

Nuclear Factor- κ B Nuclear Localization Is Predictive of Biochemical Recurrence in Patients with Positive Margin Prostate Cancer

Vincent Fradet,¹ Laurent Lessard,²
Louis R. Bégin,³ Pierre Karakiewicz,¹
Anne-Marie Mes Masson,^{2,4} and Fred Saad^{1,2}

¹Département de chirurgie (urologie), Centre Hospitalier de l'Université de Montréal–Montréal, Montreal, Quebec, Canada; ²Centre de recherche du Centre Hospitalier de l'Université de Montréal and Institut du Cancer de Montréal, Montreal, Quebec, Canada; ³Service d'anatomopathologie, Hôpital du Sacré-Cœur de Montréal, Montreal, Quebec, Canada; and ⁴Département de médecine, Université de Montréal, Montreal, Quebec, Canada

ABSTRACT

Purpose: Radical prostatectomy (RP) patients with positive surgical margins are at increased risk for recurrence, emphasizing the need for prognostic markers to stratify probable outcome for optimal patient management decisions. We tested the hypothesis that nuclear localization of nuclear factor (NF)- κ B, a transcription factor involved in the regulation of cell growth, angiogenesis, invasion, and apoptosis, is associated with an increased risk of biochemical recurrence after RP.

Experimental Design: Analyses addressed data from 42 patients (age range, 52–72 years; mean age, 63.7 years) who exhibited positive surgical margins after RP. Immunohistochemical analysis of NF- κ B (p65) was performed on the positive margin tissue. A nuclear staining cutoff of >5% was considered positive. The relation between nuclear NF- κ B expression and biochemical recurrence (prostate-specific antigen >0.3 ng/mL and rising) after RP was tested in univariate and multivariate Cox regression models.

Results: Biochemical recurrence was recorded in 23 patients (54.8%; median follow-up, 3.2 years). Univariate Cox regression demonstrated a 4.9-fold (95% confidence interval, 1.5–16.7; $P = 0.01$) higher rate of recurrence in

men with NF- κ B > 5%. In the multivariate model, after controlling for primary ($P = 0.004$) and secondary ($P = 0.7$) Gleason patterns, lymph node ($P = 0.06$) and seminal vesicle invasion ($P = 0.2$), and preoperative prostate-specific antigen ($P = 0.009$), NF- κ B > 5% was associated with a 6.2-fold higher risk of biochemical recurrence (95% confidence interval, 1.7–23.5; $P = 0.007$).

Conclusions: In univariate and multivariate analysis, NF- κ B nuclear expression was strongly predictive of biochemical recurrence in patients with positive surgical margins after RP. We propose that nuclear NF- κ B may serve as a useful independent molecular marker for stratifying patients at risk for recurrence.

INTRODUCTION

Prostate cancer is the most commonly diagnosed cancer, and the second leading cause of cancer-related deaths in North American men. Radical prostatectomy (RP) represents one of two standard treatment options for men with clinically localized prostate cancer. It is still controversial whether immediate adjuvant treatment should be instituted in the presence of pathologically confirmed positive surgical margins after RP. Several candidate molecular markers, including RNASEL, GSTP1, p21, p27, p53, NKX3.1, PTEN, and TERT, have been or are currently being studied in an effort to enhance the accuracy of pathological stage and grade and pretreatment serum prostate-specific antigen (PSA), in predicting the risk of biochemical failure after RP (1–3). To date, none have been adopted into routine clinical practice.

Classical nuclear factor (NF)- κ B transcription factor is a heterodimer formed by the p50 and RelA (p65) proteins (4–7). It is expressed in most cells but kept inactive and maintained in the cytoplasm by interaction with I κ B inhibitors. Normally, the activation of NF- κ B requires signals that lead to the phosphorylation (by the activated IKK complex), ubiquitination, and proteasome-dependent degradation of I κ B. NF- κ B then translocates to the nucleus and activates the expression of various genes involved in cell growth, differentiation, inflammatory responses, and the regulation of apoptosis. There is growing evidence that NF- κ B is implicated in oncogenesis (8, 9). NF- κ B is constitutively activated in many lymphoid and nonlymphoid cancers (10, 11). NF- κ B can prevent cell death by apoptosis in many cell types (including the human prostate cancer PC3 and DU145 cell lines) after chemotherapy, radiotherapy, or tumor necrosis factor α treatment (12, 13). More recently, constitutive activation of NF- κ B has been detected in androgen-independent prostate cancer cell lines as well as in prostate cancer tissues (14–17). In particular, strong nuclear NF- κ B staining was observed in prostate cancer lymph node metastases (18). Moreover, NF- κ B expression in RP specimen was shown to correlate with progression to bone metastasis (19). NF- κ B aberrant acti-

Received 4/20/04; revised 7/26/04; accepted 8/16/04.

Grant support: A grant from the Canadian Prostate Cancer Research Foundation and the Canadian Uro-Oncology/AstraZenca award. A-M. Mes Masson is a recipient of a Chercheur National fellowship from the Fonds de la Recherche en Santé du Québec. P. Karakiewicz receives a Chercheur-boursier clinicien Junior 1 award from the Fonds de la Recherche en Santé du Québec. L. Lessard is supported by a Canadian Institute of Health Research/Canadian Prostate Cancer Research Initiative studentship.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Requests for reprints: Fred Saad, Département d'urologie, Centre Hospitalier de l'Université de Montréal–Hôpital Notre-Dame, 1560 rue Sherbrooke E, Montreal, Quebec, H2L 4M1 Canada. E-mail: Fred.Saad.CHUM@sss.gouv.qc.ca.

©2004 American Association for Cancer Research.

vation has been shown to promote cell growth, survival, and metastasis in cell lines and xenograft models (20–25).

Whereas the role of NF- κ B in prostate carcinogenesis and cancer progression is becoming more apparent, the clinical usefulness of NF- κ B as a predictor of disease progression has not yet been fully investigated. We tested the hypothesis that NF- κ B could be a prognostic marker for eventual biochemical recurrence in men with positive pathological surgical margins after RP.

PATIENTS AND METHODS

After providing informed consent, 90 consecutive patients with positive margins were initially identified in a patient database, but 48 were excluded for the following reasons: absence of tissue blocks with true positive margins after central pathological review; detectable postoperative PSA; and neoadjuvant hormone therapy or adjuvant therapy before PSA recurrence. Tissue slides of the remaining 42 formalin-fixed paraffin-embedded specimens were immunostained for NF- κ B transcription factor by the biotin-streptavidin-peroxidase method using the NF- κ B p65 (F-6) monoclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA) that recognizes NH₂-terminal sequences of the p65 subunit.

Tissue sections were deparaffinized with toluene, rehydrated through graded EtOH, treated with 3% H₂O₂ to eliminate endogenous peroxidase activity, and incubated for 10 minutes at 95°C in citrate buffer (pH 6.0) for antigen unmasking. Nonspecific antigen was blocked with a protein blocking serum-free reagent (Dako Diagnostics Canada, Inc., Mississauga, Canada). Slides were then incubated for 60 minutes in a humidified chamber with the specific p65 antibody. The optimal concentration (1 μ g/mL) was determined by serial dilutions. Twenty-minute incubations with the secondary biotinylated antibody and streptavidin-horseradish peroxidase (Dako Diagnostics Canada Inc.) were performed sequentially. Reaction products were developed using diaminobenzidine containing 0.3% H₂O₂ as a substrate for peroxidase. Hematoxylin counterstaining was done for ease of reading. Substitution of the primary antibody with PBS served as a negative control.

Slides were examined under standard light microscopy, and

Table 1 Clinical and pathological characteristics of 42 evaluable patients who were treated with RP

	NF- κ B > 5%	NF- κ B \leq 5%	<i>P</i>
No. of patients	27	15	
Age (y)			0.6
Mean (SE)	64.1 (0.99)	63.1 (1.6)	
PSA (ng/mL)			0.2
Mean (SE)	13.9	10.96 (1.56)	
Gleason sum (%)			0.1
6	4 (14.8)	4 (26.7)	
7	17 (63.0)	10 (66.7)	
8	0	1 (6.7)	
9	6 (22.2)	0	
Seminal vesicle invasion	3	2	0.8
Lymph node metastases	4	0	0.1

NOTE. Data are tabulated according to NF- κ B nuclear staining status.

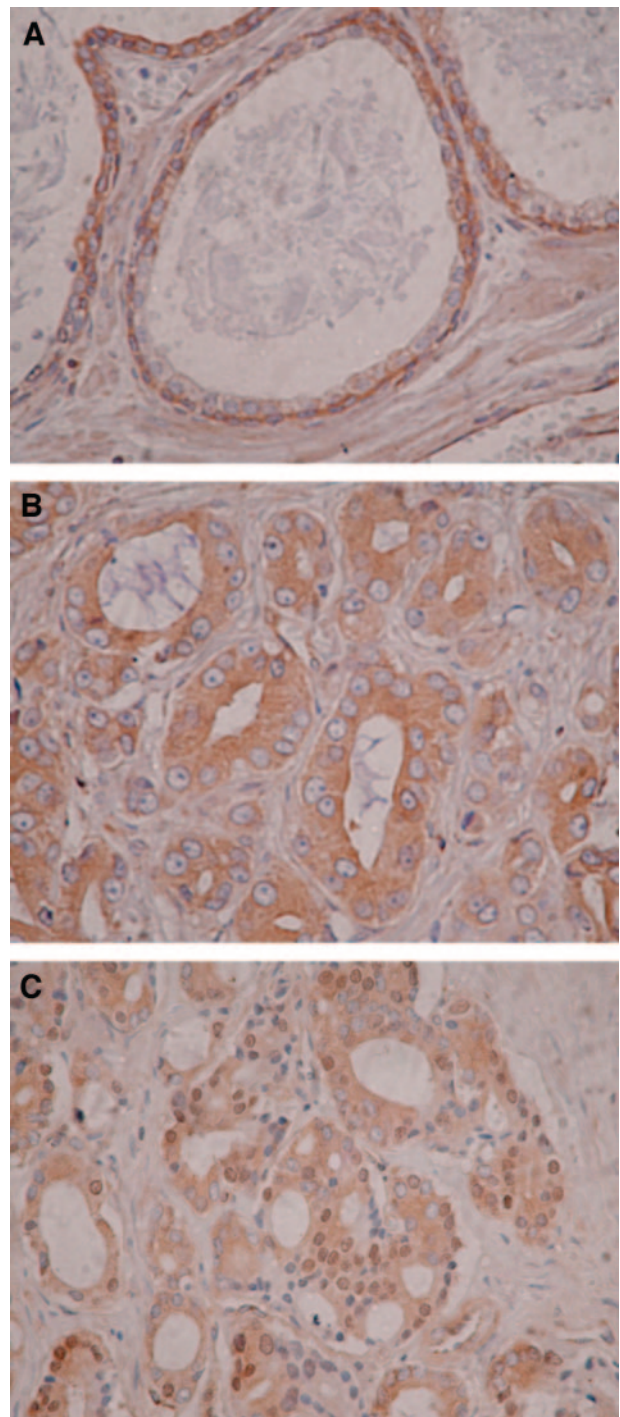


Fig. 1 Immunohistochemical detection of NF- κ B (p65 subunit) in prostate tissues. Note the strong cytoplasmic staining in normal prostate basal cells (A), whereas tumor tissues show either predominant cytoplasmic staining (B) or cytoplasmic and nuclear staining (C).

subcellular staining was identified. NF- κ B nuclear localization was evaluated as the overall proportion of tumor cells with positive nuclear staining. An average of 20 fields per tumor (100 cells per field) were analyzed at \times 400. Positive nuclear staining

Table 2 Univariate and multivariate Cox survival analyses addressing the association between NF- κ B and biochemical prostate cancer recurrence (PSA > 0.3 ng/mL and rising) after RP

Predictors variable	Biochemical recurrence after RP	
	Univariate (rate ratio and 95% CI)	Multivariate (rate ratio and 95% CI)
NF- κ B	4.9 (1.47–16.70); $P = 0.01$	6.2 (1.64–23.53); $P = 0.007$
Age (y)	0.97 (0.90–1.05); $P = 0.4$	0.95 (0.88–1.03); $P = 0.2$
pPSA (ng/mL)	0.96 (0.91–1.02); $P = 0.2$	0.91 (0.85–0.98); $P = 0.01$
Gleason sum	$P = 0.009$	$P = 0.03$
Seminal vesicle invasion	2.5 (0.83–7.46); $P = 0.1$	2.2 (0.65–7.40); $P = 0.2$
Lymph node metastases	2.2 (0.64–7.68); $P = 0.2$	4.8 (0.99–23.62); $P = 0.052$

NOTE. Covariates include age, pPSA, Gleason sum, seminal vesicle invasion, and lymph node metastases. Primary and secondary Gleason patterns were used to define the Gleason sum.

was defined as a sharp brown coloration in the nucleus of a cell. All slides were independently analyzed by two blinded observers. Interpretations diverging <10% were independently reanalyzed, resulting in concordance scores. We used >5% positive tumor cell nuclear staining as a cutoff indicating significant NF- κ B activation in cancer tissue. Univariate biochemical-free survival analyses revealed that 5% represented the most statistically significant cutoff in this data set.

Pretreatment serum prostate-specific antigen (pPSA), pathological Gleason score, presence of extracapsular extension, presence of seminal vesicle invasion, presence of pelvic lymph node metastasis, and surgical margin status were recorded for each patient. Positive surgical margins were defined by the presence of tumor cells at the inked margin of resection.

Postoperative follow-up consisted of physical examination and serial serum PSA measurements. The Hybritech PSA assay (Hybritech Tandem R, San Diego, CA) was used principally, but not exclusively, for both pretreatment evaluation and disease follow-up. PSA failures were defined as serum levels > 0.3 ng/mL and rising, and failure time was dated back to first PSA > 0.3 ng/mL. PSA assays were generally obtained at 1 month after surgery and then every 3 months for the first year, every 6 months for two subsequent years, and then annually. For statistical purposes, patients who did not relapse were censored at the date of their last undetectable PSA assay.

The probability of remaining free from recurrence [mean with 95% confidence intervals (CIs)] was calculated using the Kaplan-Meier method. Univariate and multivariate Cox proportional hazards regression models were used to test the association of clinical and pathological variables with biochemical failure. All statistical tests were performed using SPSS version 10 (SPSS, Inc., Chicago, IL) and S-PLUS Professional version 1 (MathSoft Inc., Seattle, WA). Two-sided tests with a significance level at 0.05 were used.

RESULTS

Table 1 shows clinical and pathological characteristics of our patient cohort. Age at surgery was 52 to 72 years (mean age, 63.7 years). Preoperative PSA ranged from 3 to 36 ng/mL (mean, 12.9 ng/mL). In all, 64% of patients had a Gleason sum of 7. Follow-up ranged from 0.3 to 12.2 years (mean follow-up, 3.6 years; median follow-up, 3.2 years). Biochemical recurrence occurred in 23 patients (54.8%). RP tissues were evaluated for NF- κ B staining and classified as having either >5% or <5%

nuclear staining in tumor cells. Normal prostate glands demonstrated cytoplasmic staining mainly in the basal cell layer (Fig. 1A). Cancer glands almost always demonstrated cytoplasmic staining of the NF- κ B marker, mostly of high intensity (Fig. 1B). Nuclear staining was only observed in cancerous glands, although it was quite heterogeneous within the tumor (Fig. 1C). We found no correlation between the intensity of the staining and prognosis.

Table 2 shows the univariate and multivariate Cox survival analyses addressing the association between NF- κ B and biochemical prostate cancer recurrence (PSA > 0.3 ng/mL and rising) after RP. Covariates include age, pPSA, pathological Gleason score, and seminal vesicle and lymph node metastases. Kaplan-Meier survival curves were constructed to show the biochemical recurrence-free survival difference between patients with positive and negative nuclear NF- κ B staining (Fig. 2), and these results indicate a clear difference between the two groups in terms of recurrence.

Univariate Cox regression demonstrated a 4.9-fold (95% CI, 1.5–16.7; $P = 0.01$) higher rate of recurrence in men with >5% NF- κ B nuclear staining. In the multivariate model, after controlling for pathological Gleason sum ($P = 0.03$), lymph node metastases ($P = 0.07$), seminal vesicle invasion ($P = 0.6$),

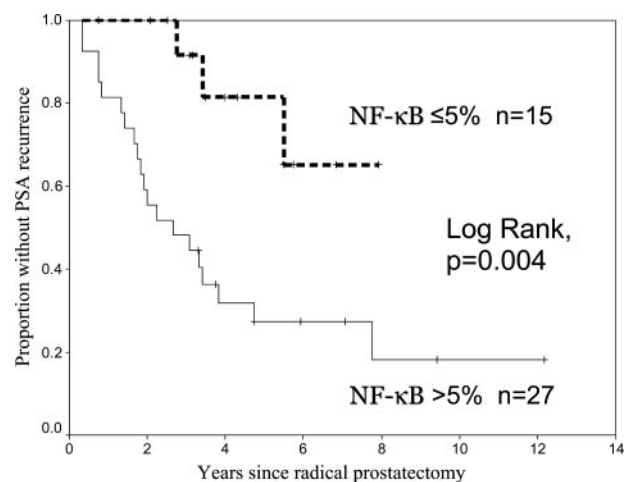


Fig. 2 Kaplan-Meier curve of biochemical recurrence-free survival according to NF- κ B status.

and pPSA ($P = 0.01$), NF- κ B nuclear staining of $>5\%$ was associated with a 6.3-fold higher risk of biochemical recurrence (95% CI, 1.7–23.5; $P = 0.007$; Table 2). These results indicate that NF- κ B status appears to represent the most statistically significant predictor of biochemical recurrence in the presence of positive surgical margins after RP.

DISCUSSION

PSA recurrence after RP is a precursor of cancer progression, with a mean time to metastases estimated at 8 years from the time of PSA elevation. Time to biochemical progression, PSA doubling time, and Gleason score have been identified as predictors of progression to metastatic disease. Once metastatic disease has occurred, the median time to death is 5 years (26). Therefore, PSA recurrence represents a valuable surrogate marker of cancer progression.

Patients with positive margins have a 2-fold higher recurrence rate when compared with patients with negative margins. The progression-free probability at 5 years for patients who have negative margins at RP is approximately 80% compared with 42% to 64% for men with positive margins (27, 28). Adjuvant radiation therapy is more effective if administered early before clinical failure. However, radiation may not be required in an appreciable proportion of patients who will not fail, despite having positive surgical margins. The identification of prognostic factors predictive of recurrence may help in developing a risk-adjusted approach to timely adjuvant therapy after RP.

Several pathological features have been shown to predict biochemical recurrence including preoperative PSA level, pathological T stage, Gleason score (biopsy and prostatectomy), margin status, lymph node and seminal vesicle involvement, perineural invasion, and tumor volume (27, 29). Although histologic grading is one of the most important predictors, its usefulness is limited because the majority of cases are moderately differentiated tumors (Gleason score of 6 or 7). The multivariate input of multiple predictors has been used in nomogram format to provide the most accurate and bias-free estimates of recurrence (30, 31). However, restrained input from predictor variables limits the accuracy of these tools. Therefore, there is a clear need for novel molecular markers that can complement standard predictors and increase the accuracy of existing predictive tools.

Several nuclear markers have been evaluated for their prognostic value in prostate cancer. Overexpression of the cell cycle inhibitor protein p21 has been shown as an independent predictor of response to salvage radiotherapy after RP (30). Progression to androgen independence has also been correlated with p21 (31). Expression of p53 and the combined loss of PTEN and p27 have been shown as independent predictors of disease-free survival in multivariate analyses (32, 33). Other promising candidate biomarkers currently under investigation and associated with disease progression include RNASEL, GSTP1, NKX3.1, and TERT (2, 3).

The results of the study we report here demonstrate that positive NF- κ B nuclear staining of cancer cells at the margin is associated with a 6-fold increase in the risk of biochemical recurrence in patients with positive RP surgical margins. In a

multivariate analysis, we confirmed the value of NF- κ B as an independent prognostic variable in this patient population. To date, none of the molecular prognostic markers have been applied to routine clinical practice, although with further confirmatory studies, we believe that the expression nuclear NF- κ B, either alone or in combination with other molecular markers, may eventually be useful in improving the value of parameters commonly used to help predict outcome in patients with prostate cancer.

ACKNOWLEDGMENTS

We thank Dr. Jean-Baptiste Lattouf, Dr. Christine Maugard, the Centre Hospitalier de l'Université de Montreal division of urology, and the pathology department of Notre-Dame Hospital for their contribution to this research.

REFERENCES

1. Miller JC, Zhou H, Kwekel J, et al. Antibody microarray profiling of human prostate cancer sera: antibody screening and identification of potential biomarkers. *Proteomics* 2003;3:56–63.
2. Nelson WG, De Marzo AM, Isaacs WB. Prostate cancer. *N Engl J Med* 2003;349:366–81.
3. Tricoli JV, Schoenfeldt M, Conley BA. Detection of prostate cancer and predicting progression: current and future diagnostic markers. *Clin Cancer Res* 2004;10:3943–53.
4. Barkett M, Gilmore TD. Control of apoptosis by Rel/NF-kappaB transcription factors. *Oncogene* 1999;18:6910–24.
5. Gilmore TD. The Rel/NF-kappaB signal transduction pathway: introduction. *Oncogene* 1999;18:6842–4.
6. Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF-kappaB activity. *Annu Rev Immunol* 2000;18:621–63.
7. Ghosh S, Karin M. Missing pieces in the NF-kappaB puzzle. *Cell* 2002;109(Suppl):S81–96.
8. Baldwin AS. Control of oncogenesis and cancer therapy resistance by the transcription factor NF-kappaB. *J Clin Invest* 2001;107:241–6.
9. Karin M, Cao Y, Greten FR, Li ZW. NF-kappaB in cancer: from innocent bystander to major culprit. *Nat Rev Cancer* 2002;2:301–10.
10. Sovak MA, Bellas RE, Kim DW, et al. Aberrant nuclear factor-kappaB/Rel expression and the pathogenesis of breast cancer. *J Clin Invest* 1997;100:2952–60.
11. Wang W, Abbruzzese JL, Evans DB, et al. The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. *Clin Cancer Res* 1999;5:119–27.
12. Sumitomo M, Tachibana M, Nakashima J, et al. An essential role for nuclear factor kappa B in preventing TNF-alpha-induced cell death in prostate cancer cells. *J Urol* 1999;161:674–9.
13. Flynn V Jr, Ramanitharan A, Moparty K, et al. Adenovirus-mediated inhibition of NF-kappaB confers chemo-sensitization and apoptosis in prostate cancer cells. *Int J Oncol* 2003;23:317–23.
14. Palayoor ST, Youmell MY, Calderwood SK, Coleman CN, Price BD. Constitutive activation of IkappaB kinase alpha and NF-kappaB in prostate cancer cells is inhibited by ibuprofen. *Oncogene* 1999;18:7389–94.
15. Chen CD, Sawyers CL. NF-kappa B activates prostate-specific antigen expression and is upregulated in androgen-independent prostate cancer. *Mol Cell Biol* 2002;22:2862–70.
16. Gasparian AV, Yao YJ, Kowalczyk D, et al. The role of IKK in constitutive activation of NF-kappaB transcription factor in prostate carcinoma cells. *J Cell Sci* 2002;115:141–51.
17. Suh J, Payvandi F, Edelstein LC, et al. Mechanisms of constitutive NF-kappaB activation in human prostate cancer cells. *Prostate* 2002;52:183–200.

18. Ismail HA, Lessard L, Mes-Masson AM, Saad F. Expression of NF-kappaB in prostate cancer lymph node metastases. *Prostate* 2004;58:308–13.
19. Lessard L, Mes-Masson AM, Lamarre L, et al. NF-kappa B nuclear localization and its prognostic significance in prostate cancer. *BJU Int* 2003;91:417–20.
20. Huang S, Pettaway CA, Uehara H, Bucana CD, Fidler IJ. Blockade of NF-kappaB activity in human prostate cancer cells is associated with suppression of angiogenesis, invasion, and metastasis. *Oncogene* 2001;20:4188–97.
21. Ling MT, Wang X, Ouyang XS, et al. Id-1 expression promotes cell survival through activation of NF-kappaB signalling pathway in prostate cancer cells. *Oncogene* 2003;22:4498–508.
22. Hodge JC, Bub J, Kaul S, Kajdacsy-Balla A, Lindholm PF. Requirement of RhoA activity for increased nuclear factor kappaB activity and PC-3 human prostate cancer cell invasion. *Cancer Res* 2003;63:1359–64.
23. Levine L, Lucci JA III, Pazdrak B, et al. Bombesin stimulates nuclear factor kappa B activation and expression of proangiogenic factors in prostate cancer cells. *Cancer Res* 2003;63:3495–502.
24. Suh J, Rabson AB. NF-kappaB activation in human prostate cancer: important mediator or epiphenomenon? *J Cell Biochem* 2004;91:100–17.
25. Cinar B, Yeung F, Konaka H, et al. Identification of a negative regulatory cis-element in the enhancer core region of the prostate-specific antigen promoter: implications for intersection of androgen receptor and nuclear factor-kappaB signalling in prostate cancer cells. *Biochem J* 2004;379:421–31.
26. Pound CR, Partin AW, Eisenberger MA, et al. Natural history of progression after PSA elevation following radical prostatectomy. *JAMA* 1999;281:1591–7.
27. Epstein JI, Partin AW, Sauvageot J, Walsh PC. Prediction of progression following radical prostatectomy. A multivariate analysis of 721 men with long-term follow-up. *Am J Surg Pathol* 1996;20:286–92.
28. Ohori M, Wheeler TM, Kattan MW, Goto Y, Scardino PT. Prognostic significance of positive surgical margins in radical prostatectomy specimens. *J Urol* 1995;154:1818–24.
29. D'Amico AV, Whittington R, Malkowicz SB, et al. A multivariate analysis of clinical and pathological factors that predict for prostate specific antigen failure after radical prostatectomy for prostate cancer. *J Urol* 1995;154:131–8.
30. Rigaud J, Tiguert R, Decobert M, et al. Expression of p21 cell cycle protein is an independent predictor of response to salvage radiotherapy after radical prostatectomy. *Prostate* 2004;58:269–76.
31. Fizazi K, Martinez LA, Sikes CR, et al. The association of p21^{WAF1/CIP1} with progression to androgen-independent prostate cancer. *Clin Cancer Res* 2002;8:775–81.
32. Bauer JJ, Sesterhenn IA, Mostofi KF, et al. p53 nuclear protein expression is an independent prognostic marker in clinically localized prostate cancer patients undergoing radical prostatectomy. *Clin Cancer Res* 1995;1:1295–300.
33. Halvorsen OJ, Haukaas SA, Akslen LA. Combined loss of PTEN and p27 expression is associated with tumor cell proliferation by Ki-67 and increased risk of recurrent disease in localized prostate cancer. *Clin Cancer Res* 2003;9:1474–9.

Clinical Cancer Research

Nuclear Factor- κ B Nuclear Localization Is Predictive of Biochemical Recurrence in Patients with Positive Margin Prostate Cancer

Vincent Fradet, Laurent Lessard, Louis R. Bégin, et al.

Clin Cancer Res 2004;10:8460-8464.

Updated version Access the most recent version of this article at:
<http://clincancerres.aacrjournals.org/content/10/24/8460>

Cited articles This article cites 32 articles, 9 of which you can access for free at:
<http://clincancerres.aacrjournals.org/content/10/24/8460.full#ref-list-1>

Citing articles This article has been cited by 23 HighWire-hosted articles. Access the articles at:
<http://clincancerres.aacrjournals.org/content/10/24/8460.full#related-urls>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link
<http://clincancerres.aacrjournals.org/content/10/24/8460>.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.