

## New Strategies in Pleural Mesothelioma: BAP1 and NF2 as Novel Targets for Therapeutic Development and Risk Assessment

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### Abstract

Malignant pleural mesothelioma (MPM) is a highly lethal cancer with limited therapeutic options. Recent work has focused on the frequent somatic inactivation of two tumor suppressor genes in MPM—*NF2* (Neurofibromatosis type 2) and the recently identified *BAP1* (BRCA associated protein 1). In addition, germline mutations in *BAP1* have been identified that define a new familial cancer syndrome, which includes MPM, ocular melanoma, and other cancers. These recent advances may allow screening of high-risk individuals and the development of new therapies that target key pathways in MPM. *Clin Cancer Res*; 18(17): 4485–90. ©2012 AACR.

### Background

Malignant pleural mesothelioma (MPM) is an almost universally fatal disease striking approximately 3,000 individuals each year in the United States (1). Currently available therapies for MPM have a limited effect on outcomes. Even with aggressive multimodality therapy, including surgery, for early-stage disease, the median overall survival is 18 months from the time of diagnosis. For patients with advanced disease, first-line chemotherapy with pemetrexed and cisplatin improves median survival by a few months, but the benefit is limited and the use of second-line chemotherapy has not been established (2).

In this context, numerous clinical trials have been conducted in MPM to evaluate a variety of targeted agents. These include angiogenesis inhibitors such as bevacizumab and thalidomide, receptor kinase inhibitors such as erlotinib and sorafenib, and histone deacetylase inhibitors such as vorinostat (3). Despite the activity of these drugs in other diseases, they have shown little activity in MPM. Ideally, treatments that more specifically target the unique biology of MPM should be more effective. Just as the identification of somatic mutations in other malignancies has led to

the development of effective disease-specific therapeutics, the potential identification of similar targetable somatic mutations in MPM would be important.

This review calls attention to emerging data on the frequent inactivation of 2 genes in MPM—neurofibromatosis type II (*NF2*) and BRCA1-associated protein 1 (*BAP1*)—that could help elucidate key pathways that drive tumorigenesis and could subsequently be exploited as rational targets for drug development in this cancer.

### NF2 Loss and mTOR Blockade

About 35% to 40% of MPMs carry inactivating mutations at the neurofibromatosis 2 (*NF2*) locus, which encodes for the FERM domain protein Merlin (moesin-ezrin-radixin-like protein; refs. 4, 5). The mechanisms by which Merlin suppresses tumorigenesis have remained poorly defined. Merlin is homologous to the ERM proteins ezrin, radixin, and moesin and, like these proteins, it associates with the cortical actin cytoskeleton. Because loss of Merlin causes activation of multiple mitogenic signaling pathways, such as HER1/2, mTOR, extracellular signal-regulated kinase (ERK), and focal adhesion kinase (FAK), it has been postulated that Merlin inhibits signaling by negatively regulating multiple cell surface receptors (6). However, Merlin lacks an actin-binding motif and its dephosphorylated conformer, which is the only one active as a tumor suppressor, is not enriched at the cell membrane (7). Recent studies have revealed that the dephosphorylated, active form of Merlin accumulates in the nucleus and interacts with DCAF1, the receptor component of the E3 ubiquitin ligase CRL4<sup>DCAF1</sup> (8). Notably, Merlin inhibits CRL4<sup>DCAF1</sup>, which in turn promotes a broad oncogenic gene expression program, presumably by ubiquitinating transcription factors, histones, or chromatin-remodeling enzymes. This deregulation of nuclear ubiquitination events is an

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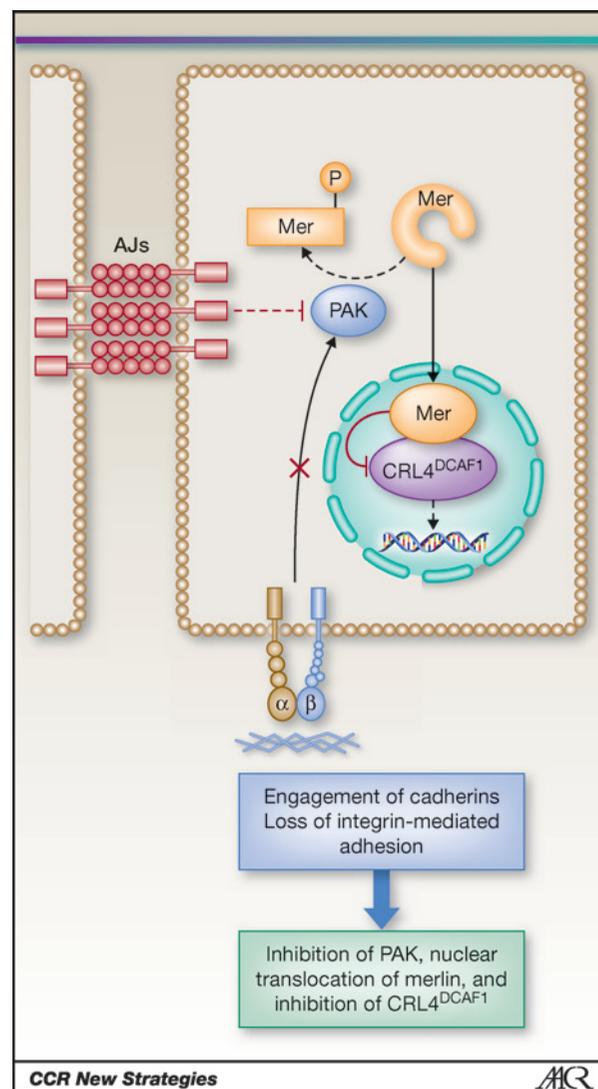
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intriguing common thread with BAP1 inactivation as described below, and the possible overlap between DCAF and BAP1 targets merits further investigation. Genetic epistasis experiments and an analysis of several Merlin missense mutations from NF2 patients support the hypothesis that the dephosphorylated CRL4<sup>DCAF1</sup> (8). These results suggest that Merlin affects multiple mitogenic signaling pathways by controlling, through CRL4<sup>DCAF1</sup>, the expression of components of regulators of these pathways (Fig. 1; ref. 9). It has also been proposed that Merlin loss in MPM, along with inactivating mutations in *LATS2* in a small percentage of cases (10, 11), may also contribute to oncogenesis through activation of the Hippo signaling cascade (12). However, it remains unclear whether loss of Merlin inactivates the Hippo kinase in the cytosol, as genetic studies in the fly suggest, or it deregulates the oncoprotein and transcriptional coactivator YAP through activation of CRL4<sup>DCAF1</sup> (9).

Merlin mediates contact-dependent inhibition of cell proliferation in normal cells, primarily through inhibition of mTOR in an AKT-independent manner (7). Without Merlin, mTOR activity is aberrantly upregulated, leading to increased cell proliferation (13). This mTOR activation has also been observed in other Merlin-deficient tumors (14). This increased proliferation can be reduced but not completely suppressed by mTOR inhibition. In the presence of Merlin, mTOR inhibition has little effect on growth. In MPM tumors and cell lines, there is an inverse relationship between phospho-mTOR expression and NF2 expression confirming the link between NF2 and the mTOR pathway, and we have found that immunohistochemical staining for NF2 and phospho-mTOR could be useful in selecting patients with MPM for trials targeting mTOR pathway activation due to NF2 loss (15). These preclinical observations have provided the rationale for studying mTOR inhibitors in patients with MPM. Everolimus, an oral derivative of the organ transplant immunosuppression drug rapamycin, which has also been approved for the treatment of advanced renal cell carcinoma, is being investigated as second-line therapy in a phase II trial run by the Southwest Oncology Group (SWOG). In addition, due to compensatory parallel pathway activation, mTOR inhibition alone may not be sufficient to suppress tumor growth. Indeed, we have found that MET, epidermal growth factor receptor, and insulin-like growth factor-1R are variably activated after mTOR inhibition in MPM cell lines, contributing to AKT feedback activation (16).

Preclinical evidence indicates that isolated mTOR inhibition alleviates feedback inhibition on PI3K and thereby allows restoration of PI3K and downstream AKT signaling (17). To address this mechanism of mTOR resistance, numerous dual PI3K/mTOR inhibitors are in early development. Two compounds, BEZ235 and GDC0980, have shown promise in MPM. BEZ235, a dual PI3K/mTOR inhibitor, is able to inhibit the growth of peritoneal mesothelioma cells (18). In addition, in a phase I trial of GDC0980, 3 of 6 enrolled patients with MPM had tumor shrinkage ranging from 23% to 28% and 1 patient had



**Figure 1.** Mechanisms of NF2 (Merlin)-mediated inhibition of proliferation. Merlin exists in an open, inactive form and a closed, active form. Matrix adhesion and the consequent activation of joint integrin receptor tyrosine kinase signaling activate the serine-threonine kinase p21-activated kinase (PAK). PAK in turn phosphorylates the C-terminus of Merlin, disrupting the intramolecular association that maintains the protein in a closed conformation. The resulting inactivation of Merlin removes a block to cell-cycle progression in normal cells. Conversely, engagement of E-cadherin and the ensuing assembly of adherens junctions (AJ) inactivate PAK, leading to an accumulation of the closed, active form of Merlin, which migrates into the nucleus to inhibit CRL4<sup>DCAF1</sup>. When it is not inhibited by Merlin, CRL4<sup>DCAF1</sup> positively regulates a broad oncogenic program of gene expression, which includes mitogenic signaling components, antiapoptotic proteins, and Hippo pathway target genes (8). CRL4, cullin-ring E3 ligase; 4; DCAF1, DDB1- and CUL4-associated factor. 1 (Figure adapted and modified from ref. 9).

stable disease on this compound for more than 1 year (19). However, cases of grade 2 and 3 pneumonitis have been reported with GDC0980, albeit at a dose above the maximum tolerated dose. It remains unclear whether the pulmonary toxicity will pose a clinical obstacle to the further application of these agents in the treatment of MPM.

### BAP1 Somatic Mutations

The frequent occurrence of *NF2* gene loss in MPM, as well as the very high prevalence of p16/*CDKN2A* deletions (20–22) have been known since the mid-1990s. However, a recent integrated genomics analysis has revealed another frequently inactivated tumor suppressor gene in this cancer. By profiling genomic copy number alterations in 53 MPM tumor samples, Bott and colleagues identified candidate regions for sequencing (10). The 3 most common deletions noted were at 22q (which includes *NF2*), 9p21 (which includes *CDKN2A*), and 3p21. Within these and other candidate regions, integration with gene expression data was used to prioritize genes for sequencing. The gene implicated by deletions in 3p21 was subsequently elucidated from sequencing of this region. Of the genes selected for sequencing, the highest rate of nonsynonymous somatic mutations was in the *BAP1* gene at 3p21.1, occurring in 23% of the samples. A high rate of somatic mutations was subsequently confirmed in 6 of 25 (24%) MPM cell lines and 12 of 68 (18%) additional MPM tumor samples. In all, *BAP1* loss, mutation, or both were identified in 42% of cases. Thirty-two different *BAP1* mutations were identified: 6 nonsense, 5 missense, 13 frameshifting, and 8 at or near splice sites. The majority of truncating mutations resulted in loss of the nuclear localization signal and/or the C-terminal protein-binding domain (Fig. 2). Several missense mutations were shown to affect the ubiquitin hydrolase activity of *BAP1*. Splice site mutations caused exon skipping leading to aberrant, out of frame transcripts. Immunohistochemistry for *BAP1* confirmed the association between *BAP1* mutation and an absence of *BAP1* protein expression. Interestingly, 25% of tumors without identified *BAP1* mutations did not display any immunohistochemical staining for *BAP1*, suggesting the possibility of another subset of tumors with functional *BAP1* loss arising by other mechanisms. A recent study has suggested that *BAP1* inactivation may be more characteristic of epithelioid MPM (23), although this was not apparent in the original study (10).

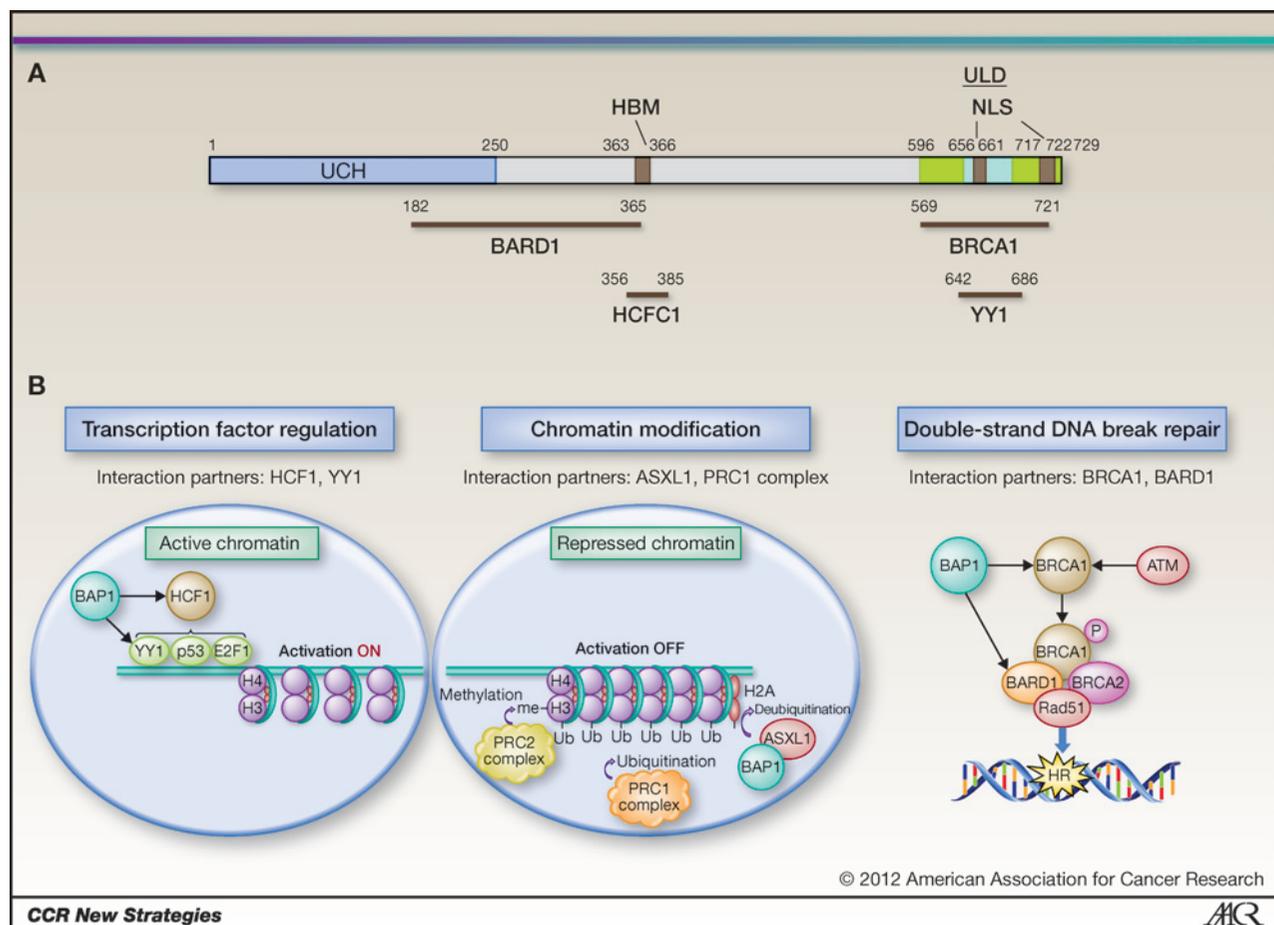
*BAP1* is a 729 amino acid nuclear ubiquitin hydrolase that has been implicated in numerous cellular processes such as cell proliferation and DNA repair (Fig. 2) as well as chromatin level control of gene expression (Fig. 2; ref. 24). For example, *BAP1* has been shown to enhance progression through the G<sub>1</sub>-S checkpoint and subsequently induce cell death by a process with similarities to both apoptosis and necrosis (24). In addition, *BAP1* regulates cell proliferation by deubiquitinating host cell factor-1 (HCF1; ref. 25), a chromatin-associated protein believed to activate and repress transcription by linking appropriate histone-modifying enzymes to a subset of transcription factors, in particular of the E2F family (26). *BAP1* knockdown using siRNA in MPM cell lines inhibited cell growth and resulted in inactivation of HCF1 and downregulation of downstream E2F-responsive genes (10). It has also been shown that *BAP1* and HCF1 can form a ternary complex with the YY1 transcription factor to regulate gene expression (27).

Inactivating somatic mutations of *BAP1* have been identified in 47% of uveal melanomas, primarily in the metastasizing subset, where the *BAP1* mutation rate is close to 80% (28). The possible involvement of *BAP1* in modulating histone modifications prompted a study of histone deacetylase inhibitors in uveal melanoma cell lines, which found that these agents seemed to counteract the expression profile of *BAP1*-deficient uveal melanomas, but the growth inhibitory effects on the cell lines were independent of *BAP1* status (29). The relevance of these results to *BAP1*-mutated MPM is unclear. We have not seen a significant effect of *BAP1* knockdown on sensitivity of *BAP1* wild-type MPM lines to the histone deacetylase inhibitor suberoylanilide hydroxamic acid [SAHA (vorinostat); R. McMillan and M. Ladanyi; unpublished data]. Furthermore, a recently completed phase III trial of vorinostat in patients with advanced MPM observed few significant responses, well below what would be expected if *BAP1*-mutated cases were preferentially sensitive (30). The relationship of *BAP1*-related histone modifications to other previously reported epigenetic changes in MPM such as hypermethylation at gene promoters (e.g., *WIF1*, *SFRP4*, and others; refs. 31–33) remains to be defined.

### BAP1 Germline Mutations

Coming on the heels of the discovery of somatic *BAP1* mutations is the recent discovery of germline *BAP1* mutations in rare families predisposed to MPM. Although the majority of MPM cases are linked to asbestos exposure with a 20- to 40-year latency, only a minority of those exposed to asbestos eventually develop malignant disease (34). For example, in the United States, it is estimated that over 30 million workers were exposed to asbestos in the second half of the 20th century, but the incidence of MPM has remained relatively stable and low in the United States over the past 15 years at 1 to 15 per 100,000. Furthermore, numerous familial clusterings of MPM both with and without exposure to asbestos or erionite have been well described with up to 50% incidence of MPM within such families (35, 36). Taken together, these observations suggest that some individuals are more susceptible to the carcinogenic effects of asbestos and erionite and some are more susceptible to the development of MPM even in the absence of asbestos or erionite exposure.

Therefore, much work has focused on identifying genetic alterations that predispose individuals to MPM. Testa and colleagues carried out array-comparative genomic hybridization on 2 MPM tumors coming from 2 different familial clusters of MPM (37). Neither family had exposure to asbestos or erionite. Alterations were noted at or near the *BAP1* locus at 3p21.1. In addition, linkage studies were conducted on germline DNA from each family, yielding a maximum lod score of 2.1 at 3p21-22. Given these observations and reports of frequent gene loss at 3p21.1 in sporadic MPM (10), germline *BAP1* sequencing was conducted in both families and revealed concordance between mutation status and linkage analysis. Furthermore, those with



**Figure 2.** A, BAP1 functional domains and mapped interacting regions. BAP1 is a nuclear deubiquitinating enzyme (DUB). DUBs catalyze the removal of single ubiquitin moieties from ubiquitin chains or cleavage of the isopeptide bond between ubiquitin and the substrate protein (45). BAP1 is composed of an N-terminal UCH domain (blue; amino acid 1–250), an HCF1-binding domain (HBM)-like motif (amino acid 363–366), a motif that shares conservation with UCH37 (ULD: UCH37-like domain; green, amino acid 634–693), and a bipartite nuclear localization signal (NLS, amino acid 656–661, amino acid 717–722). BAP1 has been reported to interact with BARD1 (amino acid 182–365), HCF1 (amino acid 365–385), BRCA1 (amino acid 596–721), and YY1 (amino acid 642–686; ref. 26). B, BAP1 functions. Role of BAP1 in transcription factor regulation (left). HCF1 is detected as a major binding partner of BAP1 by mass spectrum analysis. HCF1 interacts with specific transcription factors, including OCT1, E2F1, Kox20, Sp1, and GA-binding protein. HCF1 also associates with several chromatin methyltransferases (Set1, MLL1, MLL5), chromatin acetyltransferases (tMOF), and deacetylases (HDAC1, HDAC2). HCF1 is known to recruit LSD1 to demethylate histone H4 lysine 9 and to promote the trimethylation of histone H3 lysine 4 (27, 46). YY1 is a transcriptional factor that binds to BAP1 and HCF1. BAP1 and HCF1 are recruited by YY1 to various promoters to upregulate gene expression (27). Role of BAP1 in chromatin modifications (middle). The trimethylation of lysine 27 of histone H3 (H3K27me3) is mediated by the histone methyltransferase EZH1/2, a component of the polycomb repressive complex 2 (PRC2) complex. This triggers recruitment of the PRC1 multiprotein complex through recognition of the H3K27me3 mark by the CBX subunit of the PRC1 complex. Another subunit of the PRC1 complex, the RING1 E3 ligase, then ubiquitylates lysine 119 of histone H2A (H2AK119ub1), which fixes chromatin in a repressed state, silencing gene expression. The H2AK119ub1 mark may be in a dynamic, continuously regulated state, as another part of the PRC1 complex, containing BAP1 and ASXL1 (47), and displays an opposing, deubiquitylating activity (PR-DUB). A balance between ubiquitination and deubiquitination may be needed for appropriate PRC target gene repression. Possible role of BAP1 in DNA repair (right). Other binding partners of BAP1 include BRCA1 and its partner BARD1, which have important roles in the double strand DNA repair process (48). Currently, the possible function of BAP1 in this process is not clear. The Rad51-dependent DNA repair pathway is highly regulated and includes many proteins, some of which may be potential substrates for BAP1-mediated ubiquitin hydrolysis.

germline mutations manifested other malignancies in addition to MPM, including uveal melanoma, breast cancer, renal cancer, and skin cancer. At the same time, Wiesner and colleagues reported an association between germline mutations in *BAP1* and familial melanocytic tumors ranging from epithelioid nevi to atypical melanocytic proliferations with features of cutaneous melanoma (38). Tumors were examined from 2 families and revealed biallelic somatic inactivation of *BAP1*. Each family had 1 person with uveal

melanoma, and 1 family had multiple members with cutaneous melanoma. Currently, with only a few published pedigrees, the exact spectrum of this newly described hereditary cancer predisposition syndrome remains unclear (39). In fact, it is possible that 2 distinct cancer syndromes exist, both associated with uveal melanoma but 1 dominated by melanocytic tumors and another by MPM (40). Njauw and colleagues (41) have recently published additional melanocytic tumor-rich pedigrees of families with

germline *BAP1* mutations. They have proposed for this phenotypic complex the acronym COMMON syndrome, which we would suggest to broaden as follows: cutaneous/ocular melanoma and atypical melanocytic proliferations, mesothelioma, and other internal neoplasms. Ultimately, the phenotype associated with germline *BAP1* mutations will likely be complex and depend on a variety of factors such as the tissue in which the second *BAP1* allele is inactivated, the mechanism of inactivation for the second *BAP1* allele, the functional consequences of a particular *BAP1* mutation, coexistent mutations, and environmental exposures.

### On the Horizon

The recently published data reviewed above suggest that, together, a majority of MPMs carry either *NF2* or *BAP1* mutations. Whereas *BAP1* encodes for a ubiquitin C-terminal hydrolase [deubiquitinase (DUB)] and is therefore able to reverse the ubiquitin linkages formed by E3 ubiquitin ligases, *NF2* encodes for a negative regulator of the E3 ubiquitin ligase CRL4<sup>DCAF1</sup>. DUBs are thought to function by binding to the substrate receptor components of specific, partner E3 ligases and by reversing the linkage that the latter form. Interestingly, total loss-of-function mutations in *NF2* and *BAP1*, as assessed by immunoblotting, occur in largely nonoverlapping subsets of patients with MPM [see immunoblotting of cell lines for Merlin (13) and for *BAP1* (10)]. This raises the hypothesis that CRL4<sup>DCAF1</sup> and *BAP1* regulate in an opposing fashion a common subset of substrates, which in turn causes a protumorigenic deregulation of gene expression in MPM. Alternatively, *NF2* and *BAP1* may suppress tumorigenesis by independent mechanisms. The therapeutic implications of distinguishing between these models will be significant.

If studies define a therapeutically accessible synthetic lethal target in the setting of *BAP1* loss, this could eventually benefit the approximately 40% of patients with MPM whose tumors have *BAP1* loss or mutation. More speculatively, the same synthetic lethal target could be studied as chemoprevention drug targets in individuals (or initially, mouse models) with germline *BAP1* mutations that predispose to MPM development. More fundamentally, the biologic insights into MPM pathogenesis emerging from further work on *BAP1* could lead to novel, biologically rational treatment strategies for MPM. Finally, the impact of the work may also extend to other cancers with *BAP1* mutations, such as melanoma and, as recently reported, clear cell renal cell carcinoma (42–44).

### Disclosure of Potential Conflicts of Interest

Lee M. Krug has commercial research support from Merck, Novartis, CanBas, and Lilly and is a consultant/advisory board member of Genentech, Morphotek, and MedImmune. No potential conflicts of interest were disclosed by the other authors.

### Authors' Contributions

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**Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis):** M. Ladanyi, M. G. Zauderer, L.M. Krug, M. Bott

**Writing, review, and/or revision of the manuscript:** M. Ladanyi, M. G. Zauderer, L.M. Krug, T. Ito, F. G. Giancotti

**Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases):** T. Ito, R. McMillan, M. Bott

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### References

- Moolgavkar SH, Meza R, Turim J. Pleural and peritoneal mesotheliomas in SEER: age effects and temporal trends, 1973–2005. *Cancer Causes Control* 2009;20:935–44.
- Krug LM. An overview of chemotherapy for mesothelioma. *Hematol Oncol Clin North Am* 2005;19:1117–36.
- Zauderer MG, Krug LM. Novel therapies in phase II and III trials for malignant pleural mesothelioma. *J Natl Compr Canc Netw* 2012;10:42–7.
- Bianchi AB, Mitsunaga SI, Cheng JQ, Klein WM, Jhanwar SC, Seizinger B, et al. High frequency of inactivating mutations in the neurofibromatosis type 2 gene (*NF2*) in primary malignant mesotheliomas. *Proc Natl Acad Sci U S A* 1995;92:10854–8.
- Sekido Y, Pass HI, Bader S, Mew DJ, Christman MF, Gazdar AF, et al. Neurofibromatosis type 2 (*NF2*) gene is somatically mutated in mesothelioma but not in lung cancer. *Cancer Res* 1995;55:1227–31.
- McClatchey AI, Fehon RG. Merlin and the ERM proteins—regulators of receptor distribution and signaling at the cell cortex. *Trends Cell Biol* 2009;19:198–206.
- Okada T, Lopez-Lago M, Giancotti FG. Merlin/NF-2 mediates contact inhibition of growth by suppressing recruitment of Rac to the plasma membrane. *J Cell Biol* 2005;171:361–71.
- Li W, You L, Cooper J, Schiavon G, Pepe-Caprio A, Zhou L, et al. Merlin/NF2 suppresses tumorigenesis by inhibiting the E3 ubiquitin ligase CRL4(DCAF1) in the nucleus. *Cell* 2010;140:477–90.
- Li W, Cooper J, Karajannis MA, Giancotti FG. Merlin: a tumour suppressor with functions at the cell cortex and in the nucleus. *EMBO Rep* 2012;13:204–15.
- Bott M, Brevet M, Taylor BS, Shimizu S, Ito T, Wang L, et al. The nuclear deubiquitinase *BAP1* is commonly inactivated by somatic mutations and 3p21.1 losses in malignant pleural mesothelioma. *Nat Genet* 2011;43:668–72.
- Murakami H, Mizuno T, Taniguchi T, Fujii M, Ishiguro F, Fukui T, et al. *LATS2* is a tumor suppressor gene of malignant mesothelioma. *Cancer Res* 2011;71:873–83.
- Sekido Y. Inactivation of Merlin in malignant mesothelioma cells and the Hippo signaling cascade dysregulation. *Pathol Int* 2011;61:331–44.
- Lopez-Lago MA, Okada T, Murillo MM, Socci N, Giancotti FG. Loss of the tumor suppressor gene *NF2*, encoding merlin, constitutively activates integrin-dependent mTORC1 signaling. *Mol Cell Biol* 2009;29:4235–49.
- James MF, Han S, Polizzano C, Plotkin SR, Manning BD, Stemmer-Rachamimov AO, et al. *NF2/merlin* is a novel negative regulator of mTOR complex 1, and activation of mTORC1 is associated with

- meningioma and schwannoma growth. *Mol Cell Biol* 2009;29:4250–61.
15. Brevet M, Bott M, Zhou Q, Rusch V, Ladanyi M. Association of NF2 loss and mTOR pathway activation in pleural mesothelioma: Comprehensive genetic analysis of NF2 and immunohistochemistry in 13 cell lines and 53 tumors. *Mod Pathol* 2011;24 (suppl1):406A.
  16. Brevet M, Shimizu S, Bott MJ, Shukla N, Zhou Q, Olshen AB, et al. Coactivation of receptor tyrosine kinases in malignant mesothelioma as a rationale for combination targeted therapy. *J Thorac Oncol* 2011;6:864–74.
  17. Carracedo A, Ma L, Teruya-Feldstein J, Rojo F, Salmena L, Alimonti A, et al. Inhibition of mTORC1 leads to MAPK pathway activation through a PI3K-dependent feedback loop in human cancer. *J Clin Invest* 2008;118:3065–74.
  18. Varghese S, Chen Z, Bartlett DL, Pingpank JF, Libutti SK, Steinberg SM, et al. Activation of the phosphoinositide-3-kinase and mammalian target of rapamycin signaling pathways are associated with shortened survival in patients with malignant peritoneal mesothelioma. *Cancer* 2011;117:361–71.
  19. Wagner AJ, Bendell JC, Dolly S, Morgan JA, Ware JA, Fredrickson J, et al. A first-in-human phase I study to evaluate GDC-0980, an oral PI3K/mTOR inhibitor, administered QD in patients with advanced solid tumors. *J Clin Oncol* 2011;29:3020.
  20. Illei PB, Rusch VW, Zakowski MF, Ladanyi M. Homozygous deletion of CDKN2A and codeletion of the methylthioadenosine phosphorylase gene in the majority of pleural mesotheliomas. *Clin Cancer Res* 2003;9:2108–13.
  21. Cheng JQ, Jhanwar SC, Klein WM, Bell DW, Lee WC, Altomare DA, et al. p16 alterations and deletion mapping of 9p21-p22 in malignant mesothelioma. *Cancer Res* 1994;54:5547–51.
  22. Xio S, Li D, Vijg J, Sugarbaker DJ, Corson JM, Fletcher JA. Codeletion of p15 and p16 in primary malignant mesothelioma. *Oncogene* 1995;11:511–5.
  23. Yoshikawa Y, Sato A, Tsujimura T, Emi M, Morinaga T, Fukuoka K, et al. Frequent inactivation of the BAP1 gene in epithelioid-type malignant mesothelioma. *Cancer Sci* 2012;103:868–74.
  24. Ventii KH, Devi NS, Friedrich KL, Chernova TA, Tighiouart M, Van Meir EG, et al. BRCA1-associated protein-1 is a tumor suppressor that requires deubiquitinating activity and nuclear localization. *Cancer Res* 2008;68:6953–62.
  25. Machida YJ, Machida Y, Vashisht AA, Wohlschlegel JA, Dutta A. The deubiquitinating enzyme BAP1 regulates cell growth via interaction with HCF-1. *J Biol Chem* 2009;284:34179–88.
  26. Eletr ZM, Wilkinson KD. An emerging model for BAP1's role in regulating cell cycle progression. *Cell Biochem Biophys* 2011;60:3–11.
  27. Yu H, Mashtalir N, Daou S, Hammond-Martel I, Ross J, Sui G, et al. The ubiquitin carboxyl hydrolase BAP1 forms a ternary complex with YY1 and HCF-1 and is a critical regulator of gene expression. *Mol Cell Biol* 2010;30:5071–85.
  28. Harbour JW, Onken MD, Roberson ED, Duan S, Cao L, Worley LA, et al. Frequent mutation of BAP1 in metastasizing uveal melanomas. *Science* 2010;330:1410–3.
  29. Landreville S, Agapova OA, Matatal KA, Kneass ZT, Onken MD, Lee RS, et al. Histone deacetylase inhibitors induce growth arrest and differentiation in uveal melanoma. *Clin Cancer Res* 2012;18:408–16.
  30. Krug LM, Kindler H, Calvert H, Manegold C, Tsao AS, Fennell D, et al. VANTAGE 014: Vorinostat (V) in patients with advanced malignant pleural mesothelioma (MPM) who have failed prior pemetrexed and either cisplatin or carboplatin therapy: a phase III, randomized, double-blind, placebo-controlled trial. *Eur J Cancer* 2011;47:2–3.
  31. He B, Lee AY, Dadfarmay S, You L, Xu Z, Reguart N, et al. Secreted frizzled-related protein 4 is silenced by hypermethylation and induces apoptosis in beta-catenin-deficient human mesothelioma cells. *Cancer Res* 2005;65:743–8.
  32. Batra S, Shi Y, Kuchenbecker KM, He B, Reguart N, Mikami I, et al. Wnt inhibitory factor-1, a Wnt antagonist, is silenced by promoter hypermethylation in malignant pleural mesothelioma. *Biochem Biophys Res Commun* 2006;342:1228–32.
  33. Kohno H, Amatya VJ, Takeshima Y, Kushitani K, Hattori N, Kohno N, et al. Aberrant promoter methylation of WIF-1 and SFRP1, 2, 4 genes in mesothelioma. *Oncol Rep* 2010;24:423–31.
  34. Carbone M, Ly BH, Dodson RF, Pagano I, Morris PT, Dogan UA, et al. Malignant mesothelioma: facts, myths and hypotheses. *J Cell Physiol* 2012;227:44–58.
  35. Roushdy-Hammady I, Siegel J, Emri S, Testa JR, Carbone M. Genetic-susceptibility factor and malignant mesothelioma in the Cappadocian region of Turkey. *Lancet* 2001;357:444–5.
  36. Dogan AU, Baris YI, Dogan M, Emri S, Steele I, Elmishad AG, et al. Genetic predisposition to fiber carcinogenesis causes a mesothelioma epidemic in Turkey. *Cancer Res* 2006;66:5063–8.
  37. Testa JR, Cheung M, Pei J, Below JE, Tan Y, Sementino E, et al. Germline BAP1 mutations predispose to malignant mesothelioma. *Nat Genet* 2011;43:1022–5.
  38. Wiesner T, Obenauf AC, Murali R, Fried I, Griewank KG, Ulz P, et al. Germline mutations in BAP1 predispose to melanocytic tumors. *Nat Genet* 2011;43:1018–21.
  39. Abdel-Rahman MH, Pilarski R, Cebulla CM, Massengill JB, Christopher BN, Boru G, et al. Germline BAP1 mutation predisposes to uveal melanoma, lung adenocarcinoma, meningioma, and other cancers. *J Med Genet* 2011;48:856–9.
  40. Goldstein AM. Germline BAP1 mutations and tumor susceptibility. *Nat Genet* 2011;43:925–6.
  41. Njauw CN, Kim I, Piris A, Gabree M, Taylor M, Lane AM, et al. Germline BAP1 inactivation is preferentially associated with metastatic ocular melanoma and cutaneous-ocular melanoma families. *PLoS One* 2012;7:e35295.
  42. Guo G, Gui Y, Gao S, Tang A, Hu X, Huang Y, et al. Frequent mutations of genes encoding ubiquitin-mediated proteolysis pathway components in clear cell renal cell carcinoma. *Nat Genet* 2012;44:17–9.
  43. Duns G, Hofstra RM, Sietzema JG, Hollema H, van Duivenbode I, Kuik A, et al. Targeted exome sequencing in clear cell renal cell carcinoma tumors suggests aberrant chromatin regulation as a crucial step in ccRCC development. *Hum Mutat* 2012;33:1059–62.
  44. Pena-Llopis S, Vega-Rubin-de-Celis S, Liao A, Leng N, Pavia-Jimenez A, Wang S, et al. BAP1 loss defines a new class of renal cell carcinoma. *Nat Genet* 2012;44:751–9.
  45. Nishikawa H, Wu W, Koike A, Kojima R, Gomi H, Fukuda M, et al. BRCA1-associated protein 1 interferes with BRCA1/BARD1 RING heterodimer activity. *Cancer Res* 2009;69:111–9.
  46. Kristie TM, Liang Y, Vogel JL. Control of alpha-herpesvirus IE gene expression by HCF-1 coupled chromatin modification activities. *Biochim Biophys Acta* 2010;1799:257–65.
  47. Scheuermann JC, de Ayala Alonso AG, Oktaba K, Ly-Hartig N, McGinty RK, Fraterman S, et al. Histone H2A deubiquitinase activity of the Polycomb repressive complex PR-DUB. *Nature* 2010;465:243–7.
  48. Jensen DE, Proctor M, Marquis ST, Gardner HP, Ha SI, Chodosh LA, et al. BAP1: a novel ubiquitin hydrolase which binds to the BRCA1 RING finger and enhances BRCA1-mediated cell growth suppression. *Oncogene* 1998;16:1097–112.

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