

ColoGuidePro: A Prognostic 7-Gene Expression Signature for Stage III Colorectal Cancer Patients

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Abstract

Purpose: Improved prognostic stratification of patients with stage II and III colorectal cancer is warranted for postoperative clinical decision making. This study was conducted to develop a clinically feasible and robust prognostic classifier for these patients independent of adjuvant treatment.

Experimental Design: Global gene expression profiles from altogether 387 stage II and III colorectal cancer tissue samples from three independent patient series were included in the study. ColoGuidePro, a seven-gene prognostic classifier, was developed from a selected Norwegian learning series ($n = 95$; no adjuvant treatment) using lasso-penalized multivariate survival modeling with cross-validation.

Results: The expression signature significantly stratified patients in a consecutive Norwegian test series, in which patients were treated according to current standards [HR, 2.9 (1.1–7.5); $P = 0.03$; $n = 77$] and an external validation series [HR, 3.7 (2.0–6.8); $P < 0.001$; $n = 215$] according to survival. ColoGuidePro was also an independent predictor of prognosis in multivariate models including tumor stage in both series (HR, ≥ 3.1 ; $P \leq 0.03$). In the validation series, which consisted of patients from other populations (United States and Australia), 5-year relapse-free survival was significantly predicted for stage III patients only ($P < 0.001$; $n = 107$). Here, prognostic stratification was independent of adjuvant treatment ($P = 0.001$).

Conclusions: We present ColoGuidePro, a prognostic classifier developed for patients with stage II and III colorectal cancer. The test is suitable for transfer to clinical use and has best prognostic prediction potential for stage III patients. *Clin Cancer Res*; 18(21); 6001–10. ©2012 AACR.

Introduction

Colorectal cancer is the third most common type of cancer with a worldwide annual incidence of 1.2 million and mortality rate of approximately 50% (1). The only available curative treatment for colorectal cancer is complete surgical resection of neoplastic tissue (2). Determination of the extent of the disease by clinicopathologic tumor staging is the primary prognostic factor for patients with colorectal cancer (2). Despite the favorable outcome for patients with localized stage II tumors compared with stage III tumors, more than 20% of stage II patients suffer from recurrence (3).

Still, investigations of benefit from adjuvant chemotherapy for stage II patients show conflicting results (4–6), and surgery remains the only recommended treatment modality (7). For patients with stage III disease, large clinical trials have consistently showed improved survival with administration of adjuvant chemotherapy and this constitutes the standard of care for this group of patients (8). However, the significantly poorer survival among patients with stage IIB (T-stage 4, lymph node–negative) than with stage IIIA (T-stage 1-2, lymph node–positive) disease underlines the need for refinements to this prognostic stratification (9).

Accordingly, identification of individual patients in need of adjuvant treatment primarily by optimized prediction of prognosis for stage II and III patients remains a major clinical concern. There are currently no markers in routine clinical use for this purpose (10). Hence, identification of molecular markers for improved prognostic stratification represents a valuable step toward beneficial personalized management of patients with stage II or III disease.

Prognostic gene expression signatures have shown predictive value in cancer management, primarily for patients with breast cancer (11–13). Also for colorectal cancer, several studies have reported prognostic gene expression signatures, focusing primarily on stage II and III tumors (14–18). However, these studies have generally been limited by small sample sizes and/or lack of testing in independent sample series (19). More recently, larger studies have shown

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Translational Relevance

Colorectal cancer is a common disease with a high mortality rate. Although molecular markers have great potential for prediction of disease outcome, and thereby for guiding choice of adjuvant chemotherapy, postoperative treatment decisions for stage II and III patients are still based on clinicopathologic parameters alone. Here, we present ColoGuidePro, a prognostic test for stage II and III colorectal cancer that is independent of tumor stage in multivariate models and holds significant prognostic value for stage III patients separately. The classifier is based on the expression of only seven genes, all carrying independent prognostic information. This, and simple test result interpretations, ensures suitability for clinical use.

Stage III patients routinely receive adjuvant treatment. Accordingly, analysis of treatment-independent prognosis for these patients requires a careful study design. ColoGuidePro was developed from a historical, non-treated patient series. Treatment-independent prognostic value was shown in patient series treated according to current standards. This has great clinical implications for patients whose general health conditions cause reduced eligibility for chemotherapy.

potential prognostic predictive value for gene expression signatures of varying sizes (20–23). However, no studies have considered the abundance of prognostic information contained within the proposed signatures, possibly resulting in unnecessary large signatures and reduced robustness due to increased risk of overfitting. Furthermore, testing of independent prediction potential has generally been limited to one patient series.

In this study, we aimed to develop a nonredundant prognostic gene signature for stage II and III colorectal cancer based on the expression of only few genes using variable selection by L_1 (lasso)-penalization and cross-validation in the Cox proportional hazards model (24). This statistical model has not previously been used for the purpose of subclassification of colorectal cancer. The study was designed to clinically validate the new classifier in independent patient series across populations and technology platforms and to provide prognostic stratification independent of adjuvant treatment in stage III patients.

Materials and Methods

Material

Altogether, 172 fresh-frozen stage II and III colorectal cancer tissue samples from 2 independent Norwegian patient series were analyzed in the study. The learning series included 95 samples taken from patients treated surgically at different hospitals in the Oslo region before adjuvant chemotherapy becoming standard treatment for stage III patients to allow for treatment-independent prognostic prediction (Table 1). These patients were selected to include

approximately equal numbers of stage II and III tumors, as well as equal numbers of survival events between the stages, again to achieve independent information within each stage. Selection was also based on long-term follow-up among survivors (>10 years). The independent test series was consecutively collected (95% inclusion rate) and consisted of 77 patients treated by curative resection at one Norwegian hospital, Aker University Hospital, Oslo, Norway (Table 1). These patients received adjuvant treatment according to the current standard, that is, routine administration of chemotherapy for patients presenting with stage III tumors. RNA was extracted from all samples using the Qiagen AllPrep DNA/RNA Mini Kit (Qiagen). Microsatellite instability status of the clinical specimens has previously been determined (25, 26).

The research conformed to the Helsinki Declaration, and the research biobanks have been registered according to national legislation (numbers 2781 and 236-2005-16141). The study (amendment number 2010/1805) is part of a project approved by the Regional Committee for Medical and Health Research Ethics (numbers 1.2005.1629 and S-09282c 2009/4958), which requires that informed consent is obtained from patients being enrolled in the study.

In addition, publically available gene expression data from 2 independent series of altogether 215 patients with stage II and III colorectal cancer were accessed from NCBI's Gene Expression Omnibus (GEO), accession numbers GSE14333 and GSE17538. There was extensive overlap between samples in the 2 series [$n = 97$ stage II and III samples from the H. Lee Moffitt Cancer Center (Tampa, FL), as found by correlation analyses of the probe cell intensity (CEL) files]. Only unique samples ($n = 215$) were included in the current study and are herein referred to as the validation series. Clinical information for the patients was obtained from the respective GEO entries (Table 1). There were no significant differences in clinicopathologic characteristics between the test and validation series, which are the 2 patient series used to assess the prognostic test developed in the current study, except for patient age (mean difference, 6.9 years, $P < 0.001$; independent samples t test) and the distribution of tumor location ($P = 0.05$; χ^2 test).

Details for experimental performance of exon microarray analyses and preprocessing of gene expression data can be found in the Supplementary Material. Parts of the microarray data have previously been published (27) and can be accessed from GEO (GSE30378 and GSE24550). For the current study, 12 additional samples were included in the learning series (GSM753769-GSM753780) and are amended to the GSE30378 record.

Development of the prognostic gene expression signature

A gene expression signature for prediction of prognosis was developed from the learning series of 95 colorectal cancer samples. The gene expression data set was filtered (Supplementary Methods) to include only genes with variances in expression levels higher than 0.2, and P values

Table 1. Characteristics of the three colorectal cancer sample series

Characteristic	Learning series (n = 95)	Test series (n = 77)	Validation series ^a (n = 215)
Age at diagnosis (mean ± SD)	66 ± 11.7	73 ± 13.5	66 ± 13.3
Gender			
Male	46	33	115
Female	49	44	100
Stage			
II	52	44	108
III	43	33	107
Location			
Right	27	46	85
Left	31	20	77
Rectum	37	11	22
Unknown			31
Mean follow-up, y (min–max)			
All patients	5.9 (0.3–10) ^b	3.3 (0.2–5) ^c	3.1 (0.04–5.0) ^c
Patients with event	1.8 (0.3–7.7) ^b	2.1 (0.7–3.8) ^c	1.5 (0.1–3.4) ^c
Patients with no event	10 (10–10) ^b	3.7 (0.2–5.0) ^c	3.5 (0.04–5.0) ^c
No. of events, stage II ^d	21	9	13
No. of events, stage III ^d	27	10	32
MSI-high	7	14	NA
Adjuvant chemotherapy, stage III			
Yes	0	17	63
No	43	16	28
Unknown	0	0	16
Year of surgery/biobanking	1987–1989	2005–2007	NA
ColoGuidePro			
Positive predictions (true; false)	20; 2	6; 7	16; 18
Negative predictions (true; false)	45; 28	51; 13	152; 29

Abbreviations: MSI, microsatellite instability; NA, not available.

^aGEO accession numbers GSE14333 and GSE17538. Only nonoverlapping samples from stage II and III patients were included

^bTen-year follow-up.

^cFive-year follow-up.

^dRelapse or death from colorectal cancer.

(Wald test of predictive potential) from univariate Cox proportional hazards analysis lower than 0.5 ($n = 3,098$ genes). This gene set was subjected to penalized multivariate Cox proportional hazards survival modeling using an algorithm for variable selection based on L_1 -penalized (lasso) estimation (Supplementary Methods; ref. 24). Using this model, the genes in the prognostic expression signature were selected via cross-validation in the learning series. A penalty parameter, λ_1 , reflecting the predictive potential and calculated by cross-validation, was inflicted upon the gene expression signals during survival modeling.

Genes in the resulting prognostic expression signature were considered to be associated with poor patient survival at expression levels above the 80th percentile across the data set for genes with univariate HR > 1 and below the 20th percentile for genes with univariate HR < 1. To obtain a simple classification rule, patients were stratified into prognostic groups according to the number of genes in the

prognostic signature being expressed at levels associated with poor prognosis. For comparison, sample-wise prognostic indices (PI) were also calculated on the basis of expression values and lasso-penalized multivariate regression coefficients for genes in the signature and used for prognostic stratification (Supplementary Methods).

Further statistical analyses were done using the SPSS 16.0 software (SPSS Inc.). These include independent samples t tests, χ^2 tests, standard univariate and multivariate Cox proportional hazards analyses [estimation of HR and corresponding 95% confidence intervals (CI)], Wald test for predictive potential, generation of Kaplan–Meier survival plots, and Pearson correlation analysis. Two-sided $P \leq 0.05$ was considered significant. For survival analyses, the endpoint was relapse-free survival. Relapse or death from colorectal cancer was regarded as events, and patients with no events were censored. Generation of the correlation heatmap was done using J-Express 2011 (MolMine AS).

Results

Development of the prognostic gene expression signature

Lasso-penalized multivariate Cox proportional hazards modeling was conducted on a filtered genome-wide expression data set obtained from a learning series of 95 stage II and III colorectal cancer (Supplementary Methods). There were several survival models accommodating optimal penalty conditions from cross-validation (Supplementary Fig. S1 and Supplementary Methods). Across 1,000 iterations, 7 different gene expression signatures accommodated optimal survival prediction in the learning series more than 50 times each (size range, 1–12 genes; Fig. 1). These different

signatures expanded on the same sets of genes, and the smaller signatures were subsets of the larger. For all signatures (except the 1-gene signature), there were significant associations between patient survival and increasing numbers of genes expressed at levels associated with poor survival (HRs, 1.6–1.9; $P < 0.04$).

Dichotomization of patients to good and poor prognosis groups was tested for all possible stepwise increases in amounts of genes with associations to poor survival within each signature (Fig. 1). For the 28 possible poor prognosis groups, 22 (79%) had significant associations with poor patient survival (univariate Cox proportional hazards analyses, HR, 3.0–11.5; $P < 0.04$). To further assess which of the

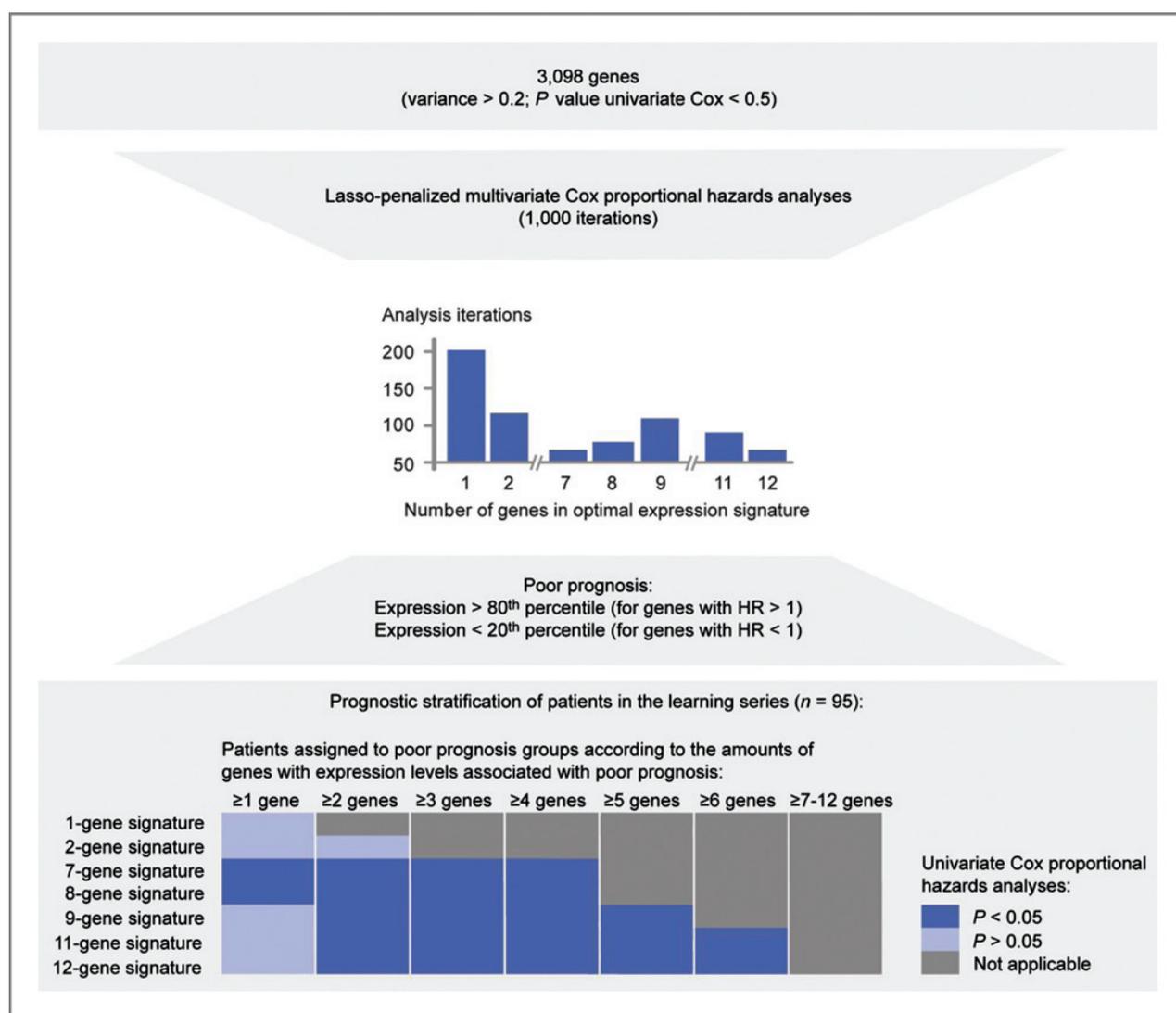


Figure 1. Workflow for development of a prognostic gene expression signature. From the learning series of 95 stage II and III colorectal cancer samples, a filtered gene expression data set was used as input for survival modeling. From 1,000 iterations of lasso-penalized multivariate modeling, 7 models were reported as optimal for survival prediction more than 50 times (middle bar plot). For each of the gene expression signatures, patients were dichotomized to good and poor prognosis groups according to all the possible stepwise increases in amounts of genes being expressed at levels associated with poor prognosis (bottom). All 28 possible stratifications (pale and dark blue boxes in heatmap) were tested for univariate associations with patient survival, yielding significant associations for 22 (dark blue).

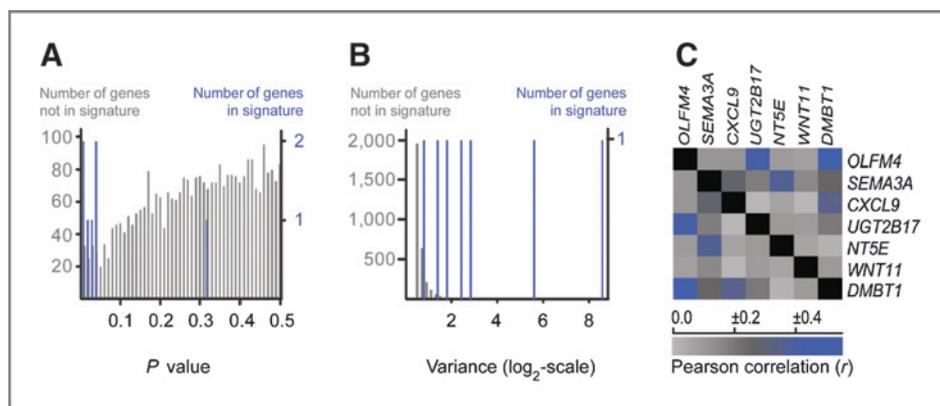


Figure 2. Statistical characteristics of genes in the identified prognostic expression signature. The 7 genes in the expression signature (blue) had (A) low P values from univariate Cox proportional hazards analyses (median $P = 0.02$, Wald test for predictive potential) and (B) high variances in gene expression signals (median 2.4, \log_2 scale), compared with the remaining 3,091 genes included for survival modeling (gray; median $P = 0.3$; median variance 0.3). C, the Pearson correlations of expression signals between the 7 genes in the signature were generally weak (absolute values, 0.006–0.55).

7 signatures had the best predictive potential in independent samples, the signatures were tested also in the test series. Here, 5 of the dichotomizing stratification rules resulted in significant prognostic stratification (univariate Cox proportional hazards analyses; HRs, 2.9–5.8; $P < 0.04$). The best conducting stratification rule across both the learning and test series (by rank of P values), assigned patients to a poor prognosis group when expressing (any) 3 or more genes in the 7-gene signature at levels associated with poor prognosis. This constitutes the ColoGuidePro prognostic test. The genes included in the signature were *OLFM4*, *CXCL9*, *DMBT1*, *UGT2B17*, *SEMA3A*, *NT5E*, and *WNT11* (further described in Supplementary Table S1 and Supplementary Fig. S2). In accordance with the initial selection criteria for genes entered into the lasso algorithm, as well as for the algorithm itself, the 7 genes had strong univariate associations with patient survival in the learning series, as well as large variation and low correlation in gene expression (Fig. 2).

Assessment of the prognostic gene expression signature

In addition to the learning and test series ($n = 95$ and 77 , respectively), the performance of the 7-gene expression signature was assessed on an external validation series of patients with stage II and III colorectal cancer ($n = 215$). In all 3 series, the sample-wise increase in amounts of genes with expression levels indicating poor survival (ranging from 0 to 5 of 7 genes) was associated with increasingly poor patient survival (HR ≥ 1.5 , $P \leq 0.04$, Wald test for predictive potential; Supplementary Methods).

According to the ColoGuidePro expression signature, all patients were significantly stratified to a poor and good survival group in all 3 series (Fig. 3). In the learning series, 23% of the patients (22 of 95) were assigned to the poor prognosis group (univariate: HR, 4.0; 95% CI, 2.2–7.2; $P < 0.001$, Wald test of predictive potential). In the test and validation series, 17% and 16% of the patients were assigned to the poor prognosis group (13 of 77 patients

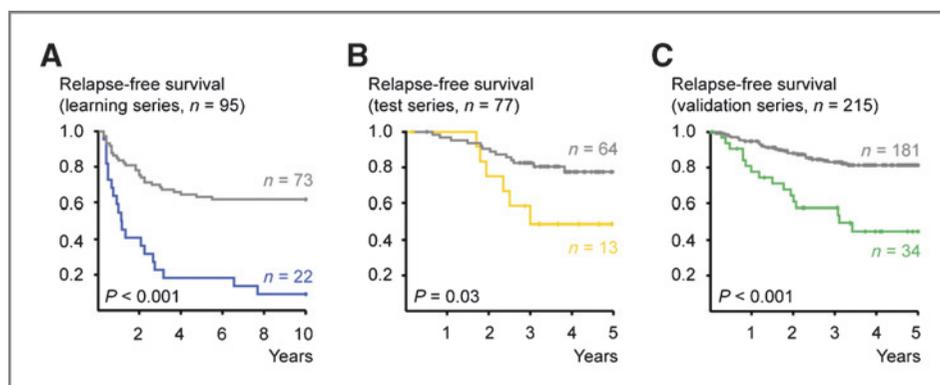


Figure 3. Survival curves for patients with stage II and III colorectal cancer in the 3 independent series stratified by the 7-gene expression signature. A, patients in the learning series assigned to the poor prognosis group had a 10-year relapse-free survival rate of 9%, significantly poorer than the 62% survival rate for patients in the good prognosis group. The corresponding 5-year survival rates for patients in (B) the test series and (C) external validation series, were 49% compared with 78% and 45% compared with 81%, respectively. All survival curves were calculated by Kaplan–Meier statistics and compared by log-rank tests. Relapse or death from colorectal cancer within the 10 or 5 years of follow-up was regarded events, and patients with no events within the indicated period of follow-up were censored.

Table 2. Prognostic stratification of stage II and III colorectal cancer patients by clinical parameters and the 7-gene expression classifier ColoGuidePro

Parameter	Test series				Validation series ^a			
	Univariate		Multivariate		Univariate		Multivariate	
	HR ^b (95% CI)	P ^c	HR ^b (95% CI)	P ^c	HR ^b (95% CI)	P ^c	HR ^b (95% CI)	P ^c
Gender (male vs. female)	1.3 (0.5–3.3)	0.5	1.3 (0.5–3.5)	0.6	1.1 (0.6–2.0)	0.8	0.9 (0.5–1.7)	0.8
Age at diagnosis (≥ 70 vs. < 70 y)	0.8 (0.3–1.9)	0.6	0.6 (0.2–2.1)	0.5	0.8 (0.4–1.4)	0.4	1.1 (0.6–2.1)	0.8
Tumor stage (III vs. II)	1.8 (0.7–4.3)	0.2	1.3 (0.4–4.1)	0.7	2.7 (1.4–5.2)	0.002	2.6 (1.3–5.4)	0.009
Tumor location (right vs. left and rectum)	0.3 (0.1–0.9)	0.03	0.4 (0.2–1.2)	0.1	0.8 (0.4–1.4)	0.4	0.8 (0.4–1.4)	0.4
MSI (MSI-high vs. MSI-low and MSS)	0.2 (0.03–1.4)	0.1	0.4 (0.04–3.1)	0.4	NA	NA	NA	NA
Adjuvant chemotherapy (yes vs. no)	1.1 (0.4–3.0)	0.9	0.6 (0.1–2.6)	0.5	1.5 (0.8–2.7)	0.2	0.9 (0.5–1.8)	0.8
ColoGuidePro	2.9 (1.1–7.5)	0.03	3.2 (1.1–9.3)	0.03	3.7 (2.0–6.8)	<0.001	3.1 (1.6–5.8)	0.001

Abbreviations: MSI, microsatellite instability; MSS, microsatellite-stable; NA, not available.

^aGEO accession numbers GSE14333 and GSE17538 ($n = 215$).

^bHRs and corresponding 95% CIs from univariate or multivariate Cox proportional hazards analysis as indicated. Event is relapse or death from colorectal cancer within 5 years. Censoring is no event or lost to follow-up within 5 years.

^cP values from Wald test of predictive potential.

and 34 of 215 patients), respectively. Poor prognosis patients in the test series had a 5-year relapse-free survival rate of 49%, compared with 78% for patients in the good prognosis group (univariate: HR, 2.9; 95% CI, 1.1–7.5; $P = 0.03$). The corresponding survival rates in the external validation series were 45% and 81%, respectively (univariate: HR, 3.7; 95% CI, 2.0–6.8; $P < 0.001$).

The validity of the stratification rule used for ColoGuidePro, that is, assigning patients to a poor prognosis group when expressing 3 or more of the 7 genes at levels associated with poor survival, was supported by evaluating the performance of the sample-wise PIs based on expression values and lasso-penalized multivariate regression coefficients for the 7 genes (Supplementary Methods). The 20% of the patients with highest PIs had markedly poorer survival than the rest of the patients. The univariate HRs for the learning series, test series, and validation series were 4.1 (2.2–7.5), 2.1 (0.8–5.5), and 2.6 (1.4–4.7), respectively ($P \leq 0.001$, ≤ 0.1 , and ≤ 0.003 , respectively).

The gene expression signature was also an independent predictor of poor patient survival in multivariate models including tumor stage and other clinicopathologic parameters. Multivariate HRs were 4.1 (2.2–7.7), 3.2 (1.1–9.3), and 3.1 (1.6–5.8), in the learning series (Supplementary Table S2), test series, and validation series (Table 2), respectively ($P \leq 0.001$, ≤ 0.03 , and ≤ 0.001 , respectively).

By analysis of patients within each tumor stage separately, prognostic stratification according to ColoGuidePro was positively validated only for stage III patients (Fig. 4). In the external validation series, 21% (23 of 107) of stage III patients were assigned to the poor prognosis group. These patients had a 5-year relapse-free survival rate of 29%, compared with 76% for patients with good prognosis [univariate: HR, 4.1 (2.0–8.2), $P < 0.001$, Wald test for predictive potential]. This survival difference was independent of the administration of adjuvant chemotherapy

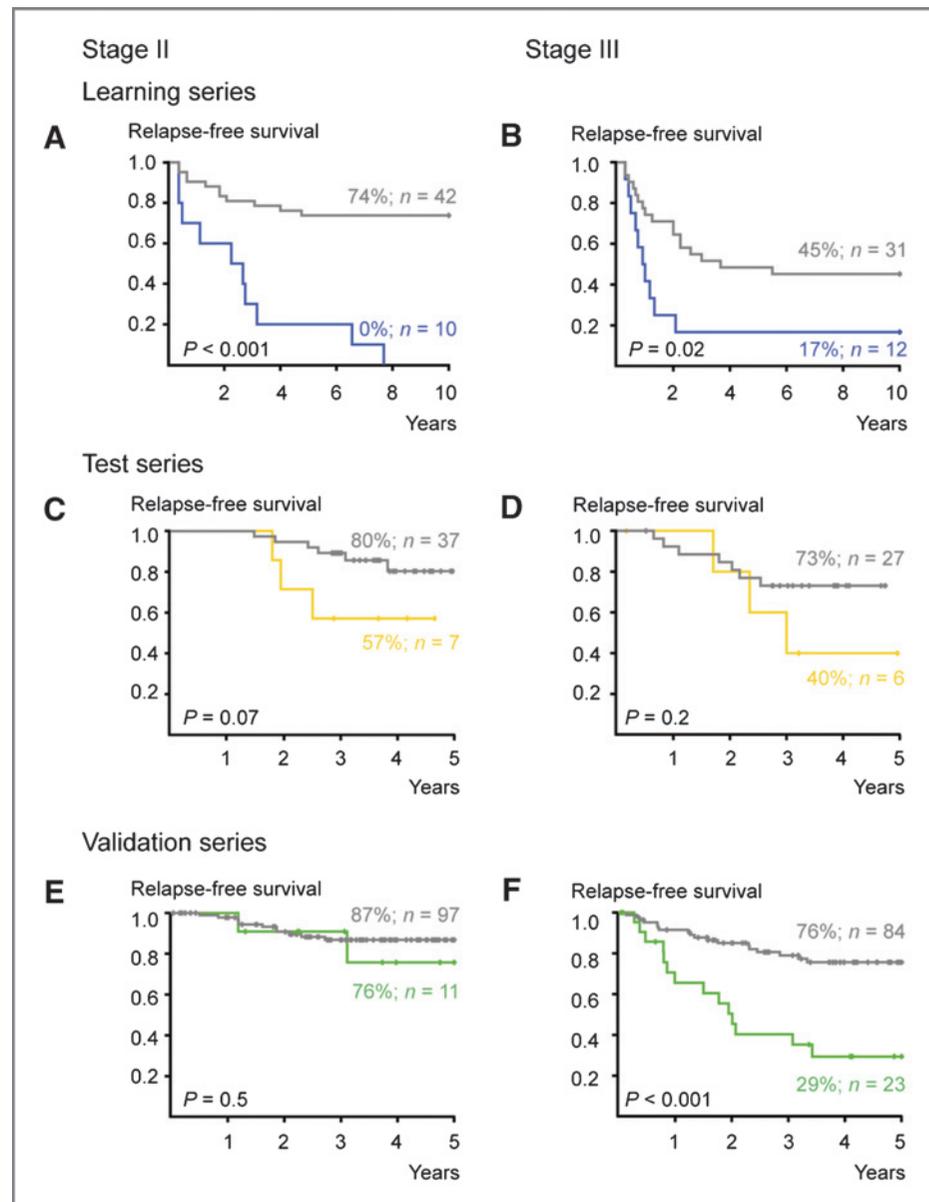
[multivariate: HR, 3.4 (1.7–7.0), $P = 0.001$; Supplementary Fig. S3]. Importantly, also stage III patients above the age of 75 years ($n = 26$ in the validation series), who are less eligible for chemotherapeutic treatment, were correctly classified according to survival by the expression signature [survival rates were 70% and 0% for the good and poor prognosis group, respectively, $P < 0.001$; HR, 13.5 (2.5–72.5); Supplementary Fig. S4]. This difference was independent of adjuvant treatment [multivariate: HR, 17.1 (2.9–102.2), $P = 0.002$]. For stage II patients in the validation series, only 10% (11 of 108) were predicted to have poor survival, and prognostic stratification of these patients was not significant [univariate: HR, 1.6 (0.3–6.9), $P = 0.5$].

Discussion

In this study, we have developed and validated ColoGuidePro, a 7-gene expression signature for prediction of prognosis for patients with stage II and III colorectal cancer. This classifier is a significant predictor of poor prognosis, both in the learning series, test series, and an external validation series of patients from different countries and analyzed on different microarray platforms, with best prognostic prediction potential for stage III patients.

Prognostic value was shown in a large number of patients (totally 387 stage II and III patients). To ensure independent prediction value additional to what can be achieved by cross-validation in the learning series alone, selection of the presented 7-gene signature from the 7 different signatures most commonly presented as optimal in the learning series was aided by testing of independent samples in the test series. Accordingly, the test series is not truly independent of the model building. However, prognostic value of ColoGuidePro was shown in an additional large and independent validation series ($n = 215$).

Figure 4. Survival curves in each stage for patients in the 3 independent series. Patients with stage II (left) and III (right) colorectal cancer were individually stratified according to the 7-gene expression signature. In the learning series, (A) 19% of stage II patients [HR, 6.6 (2.7–16.1)] and (B) 28% of stage III patients [HR, 2.6 (1.2–5.7)] were assigned to the poor prognosis group. In the test series, (C) 16% of stage II patients [HR, 3.3 (0.8–13.3)] and (D) 18% of stage III patients [HR, 2.3 (0.6–8.8)] were assigned to the poor prognosis group. In the external validation series, only (E) 10% of stage II patients were assigned to the poor prognosis group [HR, 1.6 (0.3–6.9)], compared with (F) 21% of stage III patients [HR, 4.1 (2.0–8.2)]. All survival curves were calculated by Kaplan–Meier statistics and compared by log-rank tests. Relapse or death from colorectal cancer within the 10 or 5 years of follow-up was regarded events, and patients with no events within the indicated period of follow-up were censored.



The expression signature was an independent prognostic predictor in multivariate analyses including tumor stage in all 3 series. Considering each stage individually in the independent validation series, only stage III patients were stratified according to survival (HR, 4.1). Here, prognostic stratification was independent of the administration of adjuvant chemotherapy. This may be a result of the study design, as ColoGuidePro was developed from stage III patients that were not treated in an adjuvant setting. Although constituting the standard of care for stage III patients today, adjuvant treatment was administered to only approximately half (52%) of stage III patients in the test series, which is a consecutively collected patient series. This indicates that improved rationale for selecting patients for adjuvant treatment is highly warranted also for this

group of patients, strengthening the clinical potential of ColoGuidePro. Because of poorer general health conditions and the toxic side effects of chemotherapeutic treatment, older stage III patients are less commonly treated in an adjuvant setting. In Norway, such treatment is not standard for patients older than 75 years who are individually assessed for eligibility to chemotherapy (28). Importantly, ColoGuidePro strongly stratified the small subgroup of stage III patients in the external validation series above the age of 75 years ($n = 26$) according to survival. Similar separation was indicated also in the test and learning series; however, the sample numbers ($n = 8$ and 14 , respectively) were too small for this separation to be significant (data not shown). This provides strong indications that ColoGuidePro can be valuable when deciding on treatment options for

older stage III patients. Because of small sample numbers, studies specifically aimed at investigating the potential of ColoGuidePro in this subgroup of patients are needed to conclude. For stage II patients in the validation series, the results for ColoGuidePro were indiscriminant. This may be a result of the very high survival percentage in this group of patients (5-year relapse-free survival rate of 86%). This may also explain the small percentage of stage II patients in the validation series being assigned to the poor prognosis group by ColoGuidePro (10%). Recently, we have also developed and validated ColoGuideEx, a 13-gene prognostic expression signature specific for stage II colorectal cancer (29).

The external validation series used here is a collection of data from 2 previous studies where gene expression data have been used to develop prognostic classifiers (20, 23) and include the 2 major data sets with corresponding clinical information that are available from public repositories. These samples were analyzed on gene-level microarrays (Affymetrix HG-U133 Plus2.0), whereas the 2 in-house data sets were analyzed by exon-level microarrays (Affymetrix GeneChip Human Exon 1.0 ST). The good performance of the 7-gene expression signature in the test and validation series, analyzed on both types of microarrays, indicates robustness. More reliable expression measures in the learning series, with large numbers of probes targeting each gene across the entire length of the expressed sequences, may have contributed to this.

The prognostic test described here is based on the expression levels of only 7 genes. Previously published signatures for stage II and III colorectal cancer have typically contained a rather large number of genes, ranging from 20 to 100s, and there has been little focus on the implications of this during development of the survival models (14–18, 20, 23). A major statistical concern about prognostic prediction based on gene expression profiles relates to the high dimensionality of the data. Overfitting of large and complex gene expression models to the limited heterogeneity represented within the learning set of tumors, compromises the independent predictive powers. This risk can be reduced by penalization of the gene expression data using parameters tuned during cross-validation (30). In this study, lasso was used for penalization and simultaneous variable selection (24, 31, 32). Recently, 2 promising prognostic tests based on the expression levels of a small set of genes have been reported. ColoPrint measures the expression levels of 18 genes (22), whereas Oncotype DX includes 12, 7 of which are recurrence risk genes and 5 are reference genes (21). For development of the prognostic test presented here, care has been taken not only to reduce the number of genes but also to avoid redundancy in prognostic associations between the genes. In accordance with the lasso model, the 7 proposed genes have only weak correlations in expression (median Pearson correlation 0.1). Reduced covariation may improve the independent prognostic potential of each included gene (33).

Prospective testing of prediction models in large clinical trials provides a powerful means for assessment of their prognostic value. Currently, a phase III clinical trial is

recruiting patients for assessment of the ColoPrint test in patients with stage II colorectal cancer (PARSC study; ref. 34). Prognostic value of this test has recently been validated retrospectively in fresh-frozen tissue from an independent series of patients with stage II and III colorectal cancer analyzed on the same microarray platform as the training series (univariate: HR, 2.5; $P = 0.005$; ref. 22). These data have not been made publically available, and we have accordingly not been able to compare the performance of ColoGuidePro. Oncotype DX, a recurrence predictor for stage II colorectal cancer (35), has been developed from analyses of preselected genes (from a literature search) in more than 1,800 patients across 4 studies (21). This test has the advantage of being available for formalin-fixed, paraffin-embedded tumor tissue. Its predictive value has been tested on patients recruited from the QUASAR study (3), but despite reports of positive results in this initial validation study (36), more evidence is needed for a full evaluation of its value in clinical practice (37). We think it will be of great interest to evaluate both ColoGuidePro and ColoGuideEx together with these promising signatures in the same prospective study.

Clinically useful prognostic tests should not require much resources in terms of expression measures and subsequent interpretation of results (38, 39). The 7 genes included in ColoGuidePro have high variances in expression signals, improving the reliability of differential expression measures. Also, stratification of patients is based on the simple principle of summarizing the number of genes with expression levels outside a threshold. This strategy resulted in similar prognostic stratification when comparing with computation of PIs based on multivariate regression coefficients estimated as an inherent part of the lasso survival model. This indicates that the simple stratification rule proposed here is a valid replacement for a commonly used but a more complex mathematical model. Hence, testing the performance of the proposed 7-gene expression signature should be possible for independent researchers, and we welcome such efforts.

In conclusion, we have developed and validated a clinically feasible prognostic test for stage II and III colorectal cancer, although with best prognostic prediction potential for stage III patients. This classifier is based on the expression levels of a nonredundant set of 7 genes and can be transferred to a standardized assay for validation on individual patients in a prospective study.

Disclosures of Potential Conflicts of Interest

Disclosure of invention is accepted by hospital TTO. Patent application is in progress. No potential conflicts of interest were disclosed.

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References

- Garcia M, Jemal A, Ward EM, Center MM, Hao Y, Siegel RL, et al. Global cancer facts and figures 2007. Atlanta, GA: American Cancer Society; 2007.
- Van Cutsem E, D'Hoore A, de Vleeschouwer C, Decaestecker J, Penninckx F. Colon cancer: management of locoregional disease. In: Kelsen DP, Daly JM, Kern SE, Levin B, Tepper JE, Van Cutsem E, editors. Principles and practice of gastrointestinal oncology. 2nd ed. Philadelphia, PA: Lippincott Williams & Wilkins; 2008. p. 581.
- Gray R, Barnwell J, McConkey C, Hills RK, Williams NS, Kerr DJ. Adjuvant chemotherapy versus observation in patients with colorectal cancer: a randomised study. *Lancet* 2007;370:2020-9.
- Andre T, Sargent D, Tabernero J, O'Connell M, Buyse M, Sobrero A, et al. Current issues in adjuvant treatment of stage II colon cancer. *Ann Surg Oncol* 2006;13:887-98.
- Sobrero A. Should adjuvant chemotherapy become standard treatment for patients with stage II colon cancer? For the proposal. *Lancet Oncol* 2006;7:515-6.
- Kohne CH. Should adjuvant chemotherapy become standard treatment for patients with stage II colon cancer? Against the proposal. *Lancet Oncol* 2006;7:516-7.
- Benson AB III, Schrag D, Somerfield MR, Cohen AM, Figueredo AT, Flynn PJ, et al. American Society of Clinical Oncology recommendations on adjuvant chemotherapy for stage II colon cancer. *J Clin Oncol* 2004;22:3408-19.
- Andre T, Boni C, Mounedji-Boudiaf L, Navarro M, Tabernero J, Hickish T, et al. Oxaliplatin, fluorouracil, and leucovorin as adjuvant treatment for colon cancer. *N Engl J Med* 2004;350:2343-51.
- O'Connell JB, Maggard MA, Ko CY. Colon cancer survival rates with the new American Joint Committee on Cancer sixth edition staging. *J Natl Cancer Inst* 2004;96:1420-5.
- Locker GY, Hamilton S, Harris J, Jessup JM, Kemeny N, Macdonald JS, et al. ASCO 2006 update of recommendations for the use of tumor markers in gastrointestinal cancer. *J Oncol Pract* 2006;24:5313-27.
- Glas AM, Floore A, Delahaye LJ, Witteveen AT, Pover RC, Bakx N, et al. Converting a breast cancer microarray signature into a high-throughput diagnostic test. *BMC Genomics* 2006;7:278.
- Sparano JA, Paik S. Development of the 21-gene assay and its application in clinical practice and clinical trials. *J Clin Oncol* 2008;26:721-8.
- Filipits M, Rudas M, Jakesz R, Dubsy P, Fitzal F, Singer CF, et al. A new molecular predictor of distant recurrence in ER-positive, HER2-negative breast cancer adds independent information to conventional clinical risk factors. *Clin Cancer Res* 2011;17:6012-20.
- Arango D, Laiho P, Kokko A, Alhopuro P, Sammalkorpi H, Salovaara R, et al. Gene-expression profiling predicts recurrence in Dukes' C colorectal cancer. *Gastroenterology* 2005;129:874-84.
- Barrier A, Boelle PY, Roser F, Gregg J, Tse C, Braut D, et al. Stage II colon cancer prognosis prediction by tumor gene expression profiling. *J Clin Oncol* 2006;24:4685-90.
- Bertucci F, Salas S, Eysteris S, Nasser V, Finetti P, Ginestier C, et al. Gene expression profiling of colon cancer by DNA microarrays and correlation with histoclinical parameters. *Oncogene* 2004;23:1377-91.
- Eschrich S, Yang I, Bloom G, Kwong KY, Boulware D, Cantor A, et al. Molecular staging for survival prediction of colorectal cancer patients. *J Clin Oncol* 2005;23:3526-35.
- Wang Y, Jatko T, Zhang Y, Mutch MG, Talantov D, Jiang J, et al. Gene expression profiles and molecular markers to predict recurrence of Dukes' B colon cancer. *J Clin Oncol* 2004;22:1564-71.
- Lu AT, Salpeter SR, Reeve AE, Eschrich S, Johnston PG, Barrier AJ, et al. Gene expression profiles as predictors of poor outcomes in stage II colorectal cancer: a systematic review and meta-analysis. *Clin Colorectal Cancer* 2009;8:207-14.
- Jorissen RN, Gibbs P, Christie M, Prakash S, Lipton L, Desai J, et al. Metastasis-associated gene expression changes predict poor outcomes in patients with Dukes stage B and C colorectal cancer. *Clin Cancer Res* 2009;15:7642-51.
- O'Connell MJ, Lavery I, Yothers G, Paik S, Clark-Langone KM, Lopatin M, et al. Relationship between tumor gene expression and recurrence in four independent studies of patients with stage II/III colon cancer treated with surgery alone or surgery plus adjuvant fluorouracil plus leucovorin. *J Clin Oncol* 2010;28:3937-44.
- Salazar R, Roepman P, Capella G, Moreno V, Simon I, Dreezen C, et al. Gene expression signature to improve prognosis prediction of stage II and III colorectal cancer. *J Clin Oncol* 2011;29:17-24.
- Smith JJ, Deane NG, Wu F, Merchant NB, Zhang B, Jiang A, et al. Experimentally derived metastasis gene expression profile predicts recurrence and death in patients with colon cancer. *Gastroenterology* 2010;138:958-68.
- Goeman JJ. L₁ penalized estimation in the Cox proportional hazards model. *Biom J* 2009;52:70-84.
- Lothe RA, Peltomaki P, Meling GI, Aaltonen LA, Nystrom-Lahti M, Pylkkanen L, et al. Genomic instability in colorectal cancer: relationship to clinicopathological variables and family history. *Cancer Res* 1993;53:5849-52.
- Berg M, Danielsen SA, Ahlquist T, Merok MA, Agesen TH, Vatn MH, et al. DNA sequence profiles of the colorectal cancer critical gene set KRAS-BRAF-PIK3CA-PTEN-TP53 related to age at disease onset. *PLoS One* 2010;5:e13978.
- Sveen A, Agesen TH, Nesbakken A, Rognum TO, Lothe RA, Skotheim RI. Transcriptome instability in colorectal cancer identified by exon microarray analyses: associations with splicing factor expression levels and patient survival. *Genome Med* 2011;3:32.
- Norwegian Directorate of Health. Nasjonalt handlingsprogram med retningslinjer for diagnostikk, behandling og oppfølging av kreft i tykktarm og endetarm. Oslo, Norwegian Directorate of Health. 2012 Jun 28.
- Agesen TH, Sveen A, Merok MA, Lind GE, Nesbakken A, Skotheim RI, et al. ColoGuideEx: a robust gene classifier specific for stage II colorectal cancer prognosis. *Gut* 2012 Jan 2. [Epub ahead of print].
- van Houwelingen HC, Bruinsma T, Hart AA, van't Veer LJ, Wessels LF. Cross-validated Cox regression on microarray gene expression data. *Stat Med* 2006;25:3201-16.
- Tibshirani R. Regression shrinkage and selection via the LASSO. *J R Statist Soc B* 1996;58:267-88.
- Tibshirani R. The LASSO method for variable selection in the Cox model. *Stat Med* 1997;16:385-95.
- Næs T, Mevik BH. Understanding the collinearity problem in regression and discriminant analysis. *J Chemom* 2001;15:413-26.
- PARSC study (NCT00903565). A prospective study for the assessment of recurrence risk in stage II colon cancer patients using ColoPrint

- (PARSC). 2010. [cited 2012 Sep 21]. Available from: <http://clinicaltrials.gov/ct2/show/NCT00903565>.
35. Genomic Health I. Oncotype DX colon cancer assay. 2011. [cited 2012 Sep 21]. Available from: <http://www.oncotypedx.com/en-US/Colon.aspx>.
 36. Kerr D, Gray R, Quirke P, Watson D, Yothers G, Lavery IC, et al. A quantitative multigene RT-PCR assay for prediction of recurrence in stage II colon cancer: selection of the genes in four large studies and results of the independent, prospectively designed QUASAR validation study. *J Clin Oncol* 2009;27:4000.
 37. Webber EM, Lin JS, Evelyn PW. Oncotype DX tumor gene expression profiling in stage II colon cancer. Application: prognostic, risk prediction. *PLoS Curr* 2010;2:RRN1177.
 38. Koscielny S. Why most gene expression signatures of tumors have not been useful in the clinic. *Sci Transl Med* 2010;2:14ps2.
 39. Haibe-Kains B, Desmedt C, Sotiriou C, Bontempi G. A comparative study of survival models for breast cancer prognostication based on microarray data: does a single gene beat them all? *Bioinformatics* 2008;24:2200–8.

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