Strategies for Delaying or Treating In vivo Acquired Resistance to Trastuzumab in Human Breast Cancer Xenografts


Abstract

Purpose: Acquired resistance to trastuzumab (Herceptin) is common in patients whose breast cancers show an initial response to the drug. The basis of this acquired resistance is unknown, hampering strategies to delay or treat such acquired resistance, due in part to the relative lack of appropriate in vivo tumorigenic models.

Experimental Design: We derived an erbB-2–positive variant called 231-H2N, obtained by gene transfection from the highly tumorigenic erbB-2/HER2–negative human breast cancer cell line, MDA-MB-231. Unlike MDA-MB-231, the 231-H2N variants was sensitive to trastuzumab both in vitro and especially in vivo, thus allowing selection of variant resistant to drug treatment in the latter situation after showing an initial response.

Results: The growth of established orthotopic tumors in severe combined immunodeficient mice was blocked for 1 month by trastuzumab, after which rapid growth resumed. These relapsing tumors were found to maintain resistance to trastuzumab, both in vitro and in vivo. We evaluated various therapeutic strategies for two purposes: (a) to delay such tumor relapses or (b) to treat acquired trastuzumab resistance once it has occurred. With respect to the former, a daily oral low-dose metronomic cyclophosphamide regimen was found to be particularly effective. With respect to the latter, an anti–epidermal growth factor receptor antibody (cetuximab) was effective as was the anti–vascular endothelial growth factor (anti-VEGF) antibody bevacizumab, which was likely related to elevated levels of VEGF detected in trastuzumab-resistant tumors.

Conclusions: Our results provide a possible additional rationale for combined biological therapy using drugs that target both erbB-2/HER2 and VEGF and also suggest the potential value of combining less toxic metronomic chemotherapy regimens not only with targeted antiangiogenic agents but also with other types of drug such as trastuzumab.

One of the major significant clinical advances in cancer therapeutics over the last decade has been the discovery, development, and approval of a number of drugs, both antibodies and small molecules, which target the products of oncogenes. These drugs include trastuzumab (Herceptin), the humanized antibody to the erbB-2/HER2 receptor tyrosine kinase (1); imatinib (Gleevec), a small molecule drug that targets the bcr-abl, c-kit, and platelet-derived growth factor receptor tyrosine kinases (2); and cetuximab (C225/Erbitux), the chimeric antibody to the epidermal growth factor receptor (EGFR; erbB-1) and small molecule antagonists of the EGFR (3). Unfortunately, the success of these drugs in responding patients is compromised by the same problem, which plagues other systemic therapies, such as chemotherapy, hormone receptor antagonizing drugs, or hormonal ablation therapeutics i.e., the development of acquired resistance (4). In particular, the striking responses frequently induced by imatinib in indolent chronic myelogenous leukemia or gastrointestinal stromal tumor patients followed by relapse and the discovery of mutations in the respective target genes that lead to loss of tumor response (5) have highlighted the profound effect that acquired drug resistance mechanisms can have on limiting the durability of tumor responses to such promising drugs (4).

One of the surprising aspects of this area of research is the lack of any published detailed studies devoted to defining the mechanisms of acquired resistance to trastuzumab. Despite the fact that this drug has been in development for >15 years and has by now been used in the treatment of many thousands of breast cancer patients, there are as yet no published preclinical studies on isolating and characterizing trastuzumab-resistant variants/mutants from initially erbB-2–positive...
and responsive human tumors in vivo, and only a few reports using cell lines selected in vitro for acquired resistance to trastuzumab (6–8). The study of acquired resistance to trastuzumab, especially variants derived during in vitro therapy, can be very useful for devising possible therapeutic strategies to delay acquired trastuzumab resistance, or to treat tumors that have actively acquired resistance, based on the discovery of new targets expressed in resistant variant cells that might be exploitable using second-line therapies. This is an important clinical problem because about 40% of metastatic patients with erbB-2 overexpressing breast cancer respond to first-line trastuzumab therapy as a single agent (9), but the median duration of response is only about 8 months. With respect to intrinsic resistance to trastuzumab, there have been a few recent studies in this area. For example, an important clinical study by Nagata et al. reported that activation of the PTEN tumor suppressor protein seems to contribute to trastuzumab tumor responsiveness in human breast cancer patients, such that loss of PTEN predicts trastuzumab resistance (10). However, whether this type of genetic alteration contributes to some forms of acquired trastuzumab resistance is unknown. In addition, Tanner et al. generated a breast cell line (called JIMT-1) from a patient treated with trastuzumab, but who did not respond (i.e., showed intrinsic resistance to the drug: ref. 11). The cell line showed no identifiable mutation in erbB-2. However, the accessibility of erbB-2 seems to be reduced by MUC4 overexpression (12).

There are several practical problems associated with studying acquired resistance to trastuzumab using erbB-2 breast cancer xenografts, which may help explain the lack of relevant studies in this preclinical area of research. First, there are very few human breast cancer cell lines, which overexpress erbB-2, and those few that do, such as SKBR3 and BT-474, tend to be poorly or even nontumorigenic, making it difficult to have the proper selective pressure conditions for isolating authentic variants having an acquired drug-resistant phenotype. Second, long-term therapy experiments are rare, in part because of the relatively weak responses to the drug (which are usually manifested in the form of tumor growth delays, but not frank tumor regressions), which, once again, make it difficult to isolate authentic mutants or variants that have acquired resistance to trastuzumab in a preclinical setting. To overcome these problems, we studied a variant of a highly tumorigenic human tumor cell line, MDA-MB-231 (which expresses barely detectable levels of erbB-2), from which a stable erbB-2 overexpressing variant cell population was obtained by gene transfection. The variant, called 231-H2N, was grown orthotopically in severe combined immunodeficient (SCID) mice and subjected to trastuzumab therapy alone, or combined with chemotherapy. We used chemotherapy in combination with trastuzumab for some of the experiments because trastuzumab is normally administered with chemotherapy (1, 13), and this could obviously affect the kinetics and outcome of acquired resistance to trastuzumab. We used cyclophosphamide as the chemotherapeutic for our studies, which was administered either in a daily low-dose (metronomic) fashion through drinking water (14) or alternatively, in a more conventional maximum tolerated dose (MTD) regimen with 2-week-long drug-free breaks between weekly cycles of MTD therapy (15). Part of the original rationale for the metronomic chemotherapy experiments was based on the prospect of undertaking long-term therapy studies in mice when using such relatively nontoxic chemotherapy regimens, either alone, or when these are combined with targeted therapies, such as antiangiogenic agents (16). We reasoned this paradigm might be extended to trastuzumab, especially because this drug may have antiangiogenic effects, which contribute to its antitumor efficacy (17, 18). Thus, we hypothesized that combination of this drug with a metronomic chemotherapy regimen might also result in better disease control (and overall less toxicity) compared with trastuzumab combined with MTD regimen of cyclophosphamide. We also speculated that such long-term therapy experiments in mice (assuming the combination therapy would be highly effective) could be helpful in the eventual experimental selection and overgrowth of variants having acquired resistance to trastuzumab.

Our results show that it is indeed possible to select trastuzumab-resistant variants in vivo using the aforementioned experimental approaches, whether using trastuzumab alone or in combination with chemotherapy. The resistant variants we selected were partially characterized, and the results indicated they had a significantly elevated vascular endothelial growth factor (VEGF) expression, a finding that is similar to the properties of variants of other human tumor cell lines selected for acquired resistance in vivo to erbB-1/EGFR targeting antibodies, such as C225/cetuximab (19). Consequently, addition of bevacizumab (the humanized anti-VEGF antibody) to trastuzumab, transiently restored responsiveness to trastuzumab-resistant variants, suggesting this type of approach might be of value to delay or to treat variants with acquired trastuzumab resistance, as a second-line or follow-up therapy. The resistant variants also responded to trastuzumab when combined with an EGFR targeting antibody, which may be related to an observed up-regulation of transforming growth factor-α (TGF-α) in the resistant variants. Finally, we also obtained preliminary evidence for the superiority of combining trastuzumab with metronomic versus standard MTD chemotherapy, both in terms of reduced toxicity and prolonged survival.

**Materials and Methods**

**Cells and culture conditions.** Human breast tumor cell lines, MDA-MB-361, BT-474, and SK-BR-3 were purchased from the American Type Culture Collection (Rockville, MA) and maintained as monolayers in DMEM supplemented with 10% fetal bovine serum. The human breast tumor cell line, MDA-MB-231, was kindly provided by Dr. Janet E. Price (M.D. Anderson Cancer Centre, University of Texas). MDA-MB-231 cells were transfected to stably overexpress erbB-2. Briefly, amphotropic GP+envAM12 packaging cells, cotransfected with pUMFG/her2/neu/IRES/enYFP containing the full-length human c-erbB-2 cDNA and the plasmid pGT-N28 (New England Biolabs, Beverly, MA), were used to transduce MDA-MB-231 cells. The cell line 231-H2N was derived by fluorescence-activated cell sorting to select the top 10% of erbB-2 expressors. Both MDA-MB-231 and 231-H2N were maintained as monolayers in DMEM with 5% fetal bovine serum. All cell cultures were maintained in a humidified incubator at 37°C and 5% CO2.

**Protein isolation and Western blots.** Cells were grown to ~80% confluence in normal culture conditions. Plates were placed on ice, washed with ice-cold PBS, and cold lysis buffer [20 mmol/L Tris (pH 7.5) containing 137 mmol/L NaCl, 100 mmol/L NaF, 10% glycerol, 1% NP40, 2 mmol/L NaVO4, 1 mmol/L phenylmethylsulfonyl fluoride, 10 µg/mL aprotinin, and 10 µg/mL leupeptin] was added directly to plates. Cells were scraped from plates, incubated on ice for 2 hours, and centrifuged at 10,000 rpm for 15 minutes. The pellets were resuspended in 2x SDS-PAGE loading buffer and then boiled for 5 minutes. Protein concentrations were determined using the Bio-Rad protein assay kit.

**Western blotting.** Cell lysates were subjected to SDS-PAGE on 10% gel and then electroblotted onto a nitrocellulose membrane. The membranes were blocked with 5% non-fat milk in TBS-T (20 mmol/L Tris, pH 7.8, 150 mmol/L NaCl, 0.1% Tween 20) for 2 hours at room temperature. The membranes were probed overnight at 4°C with specific antibodies, washed with TBS-T, and incubated with secondary antibodies. Signals were visualized using an enhanced chemiluminescence detection system.
40 minutes, and centrifuged at 10,000 rpm for 15 minutes. Protein concentrations of the supernatant were determined using Bio-Rad Protein Assay Dye Reagent (Bio-Rad Laboratories, Hercules, CA); 50 µg/lane of total proteins were resolved by 8% SDS-PAGE, transferred onto Immobilon-P membrane (Millipore Canada Ltd., Etobicoke, Ontario, Canada), blocked with 10% milk in TBST, and probed with primary antibody. Anti-neu, anti-β-actin (Sigma-Aldrich Canada Ltd., Oakville, Ontario, Canada) were incubated for 1 hour at room temperature, whereas Akt, p44/42 mitogen-activated protein kinase, phospho-erbB-2 Tyr1248, phospho-β-actin, phospho-EGFR, phospho-mitogen-activated protein kinase (all from Cell Signaling Technologies, Beverly, MA), PTEN (MMAC1, Ab-2, Medscorp, Montreal, Canada), PTEN (Santa Cruz Biotechnology), and extracellular erbB-2 Ab-20 (L87 + 2EBR19 from Medicorp) were incubated overnight at 4 °C. All primary antibodies were diluted 1:1,000 in 5% milk in TBST, detected with anti-rabbit IgGHRP or anti-mouse IgGHRP (Promega, Madison, WI) diluted 1:5,000, and detected with enhanced chemiluminescence reagent (Amersham Biosciences, Piscataway, NJ).

**Northern blot of total RNA.** Tumor tissues were stored at −70 °C in RNAlater. Trizol extraction (Life Technologies, Inc., Gaithersburg, MD) was carried out according to manufacturer’s instruction. VEGF Northern blots were done as previously described (20). The VEGF probe spanning the 200-bp sequence common for all VEGF isoforms was a generous gift of Drs. Brygida Berse and Harold Dvorak (Beth Israel Hospital, Boston, MA). Glyceraldehyde-3-phosphate dehydrogenase was probed as a loading control.

**In vitro determination of drug sensitivity.** Two thousand cells were plated in 96-well flat-bottomed tissue culture plates in a volume of 100 µl media per well and incubated overnight at 37 °C and 5% CO2. Drug treatments were added to wells in a volume of 100 µl media and were carried out for 72 hours. Cells were then pulsed with 2 µCi/well of methyl-3H[thymidine (Amersham Biosciences) for 6 hours. DNA was harvested onto Unifilter filtermat plates (Perkin-Elmer Life Sciences, Woodbridge, Ontario, Canada) using the Filtermate Harvester (Perkin-Elmer Life Sciences). Incorporated radioactivity was measured by scintillation counting using a Packard Topcount NXT microplate counter (Perkin-Elmer Life Sciences).

**In vivo tumor growth assessment.** Four-week-old female C.B-17 (SCID) mice were purchased from Charles River Canada (Saint-Constant, Quebec, Canada). Mice were allowed to acclimatize and mature for 2 weeks before implantation of tumor cells. To prepare cells for injection, subconfluent plates were harvested with 1% trypsin-EDTA (Life Technologies Bethesda Research Laboratories, Gaithersburg, MD), and cells were then washed and resuspended in DMEM at 4 °C until analysis. Two million cells were injected into the inguinal mammary fat pad of mice. Wounds were secured with surgical staples.

Mice were monitored twice weekly for tumor growth and fluctuations in body weight. Treatments were initiated when the average tumor volume reached 250 mm3 (unless otherwise indicated) as measured by Vernier calipers and calculated by (length × width2) / 2. Institutional guidelines were followed for end point of experiments. Plasma was collected by cardiac puncture into Microtainer Plasma Separator tubes (Becton Dickinson, Franklin Lakes, NJ) and stored at −70 °C until analysis. Tumors were removed from mice, divided into three sections, and placed into RNAlater (Ambion, Inc., Austin, TX), Prefer fixative (Anatech, Battle Creek, MI), or frozen section medium (Stephens Scientific, Riverdale, NJ).

**Drug treatments.** Cyclophosphamide was purchased from the institutional pharmacy and prepared for oral or i.p. administration as previously described (14). Oral cyclophosphamide was administered at an estimated dose of 20 mg/kg/d. Mice administered the MTX schedule were injected i.p. with 100 mg/kg, on days 1, 3, and 5 of each 21-day cycle. Trastuzumab (Herceptin) was generously provided by Genentech (San Francisco, CA). A 20 mg/mL stock solution was prepared by adding sterile PBS to the lyophilized antibody. Immediately before i.p. injection, the trastuzumab stock solution was diluted 1:5 in normal saline and was administered at 20 mg/kg twice weekly. Additional

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**Fig. 1.** A, Western blot to determine levels of erbB-2 and phospho-erbB-2 in whole-cell lysates of the 231-H2N − transfected cells compared with parental MDA-MB-231 and endogenous erbB-2 overexpressing cell lines BT-474, SK-BR-3, and MDA-MB-361. B, in vitro determination of drug sensitivity. Two thousand cells were plated in 96-well tissue culture plates per well and incubated overnight at 37 °C in 5% CO2. Drug treatments were added to wells in a volume of 100 µl media and were carried out for 72 hours. Cells were then pulsed with 2 µCi/well of [3H]thymidine for 6 hours. DNA was harvested onto Unifilter filtermat plates (Perkin-Elmer Life Sciences, Woodbridge, Ontario, Canada) using the Filtermate Harvester (Perkin-Elmer Life Sciences). Incorporated radioactivity was measured by scintillation counting using a Packard Topcount NXT microplate counter (Perkin-Elmer Life Sciences). Columns, mean (n = 8); bars, SD. *, P < 0.05.

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digested with an enzyme cocktail of serum-free DMEM containing 2 mg/mL Collagenase 3 (Worthington Biochemical Corp., Lakewood, NJ), 1 mg/mL hyaluronidase (Sigma-Aldrich), and 1 mg/mL Collagenase IV (Sigma-Aldrich). After incubation on ice for 30 minutes, followed by a 30-minute incubation at 37°C with shaking, the cells were washed and plated in DMEM supplemented with 5% FCS, Fungizone (Invitrogen Life-Technologies, Burlington, Ontario, Canada) and penicillin/streptomycin (Invitrogen, San Diego, CA). Cells were grown and passaged thrice to allow for tumor cell enrichment and also elimination of host cells before further study of the cell lines. Cells were maintained in DMEM with 5% FCS.

In vivo angiogenesis assessment by Matrigel plug and tumor perfusion assays. The perfusion assays with FITC-dextran were carried out as previously described, with minor modifications (16). Briefly, two Matrigel preparations were injected s.c. into the flanks of 8-week-old female CB-17 SCID mice, the first containing 500 ng/mL of the angiogenic factor basic fibroblast growth factor, and the second containing 3 x 10^6 C231-H2N tumors. Treatment commenced 4 days after implantation and was carried out for 7 days. For tumor perfusion assays, C231-H2N tumors were established as indicated above, and treatment commenced when tumors reached a volume of 400 mm^3. Mice were treated for 21 days.

Measurement of TGF-α and VEGF by ELISA. Cells (1 x 10^5) from parental and resistant variants were plated in 24-well plates and grown for 24 hours. Medium was replaced with DMEM containing 1% fetal bovine serum, and the cells were incubated for an additional 48 hours. Conditioned medium was then collected and assayed on the human TGF-α ELISA and human VEGF kit, respectively, following the manufacturer’s protocol (R&D Systems, Minneapolis, MN). The detected growth factor levels in the conditioned medium were corrected for final cell number per well.

Statistical analysis. Results are reported as mean ± SD or SE. Statistical significance of differences was assessed by PRISM software (version 4.00; GraphPad, San Diego, CA). The level of significance was set at P < 0.05.

Results

Establishment of a trastuzumab-responsive orthotopic breast cancer mouse model. To establish a highly tumorigenic erbB-2–positive breast tumor model in mice, the tumorigenic human breast tumor cell line MDA-MB-231, which does not normally overexpress erbB-2, was stably transfected to derive an erbB-2 overexpressing variant termed C231-H2N. MDA-MB-231 was selected as an ideal breast tumor model for these studies, due in part to the reported involvement of VEGF-dependent angiogenesis in the progression of these tumors (24). In addition, in vivo tumor growth of MDA-MB-231 mammary tumors has been shown to be effectively delayed by treatment of mice with metronomic cyclophosphamide administered through the drinking water (14), and we wanted to evaluate and compare the effects of trastuzumab alone, or combined with a chemotherapy drug (i.e., cyclophosphamide), administered either in a frequent low-dose (metronomic) fashion or in a conventional MTD manner to determine which of the two would be superior, and how each might affect kinetics of acquired resistance to trastuzumab. We wish to emphasize that some naturally erbB-2–positive breast cancer cell lines were also tested but not selected for our studies due to poor or complete lack of in vivo growth (e.g., BT-474 and SK-BR3), even when the cells were injected into the mammary fat pads of SCID mice (data not shown).

Expression levels of erbB-2 were determined by Western blot for C231-H2N cells relative to the parental MDA-MB-231 cell line and several endogenous erbB-2 overexpressing cell lines. Although the parental cell line expresses minimal levels of erbB-2, C231-H2N cells were found to express erbB-2 and, importantly, do so in the range of endogenous erbB-2 overexpressing cell lines (Fig. 1A). In addition, when the blot was probed for phospho-erbB-2, the receptor was found to be constitutively phosphorylated in C231-H2N cells (Fig. 1A), suggesting that the receptor is biologically activated. To confirm that erbB-2 was functional and involved in the activation of downstream intracellular signaling pathways within the transfected cell line, in vitro assessment of the cytostatic effects of erbB-2–targeted therapies was carried out on the C231-H2N and MDA-MB-231 cell lines. The erbB-2–targeted antibody, trastuzumab, as well as the EGFR-erbB-2 heterodimerization inhibitory antibody, pertuzumab, and the EGFR-targeted antibody cetuximab (C225) were shown to inhibit cell proliferation of C231-H2N cells by 25% at antibody concentrations as low as 1 µg/mL (Fig. 1B and C). Such modest levels of growth inhibition mediated by trastuzumab in vitro are consistent with the results of others and our own previous results using other cell lines (17). In contrast, significant inhibition of the parental cell line was observed only at very high concentrations of 50 µg/mL.

We then undertook preliminary in vivo studies, where tumor “take” in the mammary fat pad was determined. C231-H2N tumors were palpable in all mice within 7 days of injection, and tumors reached 150 to 250 mm^3 within 5 weeks with acceptable variation in tumor size (data not shown). Untreated tumors reached a volume of 1,700 mm^3 within 10 weeks and remained encapsulated within the mammary fat pad. No evidence of macroscopic metastases was observed in the lungs of these mice.

Effect of single or combined trastuzumab therapies on tumor growth. One of the main purposes of our studies was to assess the antitumor effects (and development of resistance) of trastuzumab administered alone, compared with the drug used in combination with low-dose, chronically administered cyclophosphamide, as a first step to evaluate the potential value of this type of chemotherapy treatment regimen for erbB-2 overexpressing breast cancer tumors. Two independent in vivo experiments designed to evaluate the effects of metronomic cyclophosphamide, trastuzumab, and combination therapy, compared with a standard MTD dosing schedule commonly used in mice produced similar results. In the experiment shown in Fig. 2, orthotopic C231-H2N tumors were established in the abdominal mammary fat pad of female CB-17 SCID mice and grown to ~250 mm^3 before initiation of treatment (day 0). Treatments were continued until tumors reached a volume of 1,700 mm^3 or mice experienced a loss of 15% of their initial body weight, according to institutional animal care guidelines.

Although untreated control tumors progressed rapidly, reaching a volume of 1,700 mm^3 in <40 days, daily oral metronomic cyclophosphamide effectively delayed tumor growth to this volume for 60 days (Fig. 2A), as previously reported using parental MDA-MB-231 tumors (14). A complete block of tumor growth was observed for trastuzumab-treated C231-H2N tumors for almost a month, after which time the tumors rapidly progressed, clearly suggestive of the emergence of variants with acquired resistance to the drug. The initial response of the primary tumors to trastuzumab was much greater than expected, as in vitro studies of C231-H2N monolayer cultured cells indicated only a 25% inhibition of proliferation at increasing concentrations of trastuzumab. This impressive...
**in vivo** effect of trastuzumab is likely due to one or more host dependent effects, including, potentially, antibody-dependent cell-mediated cytotoxicity (25) and the postulated antiangiogenic properties of this drug (17, 18), which, acting alone or together in concert with a direct inhibition of tumor cell proliferation, might be responsible for this surprising degree of initially potent tumor growth suppression. With respect to “relapse,” both MTD cyclophosphamide and metronomic cyclophosphamide, when combined with trastuzumab, delayed this apparent rapid onset of resistance, in the case of metronomic cyclophosphamide by an additional 75 to 90 days, illustrating the greatly improved efficacy of the combination of trastuzumab with chemotherapeutic regimens, as reported by Pegram et al. (26) and others. However, mice in the MTD cyclophosphamide + trastuzumab group had to be sacrificed due to severe toxicity after three treatment cycles (Fig. 2B). To emphasize the advantage of the metronomic cyclophosphamide and trastuzumab combination therapy, compared with the combination with standard MTD, an event-free survival curve was plotted, whereby event-free survival time was defined as a time to progression of the tumor to >1,500 mm³ or loss of >15% body weight. As seen in Fig. 2C, the combination of MTD cyclophosphamide and trastuzumab showed little benefit with respect to event-free survival compared with trastuzumab therapy alone due to the toxicity of the chemotherapy (as indicated by substantial weight loss), whereas combination therapy of metronomic cyclophosphamide and trastuzumab provided a significant survival advantage compared with trastuzumab therapy alone, due to both a prevention of tumor progression for 120 days and an absence of obvious toxic side effects. As illustrated in Fig. 2B, mice receiving trastuzumab, metronomic cyclophosphamide, or a combination of these drugs showed no significant treatment-related fluctuations in weight over the duration of the experiment. In contrast, mice administered MTD cyclophosphamide alone or in combination experienced weight loss following the administration of cyclophosphamide. The third MTD dosing cycle was followed by an irreversible weight loss of >25%, an indication of severe, cumulative toxicity. Although not carried out in these experiments, a detailed analysis of the toxic effects of low-dose metronomic versus MTD cyclophosphamide in mice was reported recently by our group (27); with the exception of lymphopenia, toxic side effects, such as myelosuppression, were not observed in low-dose metronomic cyclophosphamide regimens, despite the ultrasensitiviy of SCID mice to DNA damaging agents such as cyclophosphamide.

To determine whether the effects of metronomic chemotherapy and trastuzumab could be extended to situations involving larger, more established tumors, an experiment was carried out where treatment was commenced when tumors were ~800 mm³ in volume and continued for 3 weeks. Although single-agent therapy with either trastuzumab or metronomic cyclophosphamide

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**Fig. 2.** *In vivo* treatment of 231-H2N mammary tumors growing orthotopically in SCID mice. A, antitumor effects of low-dose metronomic cyclophosphamide (Ld CTX), MTD dosing of cyclophosphamide (MTD CTX), or trastuzumab alone, and combination regimens using low-dose metronomic cyclophosphamide plus trastuzumab, or MTD cyclophosphamide plus trastuzumab. †, time of MTD dosing. Symbols, mean (n = 7-8); bars, SD. B, corresponding mouse weight as a measurement of relative toxicity of the different therapeutic regimens.
showed some delay in tumor growth, combination therapy completely blocked tumor progression during this treatment period (Fig. 2D). In addition, MDA-MB-231 tumors were treated with trastuzumab to confirm the lack of response of the parental cell line to trastuzumab, as found above and in vitro.

Analysis of erbB-2 levels in cell lines derived from trastuzumab-resistant tumors. Because the mechanisms of resistance to trastuzumab have been rarely studied in an in vivo context, we decided to further explore the underlying causes of the development of resistance to trastuzumab following relatively long-term trastuzumab administration.

Initially, in vivo clonal selection for cells expressing lower levels of the erbB-2 target was suspected as the cause for loss of response to trastuzumab, as the cell line 231-H2N was derived from a polyclonal population of erbB-2–transfected cells. Cell lines were therefore derived from untreated primary tumors and tumors that had become unresponsive to trastuzumab therapy alone, or in combination with metronomic cyclophosphamide, and were analyzed for retention or loss of erbB-2 expression. Analysis of whole-cell lysates for total erbB-2 and EGFR protein by Western blot indicated that all cell lines maintained expression of EGFR, and five of eight of these established resistant tumor cell lines still expressed high levels of the erbB-2 protein (Fig. 3A), as has also been shown by others (6, 28). Although the remaining three cell lines were found to express lower levels of the full-length erbB-2 protein, it remains to be determined if these cell lines express a truncated or mutant form of the erbB-2 receptor that is not recognized by the antibody used to probe the membranes. In addition, blots were probed for total protein levels of Akt and p44/42 (mitogen-activated protein kinase); no significant changes in expression levels of these two proteins were observed.

Increased expression of VEGF as a potential mechanism of resistance. Previously, we reported that VEGF is up-regulated in A431 human tumor xenografts selected in vivo for acquired resistance to anti-EGFR antibody therapy and, moreover, that this alteration seems to contribute to the resistant phenotype (19). Given the similar nature of the therapy, we decided to

Fig. 2. Continued. C, effects of therapeutic regimens on event-free survival (Kaplan-Meier analysis), where duration of event-free survival is defined as time to primary tumor progression to 1,500 mm$^3$ or >15% weight loss. *, indicating time of MTD dosing. **, weight loss of >15%. Significant event-free survival was observed with low-dose metronomic cyclophosphamide plus trastuzumab (Ld CTX + Trastuzumab) compared with the other treatment regimens. D, in vivo antitumor effects of cyclophosphamide and trastuzumab therapies on well-established (800 mm$^3$) 231-H2N and MDA-MB-231 mammary tumors growing orthotopically in SCID mice. 231-H2N tumors treated with the metronomic cyclophosphamide + trastuzumab regimen remained at ~800 mm$^3$ throughout the 3-week period of this experiment. Gray bars, mean tumor volume at start of treatment; black bars, mean tumor volume after 3 weeks of treatment (symbols represent individual tumor volumes). **, $P < 0.05$. 
Acquired resistance to trastuzumab is observed in vitro and in vivo. To determine if a similar mechanism might be associated with the development of acquired resistance of 231-H2N tumors to trastuzumab. Total RNA was extracted from tumors following progression and probed for VEGF by Northern blot. Indeed, we observed an increase in the expression of VEGF mRNA in tumor resistant to combination therapy regimens (Fig. 3B) compared with control tumors.

**Analysis of cell lines derived from trastuzumab-resistant tumors.** To get an insight as to the mechanisms responsible for the evolution of trastuzumab resistance, we analyzed cell lines derived from trastuzumab-resistant tumors in more detail. Analysis of VEGF secretion into the conditioned media (measured by ELISA) showed that three of four cell lines expressed significantly higher levels compared with the controls (4.580 pg/10^6 cells/48 hours; Fig. 5A). This result suggested that perhaps one contributing factor to trastuzumab resistance could be the increased expression of this proangiogenic factor.

Overexpression of EGFR has been proposed as a mechanism of resistance to trastuzumab. Human MDA-MB-231 breast cancer cells have been shown to have autocrine activation of the EGFR/TGF-α signaling cascade, which in turn has been reported to have a role in the regulation of VEGF expression. Therefore, levels of TGF-α secreted into the conditioned media of cell lines derived from tumors resistant to trastuzumab were measured by ELISA (Fig. 5B). Cell lines derived from untreated 231-H2N xenografts (C1, C2, C3) expressed on average 6.66 pg TGF-α/10^6 cells/48 hours. Three cell lines derived from trastuzumab-resistant tumors (TrR1, TrR2, TrR3) expressed significantly elevated levels of TGF-α compared with the mean of the control group.

We next sought to determine if there was a genetic change that could explain the development of trastuzumab resistance and the concurrent increase in VEGF expression. To do so, we compared the parental cell lines with the variants TrR1 and TrR2, as these showed the highest increase in VEGF protein expression. To our surprise, we found no significant difference in VEGF mRNA expression (Fig. 5C), suggesting that the increase in VEGF protein secretion (shown in Fig. 5A) is due to post-transcriptional changes. We next tested to see if changes in the erbB-2 protein, or components of its signaling pathway, could explain the observed acquired resistance. Western blotting analysis showed that although both variants expressed the intracellular domain of erbB-2 (Fig. 5D), the expression was reduced in one variant (TrR2). This variant also showed low or barely detectable levels of phospho-HER-2 or phospho-EGFR (Fig. 5E). In contrast, no significant changes were noted between parental and trastuzumab-resistant variants in terms of phospho-AKT or phospho-mitogen-activated protein kinase levels. Therefore, although one variant (TrR2) showed changes in the target pathways of trastuzumab as a possible mechanism for resistance to this drug, no unifying event was identified to explain the observed resistance. Rather, we believe a number of mechanisms can independently contribute to trastuzumab resistance, any number of which may be observed in a trastuzumab-resistant tumor population.

![Fig. 3](image-url)
Finally, because loss of PTEN has been reported to predict intrinsic clinical resistance to trastuzumab (10), we examined various cell lines for PTEN status. However, we did not find any consistent or obvious changes in PTEN expression in the variants selected for acquired resistance (data not shown).

**Antiangiogenic activity of trastuzumab plus cyclophosphamide treatment regimens.** Because our results suggested that increased angiogenic activity might account at least in part for the acquired resistance to trastuzumab, we undertook a series of FITC-dextran perfusion studies. Our aim was to test the possible antiangiogenic effects of the various treatment regimens as used in Fig. 2A. In established orthotopic 231-H2N xenografts, all single-agent therapies produced a reduction in tumor perfusion relative to controls after one 21-day treatment cycle, ranging from 10% (trastuzumab) to 38% and 41% (metronomic and MTD cyclophosphamide groups, respectively; Fig. 6A). Combination therapies with trastuzumab resulted in additive antiangiogenic activity, reducing tumor perfusion by 50% and 55% for the metronomic and MTD cyclophosphamide combination therapies, respectively. Because trastuzumab exerts its antiangiogenic effects indirectly, by modifying the angiogenic profile of tumors (18), it was not altogether unexpected that trastuzumab alone had no effect in the basic fibroblast growth factor Matrigel plug assay due to the lack of the target (i.e., erbB-2–positive tumor cells; Fig. 6B and D). Accordingly, we noted no additive activity of the combination regimens. However, trastuzumab, metronomic cyclophosphamide, MTD cyclophosphamide, and the corresponding combination regimens noticeably reduced angiogenesis within tumor cell containing Matrigel plugs (Fig. 6C and E).

**Second-line therapies of trastuzumab-resistant tumors.** Although addition of chemotherapy, especially metronomic cyclophosphamide, to trastuzumab significantly prolonged responses, we also wanted to evaluate therapy options that might be effective in treating tumors that have already acquired overt resistance to the drug. In this regard, the aforementioned results stimulated us to undertake studies testing the anti-human VEGF antibody, bevacizumab, as one such potential second-line therapy. Another potential mechanism of resistance to trastuzumab is the overexpression of EGFR ligands (30, 32). We therefore tested the EGFR targeting antibody cetuximab (C225) and the erbB-2 heterodimerization inhibitor pertuzumab as additional means to overcome trastuzumab resistance. Mice bearing 231-H2N tumors were treated with trastuzumab and metronomic cyclophosphamide until tumors began to progress around 100 days after treatment initiation as shown in Fig. 2A. Mice were then continued on this therapy, and treatment with bevacizumab, cetuximab or pertuzumab was added. Monitoring of tumor growth illustrated that both bevacizumab and cetuximab were effective at delaying tumor growth for 25 days (i.e., around day 125), before further progression (Fig. 7). In contrast, the addition of pertuzumab provided no benefit compared with those mice maintained on trastuzumab and metronomic cyclophosphamide until tumors began to progress around 100 days after treatment initiation as shown in Fig. 2A. These results suggest that in our breast carcinoma model, VEGF...
might indeed be involved in the progression of tumors resistant to trastuzumab, suggesting that bevacizumab treatment may be a therapeutic option to consider as a second-line additional agent for the treatment of breast cancer patients who are no longer responding to trastuzumab. Whether the improved response observed by addition of bevacizumab to trastuzumab requires continuation of trastuzumab therapy is unknown and is an issue that should be explored in detail in future investigations.

Implicit as well is the possibility that upfront combination of these two drugs may be more effective than either alone, in some cases, by delaying resistance/prolonging responses. In addition, EGFR-targeted drugs, such as cetuximab, may be effective agents in breast cancer patients with both erbB-2 amplification and expression of EGFR, as suggested by the observed delay in tumor progression in our studies when cetuximab was administered to trastuzumab-resistant tumors, which express both erbB-2 and EGFR.

Discussion

The prolonged delay in human breast cancer xenograft growth followed by rapid progression in those treatment groups receiving trastuzumab therapy with or without chemotherapy (cyclophosphamide) was not unexpected due to the frequent response, and then relapse, of responding patients treated with trastuzumab for metastatic erbB-2–positive breast cancer. Nevertheless, to our knowledge, our study is the first to report such a preclinical relapse in vivo following a reasonably prolonged treatment for mice (1 month), as a review of the literature revealed that there are few examples of longer-term studies using trastuzumab therapy. Considering the clinical importance of acquired resistance to trastuzumab, it is surprising that this area is only starting to be explored preclinically in more detail. Our results suggest that the acquired resistance to trastuzumab detected in our system is likely the consequence of heritable genetic alteration(s) in the cells and not due to host-mediated effects, because the resistant phenotype was maintained both in vitro and in vivo when reinjected into mice. Also interesting is that the majority of cell lines obtained from resistant tumors in this study maintained overexpression of the erbB-2 gene, suggesting that other tumor cell alterations are required to lead to the resistant phenotype.

Although an up-regulation of VEGF (and TGF-α) in tumors resistant to trastuzumab therapy was observed in our studies,
further work is required to confirm this observation (especially at the clinical level) and identify the mechanism(s) by which this occurs. In this regard, it is perhaps noteworthy that Konecny et al. have reported an association between erbB-2/HER2 and VEGF expression, which predicts clinical outcome in the treatment of breast cancer (33). There are a number of mechanisms that could potentially be involved in an up-regulation of VEGF in the trastuzumab-resistant variants (e.g., up-regulation of insulin-like growth factor-IR signaling; refs. 34–36) or other tyrosine kinase receptors, acquisition of PTEN mutations leading to loss of function (37), constitutive activation of phosphatidylinositol-3 kinase (38), or nuclear factor-%B (39), the development of hypoxia-inducible factor-%A–independent VEGF expression (40) or changes in expression or degradation of hypoxia-inducible factor-%A (41), among others. Because the resistant phenotype of the variants was expressed not only in vivo but in vitro, the contribution of elevated VEGF expression to the observed resistance can be called into question. This assumes that VEGF can only function as a paracrine factor and affect host dependent processes, such as tumor angiogenesis. However, it is possible that VEGF may act in an autocrine fashion due to expression of VEGF binding receptors on breast cancer cells (e.g., VEGFR-1, VEGFR-2, or neuropilin-1; refs. 42–44). We are currently evaluating whether the trastuzumab-resistant variants also (over)express such receptors, and if so, whether the elevated VEGF levels in the variants might directly contribute to the resistant phenotype in vitro.

Regardless of the mechanism by which tumors become resistant to trastuzumab, addition of an anti-VEGF–directed regimen, such as the antibody bevacizumab, to trastuzumab therapy can be rationalized by both the up-regulation of VEGF through the erbB-2 oncogene (41) and our present finding that VEGF is up-regulated within the tumor microenvironment upon acquisition of a resistant phenotype. The preliminary studies presented here, using our orthotopic tumor model, indicate that resumed tumor growth due to resistance to trastuzumab can be halted for a time by the addition of bevacizumab, the anti-VEGF monoclonal antibody, to the drug cocktail. It would be interesting to determine whether or not the administration of these two drugs before the development of resistance to trastuzumab therapy, as suggested by Konecny et al. (33) and Pegram and Reese (45), could delay tumor regrowth due to drug resistance. Indeed, a clinical trial has been initiated testing the combination of bevacizumab and trastuzumab for treatment of metastatic breast cancer, and early (interim) results look extremely promising (46).

The administration of the anti-EGFR antibody cetuximab to trastuzumab-resistant tumors was also effective at delaying tumor progression compared with those mice maintained only on metronomic cyclophosphamide with trastuzumab.

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**Fig. 6.** In vivo assessment of antiangiogenic effects of trastuzumab and cyclophosphamide (CTX) therapies. A, tumor perfusion assay. The decrease in intravascular FITC-dextran fluorescence reflects the changes in tumor perfusion in established 231-H2N orthotopic tumors treated with various cyclophosphamide and trastuzumab therapies for a 3-week cycle. B, basic fibroblast growth factor Matrigel assay. C, tumor cell Matrigel assay. The decrease in FITC-dextran fluorescence in the Matrigel plugs indicates inhibition of angiogenic processes by therapeutic regimens administered for 7 days. All data reported as fluorescence relative to untreated controls. Columns, mean (n = 5); bars, SD. Appearance of (D) basic fibroblast growth factor and (E) tumor cell Matrigel plugs excised from mice after 7 days of treatment. i, control; ii, trastuzumab; iii, metronomic cyclophosphamide; iv, metronomic cyclophosphamide + trastuzumab; v, MTD cyclophosphamide; vi, MTD cyclophosphamide + trastuzumab.
Although both EGFR and erbB-2 homodimers induce the expression of VEGF, two heterodimers (erbB-2/erbB3 and EGFR/erbB-2) induce even more potently VEGF expression and enhance tumor angiogenesis (47). Therefore, by simultaneously blocking EGFR and erbB-2 signaling with cetuximab and trastuzumab, respectively, the formation of the most potent heterodimer to regulate VEGF is prevented, in addition either bevacizumab and cetuximab treatments effectively delayed further tumor growth for 25 to 30 days. Symbols, mean (n = 4); bars, SE. P < 0.001 for low-dose cyclophosphamide + trastuzumab + bevacizumab and low-dose metronomic cyclophosphamide + trastuzumab + cetuximab versus low-dose cyclophosphamide + trastuzumab + bevacizumab + cetuximab. P > 0.05 for low-dose cyclophosphamide + trastuzumab + cetuximab versus low-dose cyclophosphamide + trastuzumab. The decision to use cyclophosphamide for our studies may be questioned in view of the fact that trastuzumab is often combined with taxane- or platinum-based drugs in the clinic. There are several reasons for our choice of chronic administration of cyclophosphamide. First, we have accumulated considerable evidence of toxicity through weight fluctuations and severe weight loss, the chronic administration of oral metronomic cyclophosphamide on a daily basis had no indications of serious long-term toxicity, even for treatment periods extending for up to 140 days, using SCID mice known to be particularly sensitive to this type of drug. We suggest that the prolonged antitumor efficacy of metronomic combination therapy may in part be due to the additive antiangiogenic side effects of these two drugs. In this regard, antiangiogenic properties have been attributed to trastuzumab by several groups (17, 18). Moreover, it has been reported by others (52) that trastuzumab, when combined with weekly low-dose paclitaxel, more effectively suppresses tumor growth, metastasis, and erbB-2–mediated angiogenesis, than either therapy alone, in an orthotopic breast tumor model. Our results confirm and extend this observation with an additional low-dose metronomic chemotherapy regimen, cyclophosphamide through the drinking water, which mimics the long-term treatment of metastatic breast cancer patients with daily low-dose cyclophosphamide with no breaks (55) and supports the additive antiangiogenic effects of combination therapy.
initiated in early-stage operable breast cancer (http://www.ClinicalTrials.gov; identifier NCT00121134). Another possible methodologic concern regarding the model and approach we studied is the use of an erbB-2 transfectant. Breast cancer cell lines, which express endogenous erbB-2, may obviously behave differently with respect to how they acquire resistance to trastuzumab. However, the poor tumorigenic properties of the few such lines that are available led us to adopt the transfaction approach as an experimental alternative.

In conclusion, we have outlined an experimental approach to study acquired resistance in vivo to the erbB-2–specific anticancer drug, trastuzumab. Using this model, we have shown that eventual relapses during trastuzumab treatment can be significantly delayed by prolonged combination therapy with metronomic low-dose cyclophosphamide chemotherapy, which seems as efficacious but much less toxic compared with combination with MTD cyclophosphamide therapy. In addition to delaying acquired resistance to trastuzumab, we have outlined some potential therapeutic approaches to treating tumor variants, which have actually acquired resistance (i.e., treatment with either anti-VEGF and anti-EGFR antibodies). In this regard, our results would seem to provide further support for the rationale of treating breast cancer using combinations of the targeted therapeutics, such as trastuzumab and bevacizumab (46). Other possible drug combination may be implicated by future studies designed to uncover the biochemical/molecular basis of the resistant phenotype.

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# Strategies for Delaying or Treating *In vivo* Acquired Resistance to Trastuzumab in Human Breast Cancer Xenografts


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