Molecular Pathways: Maintaining MAPK Inhibitor Sensitivity by Targeting Nonmutational Tolerance

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Abstract

Targeting hyperactive MAPK signaling has proven to be an effective treatment for a variety of different cancers. Responses to the BRAF inhibitors vemurafenib and dabrafenib and the MEK inhibitors trametinib and cobimetinib are, however, transient, and complete remission is rarely observed; rather, outgrowth of resistant clones within progressed tumors appears inevitable. These resistant tumors display great heterogeneity, which poses a major challenge to any salvage therapy. Recent focus has, therefore, been on the early dynamics of inhibitor response during tumor regression. During this time, cells can persist in an adapted tolerant state, which results in a phase of nonmutational drug tolerance. In this article, we discuss how inhibition of the MAPK pathway leads to an adaptive rewiring that evolves from the relief of immediate negative feedback loops to short-term gene expression changes and adaptation of intracellular signaling. Tolerance can also be mediated by external signaling from the tumor microenvironment, which itself adapts upon treatment and the selection for cells with an innate drug-tolerant phenotype. In preclinical models, combination treatment with receptor tyrosine kinase (RTK) inhibitors (lapatinib and dasatinib), histone deacetylase (HDAC) inhibitors (vorinostat and entinostat), or drugs targeting cancer-specific mechanisms (nelfinavir in melanoma) can overcome this early tolerance. A better understanding of how nonmutational tolerance is created and supported may hold the key to better combinational strategies that maintain drug sensitivity.

Background

The ERK/MAPK pathway consisting of the small GTPases of the RAS family (e.g., HRAS, KRAS, NRAS) and the families of RAF, MEK, and ERK kinases (ARAF, BRAF, CRAF, MEK1/2, and ERK1/2; Fig. 1A) is notoriously deregulated in a variety of cancers, which is why it is at the center of development of small-molecule inhibitors for targeted cancer therapy (1). Under physiologic conditions, the MAPK pathway regulates growth and survival downstream of many growth factor and cytokine receptors (1, 2). Receptor activation by growth factors causes a number of events, including RAS activation through the guanine nucleotide exchange factor SOS. Active RAS can recruit RAF kinases to the membrane, thereby facilitating their dimerization, phosphorylation, and activation (3). Active RAF then phosphorylates and activates the dual-specific kinase MEK, which phosphorylates ERK on serine and tyrosine. Phosphorylated, active ERK phosphorylates substrates in the cytoplasm, but after nuclear translocation, it also phosphorylates transcription factors, often regulating genes required for survival and growth (Fig. 1A).

Because of its importance in driving growth and survival, it is not surprising that the MAPK pathway is often exploited by transformed cancer cells to proliferate without the need of external stimuli. RAS is mutated in approximately 30% of all cancers, and, more specifically, the RAF kinase BRAF is mutated in approximately 50% of melanomas, but also approximately 10% of colorectal cancers and 11% of ovarian cancers. Inhibiting mutated RAS has proved difficult, but effective drugs inhibiting BRAF\[^{V600E}\] (the most prominent constitutively active BRAF mutant) or MEK in specific BRAF-mutant melanoma have achieved FDA approval. Today, combination therapies using the BRAF inhibitor vemurafenib and the MEK inhibitor cobimetinib, or likewise dabrafenib with trametinib, are standard of care for the treatment of advanced melanoma (4, 5). The combination therapy is currently being trialed in a variety of different cancers, including colorectal cancers (6).

Despite impressive initial responses, cures using these inhibitors are challenged by the development of resistance, which occurs in a majority of patients within a year after the commencement of treatment (7, 8). Great effort has gone into characterizing the mechanisms that are involved in the development of resistance, and, to date, a catalog of genetic alterations in resistant cells has been identified (8, 9). Initially, it was thought that combination therapies tailored to the genetic alterations found in relapsed patients could be the answer to this challenge, but a major obstacle to this approach is the mutational intertumor and even intratumor heterogeneity of these mutations (8–10). Thus, understanding the response dynamics over the course of treatment with MAPK inhibitors (MAPKi) could allow identifying new opportunities to improve the outcome of these therapies.

The transient nature of the activity of MAPK pathway inhibitors in vivo has been elegantly shown using a FRET reporter for ERK activity in melanoma allografts; ERK activity had recovered within one day (11). Unfortunately, this dynamic response is the result of many different mechanisms that ultimately can counteract the effect of BRAF or MEK inhibitors, and, in patients, these mechanisms are thought to come into action during different phases of the treatment period (Fig. 1B).
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Minutes to hours—relief of negative feedback mechanisms

Like all signaling pathways under physiologic conditions, the MAPK pathway is usually kept in balance by negative feedback mechanisms driven by its own activity (Fig. 1B). After receptor stimulation and activation of the pathway, ERK directly phosphorlates and regulates the activity of many proteins in the MAPK signaling cascade (12). ERK phosphorlates RAF, which

Figure 1.
Dynamic responses to MAPK pathway inhibition. A, The MAPK pathway is controlled through membrane receptors, but in cancer mutations in RAS, BRAF or MEK leads to its deregulation. B, Following on from immediate responses to MAPK inhibitor therapy is a phase of reversible nonmutational tolerance, before mutated resistant clones irreversibly re-establish tumor growth. Within hours of MAPK inhibition, pathway rewiring will lead to the disturbance of feedback signals, allowing ERK activity to recover. Following from this, intracellular signaling pathways rewire, partly due to extracellular signals leading to the adaption of cancer cells to the drug insult. ECM, extracellular matrix.
results in its reduced interaction with RAS, and, as a consequence, prevents its activation (13). In addition, ERK can inhibit SOS, the guanine nucleotide exchange factor for RAS, thereby impeding RAS activation. ERK also phosphorylates growth factor receptors, such as EGFR, which in turn can interfere with recruitment of GRB2–SOS–RAS and regulate the endocytosis of active receptors (12). Moreover, active MAPK signaling stimulates the expression of additional genes that negatively regulate the pathway. The best characterized of these are the dual-specific phosphatases (DUSP) and the receptor activity–regulating Sprouty (SPRY) genes that antagonize the ERK kinases activity and receptor signaling, respectively (14, 15).

Because these direct negative feedback mechanisms are dependent on ERK activity, any inhibition of ERK results in a new, unrepressed steady state of the MAPK pathway, where any external stimulation would lead to hyperactivation due to a lack of feedback mechanisms (16). Indeed, an array of receptor tyrosine kinase (RTK) ligands, including HGF and EGF, is able to elicit activation of the MAPK pathway in the presence of BRAF inhibition (16). This adaptive response can be effectively suppressed by combining pan-RTK inhibitors with BRAF inhibition to stop ligand-dependent RAS activation.

**Days to weeks—adaptation by intracellular signaling and selection for drug-tolerant cells**

The immediate changes in gene expression that are induced downstream of ERK (e.g., DUSP6 or cell-cycle genes) are followed by a second wave of gene expression changes, and this can lead to major rewiring of signaling pathways in response to long-term MAPK inhibition (Fig. 1B). Importantly, this readjustment can result in adaptation of the tumor cells to the drug treatment, thereby creating a state of tolerance (Fig. 1B). This phase of tolerance is nonmutational and, therefore, reversible, and it precedes the state of acquired resistance, when mutated, fully resistant cells have re-established tumor growth.

A situation of nonmutational drug tolerance is found with MITF, a “melanoma transcription factor” that previously had been shown to confer increased tolerance to BRAF and MEK inhibitors (17–19), and whose expression is tightly regulated downstream of oncogenic BRAF (20, 21). However, this regulation is rewired after long-term pathway inhibition, and in approximately 80% of melanomas, treatment with BRAF and MEK inhibitors for 2 weeks results in upregulation of MITF (22). MITF upregulation was also seen in preclinical models, where importantly, removal of the drug caused a reversal of MITF expression, demonstrating the nonmutational nature of MITF-mediated drug tolerance (22). A screen designed to target MITF expression during the phase of upregulation identified the HIV protease inhibitor nelfinavir as a potent agent to be used in combination with BRAF or MEK inhibitors. Most strikingly, as MITF is a melanoma survival protein, this approach might also be suitable for NRAS-mutant melanoma, and, indeed, MEK inhibitor/nelfinavir combination therapy completely blocked tumor growth in an NRAS<sup>*Q61K*/BRAF<sup>*V600E* preclinical model (22).

Long-term MAPK inhibition can also induce metabolic rewiring. It is well-documented that melanoma cells treated with BRAF inhibitors become dependent on oxidative phosphorylation (23–25), which leaves the cells resistant to ROS-based therapies but vulnerable to mitochondrial uncouplers, such as 2,4-dinitrophenol (24, 25). This metabolic rewiring is governed by the transcriptional coactivator PGC1α, which intriguingly is a bona fide MITF target gene (24, 25). This indicates that MAPKi-induced MITF upregulation can directly contribute to metabolic rewiring.

Overall, MITF expression levels in patients treated with BRAF and MEK inhibitors appear to be a good surrogate marker for response, and changes in MITF expression could be easily monitored by analyzing circulating DNA during treatment.

Another common adaptive response observed in response to MAPK signaling inhibition, not only in melanoma but also in colorectal cancer, involves enhanced RTK signaling. For instance, in colorectal cancer, BRAF inhibition causes activation of EGFR due to the inhibition of CDC25C (26). In melanoma, overexpression of ERBB3 is driven by upregulation of transcription factor FOXD3 as a direct consequence of MEK or BRAF inhibition (27). ERBB3, in the presence of its ligand NRG1β, overcomes BRAF or MEK inhibition by enhanced AKT survival signaling, which can be counteracted with combination therapy with the ERBB2 inhibitor lapatinib (27).

Another route in which MAPK pathway inhibition drives increased RTK activity is through RTK stabilization, whereby MAPK pathway inhibition blocks the ADA417/10-mediated shedding of RTKs from the cell membrane (28). An example of this stabilization is AXL, which has been linked to an aggressive and resistant phenotype in melanoma (19, 28, 29). To combat this RTK stabilization, the use of selective RTK inhibitors or a neutralizing antibody to TIMP1 in combination with an MAPKi is suggested.

In some cases, it is also argued that drug treatment leads to the selection of cancer cell subpopulations differing in their transcriptional state, which are slow cycling in the absence of drug but thrive in the presence of drug. For instance, overexpression of EGFR/platelet-derived growth factor receptor (PDGFR) is an indirect consequence of MAPK pathway inhibition due to the enrichment of EGFR<sup>high</sup> cells from a heterogeneous population determined by variable levels of its transcriptional repressor SOX10 (30). Under conditions of intermittent drug dosing, this is a reversible response, but continuous drug administration might lead to the permanent establishment of RTK<sup>high</sup> clones, which initiate acquired resistance. This is, in fact, seen in patients with relapsed BRAF-mutant melanoma and colorectal cancer, whose tumors have been found to overexpress a range of RTKs, including MET, PDGFR, and IGF1R (31–34). Exposure to MAPKi for 9 to 12 days induces drug-tolerant melanoma cells that display chromatin modifications paralleled by upregulation of histone demethylases (35). Short-term exposure to MEK inhibitors in BRAF-mutant colorectal cells results in increased STAT3 activity that brings about epigenetic remodeling via histone deacetylase (HDAC) activity, and inhibition of both the epigenetic modifiers, such as HDACs (using vorinostat or entinostat) and MEK, can lead to greater treatment potency (36). Again, selection for distinct subpopulations can occur, which was seen for melanoma cells differing in the expression of the histone demethylase JARID1B; JARID1B<sup>high</sup> cells are slow cycling and are characterized by an increase in oxidative phosphorylation (37).

**Days to weeks—adaptation through the tumor microenvironment**

Because of its central role in all cells, it is not surprising to find that inhibition of the MAPK pathway eventually leads to a rewiring of intracellular signaling not only in the tumor cells but also in the nontumor cells in the microenvironment. The tumor...
microenvironment is dynamic and adapts to antagonize treatment in the context of both targeted therapy and chemo/radiotherapy (38–40). Specifically, several stromal cell types can confer tolerance to MAPK pathway inhibition–treated cancer cells (41). Stromal fibroblasts can universally antagonize different targeted treatments in several cancer types, including colorectal cancer and ERBB2+ breast cancer. Fibroblasts drive internal changes in neighboring cancer cells through a secretome rich in growth factors and cytokines. Specifically, tolerance to BRAF inhibitors in melanoma can be induced by fibroblasts through secretion of HGF, a strong inducer of the MAPK cascade that can counteract the pathway inhibition through partial reactivation (16, 42). Indeed, it was observed that in patients, high intratumor HGF expression levels were correlated with response. In addition, HGF can antagonize MEK inhibition by increasing AKT-induced HGF expression levels were correlated with response. In addition, HGF can antagonize MEK inhibition by increasing AKT-induced survival signaling, thus providing a rationale for the combination of MAPK and AKT inhibitors to overcome the adaptive fibroblast secretome (42). Fibroblasts also influence treatment response through their ability to remodel the extracellular matrix (ECM). Remodeling the ECM can drive adaptation of melanomas treated with BRAF inhibitors by creating a fibronectin- and collagen-rich matrix environment that can facilitate reactivation of the MAPK pathway through integrin/FAK signaling (11, 43).

Tumor-associated macrophages (TAM) represent another stromal cell type that can contribute to tolerance to MAPK inhibition, and the accumulation of TAMs within tumors during treatment strongly correlates with faster relapse (38, 39). As with fibroblasts, the secretome of TAMs contains growth factors, cytokines, and other signaling molecules (44). VEGF secretion by TAMs leads to partial reactivation of the MAPK pathway in the presence of a BRAF inhibitor (39), whereas TNFα enhances survival signaling through MITF (38). The adaptive inflammatory reaction induced by accumulated macrophages can be overcome by either inhibiting macrophage differentiation with colony-stimulating factor receptor inhibitors or targeting the inflammatory signaling induced by NF-κB with IkB kinase inhibitors (38, 39). Both of these proposed strategies decrease the abundance of TAMs and, thus, remove the antagonistic secretome.

Finally, cellular rewiring can promote the establishment of resistant cells in heterogeneous tumors. Thereby, cells from either melanoma of lung adenocarcinomas responding to MAPK inhibition can act in a paracrine manner to nurture resistant cells (45). A therapy-induced secretome was identified downstream of transcriptional activator FRA1 (FOSL1) that, among other growth factors, contained IGF1. IGF1 resulted in PI3K survival signaling that, if inhibited, was able to prolong the response phase when combined with a BRAF inhibitor (45).

Conclusions

It has become apparent that the mutational heterogeneity detected in tumors from patients relapsed on MAPKi presents a major challenge, and strategies to improve responses to these inhibitors must consider the dynamics of the response. When thinking about strategies to target the adaptive phase, it is important to understand the underlying mechanisms. Indeed, the rewiring that occurs after MAPK inhibition can be thought of as a temporal process, where universal immediate changes eventually evolve into more divergent changes. Although the initial stage of pathway rewiring is primarily dependent on the loss of kinase activity of ERK, the following adaptive events appear to be mediated by changes in gene expression primarily induced by transcription factors, which in turn increase survival signaling and metabolic adaptation.

This adaptive phase is further cemented with the additional stromal and cellular heterogeneity, which also add to this increased survival signaling. It remains to be seen whether targeting common processes such as increased PI3K signaling or transcriptional changes is a possible route to a successful drug combination, but it seems more promising than tackling the complexity of intra- and intertumor mutation heterogeneity at the stage of progression. In addition, the idea of drug holidays is also a plausible solution, as this tolerance stage is transient and reversible, allowing one to conceive that removing MAPKi for a period of time can allow for rewiring reversal and regained sensitivity. Indeed, there is preclinical evidence pointing in this direction (46). With the recent insight we have gained into the relevance of nonmutational tolerance, it is clear that the complexity during this early response stage needs to be better understood to capitalize on this universal phase therapeutically.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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