

Targeting the Kinesin Spindle Protein: Basic Principles and Clinical Implications

Vasiliki Sarli and Athanassios Giannis

Abstract Kinesin spindle protein (KSP), a member of the kinesin superfamily of microtubule-based motors, plays a critical role in mitosis as it mediates centrosome separation and bipolar spindle assembly and maintenance. Inhibition of KSP function leads to cell cycle arrest at mitosis with the formation of monoastral microtubule arrays, and ultimately, to cell death. Several KSP inhibitors are currently being studied in clinical trials and provide new opportunities for the development of novel anti-cancer therapeutics alternative from the available microtubule targeting drugs.

Background

The basics of kinesin spindle protein signaling. Several clinically important antimetabolites, for example, taxanes, epothilone derivatives, and *Vinca* alkaloids target tubulin, the basic microtubule subunit. This heterodimeric protein is an important constituent of the cytoskeleton and of the mitotic spindle. Microtubule-directed agents belong to the most successful anticancer agents, and since the introduction of paclitaxel (Taxol) in 1992, these drugs have had a central role in the therapy of breast, lung, ovarian, bladder, and head and neck cancers (1). They also contributed dramatically to a better quality of life of cancer patients and represent milestones in modern chemotherapy (2). However, the use of these agents is associated with serious side effects, such as neurotoxicity, related to the central role tubulin plays in cellular transport processes.

A few years ago, Mayer et al. discovered monastrol, the first small molecule able to inhibit the mitotic kinesin Eg5 (kinesin spindle protein, KSP) using a phenotype-based assay and methods of chemical genetics combined with advances in high-throughput screening (3). This discovery may be regarded as a paradigm shift in anticancer drug development. Monastrol inhibits a completely new target, i.e., a protein acting as a cellular motor (mitotic kinesin), expressed during mitosis and involved in cell division. Inhibitors of mitotic kinesins are now being investigated for their therapeutic potential and may represent the next generation of antimetabolites.

Kinesins are a large superfamily of motor proteins that participate in various biological phenomena including mitosis and intracellular transport of vesicles and organelles (4). Kinesins consists of a long coiled-coil stalk with a cargo-binding tail at one end and a globular tail domain, usually

called the head, at the other (Fig. 1A). The highly conserved motor domain, ~320 residues in size, contains both microtubule and nucleotide binding sites. This domain cooperates with the neck linker ~40 amino acids, to enable motor binding and stepping across the microtubules by converting the chemical energy of ATP hydrolysis to a mechanical force. To date, there are at least 12 kinesins involved in mitosis and meiosis which are responsible for spindle and chromosomal movement. Among them, KSP (HsEg5) is a slow, plus end-directed motor of the kinesin-5 subfamily (5–8). KSP forms a homotetrameric structure capable of binding antiparallel microtubules and sliding them apart (Fig. 1B and C; ref. 9). It acts during the early stages of mitosis and is responsible for centrosome separation and bipolar spindle assembly, which are essential for proper segregation of chromosomes (10). Failure of KSP function, by immunodepletion or knockdown of KSP mRNA by small interfering RNA, leads to cell cycle arrest in mitosis with monoastral microtubule arrays (11, 12). The important role of KSP in mitotic progression makes it an ideal candidate for drug discovery. Furthermore, it is most abundant in proliferating human tissues and is highly expressed in tumors of the breast, colon, lung, ovary, and uterus.

Targeting KSP provides a novel route for the manipulation of the cell cycle, alternative to antimetabolite agents that target tubulin. Tubulin, a heterodimeric protein, is an important constituent of the cytoskeleton and of the mitotic spindle. Microtubule-directed agents belong to the most successful anticancer agents, and since the introduction of paclitaxel (Taxol) in 1992, these drugs have had a central role in the therapy of breast, lung, ovarian, bladder, and head and neck cancers (13). They also contributed dramatically to a better quality of life of cancer patients and represent milestones in modern chemotherapy (2). Agents which target KSP selectively would be expected to act only on cells undergoing cell division, thus making KSP inhibitors mitosis-specific drugs, likely to have fewer side effects than drugs inhibiting other essential microtubule-based processes.

KSP inhibitors and mechanism of action. A few years ago, Mayer et al. discovered monastrol (1), the first small molecule able to inhibit the mitotic kinesin Eg5 (KSP) using a phenotype-based assay and methods of chemical genetics combined with advances in high-throughput screening (14).

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Monastrol is an allosteric inhibitor that binds to an induced-fit pocket 12 Å away from the catalytic center by helices $\alpha 2$ and $\alpha 3$ and loop L5 (15). It binds to KSP and causes both local and distal conformational changes that allow ATP binding, but prevent ADP release. The weak inhibitory activity of monastrol and its non-drug-like properties led to the synthesis of a series of second-generation derivatives with improved cellular potency and solubility (16, 17). Moreover, several other inhibitors that exhibit great chemical diversity have now been reported including adociasulfate-2 (2, 18), terpendole E (3, 19), HR22C16 (4, 20), CK0106023 (5, 21), dihydropyrazoles (6, 22), S-trityl-L-cysteine (7, 23), dihydropyrroles (8, 24), thiazoles (9, 25), and many more (Fig. 2; ref. 26, 27).

Most of the inhibitors mentioned above are highly selective and share a common mode of action as monastrol, by binding

to the same allosteric pocket of KSP. Others, like the thiazole-containing inhibitors, are competitive with ATP and uncompetitive with microtubules. Interestingly, biaryl compounds, such as GSK-1 and GSK-2, do not bind in the nucleotide-binding pocket, but they antagonize ATP interactions through an allosteric mechanism, proven by site-directed mutagenesis and photo-affinity labeling (28, 29). The natural product, adociasulfate-2, also acts in a unique manner, as it is not competitive with ATP binding, but interferes with the microtubule-binding site (30). The distinct mechanism of action that KSP inhibitors display could prove especially useful in the clinic for avoiding KSP mutation-mediated resistance.

Mechanisms of cell death induced by KSP inhibitors. Although it is known that KSP inhibitors induce apoptosis after prolonged mitotic arrest, the underlying mechanism is not completely understood. Leizerman et al. have shown that monastrol causes mitotic arrest and induces early apoptosis through mitochondrial membrane depolarization, caspase-8 and caspase-3 activation, and cleavage of poly-ADP-ribose polymerase 1 with different sensitivity in human AGS and HT29 cell lines from gastric and colon carcinoma (31). Similar to other antimetotics that target tubulin, KSP inhibitors activate the spindle assembly checkpoint. The spindle checkpoint ensures the correct segregation of chromosomes by inhibition of cell cycle progression until all chromosome kinetochores are properly attached to the bipolar spindle and chromosomes are aligned at the metaphase plate (32). Well-characterized components of the spindle checkpoint include Mad1, Mad2, Mad3 (BubR1), Bub1, Bub3, and Mps1. The metaphase-anaphase transition occurs as a result of the Cdc20-dependent activation of the anaphase-promoting complex or cyclosome (APC/C). APC/C is a multisubunit E3 ubiquitin ligase that triggers the ubiquitination of a number of mitotic substrates targeting them for destruction by the proteasome.

One hypothesis is that, in response to KSP inhibitors, cells eventually die as a result of overriding the checkpoint. Cells that exit mitosis with unaligned chromosomes will become aneuploid and die. Tao et al. (33) reported that induction of apoptosis in cells treated with a KSP inhibitor happens after long-term mitotic arrest, followed by adaptation and slippage into the next G_1 phase. In their studies, KSP-IA (8), a dihydropyrrole small molecule arrests cells in mitosis and induces apoptosis by caspase-dependent death. Cells refractory to slippage or cells with a weakened mitotic checkpoint showed a diminished apoptotic response. It seems that activated checkpoint components such as BubR1 (a protein kinase that is active at unattached kinetochores and contributes to the signal that stops the progression of anaphase), may be required for the induction of apoptosis by KSP-targeted drugs following exit from mitosis. Moreover, KSP-IA was able to induce apoptotic cell death in a p53-independent manner, suggesting that KSP inhibitors could be proved active in p53-deficient tumors.

On the other hand, Chin et al. showed that induction of apoptosis by monastrol is independent of the spindle checkpoint and inhibition of KSP leads to caspase activation and apoptosis in the absence of critical checkpoint proteins such as BubR1 and Mad2 (34). Vijapurkar et al. further elucidated the cellular responses following monastrol-induced mitotic arrest (35). They suggest that the cellular responses induced by monastrol are correlated with overexpression of

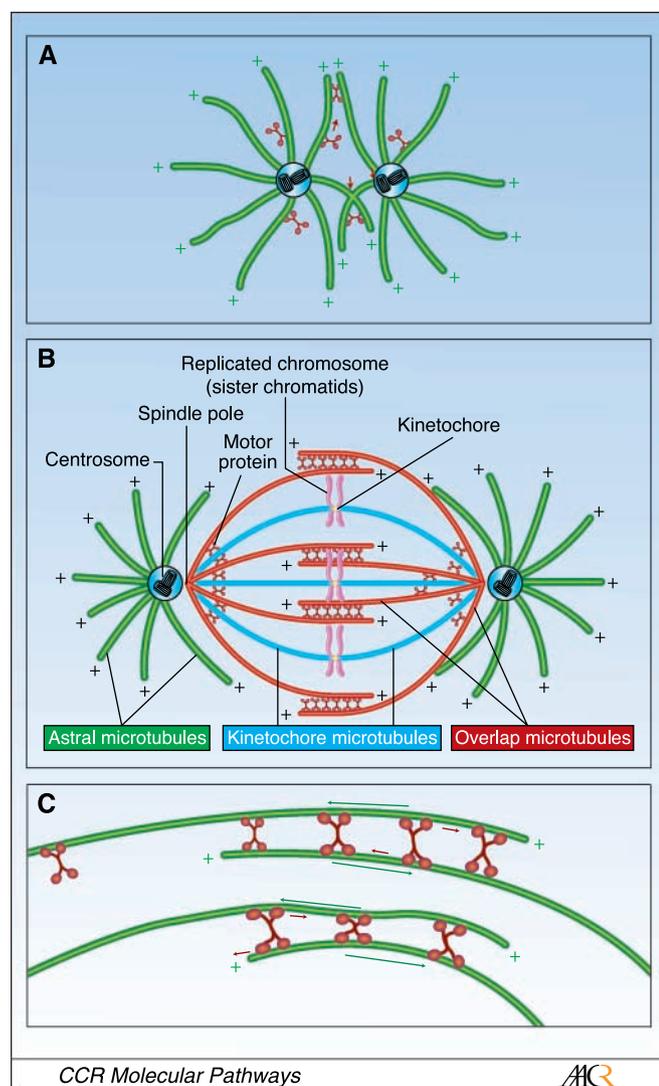


Figure 1. KSP motors (red) enable microtubules (green) to form the mitotic spindle. *A*, at the onset of mitosis, the duplicated centrosomes (blue) separate and generate two star-like structures (microtubule asters). KSP motors are moving to the plus-ends of microtubules, and promote bipolarity. *B*, during metaphase, a stable bipolar spindle is formed and KSP generate a poleward flux. *C*, KSP motors are shown walking to the plus-ends of antiparallel microtubules, moving both poleward simultaneously. *A* and *C* adapted with permission from Valentine et al. (52); *B*, adapted with permission from Albers et al. (53).

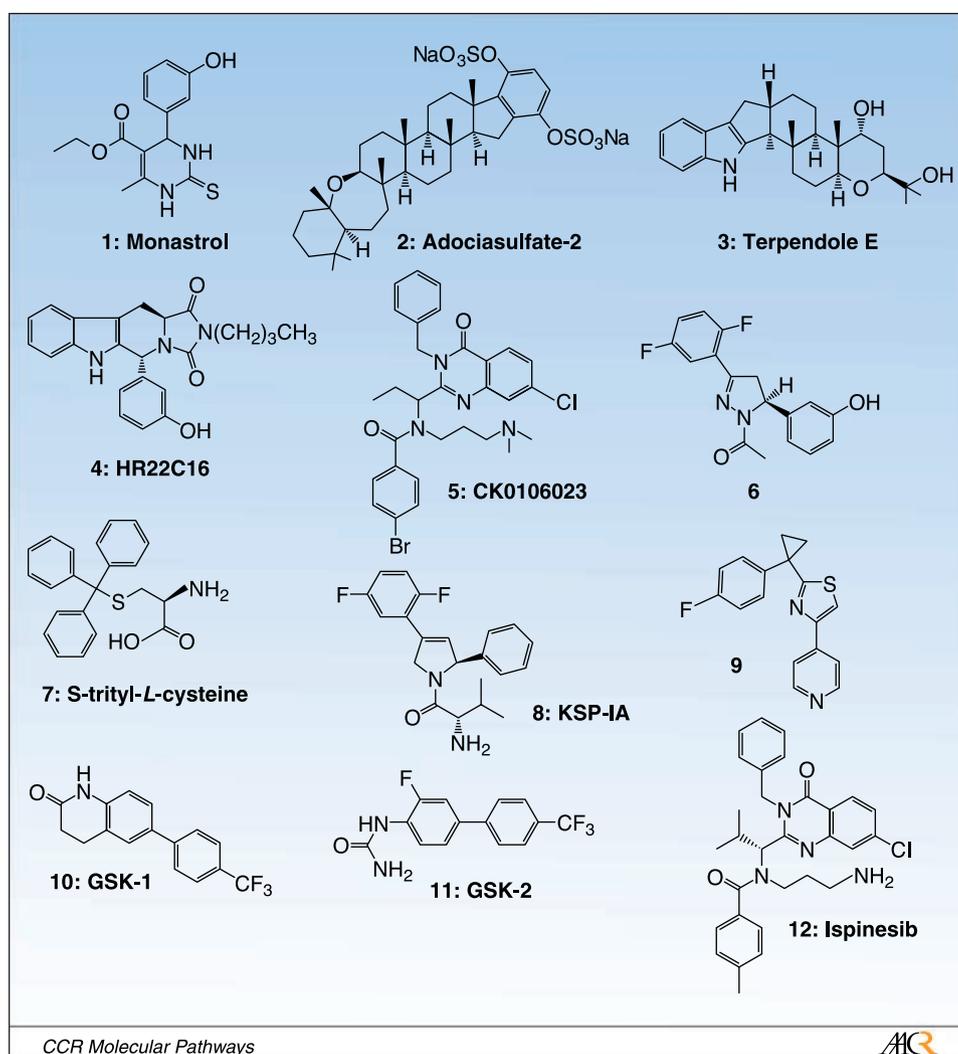


Figure 2. Structures of KSP inhibitors (see also text).

BclXL, the antiapoptotic Bcl-2 family protein. Overexpression of BclXL provides a protective mechanism, and its depletion rescues the apoptotic response to monastrol. Furthermore, activation of the death receptor pathway by treatment with Fas receptor agonists sensitized the cells to monastrol-induced cell death, following exit from mitosis.

In another study (36), it was shown that dimethylnastron (a KSP inhibitor) induced apoptosis and simultaneously up-regulated Hsp70 in human multiple myeloma cells. Dimethylnastron-mediated Hsp70 up-regulation is cytoprotective because blocking Hsp70 induction directly by antisense or small interfering RNA or indirectly by inhibitors of the phosphatidylinositol 3-kinase/Akt pathway dramatically increased KSP inhibitor-induced apoptosis. In addition, FTI277, a specific small-molecule inhibitor of farnesyltransferase, interacted synergistically with dimethylnastron in inducing apoptosis through disrupting the Akt/Hsp70 signaling axis. These findings provided the first evidence for KSP inhibitor activity in hematologic malignancy and identify Hsp70 up-regulation as a critical mechanism responsible for modulating myeloma cell sensitivity to KSP inhibitors. These results suggest that a combination of KSP inhibitors with agents abolishing Hsp70 induction would be useful for myeloma therapy and

may be of importance for future combination therapies in other malignancies.

Clinical-Translational Advances

Several KSP kinesin inhibitors are currently under development by pharmaceutical or biotechnology companies for cancer treatment. Ispinesib (SB-715992; ref. 12) was the first kinesin inhibitor to enter clinical trials by Cytokinetics and GlaxoSmithKline (37). At present, it represents the most advanced and best studied KSP inhibitor. Below, we summarize and discuss the results from clinical trials involving this quinazolinone derivative, which is now in phase II clinical trials. Phase I studies of ispinesib in patients with solid tumors have been completed. Generally, ispinesib was well-tolerated with an acceptable safety profile with no indications of neurotoxicity. The most common adverse effects were neutropenia, fatigue, anemia, leukopenia, thrombocytopenia, diarrhea, nausea, and vomiting (38–40). The most promising results have been observed in patients with locally advanced or metastatic breast cancer (41). In this study, the predetermined response criteria to progress from stage 1 to stage 2 of the clinical trial were achieved, and patients are currently being

enrolled in stage 2, in which an additional 25 patients are planned to be enrolled and evaluated. Furthermore, preliminary results from ongoing trials in patients with breast cancer show that ispinesib, in conjunction with capecitabine, indicate an acceptable tolerability profile, and that 8 of 16 patients experienced stable disease for 2 to 6.5 months (42).

Recently, results from the phase II trial of ispinesib in 15 patients with metastatic hepatocellular carcinoma were published (43). Ispinesib was administered as a 1-hour i.v. infusion at a dose of 18 mg/m² once every 3 weeks. In this study, no conclusive evidence of benefit was seen with ispinesib monotherapy. Similar results were obtained from the phase II study in patients with metastatic or recurrent malignant melanoma (44). Although KSP expression seems to be common in melanoma and KSP would be an appropriate target for inhibition, no significant responses were observed in response to ispinesib and further development in malignant melanoma is not recommended. Other preliminary reports of ispinesib in recurrent or metastatic squamous cell carcinoma of the head and neck (45), colorectal cancer, ovarian cancer, and renal cell carcinoma have not indicated significant response rates (46, 47).

Taken together, the results from the first clinical trials of ispinesib are rather disappointing. Nevertheless, it should be taken into account that in these trials, ispinesib was used as monotherapy and further trials are necessary for the evaluation of safety, tolerability, and efficacy of ispinesib in combination with other anticancer therapeutics. For example, cisplatin, a DNA-damaging agent, has been shown to enhance the activity of ispinesib against murine P388 lymphocytic leukemia. In general, it was shown that ispinesib in combination with these standard chemotherapeutic agents has an acceptable tolerability profile with neutropenia as dosing-limiting toxicity (48).

Second-generation KSP inhibitors are now in the clinic. SB-743921, a derivative of ispinesib, is 5-fold more potent against KSP ATPase activity and is currently being evaluated in a phase I/II clinical trial in non-Hodgkin's lymphoma. MK-0731 (Merck) progressed in clinical development has shown antiproliferative activity in many tumor cell lines and significant efficacy in

several murine tumor models. In addition, ARRY-520 (Array Biofarma) is currently in a phase I trial in advanced cancer patients and has shown remarkable efficacy in preclinical models of human solid tumors and human leukemias.

The next challenge for the development of KSP inhibitors in the clinic will be multidrug resistance. Resistance to chemotherapeutic drugs is a principal problem in the treatment of cancer that limits the effectiveness of cytostatic drugs, through the expression of efflux pumps, such as P-glycoprotein (49, 50). It is known that ispinesib resistance, observed in some clinical trials, may be due to multidrug resistance. In fact, preclinical studies indicated that ispinesib may be a substrate for MDR-1, resulting in a variation in response (37). Furthermore, mutations that attenuate ispinesib binding to KSP have already been identified in resistant HCT116 colorectal tumor cells. Two distinct point mutations have been observed resulting in amino acid substitutions within the loop 5 region of the KSP motor domain (51).

Taken together, clinical trials evaluating the efficacy and safety of KSP inhibitors, both as single and in combination with other agents, are currently ongoing. A better understanding of the molecular pathways that KSP inhibitors induce cell death will generate new insights into future therapy. Although ispinesib used as cancer monotherapy proved to be of limited efficacy, its combination with, for example, modulators of the Akt/Hsp70 signaling axis or Fas receptor agonists may be a promising alternative. The cytostatic activity of KSP inhibitors may be augmented by their inhibitory action toward endothelial cell proliferation, resulting in the inhibition of angiogenesis. Furthermore, it should be mentioned that inhibitors of mitotic kinesins may find broad application for the treatment of other proliferative diseases like diabetic retinopathy, pulmonary and liver fibrosis, Sjögren's syndrome, lupus erythematosus, and lymphoproliferative disorders that develop in patients with a history of autoimmune disease.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

References

- Chabner BA, Amrein PC, Druker, et al. Antineoplastic agents. In: Brunton LL, Lazo JS, Parker KL, editors. Goodman & Gilman's the pharmacological basis of therapeutics. 11th ed. McGraw-Hill, New York; 2006.
- Chabner BA, Roberts TG. Chemotherapy and the war on cancer. *Nat Rev Cancer* 2005;5:65–72.
- Mayer TU, Kapoor TM, Haggarty SJ, King RW, Schreiber SL, Mitchison TJ. Small molecule inhibitor of mitotic spindle bipolarity identified in a phenotype-based screen. *Science* 1999;286:971–4.
- Vale RD, Fletterick RJ. The design plan of kinesin motors. *Annu Rev Cell Dev Biol* 1997;13:745–77.
- Kapitein LC, Peterman E, Kwok BH, et al. The bipolar mitotic kinesin Eg5 moves on both microtubules that it crosslinks. *Nature* 2005;435:114–8.
- Cameron LA, Yang G, Cimini D, et al. Kinesin 5-independent poleward flux of kinetochore microtubules in PtK1 cells. *J Cell Biol* 2006;173:173–9.
- Valentine MT, Fordyce PM, Krzyziak TC, et al. Individual dimers of the mitotic kinesin motor Eg5 step processively and support substantial loads *in vitro*. *Nat Cell Biol* 2006;8:470–6.
- Kashina AS, Baskin RJ, Cole DG, et al. A bipolar kinesin. *Nature* 1996;379:270–2.
- Cole DG, Saxton WM, Sheehan KB, et al. A "slow" homotetrameric kinesin-related motor protein purified from *Drosophila* embryos. *J Biol Chem* 1994;269:22913–6.
- Sawin KE, LeGuellec K, Philippe M, et al. Mitotic spindle organization by a plus-end-directed microtubule motor. *Nature* 1992;359:540–3.
- Blangy A, Lane HA, d'Herin P, et al. Phosphorylation by p34cdc2 regulates spindle association of human Eg5, a kinesin-related motor essential for bipolar spindle formation *in vivo*. *Cell* 1995;83:1159–69.
- Weil D, Garçon L, Harper M, et al. Targeting the kinesin Eg5 to monitor siRNA transfection in mammalian cells. *Biotechniques* 2002;33:1244–8.
- Chabner BA, Amrein PC, Druker, et al. Antineoplastic agents. In: Brunton LL, Lazo JS, Parker KL, editors. Goodman & Gilman's the pharmacological basis of therapeutics. 11th ed. McGraw-Hill, New York; 2006.
- Mayer TU, Kapoor TM, Haggarty SJ, et al. Small molecule inhibitor of mitotic spindle bipolarity identified in a phenotype-based screen. *Science* 1999;286:971–4.
- Yan Y, Sardana V, Xu B, et al. Inhibition of a mitotic motor protein: where, how, and conformational consequences. *J Mol Biol* 2004;335:547–54.
- Gartner M, Sunder-Plassmann N, Seiler J, et al. Development and biological evaluation of potent and specific inhibitors of mitotic kinesin Eg5. *Chembiochem* 2005;6:1173–7.
- Sarli V, Huemmer S, Sunder-Plassmann N, et al. Synthesis and biological evaluation of novel EG5 inhibitors. *Chembiochem* 2005;6:2005–13.
- Sakowicz R, Berdelis MS, Ray K, et al. A marine natural product inhibitor of kinesin motors. *Science* 1998;280:292–5.
- Nakazawa J, Yajima J, Usui T, et al. A novel action of terpendole E on the motor activity of mitotic kinesin Eg5. *Chem Biol* 2003;10:131–7.
- Hotha S, Yarrow JC, Yang JG, et al. HR22C16: a potent small-molecule probe for the dynamics of cell division. *Angew Chem Int Ed Engl* 2003;42:2379–82.
- Sakowicz R, Finer JT, Beraud C, et al. Antitumor activity of a kinesin inhibitor. *Cancer Res* 2004;64:3276–80.
- Cox CD, Breslin MJ, Mariano BJ, et al. Kinesin spindle protein (KSP) inhibitors. Part 1: the discovery of 3,5-diaryl-4,5-dihydropyrazoles as potent and selective inhibitors of the mitotic kinesin KSP. *Bioorg Med Chem Lett* 2005;15:2041–5.
- DeBonis S, Skoufias D, Robin G, et al. *In vitro* screening for inhibitors of the human mitotic kinesin

- Eg5 with antimetabolic and antitumor activities. *Mol Cancer Ther* 2004;3:1079–90.
24. Tao W, South VJ, Zhang Y, et al. Induction of apoptosis by an inhibitor of the mitotic kinesin KSP requires both activation of the spindle assembly checkpoint and mitotic slippage. *Cancer Cell* 2005;8:49–59.
 25. Rickert KW, Schaber M, Torrent M, et al. Discovery and biochemical characterization of selective ATP competitive inhibitors of the human mitotic kinesin KSP. *Arch Biochem Biophys* 2008;469:220–31.
 26. Bergnes G, Brejc K, Belmont L. Mitotic kinesins: prospects for antimetabolic drug discovery. *Curr Top Med Chem* 2005;5:127–45.
 27. Sarli V, Giannis A. Inhibitors of mitotic kinesins: next-generation antimetabolites. *ChemMedChem* 2006;13:293–8.
 28. Parrish CA, Adams ND, Auger KR, et al. Novel ATP-competitive kinesin spindle protein inhibitors. *J Med Chem* 2007;50:4939–52.
 29. Luo L, Parrish CA, Nevins N, et al. ATP-competitive inhibitors of the mitotic kinesin KSP that function via an allosteric mechanism. *Nat Chem Biol* 2007;3:722–6.
 30. Brier S, Carletti E, DeBonis S, et al. The marine natural product adociasulfate-2 as a tool to identify the MT-binding region of kinesins. *Biochemistry* 2006;45:15644–53.
 31. Leizerman I, Avunie-Masala R, Elkabets M, et al. Differential effects of monastrol in two human cell lines. *Cell Mol Life Sci* 2004;61:2060–70.
 32. Kops GJ, Weaver BA, Cleveland DW. On the road to cancer: aneuploidy and the mitotic checkpoint. *Nat Rev Cancer* 2005;5:773–85.
 33. Tao W, South VJ, Diehl RE, et al. An inhibitor of the kinesin spindle protein activates the intrinsic apoptotic pathway independently of p53 and *de novo* protein synthesis. *Mol Cell Biol* 2007;27:689–98.
 34. Chin GM, Herbst R. Induction of apoptosis by monastrol, an inhibitor of the mitotic kinesin Eg5, is independent of the spindle checkpoint. *Mol Cancer Ther* 2006;5:2580–91.
 35. Vijapurkar U, Wang W, Herbst R. Potentiation of kinesin spindle protein inhibitor-induced cell death by modulation of mitochondrial and death receptor apoptotic pathways. *Cancer Res* 2007;67:237–45.
 36. Liu M, Aneja R, Liu C, et al. Inhibition of the mitotic kinesin Eg5 up-regulates Hsp70 through the phosphatidylinositol 3-kinase/Akt pathway in multiple myeloma cells. *J Biol Chem* 2006;281:18090–7.
 37. Johnson RK, McCabe FL, Cauder E, et al. SB-715992, a potent and selective inhibitor of KSP mitotic kinesin, demonstrates broad-spectrum activity in advanced murine tumors and human tumor xenografts. *Proc Am Assoc Cancer Res* 2002;43:269.
 38. Burris HA, Lorusso P, Jones S, et al. Phase I trial of novel kinesin spindle protein (KSP) inhibitor SB-715992 IV days 1, 8, 15 q 28 days (this is an abstract, only one page). *J Clin Oncol* 2004;22:2004.
 39. Heath EI, Alouisi A, Eder JP, et al. A phase I dose escalation trial of ispinesib (SB-715992) administered days 1-3 of a 21-day cycle in patients with advanced solid tumors. *Proc Am Soc Clin Oncol* 2006 24:A2026.
 40. Chu QS, Holen KD, Rowinsky EK, et al. Phase I trial of novel kinesin spindle protein (KSP) inhibitor SB-715992 IV Q 21 days. *Proc Am Soc Clin Oncol* 2004;22:A2078.
 41. Miller K, Ng C, Ang P, et al. PTC phase II, open label study of SB-715992 (Ispinesib) in subjects with advanced or metastatic breast cancer. *Breast Cancer Res Treat* 2005;94 (suppl 1): abstract 1089.
 42. Tripathy D. Capecitabine in combination with novel targeted agents in the management of metastatic breast cancer: underlying rationale and results of clinical trials. *Oncologist* 2007;12:375–89.
 43. Knox JJ, Gill S, Synold TW, et al. A phase II and pharmacokinetic study of SB-715992, in patients with metastatic hepatocellular carcinoma: a study of the National Cancer Institute of Canada Clinical Trials Group (NCIC CTG IND.168). *Invest New Drugs* 2008; 26:265–72.
 44. Lee CW, Bélanger K, Rao SC, et al. Phase II study of ispinesib (SB-715992) in patients with metastatic or recurrent malignant melanoma: a National Cancer Institute of Canada Clinical Trials Group trial. *Invest New Drugs* 2008;26:249–55.
 45. Tang PA, Siu LL, Chen EX, et al. Phase II study of ispinesib in recurrent or metastatic squamous cell carcinoma of the head and neck. *Invest New Drugs* 2008;26:257–64.
 46. El-Khoueiry AB, Iqbal S, Singh DA, et al. A randomized phase II non-comparative study of Ispinesib given weekly or every three weeks in metastatic colorectal cancer. A California Cancer Consortium Study (CCC-P). *ASCO Annual Meeting Proceedings* 2006, Part 1 24 (18S) *Invest New Drugs*: abstract 3595.
 47. Beekman KW, Dunn R, et al. University of Chicago Consortium phase II study of ispinesib (SB-715992) in patients (pts) with advanced renal cell carcinoma (RCC). *ASCO Annual Meeting Proceedings Part 1* 25(18S): abstract 15573.
 48. Calvo E, Chu Q, Til E, et al. Phase I study of ispinesib in combination with capecitabine in patients with advanced solid tumors. *AACR-NCI-EORTC*, November 2005.
 49. Dean M, Fojo T, Bates S. Tumor stem cells and drug resistance. *Nat Rev Cancer* 2005;5:275–84.
 50. Mimeault M, Hauke R, Batra SK. Recent advances on the molecular mechanisms involved in the drug resistance of cancer cells and novel targeting therapies. *Clin Pharmacol Ther* 2008;83:673–91.
 51. Jackson JR, Auger KR, Gilmartin A, et al. A resistance mechanism for the KSP inhibitor Ispinesib implicates point mutations in the compound-binding site. *Poster Meeting C207, AACR-NCI-EORTC International Conference on Molecular Targeted Cancer Therapeutics: Discovery, Biology and Clinical Applications*, November 14-18, 2005; Philadelphia.
 52. Valentine MT, Fordyce PM, Block SM. Eg5 steps it up! *Cell Div* 2006;1:31.
 53. Albers B, et al. *Molecular biology of the cell*. Walter, 4th ed. Garland Science; 2002. p. 1037.

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