

Prognostic Significance of Transforming Growth Factor β Receptor II in Estrogen Receptor-Negative Breast Cancer Patients

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ABSTRACT

Purpose: The role of transforming growth factor β (TGF- β) in breast cancer is ambiguous; it can display both tumor suppressing and enhancing effects. Activation of the TGF- β signal transduction system is subject to hormonal regulation. This study was conducted to further analyze the role of TGF- β receptors in breast cancer and to evaluate their significance as prognostic markers.

Experimental Design: Expression of TGF- β receptor I (T β RI) and TGF β receptor II (T β RII) was retrospectively analyzed by immunohistochemistry in 246 breast cancer patients.

Results: Expression of T β RI was strongly correlated with tumor size ($P < 0.001$) and nodal status ($P = 0.012$) but only weakly with overall survival ($P = 0.056$). In contrast, T β RII was prognostic for overall survival in univariate analysis ($P = 0.0370$). In estrogen receptor (ER) -negative patients T β RII expression was correlated with highly reduced overall survival ($P = 0.0083$). In multivariate analysis T β RII proved to be an independent and highly significant prognostic marker with a hazard ratio of 6.8. Simultaneous loss of both ER and T β RII was associated with longer overall survival times comparable with those of ER-positive patients.

Conclusions: The results of this exploratory study show that T β RII is an independent, highly significant prognostic indicator for overall survival in ER-negative patients. In addition our results are supportive of a mechanism of breast cancer progression in which a selective loss of the tumor inhibitory action of TGF β takes place, whereas tumor-promoting aspects remain intact.

INTRODUCTION

Breast cancer is the most common malignancy in women of the Western world. Generally accepted prognostic factors are nodal status, tumor size, and tumor grading. Moreover, breast tumors can be classified as either estrogen receptor (ER) positive or ER negative. The presence of ER is correlated with a better prognosis and predicts for response to antiestrogen treatment. Tamoxifen is currently the most frequently used endocrine therapy for ER-positive breast cancer patients (1). We have shown previously that the action of tamoxifen is at least partially mediated through activation of transforming growth factor- β (TGF- β ; Refs. 2–6).

TGF- β is a pleiotropic growth factor, which affects many different cell functions such as proliferation, extracellular matrix synthesis, and immune responses (7).

TGF- β signals are mediated by specific transmembrane receptors. TGF- β receptor type I and II (T β RI, T β RII) are serine-threonine kinases, which form a heteromeric signal transduction complex upon ligand binding. T β RII phosphorylates T β RI, which activates T β RI kinase and initiates downstream signaling (8). Expression of T β RII is hormonally regulated, but expression of T β RI is not (9).

Conflicting data exist about the influence of TGF- β on the development and progression of breast cancer. TGF- β is a very potent inhibitor of primary human mammary epithelial cells, and most human breast cancer cell lines are growth inhibited by TGF- β as well (10, 11). However, in later tumor stages TGF- β appears to become a promoter of progression (12, 13), and stimulation of angiogenesis, induction of extracellular matrix degradation, or the inhibition of antitumor immune responses prevail over the inhibitory effects on proliferation.

Loss of expression or functional inactivation of T β RI or T β RII leads to resistance against TGF- β (14, 15). Furthermore, defects in downstream signaling components have been associated with altered sensitivity to TGF- β and tumor progression in different tumor types (16, 17).

However, the clinical implications of such findings are still unclear. Therefore, in a retrospective immunohistochemical study we analyzed the correlation of T β RI and T β RII expression with overall survival in 246 breast cancer patients with a median follow-up of 5.7 years. To discriminate between hormone-dependent and -independent effects patients were additionally stratified for their ER status.

Our results show that expression of T β RII is correlated with poor prognosis and represents an independent prognostic marker in the subgroup of ER-negative patients.

MATERIALS AND METHODS

Patients. Paraffin-embedded primary tumors of 246 breast cancer patients were obtained from the archives of surgical pathology of the Robert Bosch Hospital (Stuttgart, Germany). The median age was 55 years (range, 26–88). Histolog-

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ical typing showed 196 ductal invasive breast tumors, 25 lobular invasive and 25 tumors of miscellaneous type (medullar, inflammatory, or mucinous). The median follow-up was 5.7 years (range, 2 months to 15.3 years). Tumor staging was performed according to the Tumor-Node-Metastasis classification system, and histological typing was done according to WHO guidelines (18). ER and progesterone receptor (PR) were analyzed by either immunohistochemistry or the charcoal dextran method. ER and PR were classified as positive when either 15 fmol/mg protein or an immunoreactive score ≥ 2 was reached (19). Of the 246 patients 148 were classified as ER positive and 87 as ER negative. There were 142 PR-positive patients and 92 PR negative.

All of the patients had initially undergone either mastectomy or a breast-conserving resection of their primary carcinomas. There were 166 patients being treated in the adjuvant setting, and 44 received palliative treatment after relapse. Adjuvant treatment with tamoxifen was given to 71 patients, 49 patients received adjuvant treatment with cyclophosphamide, methotrexate and fluorouracil, 46 with anthracycline regimens (either mitoxantrone and cyclophosphamide; fluorouracil, doxorubicin and cyclophosphamide; or bonnadonna regimen), and 91 patients were treated with radiation. The sum of all treatments is >166 , as some patients received more than one treatment.

Antibodies and Immunohistochemical Method. Affinity-purified rabbit polyclonal antibodies raised against T β RI (R-20) and T β RII (C-16; Santa Cruz Biotechnology, Inc., Heidelberg, Germany) were used for immunostaining of the tissues. Both antibodies were used previously for immunohistochemical studies by other groups (20–23).

Sections of 3 μ m were prepared from the paraffin block of each patient tumor. The tissue sections were deparaffinized and rehydrated in descending alcohol dilutions. Before staining on a TechMate instrument (Dako, Glostrup, Denmark) the tissue sections were subjected to antigen retrieval in microwave oven using a citrate buffer solution (Dako). Endogenous biotin, biotin receptors, or avidin binding sites present in the tissue were blocked using an Avidin/Biotin Blocking kit (Vector Laboratories, Inc., Burlingame, CA). Endogenous peroxidase activity was blocked by treatment with hydrogen peroxide. Staining was performed with the Dako ChemMate Detection kit, Peroxidase/3,3'-diaminobenzidine. The slides were incubated with primary antibodies for 25 min at room temperature. The optimum concentration of antibodies for staining, as determined by previous experiments, was 5 μ g/ml for anti-T β RI and 2 μ g/ml for anti-T β RII. Tissues were then incubated with biotinylated goat anti-rabbit IgG (diluted 1:100) for 25 min at room temperature, followed by incubation with peroxidase-conjugated streptavidin for 25 min. The immunoreaction was visualized by using diaminobenzidine in the presence of H₂O₂ resulting in brown colored final reaction products. Tissues were counterstained with hematoxylin.

Skin tissue was used as positive control. Negative controls were obtained by omission of the primary antibody.

The stained tumor tissues and surrounding normal tissues were scored blindly with respect to clinical patient data.

Staining intensity was visually scored in four degrees: absent (0), weak (1), moderate (2), and strong (3). The percentage of T β RI- or T β RII-positive tumor cells was graded as

absent (0), 1–20% (1), 21–50% (2), 51–80% (3), and 81–100% (4). An immunoreactive score (IRS) index was calculated as the product of both values. With respect to the well-known heterogeneity of breast tumors initially the IRS was used for analysis of T β RI or T β RII expression. However, further analysis revealed that heterogeneity in the samples was not sufficient to justify the continuous use of IRS for clinical application. Therefore, we reduced our assessment to two grades, because it has been already shown for scoring of erbB-2 staining in terms of the decision of treatment with Herceptin (24). For these reasons tissue samples were classified into either IRS = 0 (negative) or IRS >0 (positive) staining for TGF- β receptors.

Statistical Methods. Data assessment was made using the statistics software program SPSS (SPSS Software GmbH, Munich, Germany). Survival curves were established by the Kaplan-Meier method, and comparisons between survival curves were performed by the log-rank test. Patients who died from unrelated causes were considered as censored by the time of their death.

Multivariate analysis in the subgroup of ER-negative patients was performed using Cox regression analysis in a model, which included tumor size, nodal status, distant metastases, and grading.

Differences in the TGF- β receptor expression between normal and tumor tissues, and associations between TGF- β receptor expression and other parameters such as tumor size, nodal status, grading, and hormonal status were assessed by χ^2 - or Fisher's exact-test.

RESULTS

Characteristics of the Patients. Kaplan-Meier survival curves were calculated to evaluate the prognostic value of clinical factors and biological markers for overall survival. The median follow-up was 5.7 years (range, 2 months to 15.3 years). The results of the univariate analyses are shown in Table 1. As expected there was a significant association between the classical prognostic factors (tumor size, nodal status, distant metastases, grading, and ER) and outcome. No correlation was observed for menopausal and PR status.

Expression of T β RI and T β RII in Breast Cancer Tissue and Adjacent Normal Tissue. The tissue samples were classified into either positive or negative (not detectable) staining for TGF- β receptors. As far as possible normal tissue adjacent to the tumor tissue was analyzed as well. Expression of T β RI and T β RII was evaluated in 39 normal tissues.

T β RI was detected in 54% of the normal tissue samples. In tumor tissue expression of T β RI was significantly more frequent. Of the tumor samples, 78% showed positive staining for the receptor ($P = 0.0012$). T β RII on the other hand was detected in the majority of the samples, and no significant differences between normal (64%) and tumor (72%) tissues were observed (Table 2).

Fig. 1 shows a representative immunohistochemical staining for T β RI and T β RII in tumor and adjacent normal tissue.

Correlation between T β RI and T β RII Expression and Prognostic Markers. The correlation of T β RI and T β RII expression with tumor size, nodal status, distant metastases, histological grading, tumor stage, ER, PR, and menopausal

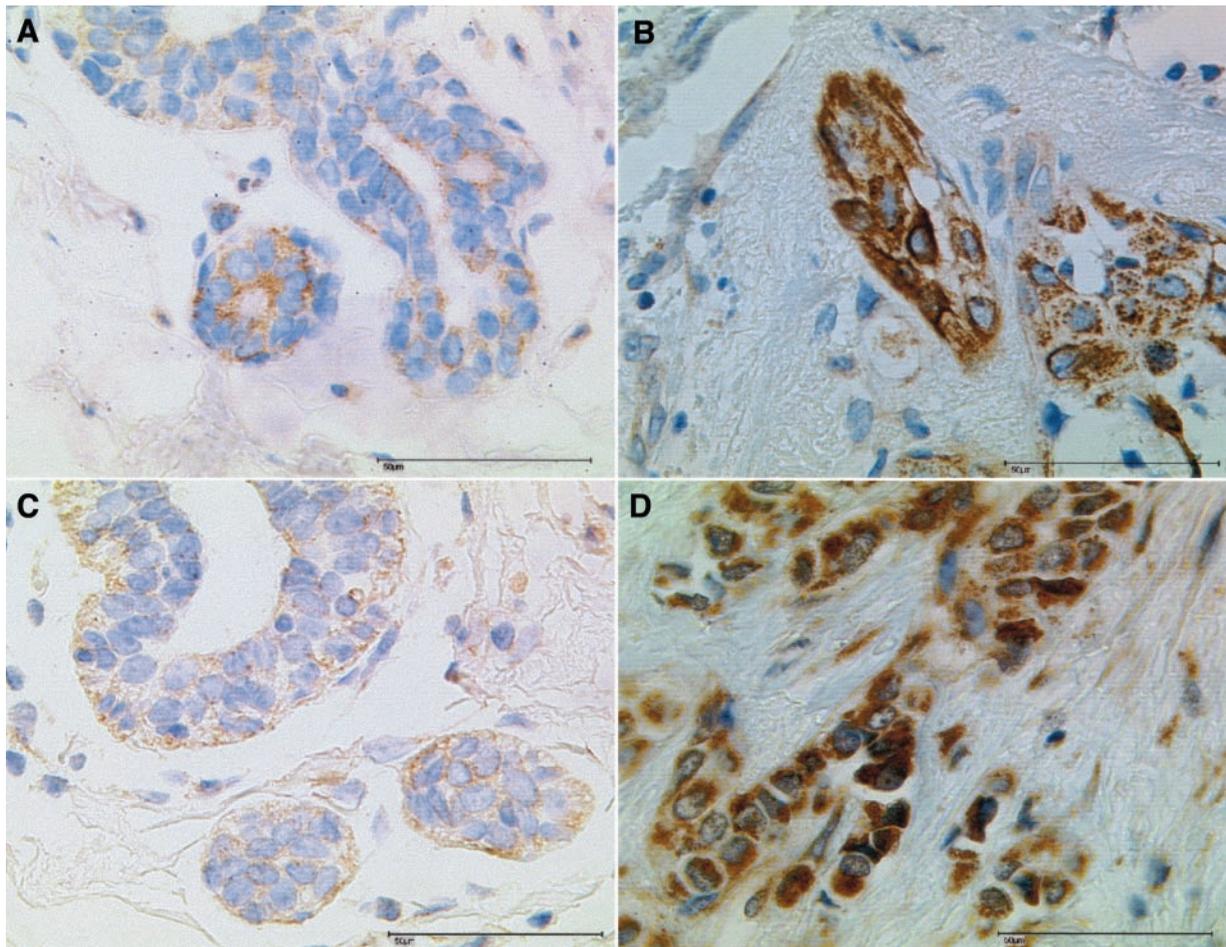


Fig. 1 Immunohistochemical analysis of transforming growth factor- β receptor (TBR) I and TBR II expression in breast cancer tissue and adjacent normal tissue. Immunohistochemical staining was performed using polyclonal antibodies (TBR I, R-20; TBR II, C-16; both Santa Cruz Biotechnology) and a modified avidin-biotin-peroxidase complex technique. **A**, TBR I, normal breast tissue, staining intensity: 1, positive staining in 50% of cells, immunoreactive score (IRS): 2. **B**, TBR I, breast cancer tissue, staining intensity: 2, positive staining in 80% of cells, IRS: 6. **C**, TBR II, normal breast tissue, staining intensity: 1, positive staining in 50% of cells, IRS: 2. **D**, TBR II, breast cancer tissue, staining intensity: 3, positive staining in 80% of cells, IRS: 9. Specimens were counterstained with hematoxylin.

status of the patients was analyzed. The results are presented in Table 3.

TBR I showed a very strong correlation with tumor size and nodal status. The receptor was most frequently expressed in tumors >2 cm in diameter ($P > 1$; $P < 0.001$) and in tumors of node-positive patients ($P = 0.012$). No correlations were observed with distant metastases, grading, hormone receptor, or menopausal status.

TBR II showed no significant correlations with any of the parameters analyzed.

Expression of the TGF- β Receptors and Overall Survival. Loss of TGF- β receptor expression has been associated with loss of TGF- β growth-inhibitory effects and progression to more aggressive tumor types (21, 25). However, these studies did not consider overall patient survival. To assess the influence of TGF- β receptor expression on prognosis of breast cancer patients Kaplan-Meier survival curves were calculated and log-rank analysis performed.

Surprisingly, patients with detectable expression of TBR II

had significantly shorter overall survival times in comparison with patients with undetectable receptor expression ($P = 0.0370$; Fig. 2B). The mean overall survival time was 11.5 years [95% confidence interval (CI), 10.3–12.7] in patients without TBR II expression and 10.4 years (95% CI, 9.5–11.4) in patients with detectable expression of TBR II.

Similar results were obtained for TBR I. The effect of TBR I expression on the overall patient survival, however, was only nearly statistically significant ($P = 0.0560$; Fig. 2A) and probably due to the strong correlation of TBR I expression with tumor size and nodal status (Table 3).

ER Subgroup Analysis. To take hormonal influences into consideration the tissue specimen were additionally subgrouped into either ER positive or ER negative, and Kaplan-Meier survival curves were calculated.

Expression of TBR II defined a subset of patients in the ER-negative subgroup with strongly reduced overall survival times ($P = 0.0083$; Fig. 3B). The mean overall survival time was 11.3 years (95% CI, 10.2–12.4) for TBR II-negative patients

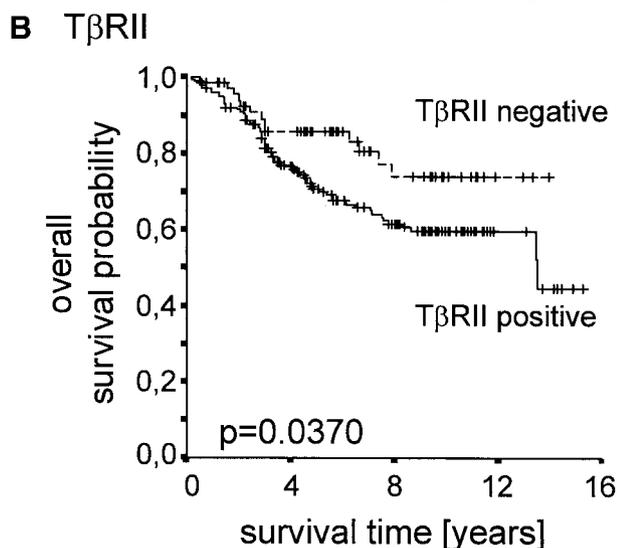
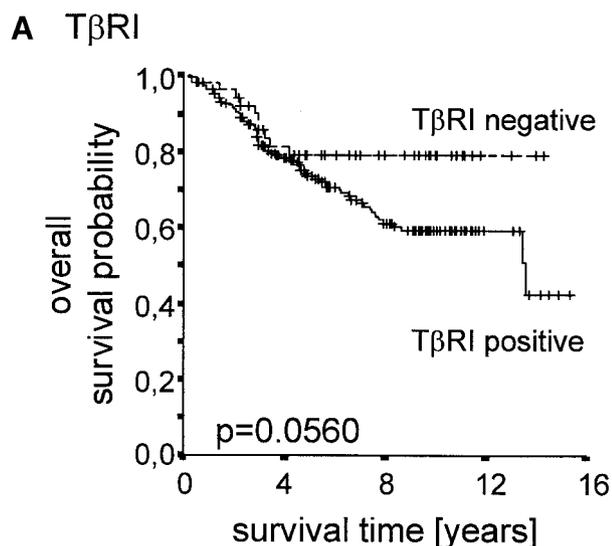


Fig. 2 Kaplan-Meier overall survival analysis for transforming growth factor- β receptor (T β R) I (A) or T β RII (B).

compared with only 8.3 years (95% CI, 6.8–9.7) for T β RII-positive patients. Additional stratification for treatment regimens (cyclophosphamide, methotrexate, and fluorouracil; and anthracyclin containing) gave similar results. In both treatment groups patients with detectable expression of T β RII had a considerably worse prognosis than patients without detectable expression of T β RII (data not shown).

In the ER-positive subgroup, on the other hand, expression of T β RII was without influence on the overall survival ($P = 0.7035$; Fig. 3A). The mean overall survival time was 11.1 years (95% CI, 9.6–12.6) for T β RII-negative patients and 11.4 years (95% CI, 10.3–12.6) for T β RII-positive patients.

T β RI expression had no effect on overall survival after stratification for the ER status. In the ER-positive as well as in the ER-negative subgroup patients with detectable T β RI expression had shorter survival times but differences to T β RI-negative patients were not significant (data not shown).

Multivariate Analysis. A multivariate Cox proportional hazards regression analysis was carried out to establish if expression of T β RII was an independent prognostic marker in the subgroup of ER-negative patients. The model initially included all of the parameters that were predictive of overall survival in the univariate analysis of the entire study group as presented in Table 1 (tumor size, nodal status, distant metastases, and tumor grading). A forward stepwise procedure was adopted to obtain the final model of significant predictors for overall survival consisting of the factors distant metastases, nodal status, and expression of T β RII. Inclusion of tumor size or grading into this model did not improve the log partial likelihood significantly. In the subgroup of ER-negative patients expression of T β RII was strongly associated with poor outcome, with a hazard ratio of 6.8 (Table 4).

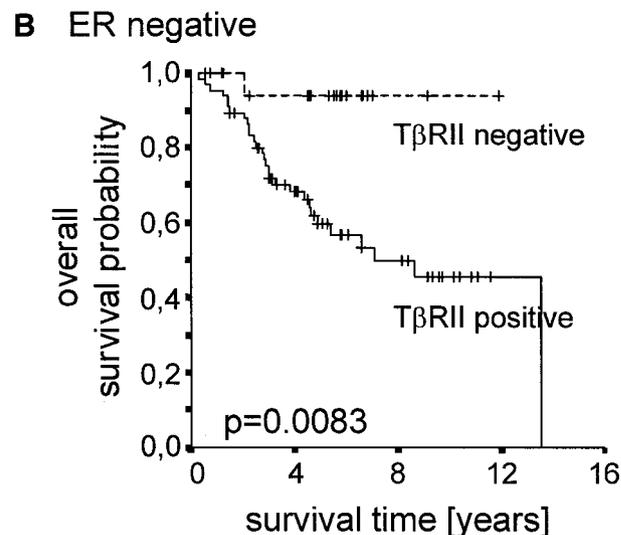
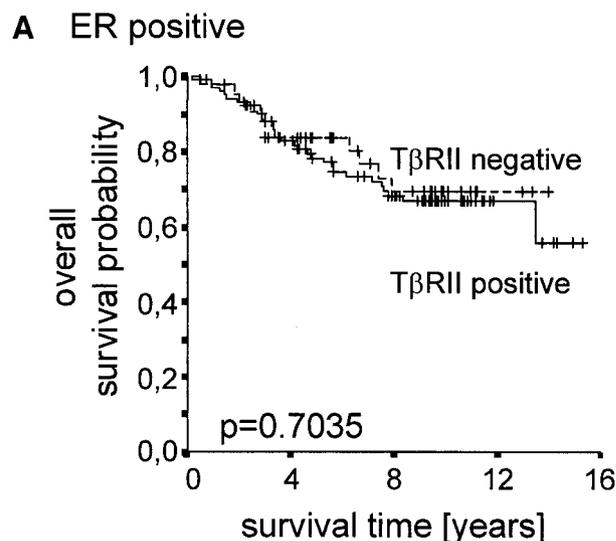


Fig. 3 Kaplan-Meier overall survival analysis for transforming growth factor- β receptor (T β R) II after stratification for estrogen receptor (ER). A, ER-positive patients; B, ER-negative patients.

Table 1 Summary statistics on patients clinical data and results of univariate analysis of classical prognostic factors ($n = 246$)

Factor	No. of patients	Overall survival			
		Years (mean)	95% confidence interval (for the mean)	P^a	
tumor size	pT = 1	70	13.5	12.4–14.5	0.0002
pT	pT > 1	174	9.5	8.6–10.3	
nodal status	T1/T2/T3/T4/not known	70/119/22/33/2			<0.0001
	pN = 0	106	12.8	11.9–13.7	
pN	pN > 0	138	9.3	8.1–10.4	<0.0001
	N0/N1/N2/N3/not known	106/119/12/7/2			
distant metastases	no	229	11.4	10.6–12.2	<0.0001
	yes	10	2.5	1.4–3.7	
	not known	7			
grading	G1 + G2	164	11.7	10.8–12.6	0.0050
	G3	78	8.7	7.3–10.1	
	G1/G2/G3/not known	9/155/78/4			
ER ^b	positive	148	11.6	10.6–12.5	0.0281
	negative	87	9.2	7.9–10.5	
	not known	11			
PR	positive	142	11.2	10.2–12.2	0.4725
	negative	92	10.1	8.9–11.3	
	not known	12			
menopausal status	pre	48	11.3	9.7–12.9	0.3278
	post	127	10.1	9.0–11.2	
	not known	71			

^a Log-rank test.^b ER, estrogen receptor; PR, progesterone receptor.

DISCUSSION

The role of TGF- β in breast cancer progression is unclear. TGF- β can display both tumor-suppressive and tumor-promoting effects. The hormonal influence on activation of the TGF- β system adds an additional layer of complexity. A central role in TGF- β signal transduction is played by the TGF- β receptors. TGF- β signals are mediated by an activated complex of T β RI and T β RII (8). Downstream of the receptors different signal transduction pathways have been implicated in TGF- β signaling (26–30).

Thus far only a few studies have examined the role of the TGF- β receptors in breast cancer tissues; none of these studies considered the influence of receptor expression on disease outcome. In this retrospective exploratory study we have therefore analyzed the expression of T β RI and T β RII in 246 human breast cancer tissues and adjacent normal tissues, and evaluated their association with prognosis. To assess hormonal influences on the TGF- β system we additionally included the ER status of the tumors into the analysis.

It has been suggested previously that loss of T β RII expression may contribute to breast cancer progression and may be associated with a more aggressive phenotype (21). Our data show only a slight loss of T β RII expression with increasing tumor grade, which does not reach statistical significance. T β RII was expressed in a large part of the normal (64%) as well as the tumor tissue (72%; Table 2). The number of tumors with detectable expression of T β RII decreased only slightly with high tumor grading (G₁ + G₂ 75%, G₃ 64%). T β RII was not correlated with any of the clinical parameters analyzed, including tumor size, node status, distant metastasis, grading, ER, PR, and menopausal status (Table 3). However, expression of T β RII had a strong negative influence on prognosis. Patients with

detectable expression of T β RII had significantly decreased survival times ($P = 0.0370$; Fig. 2B). These data indicate that in breast cancer, loss of TGF- β growth-inhibitory effects is not caused by loss of T β RII expression. Similar results were obtained previously for pancreatic cancer in which enhanced expression of T β RII was significantly correlated with reduced overall survival. In pancreatic cancer increased expression of T β RII was associated with high expression of plasminogen activator inhibitor 1 and matrix metalloproteinase 9 indicating a dissociation between TGF- β signals leading to growth inhibition and extracellular matrix production (31).

Because TGF- β signals are mediated by a heteromeric complex of T β RI and T β RII another possible explanation for the absence of TGF- β growth-inhibitory effects and tumor progression in breast cancer could be a loss of T β RI expression. In colon and prostate cancer loss of T β RI expression was associated with increased malignant potential (25, 32, 33). No comparable studies exist for breast cancer. In our patient collective, T β RI was expressed in most of the tumor tissues (78%; Table 2), and expression was more often detected in tumors of lower

Table 2 Transforming growth factor- β receptor (T β R) I and T β RII expression in normal and tumor tissue percentages in parenthesis

	T β RI		T β RII	
	Normal	Tumor	Normal	Tumor
Negative	18 (46)	54 (22)	14 (36)	69 (28)
Positive	21 (54)	192 (78)	25 (64)	177 (72)
Total	39	246	39	246
P^a	0.0012		0.3162	

^a χ^2 test.

Table 3 Cross-tabulation of transforming growth factor- β receptor (T β R) I and T β RII with other tumor variables in breast cancer, percentages in parenthesis

		T β RI		<i>P</i> ^a	T β RII		<i>P</i> ^a
		Negative	Positive		Negative	Positive	
Tumor size	pT = 1	26 (37)	44 (63)	<0.001	19 (27)	51 (73)	0.803
	pT > 1	28 (16)	146 (84)		50 (29)	124 (71)	
Nodal status	pN = 0	31 (29)	75 (71)	0.012	32 (30)	74 (70)	0.562
	pN > 0	22 (16)	116 (84)		37 (27)	101 (73)	
Distant metastasis	no	52 (23)	177 (77)	0.081	65 (28)	164 (72)	0.578
	yes	0 (0)	10 (100)		3 (30)	7 (70)	
Grading	G1–G2	39 (24)	125 (76)	0.427	41 (25)	123 (75)	0.079
	G3	15 (19)	63 (81)		28 (36)	50 (64)	
ER ^b	positive	32 (22)	116 (78)	0.866	44 (30)	104 (70)	0.355
	negative	18 (21)	69 (80)		21 (24)	66 (76)	
PR	positive	31 (22)	111 (78)	0.677	40 (28)	102 (72)	0.868
	negative	18 (20)	74 (80)		25 (27)	67 (73)	
Menopausal status	pre	6 (13)	42 (87)	0.069	13 (27)	35 (73)	0.883
	post	32 (25)	95 (75)		33 (26)	94 (74)	

^a χ^2 test or Fisher's exact test.

^b ER, estrogen receptor; PR, progesterone receptor.

grade (81%; Table 3). Furthermore, expression of T β RI had a weak negative effect on prognosis. However, this correlation with survival is probably because T β RI expression was significantly more often detected in larger and node-positive tumors (Table 3), two factors known to have a strong negative impact on survival (1).

In summary our results suggest that loss of TGF- β growth-inhibitory effects in breast cancer cannot be attributed to either loss of T β RI or T β RII expression.

It has been implicated before that in advanced tumor stages the growth-inhibitory component of TGF- β signaling selectively gets lost, whereas tumor promoting effects gain importance (13, 31). In some tumor types active TGF- β signaling seems to play an important role in the progression to more aggressive phenotypes. Several reports exist that link TGF- β signaling to cell invasiveness and formation of metastasis (13, 34–36).

Our data support a role for TGF- β in breast cancer progression, because both receptors were expressed in the majority of tumor tissues and expression was associated with poor outcome. It can be assumed that the TGF- β receptors detected in this study are able to transduce signals, because mutational inactivation of T β RI or T β RII appears to be a rare event in breast cancer. Thus far the only mutation in T β RI has been found in breast cancer metastases, but this mutation does not seem to occur frequently (37). Four inactivating mutations in T β RII have been found recently in recurrent breast tumors, but

no T β RII mutations have been detected in primary breast cancers (38). In addition, an immunohistochemical study on the expression of Smad2, phosphorylated Smad2, and Smad4 suggests, that the majority of invasive breast carcinomas are able to actively mediate TGF- β signals (17).

The present study indicates that the ER status of a tumor is an important marker for the transition of TGF- β from tumor suppressor to tumor promoter. In ER-negative tumors expression of T β RII was associated with a subset of tumors that seemed to be highly aggressive leading to strongly reduced overall survival times (8.3 years; Fig. 3B). Simultaneous loss of both ER and T β RII, on the other hand, was associated with longer overall survival times (11.3 years) comparable with those of ER-positive patients with and without T β RII expression (11.4 years and 11.1 years, respectively). Expression of T β RII proved to be an independent prognostic marker in the subgroup of ER-negative patients (Table 4).

Differences in treatment appear not to influence the impact of T β RII expression on overall survival in ER-negative patients. We distinguished between the two most commonly used treatment regimens, namely cyclophosphamide, methotrexate, and fluorouracil, and anthracycline-containing regimens. In each treatment group prognosis was worse for patients with detectable expression of T β RII.

It has already been shown that a cross-talk between growth factors and steroid hormone receptors may be relevant to the regulation of growth and differentiation processes in hormone

Table 4 Cox proportional hazard model for overall survival of estrogen receptor-negative patients

Sequential inclusion of factors in the model	Improvement of $-2 \log$ partial likelihood	<i>P</i> ^a	Hazard ratio	(95% confidence interval)
Distant metastases	5.059	0.025	22.3	(2.0–247.9)
Positive nodes	9.927	0.002	3.6	(1.4–9.1)
Expression of T β RII	6.517	0.011	6.8	(0.9–50.6)
pT >1/grading >1	0.204/0.118	0.652/0.732	–	–

^a *P*s were derived from the Cox proportional hazards model, with inclusion of all factors shown.

responsive tissues: the effect of epidermal growth factor seems to be partially mediated through the ER even in the absence of estradiol (39, 40). A similar cross-talk appears to exist for TGF- β signaling and the ER. The transcriptional activity of Smad3 can be suppressed by ER, whereas ER-mediated transcriptional activity can be increased by activation of TGF- β signaling (41).

In ER-positive tumors, TGF- β seems to act in an autocrine inhibitory fashion because a rapid increase in TGF- β 2 levels under treatment with tamoxifen was correlated previously with clinical remission in patients with metastatic breast cancer (6). Loss of ER could evoke distinct changes in the cellular response to TGF- β and represent the starting point for a loss of TGF- β growth-inhibitory effects.

Therefore, a therapeutic approach that inhibits TGF- β signal transduction might turn out to be specifically effective for ER-negative patients with detectable expression of T β RII. A soluble TGF- β receptor type II protein that interferes with TGF- β binding to endogenous TGF- β receptors has been shown previously to reduce tumor cell motility, intravasation, and distant metastasis in a mouse model (34).

In conclusion, in our study T β RII proved to be an independent prognostic marker in the subgroup of ER-negative patients. According to the criteria of the American Society of Clinical Oncology (1) this study can be classified as evidence level III. Our results support a mechanism for breast cancer progression in which a selective loss of the growth-inhibitory action of TGF- β takes place, whereas tumor-promoting aspects of TGF- β signaling remain intact. We identified the ER as an important marker for changes in the tumor response to TGF- β . The precise interactions between ER and TGF- β signaling on the molecular level remain to be clarified. However, analysis of T β RII expression in ER-negative patients could be relevant for treatment decisions, as patients without detectable expression of T β RII had a prognosis similar to that of ER-positive patients.

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REFERENCES

- Souhami, R. L., Tannock, I., Hohenberger, P., and Horiot, J. C. *Oxford Textbook of Oncology*. Oxford: New York Oxford University Press, 2002.
- Knabbe, C., Lippman, M. E., Wakefield, L. M., Flanders, K. C., Kasid, A., Derynck, R., and Dickson, R. B. Evidence that transforming growth factor- β is a hormonally regulated negative growth factor in human breast cancer cells. *Cell*, *48*: 417–428, 1987.
- Knabbe, C., Zugmaier, G., Schmahl, M., Dietel, M., Lippman, M. E., and Dickson, R. B. Induction of transforming growth factor β by the antiestrogens droloxifene, tamoxifen, and toremifene in MCF-7 cells. *Am J. Clin. Oncol.*, *14*: S15–S20, 1991.
- Knabbe, C., Kopp, A., Hilgers, W., Lang, D., Muller, V., Zugmaier, G., and Jonat, W. Regulation and role of TGF β production in breast cancer. *Ann. N. Y. Acad. Sci.*, *784*: 263–276, 1996.
- Muller, V., Jensen, E. V., and Knabbe, C. Partial antagonism between steroidal and nonsteroidal antiestrogens in human breast cancer cell lines. *Cancer Res.*, *58*: 263–267, 1998.
- Kopp, A., Jonat, W., Schmahl, M., and Knabbe, C. Transforming growth factor β 2 (TGF- β 2) levels in plasma of patients with metastatic breast cancer treated with tamoxifen. *Cancer Res.*, *55*: 4512–4515, 1995.
- Roberts, A. B. Molecular and cell biology of TGF- β . *Miner. Electrolyte Metab.*, *24*: 111–119, 1998.
- Wrana, J. L., Attisano, L., Wieser, R., Ventura, F., and Massague, J. Mechanism of activation of the TGF- β receptor. *Nature (Lond.)*, *370*: 341–347, 1994.
- Buck, M., von der Fecht, J., and Knabbe, C. Antiestrogenic regulation of transforming growth factor β receptors I and II in human breast cancer cells. *Ann. N. Y. Acad. Sci.*, *963*: 140–143, 2002.
- Basolo, F., Fiore, L., Ciardiello, F., Calvo, S., Fontanini, G., Conaldi, P. G., and Toniolo, A. Response of normal and oncogene-transformed human mammary epithelial cells to transforming growth factor β 1 (TGF- β 1): lack of growth-inhibitory effect on cells expressing the simian virus 40 large-T antigen. *Int. J. Cancer*, *56*: 736–742, 1994.
- Zugmaier, G., Ennis, B. W., Deschauer, B., Katz, D., Knabbe, C., Wilding, G., Daly, P., Lippman, M. E., and Dickson, R. B. Transforming growth factors type β 1 and β 2 are equipotent growth inhibitors of human breast cancer cell lines. *J. Cell Physiol.*, *141*: 353–361, 1989.
- Gorsch, S. M., Memoli, V. A., Stukel, T. A., Gold, L. I., and Arrick, B. A. Immunohistochemical staining for transforming growth factor β 1 associates with disease progression in human breast cancer. *Cancer Res.*, *52*: 6949–6952, 1992.
- McEarchern, J. A., Kobie, J. J., Mack, V., Wu, R. S., Meade-Tollin, L., Arteaga, C. L., Dumont, N., Besselsen, D., Seftor, E., Hendrix, M. J., Katsanis, E., and Akporiaye, E. T. Invasion and metastasis of a mammary tumor involves TGF- β signaling. *Int. J. Cancer*, *91*: 76–82, 2001.
- Kalkhoven, E., Roelen, B. A., de Winter, J. P., Mummery, C. L., van den Eijnden-van Raaij, A. J., van der Saag, P. T., and van der Burg, B. Resistance to transforming growth factor β and activin due to reduced receptor expression in human breast tumor cell lines. *Cell Growth Differ.*, *6*: 1151–1161, 1995.
- Laiho, M., Weis, M. B., and Massague, J. Concomitant loss of transforming growth factor (TGF)- β receptor types I and II in TGF- β -resistant cell mutants implicates both receptor types in signal transduction. *J. Biol. Chem.*, *265*: 18518–18524, 1990.
- Chen, C. R., Kang, Y., and Massague, J. Defective repression of c-myc in breast cancer cells: A loss at the core of the transforming growth factor β growth arrest program. *Proc. Natl. Acad. Sci. USA*, *98*: 992–999, 2001.
- Xie, W., Mertens, J. C., Reiss, D. J., Rimm, D. L., Camp, R. L., Haffty, B. G., and Reiss, M. Alterations of Smad signaling in human breast carcinoma are associated with poor outcome: a tissue microarray study. *Cancer Res.*, *62*: 497–505, 2002.
- TNM Classification of Malignant Tumors, 5 edition: J. Wiley & Sons, New York, 1997.
- Fritz, P., Murdter, T. E., Eichelbaum, M., Siegle, I. I., Weissert, M., and Zanger, U. M. Microsomal epoxide hydrolase expression as a predictor of tamoxifen response in primary breast cancer: a retrospective exploratory study with long-term follow-up. *J. Clin. Oncol.*, *19*: 3–9, 2001.
- Zwaagstra, J. C., Guimond, A., and O'Connor-McCourt, M. D. Predominant intracellular localization of the type I transforming growth factor- β receptor and increased nuclear accumulation after growth arrest. *Exp. Cell Res.*, *258*: 121–134, 2000.
- Gobbi, H., Arteaga, C. L., Jensen, R. A., Simpson, J. F., Dupont, W. D., Olson, S. J., Schuyler, P. A., Plummer, W. D., Jr., and Page, D. L. Loss of expression of transforming growth factor β type II receptor correlates with high tumour grade in human breast *in-situ* and invasive carcinomas. *Histopathology*, *36*: 168–177, 2000.
- Gobbi, H., Dupont, W. D., Simpson, J. F., Plummer, W. D., Jr., Schuyler, P. A., Olson, S. J., Arteaga, C. L., and Page, D. L. Transforming growth factor- β and breast cancer risk in women with mammary epithelial hyperplasia. *J. Natl. Cancer Inst.*, *91*: 2096–2101, 1999.
- Guo, Y., and Kyprianou, N. Overexpression of transforming growth factor (TGF) β 1 type II receptor restores TGF- β 1 sensitivity and signaling in human prostate cancer cells. *Cell Growth Differ.*, *9*: 185–193, 1998.
- Schaller, G., Evers, K., Papadopoulos, S., Ebert, A., and Buhler, H. Current use of HER2 tests. *Ann. Oncol.*, *12* (Suppl. 1): S97–S100, 2001.

25. Kim, I. Y., Ahn, H. J., Zelner, D. J., Shaw, J. W., Lang, S., Kato, M., Oefelein, M. G., Miyazono, K., Nemeth, J. A., Kozlowski, J. M., and Lee, C. Loss of expression of transforming growth factor β type I and type II receptors correlates with tumor grade in human prostate cancer tissues. *Clin. Cancer Res.*, 2: 1255–1261, 1996.
26. Bakin, A. V., Tomlinson, A. K., Bhowmick, N. A., Moses, H. L., and Arteaga, C. L. Phosphatidylinositol 3-kinase function is required for transforming growth factor β -mediated epithelial to mesenchymal transition and cell migration. *J. Biol. Chem.*, 275: 36803–36810, 2000.
27. Frey, R. S., and Mulder, K. M. Involvement of extracellular signal-regulated kinase 2 and stress-activated protein kinase/Jun N-terminal kinase activation by transforming growth factor β in the negative growth control of breast cancer cells. *Cancer Res.*, 57: 628–633, 1997.
28. Hanafusa, H., Ninomiya-Tsuji, J., Masuyama, N., Nishita, M., Fujisawa, J., Shibuya, H., Matsumoto, K., and Nishida, E. Involvement of the p38 mitogen-activated protein kinase pathway in transforming growth factor- β -induced gene expression. *J. Biol. Chem.*, 274: 27161–27167, 1999.
29. Heldin, C. H., Miyazono, K., and ten Dijke, P. TGF- β signalling from cell membrane to nucleus through SMAD proteins. *Nature (Lond.)*, 390: 465–471, 1997.
30. Petritsch, C., Beug, H., Balmain, A., and Oft, M. TGF- β inhibits p70 S6 kinase via protein phosphatase 2A to induce G(1) arrest. *Genes Dev.*, 14: 3093–3101, 2000.
31. Wagner, M., Kleeff, J., Friess, H., Buchler, M. W., and Korc, M. Enhanced expression of the type II transforming growth factor- β receptor is associated with decreased survival in human pancreatic cancer. *Pancreas*, 19: 370–376, 1999.
32. Matsushita, M., Matsuzaki, K., Date, M., Watanabe, T., Shibano, K., Nakagawa, T., Yanagitani, S., Amoh, Y., Takemoto, H., Ogata, N., Yamamoto, C., Kubota, Y., Seki, T., Inokuchi, H., Nishizawa, M., Takada, H., Sawamura, T., Okamura, A., and Inoue, K. Down-regulation of TGF- β receptors in human colorectal cancer: implications for cancer development. *Br. J. Cancer*, 80: 194–205, 1999.
33. Wang, J., Han, W., Zborowska, E., Liang, J., Wang, X., Willson, J. K. V., Sun, L., and Brattain, M. G. Reduced expression of transforming growth factor β type I receptor contributes to the malignancy of human colon carcinoma cells. *J. Biol. Chem.*, 271: 17366–17371, 1996.
34. Muraoka, R. S., Dumont, N., Ritter, C. A., Dugger, T. C., Brantley, D. M., Chen, J., Easterly, E., Roebuck, L. R., Ryan, S., Gotwals, P. J., Kotliansky, V., and Arteaga, C. L. Blockade of TGF- β inhibits mammary tumor cell viability, migration, and metastases. *J. Clin. Investig.*, 109: 1551–1559, 2002.
35. Welch, D. R., Fabra, A., and Nakajima, M. Transforming growth factor β stimulates mammary adenocarcinoma cell invasion and metastatic potential. *Proc. Natl. Acad. Sci. USA*, 87: 7678–7682, 1990.
36. Oft, M., Heider, K. H., and Beug, H. TGF β signaling is necessary for carcinoma cell invasiveness and metastasis. *Curr. Biol.*, 8: 1243–1252, 1998.
37. Anbazhagan, R., Bornman, D. M., Johnston, J. C., Westra, W. H., and Gabrielson, E. The S387Y mutations of the transforming growth factor- β receptor type I gene is uncommon in metastases of breast cancer and other common types of adenocarcinoma. *Cancer Res.*, 59: 3363–3364, 1999.
38. Lucke, C. D., Philpott, A., Metcalfe, J. C., Thompson, A. M., Hughes-Davies, L., Kemp, P. R., and Hesketh, R. Inhibiting mutations in the transforming growth factor β type 2 receptor in recurrent human breast cancer. *Cancer Res.*, 61: 482–485, 2001.
39. Curtis, S. W., Washburn, T., Sewall, C., DiAugustine, R., Lindzey, J., Couse, J. F., and Korach, K. S. Physiological coupling of growth factor and steroid receptor signaling pathways: estrogen receptor knockout mice lack estrogen-like response to epidermal growth factor. *Proc. Natl. Acad. Sci. USA*, 93: 12626–12630, 1996.
40. Ignar-Trowbridge, D. M., Nelson, K. G., Bidwell, M. C., Curtis, S. W., Washburn, T. F., McLachlan, J. A., and Korach, K. S. Coupling of dual signaling pathways: epidermal growth factor action involves the estrogen receptor. *Proc. Natl. Acad. Sci. USA*, 89: 4658–4662, 1992.
41. Matsuda, T., Yamamoto, T., Muraguchi, A., and Saatcioglu, F. Cross-talk between transforming growth factor- β and estrogen receptor signaling through Smad3. *J. Biol. Chem.*, 276: 42908–42914, 2001.

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