the E1A and E1B genes linked with an internal ribosomal entry site (IRES) are located downstream of the hTERT promoter, creating pSh-AIB-143T.

For the construction of vector plasmids for TRADs, I-CeuI/Pmel-digested pSh-AIB-143T was ligated with the I-CeuI/Pmel-digested pAdHM3 (21), resulting in pAdHM3-AIB-143T. To generate TRADs, pAdHM3-AIB-143T was digested with PacI and was transfected into 293 cells using Superfect transfection reagent (Qiagen). All TRADs were propagated in 293 cells, purified by 2 rounds of cesium chloride gradient ultracentrifugation, dialyzed, and stored at −80°C. TRADs containing other miRNA complementary sequences were similarly constructed using the corresponding oligonucleotides (Supplementary Table S1). The parental TRAD was similarly prepared using pSh-AIB and pAdHM3. The virus particles (VP) and biological titers were determined by a spectrophotometrical method (24) and by using an Adeno-X rapid titer kit (Clontech), respectively. The ratio of particle-to-biological titer was between 6 and 9 for each TRAD used in this study.

**Determination of miRNA expression levels in human normal and tumor cells**

Total RNA, including miRNAs, was isolated from cells using Isogen (Nippon Gene). After quantification of the RNA concentration, miRNA levels were determined using a TaqMan MiRNA reverse transcription kit, TaqMan miRNA assay kit, and ABI Prism 7000 system (Applied Biosystems). Amplification of U6 served as an endogenous control to normalize the miRNA expression data.

**Infection with TRADs**

Cells were seeded into 24-well plates at 5 × 10^4 cells/well. On the following day, cells were infected with TRADs at a multiplicity of infection (MOI) of 0.4 or 2 (for cancer cell lines), or of 10 (for normal cells), for 2 hours. Following incubation for 3 (for cancer cell lines) or 5 days (for normal cells), total DNA, including viral genomic DNA, was isolated from the cells using a DNeasy Blood & Tissue Kit (Qiagen). After isolation, the Ad genomic DNA contents were quantified using an ABI Prism 7000 system (Applied Biosystems) as previously described (25). The Ad genome copy numbers were normalized by the copy numbers of glycerolaldehyde-3-phosphate-dehydrogenase (GAPDH). Cell viability was also examined by crystal violet staining and Alamar blue assay at the indicated time points. To examine the miRNA-specific suppression of TRAD replication in normal human cells, 50 nmol/L of 2′-O-methylated antisense oligonucleotide complementary to miR-143 or miR-199a (Gene Design Inc.) was transfected into normal cells using Lipofectamine 2000 (Invitrogen). Twenty-four hours after transfection, the cells were infected with TRADs and replication of TRADs was evaluated as described above.

**Real-time reverse transcriptase PCR analysis for E1A gene expression**

Cells were seeded as described above and were infected with TRADs at an MOI of 2 (for cancer cells) or 10 (for normal cells) for 1.5 hours. After a 24-hour incubation, total RNA was isolated, and reverse transcription reaction was carried out using a SuperScript II First-Strand Synthesis System (Invitrogen). E1A mRNA levels were determined with the E1A-specific primers and probe using an ABI prism 7000 system (26). The E1A mRNA levels were normalized by the GAPDH mRNA levels.

**Statistical analysis**

Statistical significance (P < 0.05) was determined using Student’s t test. Data are presented as means ± SD.

**Results**

**Replication of the conventional TRAD in normal human cells**

First, to examine replication of the conventional TRAD in normal human cells, WI38 cells, which are human embryonic lung fibroblasts, were infected with the conventional TRAD at an MOI of 2 or 10 (Fig. 1). The conventional TRAD did not highly replicate in WI38 cells at an MOI of 2; however, an almost 500-fold increase in the Ad genome
was found 3 days after infection at an MOI of 10. These data indicate that the conventional TRAD replicates in normal human cells at a high MOI, even though tumor-specific hTERT promoters are used for the E1 gene expression.

MiRNA expression levels in human tumor and normal cells

To examine the expression levels of miR-143, -145, -199a, and let-7a in the human normal and tumor cells, reverse transcriptase PCR (RT-PCR) analysis was carried out. Several studies have shown that these miRNAs are downregulated in various types of tumor cells isolated from cancer patients, compared with the corresponding normal tissues (16–18, 27). The expression levels of miR-143, -145, and -199a in the tumor cells were approximately 2- to 100-fold lower than those in the normal cells, although SAECs expression levels of miR-143, -145, and -199a were comparable or lower than those in the tumor cells (Fig. 2). In particular, a large reduction was found for miR-199a expression in all tumor cells, compared with the normal cells. On the other hand, the expression levels of let-7a in HT29 and H1299 cells were higher than those in the normal cells, although HepG2 cells expressed lower levels of let-7a than the normal cells. The absolute amounts of let-7a were more than 10-fold higher than those of the other miRNAs in all tumor and normal cells, except for NHLE, NHep, and HepG2 cells (data not shown).

Development of TRADs carrying an miRNA-regulated E1 gene expression system

Next, to develop TRADs carrying a miRNA-regulated E1 gene expression cassette (TRAD-miRT), we incorporated 4 copies of the perfectly complementary sequences for miR-143, -145, -199a, or let-7a into the 3′-UTR of the E1 gene expression cassette (Fig. 3A). In TRADs, the E1A gene was connected with the E1B gene via IRES. We found that the expression of both the first and second gene in the IRES-containing expression cassette was suppressed in an miRNA-dependent manner by insertion of the miR-122a complementary sequences into the region downstream of the second gene in miR-122a-expressing Huh-7 cells, not in HepG2 cells, which express a low level of miR-122a (Supplementary Fig. S1), although it remains controversial whether miRNA-mediated posttranscriptional regulation can occur in an IRES-containing expression cassette (28–30). All TRADs were efficiently grown in normal 293 cells, and the ratios of infectious titers to physical titers were comparable among all the TRADs, including the parental TRAD.

Tumor cell lysis activity and replication of TRAD-miRT in tumor cells

To examine whether or not the inclusion of the sequences complementary to the miRNAs downregulated in tumor cells would inhibit the tumor cell lysis activity of TRADs, the viability of tumor cells was evaluated after infection with the TRADs. Almost all tumor cells were lysed by TRAD-143T, -145T, and -199aT at 3 days after infection, although cell lysis by TRAD-let7aT was largely inhibited (Fig. 3B). Furthermore, time-course studies of cell viability showed that TRAD-143T, -145T, and -199aT exhibited cytopathic efficacies comparable to that of the parental TRAD in the tumor cells at an MOI of 0.4 (Fig. 3C). Similar results were obtained at an MOI of 2 (data not shown).
We next examined the replication ability of the TRADs in the tumor cells by determining the viral genome copy numbers. TRAD-143T, -145T, and -199aT efficiently replicated in the tumor cells, and the viral genome copy numbers of TRAD-143T, -145T, and -199aT in the tumor cells were more than 500-fold higher than those in the normal cells (data not shown). In addition, TRAD-143T, -145T, and -199aT exhibited viral genome copy numbers similar to that of the conventional TRAD in all tumor cells (Fig. 3D). All TRADs except for TRAD-let7aT also expressed similar levels of E1A mRNA (Fig. 3E). In contrast, insertion of let-7a complementary sequences largely inhibited the replication in all tumor cells. The E1A mRNA level was also reduced by 42% in H1299 cells infected with TRAD-let7aT. Inefficient replication of TRAD-let7aT in the tumor cells corresponded to the low cytopathic effects described above. These results indicate that TRADs containing the complementary sequences for miR-143, -145, or -199a exhibit efficient E1 gene expression in the tumor cells and tumor cell lysis activity comparable to those of the conventional TRAD.

Reduced replication of TRAD-miRT in normal cells

To examine whether replication of TRADs in normal cells is suppressed by incorporation of the sequences complementary to the miRNAs downregulated in tumor cells, normal human cells were infected with the TRADs. The virus genome copy numbers of TRAD-143T, -145T, and -199aT were 5- to 1,000-fold reduced, compared with the conventional TRAD at 5 days following infection in WI38 cells (Fig. 4A). An approximately 3- to 300-fold reduction in the genome copy numbers of TRAD-143T, -145T, and -199aT was also observed in NHLF and PrSC. The replication of TRADs was also suppressed in SAEC by the insertion of the miRNA complementary sequences, although the expression levels of miR-143, -145, and -199a in SAEC were much lower than those in the other normal cells (Fig. 2). The suppressive effects of insertion of the miRNA target sequences were different among the cells; however, overall, the insertion of miR-199a complementary sequences mediated similar or higher suppressive effects on the replication of TRADs in all the normal cells examined, compared with insertion of the sequences complementary to miR-143 and -145. Replication of TRAD-199aT was inhibited by more than 10-fold in all the normal cells except for SAEC. We also examined the viabilities of the normal cells after infection with the TRADs. No apparent differences in cell viabilities were found among the TRADs by crystal violet staining (data not shown); however, Alamar blue assay showed that the average values of the normal cell viabilities were higher after infection with TRAD-miRT than after infection with the conventional TRAD (Fig. 4B). These results suggest that the suppression of TRAD replication by insertion of the miRNA complementary sequences results in the improvement of the TRAD safety profile in normal cells.

Next, to evaluate whether the reduction in replication of TRAD-miRT was miRNA-dependent, miRNAs were inhibited by a 2’-O-methylated antisense oligonucleotide. NHLF and PrSC cells were transfected with the 2’-O-methylated antisense oligonucleotide against miR-143 or -199a, and then the cells were infected with the TRADs, 24 hour after transfection. In the cells transfected with the 2’-O-methylated antisense oligonucleotide against miR-143 or -199a, the reduction in the replication of TRAD-miRT was significantly restored, but the scramble 2’-O-methylated oligonucleotide did not significantly affect the replication of TRAD-miRT (Fig. 4C). These results indicate that the reduction in the replication of TRAD-miRT in the normal cells was miRNA-dependent.
Figure 3. (Continued)

C

Days after infection

Days after infection

Cell viability (% of control)

Cell viability (% of control)

Mock
The conventional TRAD
TRAD-143T
TRAD-145T
TRAD-199aT

D

Ad genome copy number (% of control)

Days after infection

Ad genome copy number (% of control)

Ad genome copy number (% of control)

Ad genome copy number (% of control)

Figure 3.

Sugio et al.

Clin Cancer Res; 17(9) May 1, 2011

Clinical Cancer Research

Published OnlineFirst February 23, 2011; DOI: 10.1158/1078-0432.CCR-10-2008

Downloaded from clincancerres.aacrjournals.org on July 16, 2017. © 2011 American Association for Cancer Research.
Figure 4. Reduced replication of TRADs in normal human cells by insertion of the miRNA complementary sequences. A, the viral genome copy numbers of TRADs in normal cells. The cells were infected with the TRADs at an MOI of 10 for 2 hours. Five days after infection, the viral genome copy numbers were determined by real-time PCR. B, time-course study of the normal human cell viabilities after infection with TRADs by Alamar blue assay. The cells were infected with the TRADs at an MOI of 10 for 2 hours. At the indicated time points, the viability of the cells was analyzed by Alamar blue assay. The data were normalized by the data of the mock-infected group. C, restoration of TRAD replication in human normal cells by 2'-O-methylated antisense oligonucleotides. The cells were transfected with 50 nmol/L of 2'-O-methylated antisense oligonucleotides for miR-143 or -199a. Twenty-four hours after transfection, the cells were infected with the TRADs at an MOI of 10, and the viral genome copy numbers were determined 5 days after infection with the TRADs. D, the E1A mRNA levels in normal human cells. The cells were infected with the TRADs at an MOI of 10 for 1.5 hours. Twenty-four hours after infection, the E1 mRNA levels were determined by real-time RT-PCR. The data was normalized by the data of the conventional TRAD group. All the data are shown as the means ± SD (n = 3–4). *, P < 0.05; **, P < 0.005.
**E1A expression by TRAD-miRT in normal cells**

To determine whether incorporation of the miRNA complementary sequences into the E1 gene expression cassette decreases the E1 mRNA levels in normal human cells, real-time RT-PCR analysis for the E1A mRNA levels was carried out. The E1A mRNA levels were reduced by more than 30% for TRAD-143T, -145T, and -199aT, compared with the parent TRAD, in NHLF (Fig. 4D). The reduction in the E1A mRNA levels corresponded to the suppression in replication of TRAD-miRT, indicating that miRNA-mediated reduction in the E1 gene expression resulted in a reduced replication of TRAD-miRT.

**Development of TRADs containing the complementary sequences for liver-specific miRNA**

To prevent the replication of TRADs in liver hepatocytes as well as other normal cells, we incorporated not only miR-199a complementary sequences but also sequences complementary to liver-specific miR-122a into the E1 gene expression cassette, resulting in TRAD-122a/199aT (Fig. 5A). It is well known that Ads have high hepatic tropism, leading to efficient liver accumulation even after local administration. MiR-122a was expressed approximately 100- to 20-fold more abundantly in NHep and Huh-7 cells, respectively, than in the other normal human cells and tumor cells (Fig. 5B); conversely, the other normal cells expressed more than 10-fold lower levels of miR-122a than miR-143, -145, and -199a (data not shown). Incorporation of miR-122a complementary sequences alone significantly reduced the virus genome copy number of TRAD-122aT in NHLF and NHep; however, no statistically significant decrease in the genome copy number of TRAD-122aT was found in PrSC (Fig. 5C). On the other hand, insertion of miR-199a target sequences alone was less efficient than insertion of miR-122a target sequences in NHep, probably due to the lower expression of miR-199a than miR-122a in NHep. By contrast, insertion of both miR-122a and miR-199a target sequences into the E1 gene expression cassette efficiently reduced the replication of TRAD-122a/199aT by 10- to 50-fold in all normal cells examined. Significantly reduced replication of TRAD-122a and TRAD-122a/199aT was also found in Huh-7 cells, which are a hepatoma cell line highly expressing miR-122a and are often used as a model of hepatocytes (Supplementary Fig. S2). The incorporation of the miR-145 complementary sequences was also effective for suppressing the TRAD replication in NHep (Supplementary Fig. S3). The E1A mRNA levels were reduced for TRAD-122aT and -122a/199aT in NHep (Fig. 5D). In addition, TRAD-122a/199aT efficiently replicated in the tumor cells, resulting in efficient tumor cell lysis (Fig. 5E and F). These results indicate that replication of the TRADs in various types of normal human cells, including liver hepatocytes, is significantly reduced by insertion of the multiple target sequences to both miR-122a and -199a, without influencing the tumor cell lysis activity.

**Discussion**

The aim of this study was to prevent the replication of TRADs in normal human cells by incorporation of sequences complementary to miRNAs that are selectively downregulated in tumor cells, without altering the tumor cell lysis activity. Currently, there is no appropriate animal model which fully supports the *in vivo* replication of Ads and evaluation of the *in vivo* toxicity caused by oncolytic Ads, and thus it is important to be cautious in regard to oncolytic Ad-induced toxicity. To prevent the E1 gene expression and replication of oncolytic Ads in normal cells as much as possible, an miRNA-mediated posttranscriptional detargeting system was included in TRADs, in
addition to the transcriptional targeting system via tumor-specific promoters.

As described above, TRAD replicates in the injected tumors and is disseminated from the injected tumors into the systemic circulation, leading to infection of distant, uninjected tumors (11, 13, 14). This property of TRAD had led to a concern that TRAD could infect normal cells over the whole body, including the hepatocytes, after dissemination from the injected tumors. It is crucial that such unexpected infection of normal cells by TRAD is prevented. Previous studies have shown that insertion of sequences complementary to liver-specific miR-122a reduced the replication of oncolytic Ads in Huh-7 cells, which are a model cell for hepatocytes (31–33). It is especially crucial...
to prevent the replication of TRAD in the liver, because Ad vectors have strong hepatotropism. However, TRAD also might infect normal cells other than hepatocytes, indicating that replication of oncolytic Ads in normal cells other than hepatocytes should also be suppressed. To prevent the replication of TRADS in other normal cells, we incorporated the sequences complementary to miR-143, -145, -199a, or let-7a, which are downregulated in the tumors and widely expressed in normal cells. The expression levels of these miRNAs in the tumor cells were lower than those in the normal cells in this study, and insertion of sequences complementary to miR-143, -145, or -199a significantly reduced the E1A mRNA levels and the replication of TRADS in the normal cells.

Overall, among the miRNA complementary sequences, the miR-199a complementary sequences appeared to be the most effective at suppressing the replication of TRADS across all the normal cells except for hepatocytes; however, insertion of miR-199a target sequences alone failed to significantly reduce the replication of TRADS in the hepatocytes. To simultaneously prevent the replication of TRADS in various types of normal cells, including hepatocytes, we incorporated sequences complementary to miR-122a, which is abundantly expressed in hepatocytes, in addition to miR-199a target sequences. Brown and colleagues reported that a desired transgene expression pattern was achieved, depending on the miRNA expression profile, by incorporation of target sequences for 2 distinct miRNAs (34). TRAD-122aT/199aT exhibited more than 10-fold reduction in the replication in all the normal cells except for SAEC, although insertion of target sequences for miR-122a or miR-199a alone failed to suppress the replication of TRADS in either of the normal cells. Furthermore, TRAD-122aT/199aT and the parental TRAD mediated similar cytopathic efficacies in the tumor cells. These results indicate that replication of TRADS in not only hepatocytes but also other normal cells is simultaneously reduced by insertion of both miR-122a complementary sequences and sequences complementary to miRNAs highly expressed in normal cells, without altering the tumor cell lysis activity.

TRADS containing miR-122a complementary sequences are also considered to be promising for the treatment of liver cancer because miR-122a is significantly downregulated in liver cancer cells (35–37) leading to efficient replication and lytic activity of TRADS containing miR-122a complementary sequences in liver cancer cells. This study has shown that TRAD-122aT/199aT caused efficient cell lysis in a hepatocellular carcinoma cell line, HepG2 cells, while the replication of TRADS containing the miR-122a complementary sequences in normal hepatocytes, which highly express miR-122a, was significantly inhibited.

The expression levels of miRNAs are a crucial factor to suppress the gene expression by miRNAs. Brown and colleagues showed that miRNAs should be expressed at a concentration above the threshold (>100 copies/pg small RNA) to induce miRNA-regulated suppression of transgene expression (34). We were not able to precisely show the expression levels of miRNAs as the ratio of copies/pg small RNA in this study; however, comparing the miRNA levels in this study with those reported by Brown and colleagues (34), we consider that the expression levels of miR-143, -145, and -199a in the normal cells were higher than 100 copies/pg small RNA, leading to efficient suppression of the replication of TRADS.

Several studies have shown that let-7, including let-7a, is significantly downregulated in tumor cells (16, 19, 20). Edge and colleagues reported that insertion of let-7a complementary sequences into the matrix protein expression cassette of the vesicular somatitis virus (VSV) suppressed the replication of VSV in human primary fibroblast MG38 cells; on the other hand, VSV carrying let-7a target sequences efficiently replicated in A549 cells (38). However, our data showed that cancer cell lines other than HepG2 cells expressed similar or higher levels of let-7a than the normal cells. In addition, the expression levels of let-7a were more than 10-fold higher than those of the other miRNAs in the tumor cells. Abundant let-7a expression leads to a reduction in the replication of TRAD-let7aT in tumor cells. Furthermore, the members of the let-7 family, including let-7b and let-7c, have the same seed sequence, suggesting that let-7 family members other than let-7a would also contribute to the significant suppression of replication of TRAD-let7aT. These results suggest that not only expression profiles of miRNAs but also absolute amounts of miRNA expression in the cells are of great importance for miRNA-regulated gene expression.

Our data showed that the E1A mRNA levels were reduced by approximately 30% to 50% for TRAD-143T, -145T, and -199aT, compared with the conventional TRAD 24 hour after infection with the normal cells. These reduction levels in the E1A mRNA were much smaller than those in the Ad genome copy numbers at 5 days after infection; however, these reductions in the E1A mRNA levels would lead to large differences in the Ad genome copy numbers after several virus replication cycles. More than 5-fold reductions in the E1A mRNA were found for TRAD-143T, -145T, and -199aT, compared with the parental TRAD, 5 days after infection with the normal cells (data not shown).

A phase I clinical trial of the parental TRAD was conducted, and serious adverse events were not observed (3). In this study, efficient replication of the conventional TRAD in WI38 cells was found at an MOI of 10; however, the conventional TRAD did not exhibit a high level of replication at an MOI of 2. It might be unlikely that such a high titer (MOI 10) of oncolytic Ad would infect organs distal from the injection points in clinical trials; however, normal cells around the injection points might be infected with a high titer of oncolytic Ad. In addition, even though no apparent replication of TRADS is observed in normal cells after infection of TRADS, the expression of Ad proteins, including E1A and E4 proteins, affects the cellular functions via various mechanisms (39–41). This study indicates that inclusion of an miRNA-regulated E1 gene expression system in oncolytic Ads enhances the safety of oncolytic Ads and makes it possible to increase the injection doses, leading to superior therapeutic effects.
In summary, we developed TRADs in which the E1 gene expression is controlled by miRNAs more highly expressed in normal cells than tumor cells. The TRADs containing the sequences complementary to miR-143, -145, or -199a exhibited reduced replication in the normal cells without altering the tumor cell lysis activity. Furthermore, incorporation of both miR-199a and miR-122a target sequences significantly suppressed the replication in all human primary cells examined, including hepatocytes. TRAD-miRT has enhanced both the safety profiles and comparable tumor cell lysis activity to the parental TRAD, suggesting that TRAD-miRT offers great potential for the treatment of tumors.

Disclosure of Potential Conflicts of Interest

Toshiyoshi Fujiwara and Hiroyuki Mizuguchi are consultants to Oncolys BioPharma, Inc. No other potential conflicts of interest were disclosed.

References

15. Hitt MM, Graham FL. Adenovirus E1A under the control of heterologous promoters: wide variation in E1A expression levels has little effect on virus replication. Virology 1990;179:667–78.

Acknowledgments

We thank Takako Ichinose, Koyori Yano (National Institute of Bio- medical Innovation, Osaka, Japan), and Sayuri Okamoto (Graduate School of Pharmaceutical Sciences, Osaka University, Osaka, Japan) for their help.

Grant Support

Support was received from a grant-in-aid for Young Scientists (A) F. Sakurai from the Ministry of Education, Culture, Sports, Science, and Technology (MEXT) of Japan (F. Sakurai), and a grant from the Takeda Science Foundation (H. Mizuguchi).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received July 28, 2010; revised November 18, 2010; accepted December 14, 2010; published OnlineFirst February 23, 2011.
Enhanced Safety Profiles of the Telomerase-Specific Replication-Competent Adenovirus by Incorporation of Normal Cell-Specific microRNA-Targeted Sequences

Kumiko Sugio, Fuminori Sakurai, Kazufumi Katayama, et al.

Clin Cancer Res Published OnlineFirst February 23, 2011.

Updated version
Access the most recent version of this article at:
doi:10.1158/1078-0432.CCR-10-2008

Supplementary Material
Access the most recent supplemental material at:
http://clincancerres.aacrjournals.org/content/suppl/2011/05/05/1078-0432.CCR-10-2008.DC1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.