

Significant Association of Rho/ROCK Pathway with Invasion and Metastasis of Bladder Cancer¹

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ABSTRACT

Purpose: The small GTP-binding protein Rho and its best-characterized downstream effector Rho-associated serine-threonine protein kinase, ROCK, participate in actin cytoskeleton organization, and are linked to pathogenesis and progression of several human tumors. We investigated the roles of Rho and ROCK in bladder cancer.

Experimental Design: Using Western blotting, we quantitated Rho and ROCK protein expression in paired tumor and nontumor surgical samples from 107 consecutive Japanese patients with bladder cancer.

Results: RhoA, RhoC, and ROCK were more abundant in tumors and metastatic lymph nodes than in nontumor bladder and uninvolved lymph nodes ($P < 0.0001$). Amounts of RhoA and RhoC protein, and ROCK protein expression correlated positively with one another ($P < 0.0001$). High RhoA, RhoC, and ROCK expression were related to poor tumor differentiation ($P < 0.05$, $P < 0.01$, and $P < 0.01$, respectively), muscle invasion ($P < 0.001$), and lymph node metastasis ($P < 0.05$). Kaplan-Meier plots linked high RhoA, RhoC, and ROCK protein expression to shortened disease-free and overall survival ($P < 0.0001$). By univariate analysis, high RhoA, RhoC, and ROCK protein expression predicted shortened disease-free and overall survival ($P < 0.0001$). By multivariate analysis, only RhoC was independently influenced in disease-free survival ($P < 0.05$), and RhoA and RhoC in overall survival ($P < 0.001$). In contrast, RhoB expression was inversely related to the grade and stage ($P < 0.05$), and its higher expression is associated with

better overall survival ($P < 0.05$). In superficial tumors (Ta or T1; 63 patients), RhoA, RhoC, and ROCK were unrelated with recurrence-free survival. Overall survival in tumors invading muscle (T2 to T4; 44 patients) was significantly influenced by RhoA, RhoC, and ROCK in a Kaplan-Meier analysis ($P < 0.0001$, $P < 0.0001$, and $P < 0.01$, respectively). Whereas RhoA, RhoC, and ROCK independently predicted shortened overall survival in patients with invasive tumor by univariate analysis ($P < 0.0001$, $P < 0.0001$, and $P < 0.01$, respectively), only RhoC did so by multivariate analysis ($P < 0.05$).

Conclusion: Rho/ROCK pathway apparently involved in occurrence and progression of bladder cancer may be valuable prognostic markers.

INTRODUCTION

Members of the Rho family of small GTPases are involved in regulation of a variety of cellular processes, including organization of the microfilament network, intercellular contact, and malignant transformation (1). Indeed, these events are interrelated. Specifically, subfamily of Rho family proteins have specialized action on the actin cytoskeleton. The Rho subfamily regulates formation of stress fibers and focal adhesions within cells. The Rac subfamily regulates formation of lamellipodia and membrane ruffling, whereas the Cdc42 subfamily regulates formation of filopodia (1, 2). Lamellipodia and filopodia are seen at the advancing aspect of motile cells, whereas retraction is seen on the opposite side (3). These alterations are accompanied by reorganization of the actin cytoskeleton within the cell. Rho-associated serine-threonine protein kinase, ROCK (4, 5), one of the best characterized downstream effectors of Rho, is activated when it selectively binds to the active GTP-bound form of Rho. Activated ROCK interacts with the actin cytoskeleton to promote stress-fiber formation and assembly of focal contacts (6).

Cancer cell migration is central to the process of metastasis. Here too, rearrangements of the actin cytoskeleton are involved. Rearranging the actin cytoskeletal proteins in response to Rho is important for the ability of tumor cells to metastasize (7). Overexpression of Rho has been linked to progression of human cancers (8–10). Thus, action of the Rho/ROCK pathway takes part in cancer progression by regulating actin cytoskeleton reorganization; indeed, a specific ROCK inhibitor was found to suppress the tumor growth and metastasis (11, 12). These observations suggest that the Rho/ROCK pathway may be a molecular target for prevention of cancer invasion and metastasis. As no data have been reported concerning the Rho/ROCK pathway in progression of bladder cancer, we used Western blotting to quantify protein expression for Rho and ROCK in bladder cancer tissue and non-neoplastic portions of the same resected specimen. Relationships between Rho and ROCK expression, and pathologic features of the tumors were examined,

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as were influences of Rho and ROCK expression on patient survival. Such information might lead to improved protocols and case selection for adjuvant therapies.

MATERIALS AND METHODS

Patients and Tissue Preparation. Surgical specimens of bladder cancer obtained between 1994 and 2000 from 107 consecutive Japanese patients with newly diagnosed primary transitional cell carcinoma of the bladder were examined: 68 men and 39 women; 41 to 89 years old; mean age, 67.7 years. All of the patients routinely underwent imaging studies (computed tomography and/or magnetic resonance imaging) before surgery of disease for preoperative staging. Postoperative follow-up ranged from 3 to 84 months (median, 45 months). Patients underwent surgery before receiving any other therapy. Transurethral resection was performed for superficial bladder tumors and a couple of nontumor epitheliums. During partial or total cystectomy, lymph nodes were resected. In all of the cystectomy cases, three sites of tumor, various portions of adjacent non-neoplastic bladder, and lymph nodes were resected for the study. Grading and staging were carried out according to the criteria of the Tumor-Node-Metastasis classification (13). The study was conducted in accord with the Helsinki Declaration. Institutional Review Board approval was obtained for this investigation. Each patient signed a consent form approved by the Committee on Human Rights in Research of our institution. Intravesical instillation of chemotherapeutic agents (mitomycin C, doxorubicin) and/or immunotherapy with *Bacillus Calmette-Guerin* was performed in cases where transurethrally resected superficial bladder tumors showed a high histopathologic grade. Systemic postoperative combination chemotherapy using methotrexate, vinblastine, Adriamycin, and cisplatin was administered for invasive disease that showed a high histopathologic grade and/or lymph node involvement.

Western Blotting. We dissected tumors and epitheliums by omitting stromal tissue. Western blotting was carried out as described previously (8). Briefly, 50 μ g of cytosolic proteins were separated by SDS-PAGE (12.5% gel) for immunological detection of proteins. After transfer blotting from the gel to a nitrocellulose membrane, bound proteins were stained by Ponceau S to confirm that identical amounts of protein had been transferred. Rho and ROCK protein expression each was analyzed using specific antibodies (each diluted 1:2000; Santa Cruz Biotechnology, Santa Cruz, CA). After incubation with peroxidase-conjugated antirabbit IgG, these proteins were visualized by chemiluminescence. The blotted membrane was scanned densitometrically with a PDI imaging scanner (Agfa Japan, Tokyo, Japan) and analyzed with NIH Image software. For quantitation of proteins, relative amounts of Rho and ROCK in tumors were expressed as a ratio of absorbance of bands from the tumor specimen to those from the corresponding normal tissue, which was set to 1.0, by densitometrical analysis as described previously (8). Mean values from three experiments were obtained for tumor, nontumor, and lymph node tissues, respectively (9).

Immunohistochemistry. Immunohistochemistry, using the same specific antibodies as antibodies for Western blotting,

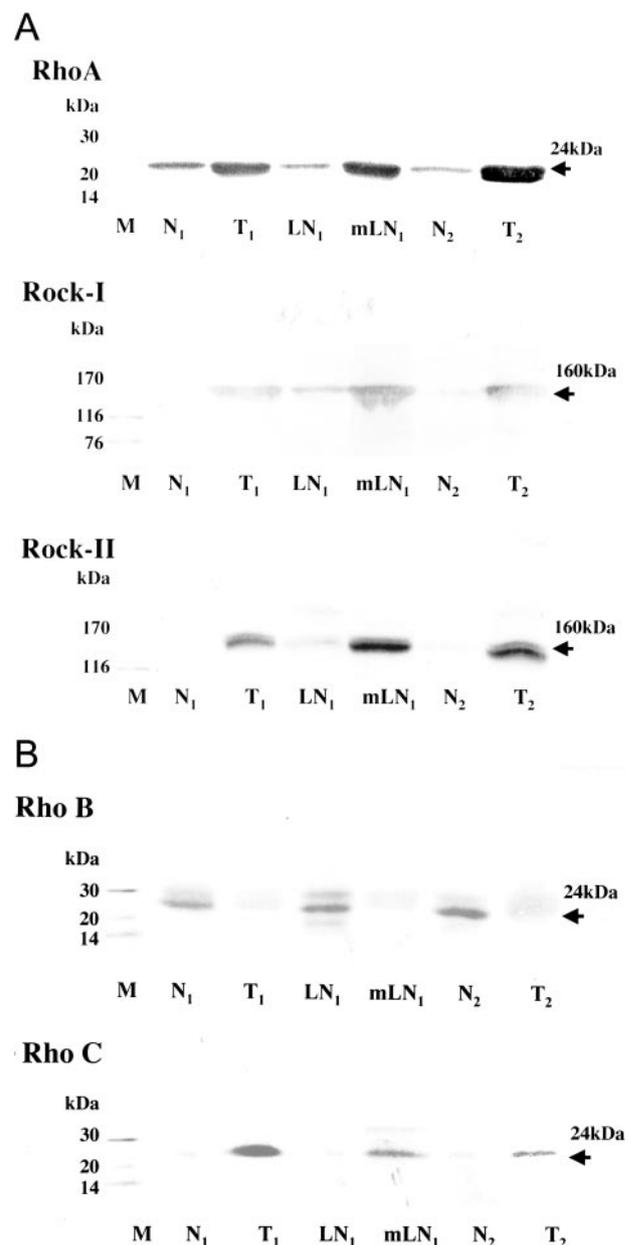


Fig. 1 Expression of Rho and ROCK proteins using Western blotting. *M*, marker; *N*, nontumor tissue; *T*, bladder tumor tissue; *LN*, normal lymph node; *mLN*, metastatic lymph node. Each number corresponds to a case number.

was performed to support the data obtained by Western blotting as described previously (8).

Statistical Analysis. Results of Western blotting were analyzed statistically using the Mann-Whitney *U* test as described previously (9, 10). The Spearman rank correlation coefficient was used to determine the relationship between protein expression for Rho and ROCK (14). Expression of Rho and ROCK protein, as well as tumor grade and stage of disease, were assessed in terms of survival by the Cox proportional hazards model using univariate and multivariate analysis. The

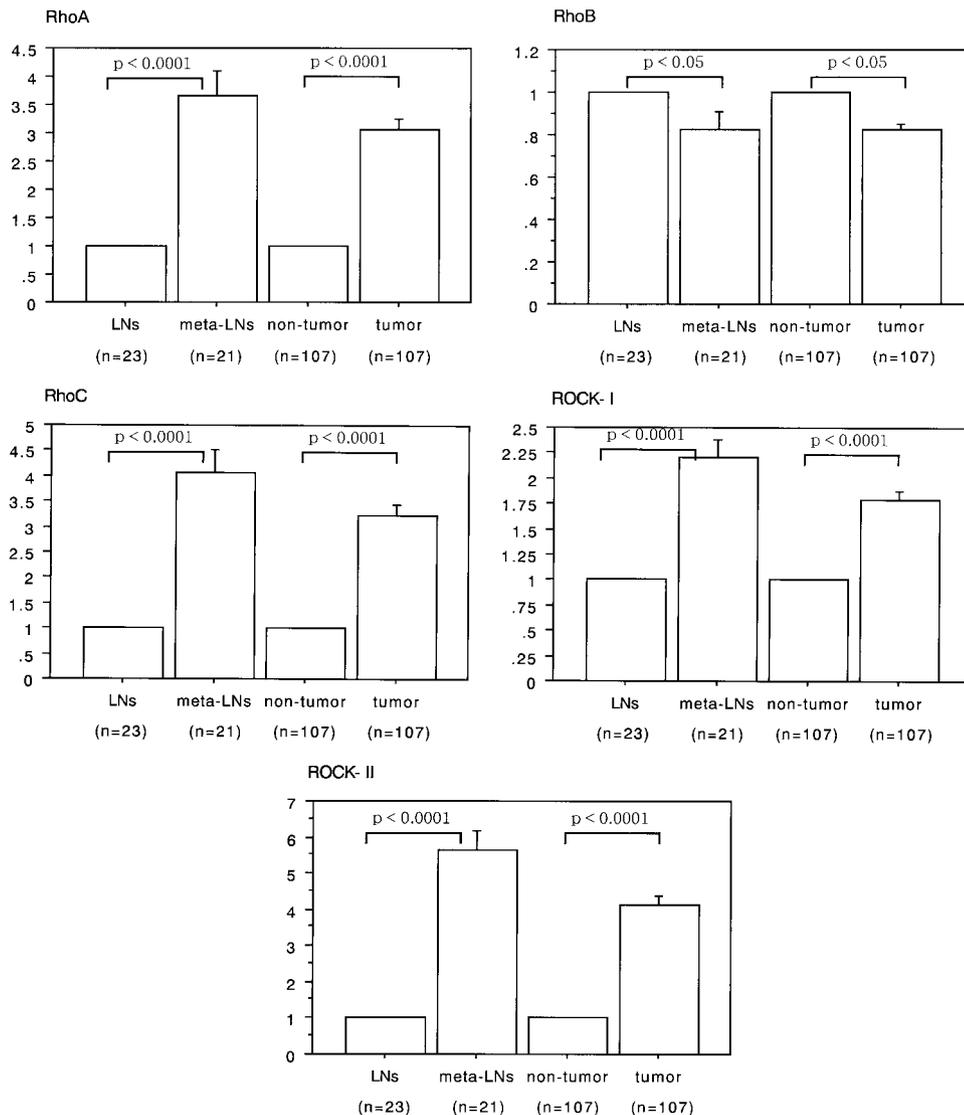


Fig. 2 The relative expression levels of Rho and ROCK protein in tumor to those in corresponding nontumor portion, which was set to 1.0, according to the method described previously (8). The data show the 95% confidential interval; bars, \pm SD.

Kaplan-Meier method was used to estimate survival as a function of time, and survival differences were assessed by the log-rank test. P s < 0.05 were considered significant. Data were analyzed using commercially available software.

RESULTS

Rho/ROCK Expression and Pathologic Characteristics.

RhoA protein was detected in tumor, nontumor bladder, lymph nodes with metastasis, and normal lymph nodes (Fig. 1). The amounts of protein for RhoA were significantly greater in cancerous (mean \pm SD = 3.07 ± 0.94) than in nontumor bladder tissue, defined as 1.0 ($P < 0.0001$; Fig. 2). Expression of protein for RhoA was greater in lymph nodes with metastases (3.66 ± 0.98) than in uninvolved lymph nodes, defined as 1.0 ($P < 0.0001$; Fig. 2). Increased RhoA protein was associated with high-grade poor differentiation: grade 1, 2.53 ± 0.37 ; grade 2, 2.91 ± 0.79 ; and grade 3, 3.64 ± 1.07 ($P < 0.05$, Fig. 3A). RhoA protein also was related to higher stage: pTa, 1, $2.61 \pm$

0.41; pT2, 3.01 ± 0.64 ; and pT3,4, 4.24 ± 1.05 ($P < 0.001$; Fig. 3F).

RhoC, ROCK-I, and ROCK-II expression was significantly greater in primary tumor tissue (3.22 ± 1.22 , 1.79 ± 0.49 , and 4.13 ± 1.34 , respectively) than in nontumor tissue, defined as 1.0 ($P < 0.0001$; Fig. 2). RhoC, ROCK-I, and ROCK-II proteins were expressed more strongly in lymph nodes with metastases (4.08 ± 0.95 , 2.20 ± 0.38 , and 5.64 ± 1.16 , respectively) than in normal lymph node tissue, defined as 1.0 ($P < 0.0001$; Fig. 2). Higher RhoC and ROCK expression in tumors was associated with poor differentiation (RhoC, in grade 1 lesion, 2.47 ± 0.26 ; grade 2, 3.14 ± 1.19 ; and grade 3, 3.85 ± 2.04 ; $P < 0.01$, Fig. 3C; ROCK-I, in grade 1 lesions, 1.42 ± 0.28 ; grade 2, 1.77 ± 0.43 ; and grade 3, 2.08 ± 0.49 ; $P < 0.01$, Fig. 3D; and ROCK-II, in grade 1 lesions, 3.06 ± 0.48 ; grade 2, 4.01 ± 1.25 ; and grade 3, 5.04 ± 1.35 ; $P < 0.001$, Fig. 3E). High expression also was related to muscle invasion (for RhoC, pTa and 1, 2.53 ± 0.38 ; pT2, 3.31 ± 0.84 ; pT3 and 4, 4.88 ± 1.23 ; $P <$

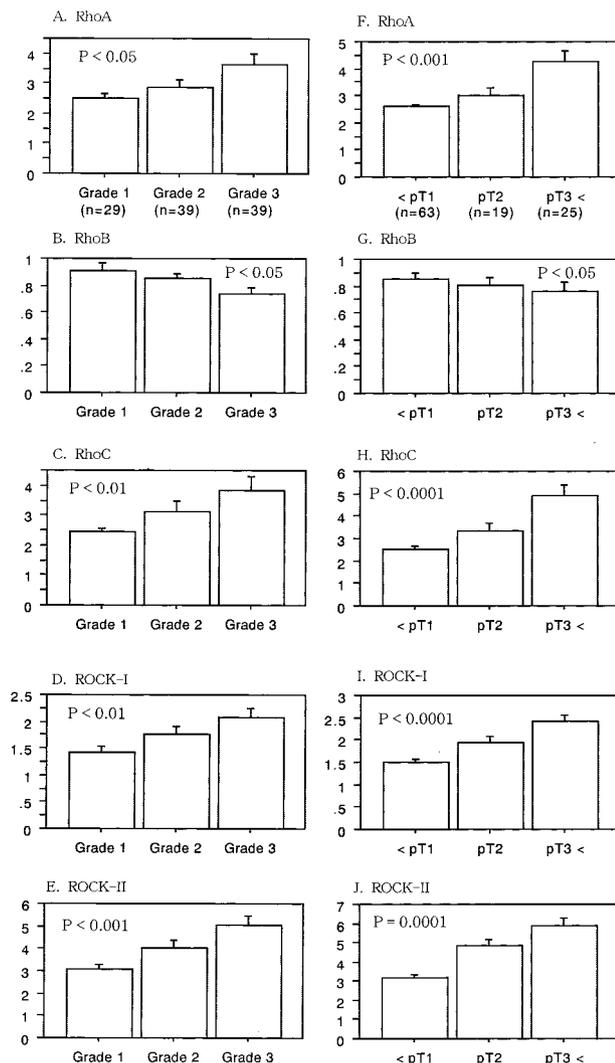


Fig. 3 Rho and ROCK expression in tumor. A–E, in grade 1 to 3 tumors. F–J, In < pT1, pT2 and pT3 < tumors. The data show the 95% confidential interval; bars, \pm SD.

0.0001, Fig. 3H; for ROCK-I, pTa and 1, 1.48 ± 0.29 ; pT2, 1.94 ± 0.30 ; pT3 and 4, 2.43 ± 0.32 ; $P < 0.0001$; Fig. 3I; for ROCK-II, pTa and 1, 3.20 ± 0.49 ; pT2, 4.82 ± 0.70 ; and pT3 and 4, 5.93 ± 0.94 ; $P < 0.001$; Fig. 3J).

RhoA, RhoC, ROCK-I, and ROCK-II protein were more abundant in primary tumors that invaded bladder muscle and involved lymph nodes (4.15 ± 1.12 , 4.93 ± 1.32 , 2.41 ± 0.35 , and 5.82 ± 0.91 , respectively) than in those that did not (3.42 ± 0.93 , $P = 0.0251$; 3.71 ± 0.94 , $P = 0.001$; 2.01 ± 0.36 , $P = 0.0008$; and 5.05 ± 1.02 , $P = 0.013$, respectively; Fig. 4). RhoB expression did not show a statistical difference between them (data not shown). Only RhoC protein level in tumors with muscle invasion and lymph node involvement (4.93 ± 1.32) were greater than in involved lymph nodes (4.08 ± 0.97 ; $P = 0.029$; Fig. 4).

Expression of protein for RhoA correlated positively with expression of ROCK-I and ROCK-II proteins in tumor tissue

(respective correlation coefficients, $r = 0.572$, $P < 0.0001$, and $r = 0.643$, $P < 0.0001$; Fig. 5), as well as for RhoC ($r = 0.598$, $P < 0.0001$, and $r = 0.681$, $P < 0.0001$, respectively; Fig. 5), but RhoB did not (data not shown).

Whereas RhoA, RhoC, and ROCK proteins were abundantly expressed in tumor cells, these were very weakly expressed in nontumor cells in immunohistochemistry (Fig. 6). RhoB was more abundantly expressed in nontumor cells than in tumor cells (Fig. 6), and inversely related to the grade and stage (Fig. 3).

Rho/ROCK and Survival. The mean value for protein expression of RhoA in tumor samples was $3.07 (\pm 0.94)$. As described in a previous study (10), cases were divided into two groups with expression above and below this mean, high expression (34 patients), and low expression (73 patients). Similarly, we divided the cases into groups by the mean values for RhoC and ROCK protein expression, for RhoC (3.22 ± 1.22), including 36 patients with high expression and 71 patients with low expression; for ROCK-I (1.79 ± 0.49), including 45 patients with high expression and 62 patients with low expression; for ROCK-II (4.13 ± 1.34), including 41 patients with high expression and 66 patients with low expression.

Univariate disease-free survival according to the Cox proportional hazards model was influenced significantly by RhoA, RhoC, ROCK-I, ROCK-II, stage, and grade (Table 1). By multivariate analysis, RhoC was an independent prognostic factor for disease-free survival. With regard to overall survival, whereas all of the factors analyzed were statistically significant by univariate analysis, only RhoC, RhoA, stage, and grade were significant by multivariate analysis (Table 1).

Comparison of Kaplan-Meier survival rate plots in patients with low versus high expression of RhoA protein linked high RhoA expression with shortened disease-free and overall survival ($P < 0.0001$; Fig. 7). Similarly, high expression of RhoC, ROCK-I, and ROCK-II was associated with poorer disease-free survival than low expression ($P < 0.0001$; Fig. 7). This also was true for overall survival ($P < 0.0001$; Fig. 7).

High expression of RhoB in 54 patients with a mean of $0.81 (\pm 0.15)$ was related to the better overall survival (Table 1; Fig. 7).

Cases of superficial disease (Ta and T₁; 63 patients) were divided into those with RhoA expression above (27 patients) or below (36 patients) based on a mean of $2.61 (\pm 0.41)$. In these superficial cases, a mean of RhoC was $2.53 (\pm 0.38)$ with high expression in 27 patients and low expression in 36 patients. ROCK-I showed a mean of $1.48 (\pm 0.29)$ with high expression in 35 patients and low expression in 28 patients, whereas for ROCK-II the mean was $3.20 (\pm 0.49)$ with high expression in 29 patients and low expression in 34 patients. Expression of these proteins failed to affect recurrence-free survival in superficial tumors according to Cox and Kaplan-Meier analyses (data not shown).

RhoB, with a mean $0.85 (\pm 0.16)$, low expression in 28 patients, and high expression in 35 patients, was not related to disease-free survival in superficial tumors (data not shown).

In tumor-invading muscle (T₂ and higher; 44 patients), mean RhoA expression was $3.73 (\pm 1.08)$, with high expression in 21 patients and low expression in 23 patients. For RhoC, the invasive cases showed a mean of $4.20 (\pm 1.33)$, with high

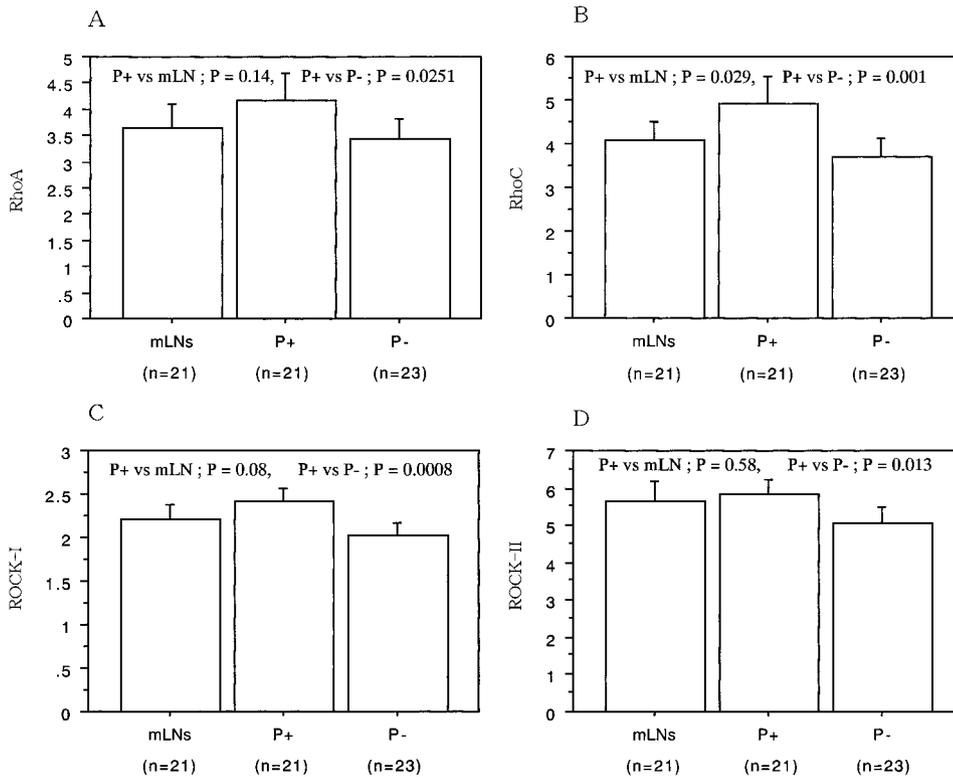


Fig. 4 RhoA, RhoC, ROCK-I, and ROCK-II expression in metastatic lymph node tissues (mLNs) and its primary muscle invasion tumor (P+), and muscle invasion tumor without lymph node involvement (P-). The data show the 95% confidential interval.

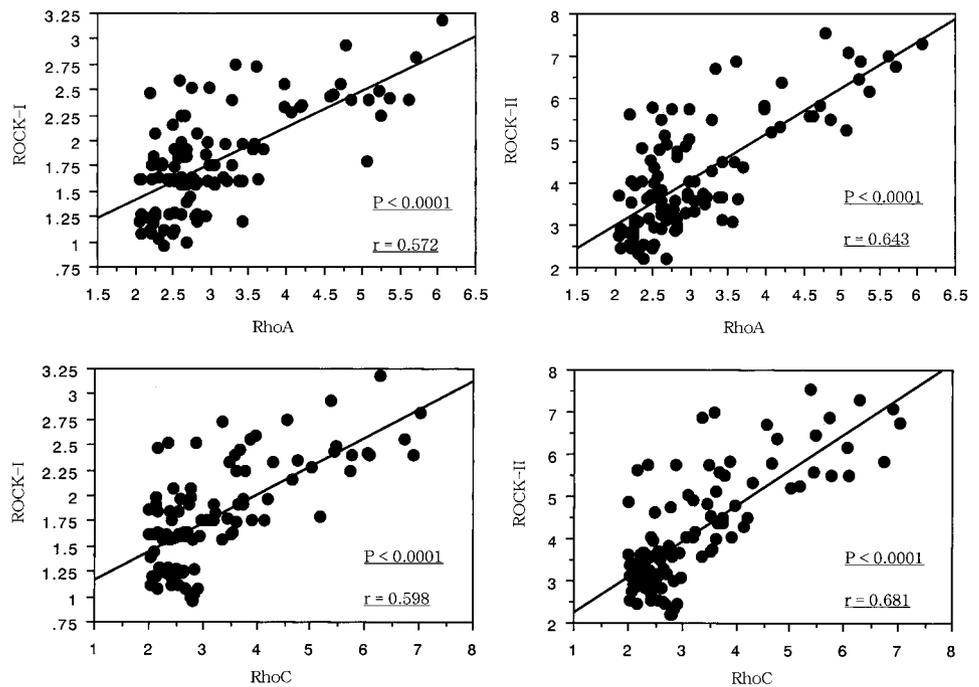


Fig. 5 Spearman rank correlation coefficient relationship between expression levels of proteins for RhoA/RhoC and ROCK.

expression in 24 patients and low expression in 20 patients; for ROCK-I, the mean was $2.22 (\pm 0.39)$, with high expression in 26 patients and low expression in 18 patients; for ROCK-II, the mean was $5.45 (\pm 1.29)$, with high expression in 30 patients and

low expression in 14 patients. Kaplan-Meier plots showed that high expression of RhoA, RhoC, ROCK-I, or ROCK-II protein each had a significant influence on disease-free survival in the invasive tumor group ($P = 0.0112$, $P = 0.0109$, $P = 0.0129$,

Fig. 6 Immunohistochemical staining using anti-RhoA (top left), anti-RhoB (bottom left), anti-RhoC (top right), and anti-ROCK-II (bottom right) monoclonal antibody in nonpapillary type, grade 3 and pT3 bladder tumor ($\times 400$ magnification). Panels, except for bottom left, show intensely brown staining in most of the cytoplasm of the cancer cells, displaying high RhoA, RhoC, and ROCK-II protein levels. Bottom left panel shows negative reaction, displaying low RhoB protein levels.

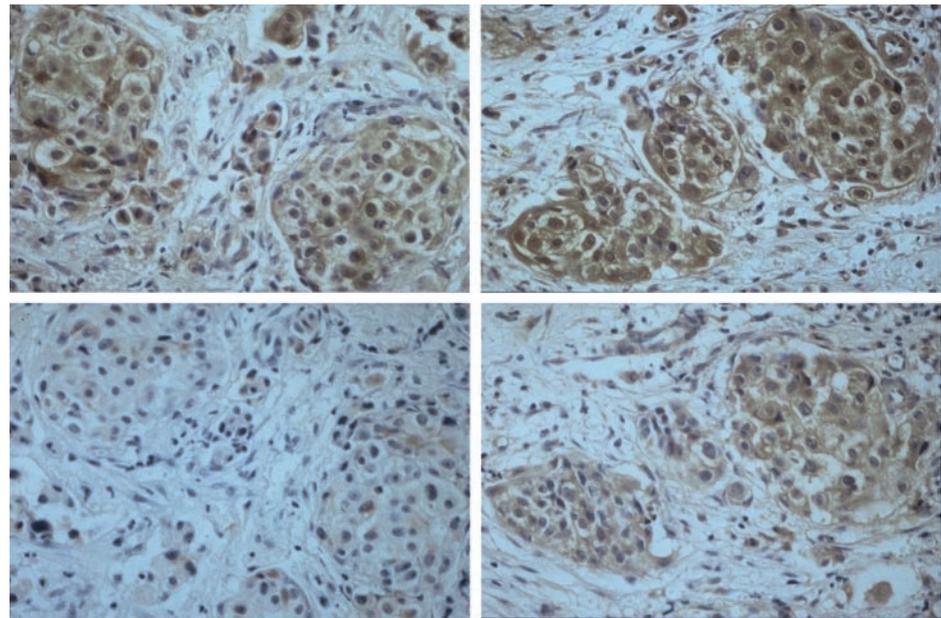


Table 1 Cox regression analysis for various potential prognostic factors in survival

Variable	Unfavorable/favorable characteristics	No. of patients	Analysis	Disease-free survival			Overall survival		
				Relative risk	95% Confidential interval	P	Relative risk	95% Confidential interval	P
Grade	3/2/1	39/39/29	U ^a	1.775	1.243–2.535	0.0016	3.958	1.964–7.974	0.0001
			M ^b	1.106	0.539–1.541	0.7104	2.141	1.017–3.253	0.0478
pT	>3/2/1, a	25/19/63	U	2.213	1.632–3.011	<0.0001	5.426	3.056–9.634	<0.0001
			M	1.191	0.699–1.766	0.2452	1.618	1.252–3.845	0.0361
RhoA	High/low	34/73	U	3.381	1.943–5.884	<0.0001	37.931	8.909–161.493	<0.0001
			M	1.681	0.787–3.591	0.1081	10.956	2.067–58.084	0.0049
RhoB	Low/high	53/54	U	1.612	0.925–2.811	0.0923	2.334	1.007–5.413	0.0482
			M	1.272	0.661–2.441	0.4729	1.019	0.241–4.313	0.9798
RhoC	High/low	36/71	U	4.484	2.508–8.017	<0.0001	70.583	9.427–228.496	<0.0001
			M	2.642	1.136–6.142	0.0241	24.302	2.765–113.602	0.004
ROCK-I	High/low	45/62	U	3.243	1.845–5.701	<0.0001	14.463	4.311–48.528	<0.0001
			M	1.281	0.541–3.031	0.5737	1.325	0.162–4.811	0.8852
ROCK-II	High/low	41/66	U	4.248	2.405–7.502	<0.0001	18.132	5.389–61.013	<0.0001
			M	2.054	0.532–7.926	0.2961	2.059	0.294–21.847	0.4491

^a U, univariate.

^b M, multivariate.

and $P = 0.0295$, respectively; Fig. 8, A–D), as well as on overall survival ($P < 0.0001$, $P < 0.0001$, $P = 0.0002$, and $P = 0.002$, respectively; Fig. 8, E–H). Although all four of the proteins predicted disease-free and overall survival of invasive cases in univariate analyses, only RhoC remained significant for overall survival in a multivariate analyses (Table 2).

RhoB expression, with a mean $0.77 (\pm 0.14)$, low expression in 25 patients, and high expression in 17 patients, was not linked to survival in the invasive tumor (data not shown).

DISCUSSION

Rho proteins act as intracellular molecular switches that transduce signals from extracellular stimuli to the actin cytoskeleton and the nucleus, regulating cell migration and malignant transformation. To take into account the possibility of

interindividual variation in expression of proteins, we compared protein expression of Rho and ROCK between paired tumor and nontumor tissue samples from bladder and from lymph node. Protein expression for RhoA, RhoC, and ROCK were significantly greater in primary tumors and lymph node with metastases than in non-neoplastic bladder and normal lymph node samples. High RhoA, RhoC, and ROCK protein expression in tumors was associated with poor differentiation, muscle invasion, lymph node metastasis, and shortened survival. In contrast, RhoB expression was inversely related to grade and stage. To our knowledge, this is the first report concerning the relationship between Rho/ROCK and bladder cancer. The data suggest that the Rho/ROCK pathway may be associated with progression of bladder cancer.

Overexpression of Rho has been reported in a number of

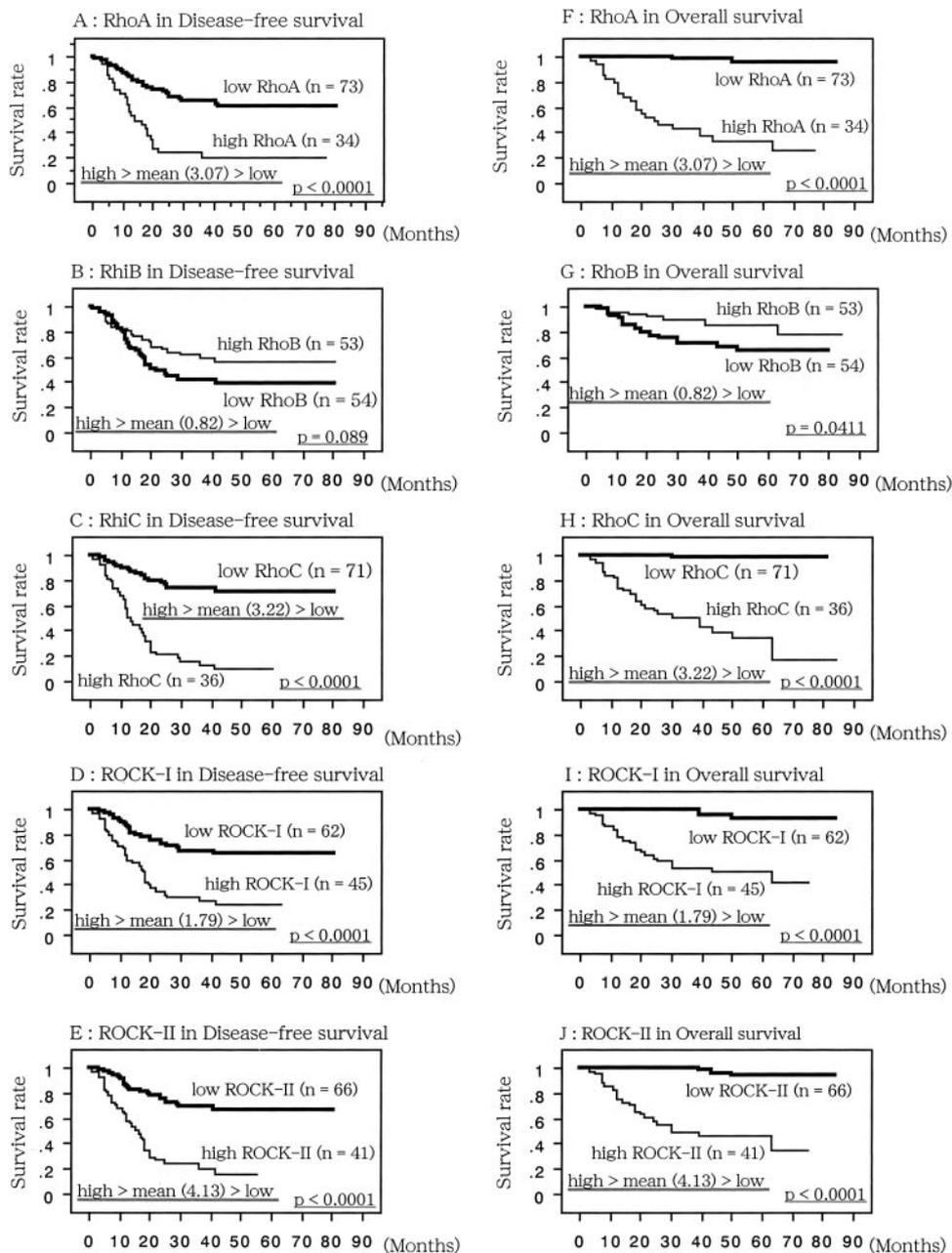


Fig. 7 Survival curve in patients with bladder cancer based on the mean values of protein expression of Rho and ROCK in tumor tissues, the cases were divided into two groups at this level: high and low expression. Disease-free and overall survival curve in total 107 tumors. *P* was analyzed by log-rank test.

human cancers (8–10, 15). Furthermore, several lines of evidence directly link Rho to acquisition of a migratory, invasive, and metastatic phenotype (16). Rho is important for the ability of tumor cells to metastasize (7). We showed that RhoA, RhoC, and ROCK, but not RhoB protein expressions were greater in bladder tumors than in parts of the same bladder resection specimen containing no tumor.

RhoA, the prototype of Rho, and ROCK are best characterized in human cancers (11, 12, 14). High expression of RhoA facilitates translocation of this protein from the cytosol to the cell membrane, where it becomes activated and promotes invasion (17). Increased expression of Rho has been linked to higher

stage in human cancers (8–10). ROCK mediates responses in the pathway initiated by Rho, and regulates dynamic reorganization of cytoskeletal proteins such as in formation of stress fibers and focal adhesions (18). Although at least two ROCK isoforms, ROCK-I and ROCK-II, may be expressed in the same species (19), differences in their roles in Rho-mediated signaling remain unclear. In the present study, a positive correlation was seen in tumor tissues between expression of RhoA and ROCK. Moreover, higher expressions of ROCK-I/ROCK-II, as well as RhoA in the primary lesion was associated not only with higher stages of disease, but also with poorer survival, in particular in tumors invading muscle. Whereas only weakly detectable in

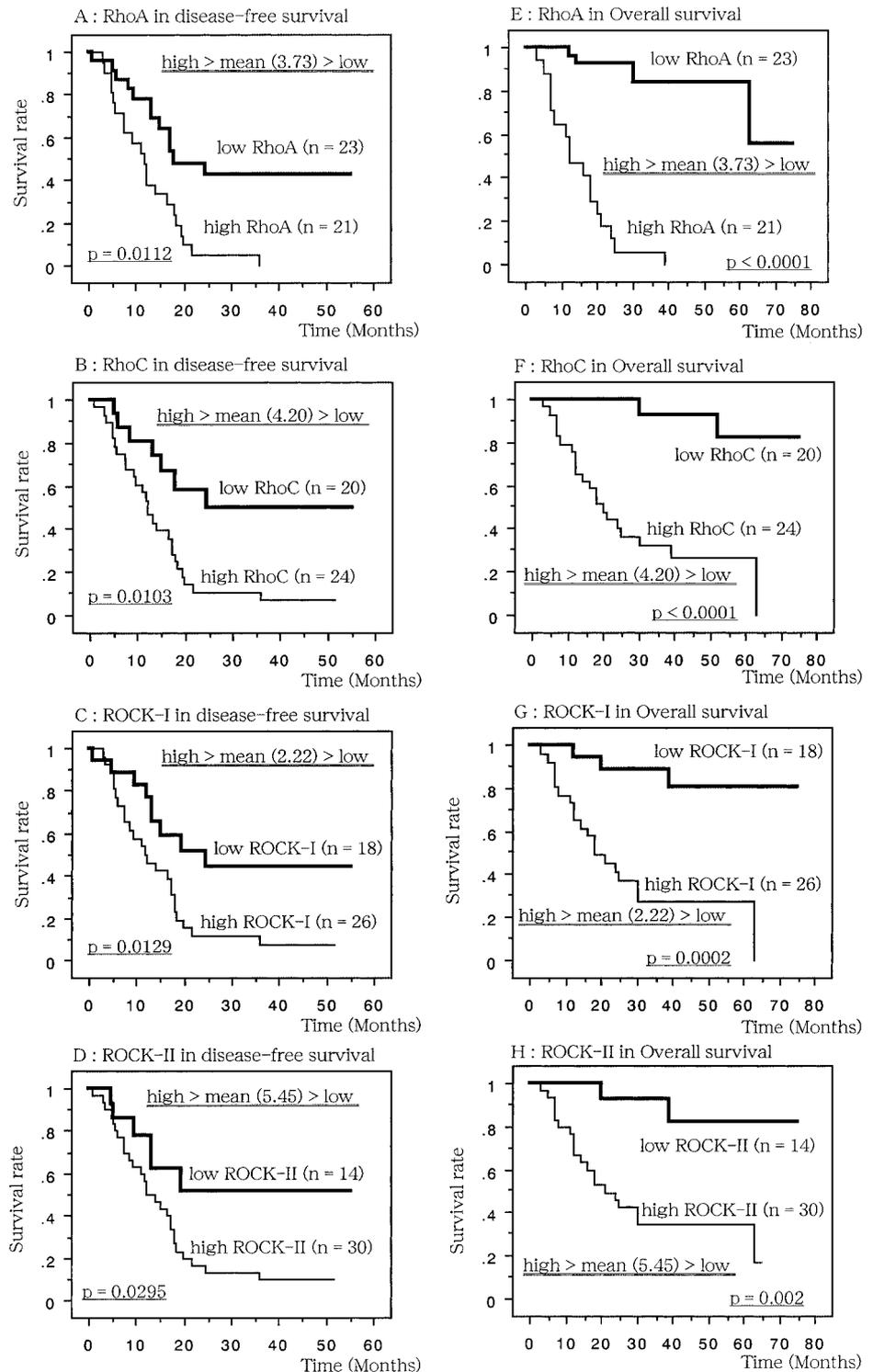


Fig. 8 Survival curve in muscle invasive bladder cancer based on the mean values of protein expression of Rho and ROCK in tumor tissues, the cases were divided into two groups at this level: high and low expression. Disease-free and overall survival curve in 44 tumors. *P* was analyzed by log-rank test.

normal lymph nodes tissue, RhoA and ROCK were abundant in lymph node with metastasis. Furthermore, both were significantly more abundant in tumors invading muscle that showed lymph node involvement than in those without nodal involvement. Ability to enter either blood or lymphatic vasculature is

necessary for tumor cells to metastasize to distant sites (20). RhoA and ROCK reportedly are required in both endothelial and the migrating cells for them to cross the vascular endothelium (21, 22).

RhoC, high homology to RhoA, has been shown to be the

Table 2 Cox regression analysis for survival in muscle invasive tumor

Variable	Unfavorable/favorable characteristics	No. of patients	Analysis	Disease-free survival			Overall survival		
				Relative risk	95% Confidential interval	P	Relative risk	95% Confidential interval	P
RhoA	High/low	21/23	U ^a	2.415	1.206–4.837	0.0128	8.125	3.113–21.204	<0.0001
			M ^b	1.387	0.523–3.678	0.5105	1.496	0.485–4.619	0.4835
RhoC	High/low	24/20	U	2.522	1.232–5.161	0.0113	18.474	4.269–79.935	<0.0001
			M	1.537	0.542–4.354	0.4189	9.339	1.691–51.580	0.0104
ROCK-I	High/low	26/18	U	2.574	1.188–5.577	0.0166	7.287	2.139–24.823	0.0015
			M	1.652	0.508–5.378	0.4042	1.889	0.459–7.775	0.378
ROCK-II	High/low	30/14	U	2.58	1.062–6.267	0.0363	7.09	1.649–30.478	0.0085
			M	1.255	0.320–4.932	0.7446	2.624	0.502–13.723	0.2531

^a U, univariate.

^b M, multivariate.

most associated with metastasis of cancer cells among Rho subfamily using high-density DNA microassays (23). Moreover, overexpression of RhoC has been associated with the progression of pancreatic ductal adenocarcinoma (10) and inflammatory breast cancer (24). It has been reported that overexpression of RhoC leads to increased expression of angiogenic factors (25), and that RhoC overexpression also promotes the ability of melanoma cells to exit the blood and colonize the lung (23). Therefore, taken together, the RhoA and RhoC/ROCK pathway is likely to be involved in local invasion and lymph node metastasis of tumor cells. Expression of pathway components may be a useful prognostic marker in bladder cancer.

RhoB also regulates actin organization. It is required for signaling apoptosis in transformed cells that are exposed to farnesyltransferase inhibitors, DNA-damaging agents, or Taxol. Genetic analysis in mice indicates that RhoB is dispensable for normal cell physiology, but that it has a suppressor or negative modifier function in stress-associated process, including cancer (26). As well as in bladder cancer shown in the present study, it has been reported that RhoB expression is readily detected in normal epithelium, carcinomas *in situ*, and well-differentiated tumors, but it becomes weak to undetectable as tumors become deeply invasive and poorly differentiated in human head and neck squamous cell carcinomas, and RhoA protein levels increase with tumor progression (27). Furthermore, we showed here that higher expression of RhoB was associated with favorable prognosis in bladder cancer. A negative role in growth control and/or transformation would contrast with the positive effects of RhoA and RhoC in these processes (26). These findings give additional support to the notion that RhoB may play a tumor-suppressive role in human cancers.

Although it is likely that RhoA, RhoB, and RhoC share common functions in regulating stress fiber formation (28), these proteins differ in subcellular location (29) and regulation of expression (30, 31). Therefore, abnormal regulation of expression patterns between members of the Rho subfamily in neoplastic cells may be involved in carcinogenesis and cancer progression.

We did not analyze the effect of treatment modalities on outcome of patients in the current study. Despite reports that tumor burden can be reduced for superficial tumors by intravesical chemotherapy and *Bacillus Calmette-Guerin* immunotherapy or systemic combined methotrexate, vinblastine, Adri-

mycin, and cisplatin chemotherapy in the case of invasive tumors, these outcomes were poorer than expected (32, 33). In the present study, higher expression of RhoA, RhoC, and ROCK was associated with shorter survival in bladder cancer. New treatment modalities such as cell migration inhibitors may be important possibilities to consider.

A specific ROCK inhibitor, Y-27632 (34, 35), has been reported to block both Rho-mediated activation of actomyosin and invasive activity of cultured rat MMI hepatoma cells (12). Continuous treatment with this inhibitor reduced dissemination of MMI cells implanted into the peritoneal cavity of syngeneic rats (11). These reports suggested that ROCK inhibition may represent a way to prevent cancer invasion and metastasis by inhibiting cell migration and morphological alterations. We will need to study the effect of ROCK inhibitors on bladder cancer cells *in vitro* and *in vivo*. These results may suggest improved treatment strategies.

Mutations of Rho have not been identified in human tumors (10, 36, 37). Although mutational change in Rho genes was not within the scope of the current study, this should be investigated in the future.

Many downstream effectors are likely to be involved in the Rho signaling pathway (4). p140mDia, a mammalian homologue of the *Drosophila* diaphanous, controls actin polymerization (38). ROCK and p140mDia act cooperatively in the stress fiber formation to mediate Rho effects (39). Other possible downstream effectors and their roles need to be explored in bladder cancer. Rho family GTPases, including Rho, Rac, and Cdc42, have been shown to differentially and cooperatively contribute to triggering of invasive behavior in tumor cells (40). More complete understanding of how Rho family GTPases mutually and specifically interact in bladder cancer may throw light on the therapeutic approaches.

REFERENCES

- Hall, A. Rho GTPases and the actin cytoskeleton. *Science* (Wash. DC), 279: 509–514, 1998.
- Van Aelst, L., and D'Souza-Schorey, C. Rho GTPases and signaling networks. *Genes Dev.*, 11: 2295–2322, 1997.
- Nobes, C. D., and Hall, A. Rho, Rac, and Cdc42 GTPases regulate the assembly of multimolecular focal complexes associated with actin stress fibers, lamellipodia, and filopodia. *Cell*, 81: 53–62, 1995.

4. Bishop, A. L., and Hall, A. Rho GTPases and their effector proteins. *Biochem. J.*, *348*: 241–255, 2000.
5. Ishizaki, T., Maekawa, M., Fujisawa, K., Okawa, K., Iwamatsu, A., Fujita, A., Watanabe, N., Saito, Y., Kakizuka, A., Morii, N., and Narumiya, S. The small GTP-binding protein Rho binds to and activates a 160 kDa Ser/Thr protein kinase homologous to myotonic dystrophy kinase. *EMBO. J.*, *15*: 1885–1893, 1996.
6. Ishizaki, T., Naito, M., Fujisawa, K., Maekawa, M., Watanabe, N., Saito, Y., and Narumiya, S. p160ROCK, a Rho-associated coiled-coil forming protein kinase, works downstream of Rho and induces focal adhesions. *FEBS Lett.*, *404*: 118–124, 1997.
7. del Peso, L., Hernandez-Alcoceba, R., Embade, N., Carnero, A., Esteve, P., Paje, C., and Lalca, J. C. Rho proteins induce metastatic properties *in vivo*. *Oncogene*, *15*: 3047–3057, 1997.
8. Fritz, G., Just, I., and Kaina, B. Rho GTPase over-expressed in human tumors. *Int. J. Cancer*, *81*: 682–687, 1999.
9. Kamai, T., Arai, K., Tsujii, T., Honda, M., and Yoshida, K-I. Over-expression of RhoA mRNA is associated with advanced stage in testicular germ cell tumour. *BJU Int.*, *87*: 227–231, 2001.
10. Suwa, H., Ohshio, G., Imamura, T., Watanabe, G., Arai, S., Imamura, M., Narumiya, S., Hiai, H., and Fukumoto, M. Overexpression of the *rho C* gene correlates with progression of ductal adenocarcinoma of the pancreas. *Br. J. Cancer*, *77*: 147–152, 1998.
11. Itoh, K., Yoshioka, K., Akedo, H., Uehata, M., Ishizaki, T., and Narumiya, S. An essential part for Rho-associated kinase in the transcellular invasion of tumor cells. *Nat. Med.*, *5*: 221–225, 1999.
12. Imamura, F., Mukai, M., Ayaki, M., and Akedo, H. Y-27632, an inhibitor of Rho-associated protein kinase, suppresses tumor cell invasion via regulation of focal adhesion and focal adhesion kinase. *Jpn. J. Cancer Res.*, *91*: 811–816, 2000.
13. Sobin, L. H. (ed.). International union against cancer. *In: TNM Classification of Malignant Tumours*, 5th ed. Geneva: UICC, 1997.
14. Kamai, T., Arai, K., Sumi, S., Tsujii, T., Honda, M., Yamanishi, H., and Yoshida, K-I. The rho/rho-kinase pathway is involved in the progression of testicular germ cell tumour. *BJU Int.*, *89*: 449–453, 2002.
15. Sahai, E., and Marshall, C. J. Rho-GTPases and cancer. *Nat. Rev. Cancer*, *2*: 133–142, 2002.
16. Schmitz, A. A., Govek, E. E., Bottner, B., and Van Aelst, L. Rho GTPases: signaling, migration, and invasion. *Exp. Cell Res.*, *261*: 1–12, 2000.
17. Yoshioka, K., Nakamori, S., and Itoh, K. Overexpression of small GTP-binding protein RhoA promotes invasion of tumor cells. *Cancer Res.*, *59*: 2004–2010, 1999.
18. Amano, M., Fukata, Y., and Kaibuchi, K. Regulation and functions of Rho-associated kinase. *Exp. Cell Res.*, *261*: 44–51, 2000.
19. Nakagawa, O., Fujisawa, K., Ishizaki, T., Saito, Y., Nakao, K., and Narumiya, S. ROCK-I and ROCK-II, two isoforms of Rho-associated coiled-coil forming protein serin/threonine kinase in mice. *FEBS Lett.*, *392*: 189–193, 1996.
20. Price, J. T., Bonovich, M. T., and Kohn, E. C. The biochemistry of cancer dissemination. *Crit. Rev. Biochem. Mol. Biol.*, *32*: 175–253, 1997.
21. Worthylake, R. A., Lemoine, S., Watson, J. M., and Burridge, K. RhoA is required for monocyte tail retraction during transendothelial migration. *J. Cell Biol.*, *154*: 147–160, 2001.
22. Adamson, P., Etienne, S., Couraud, P. O., Calder, V., and Greenwood, J. Lymphocyte migration through brain endothelial cell monolayers involves signaling through endothelial ICAM-1 via a Rho-dependent pathway. *J. Immunol.*, *162*: 2964–2973, 1999.
23. Clark, E. A., Golub, T. R., Lander, E. S., and Hynes, R. O. Genomic analysis of metastasis reveals an essential role for RhoC. *Nature (Lond.)*, *406*: 532–535, 2000.
24. van Golen, K. L., Wu, Z-F., Qiao, X. T., Bao, L. W., and Merajver, S. D. RhoC GTPase, a novel transforming oncogene for human mammary epithelial cells that partially recapitulates the inflammatory breast cancer phenotype. *Cancer Res.*, *60*: 5832–5838, 2000.
25. van Golen, K. L., Wu, Z-F., Qiao, X. T., Bao, L. W., and Merajver, S. D. RhoC GTPase overexpression modulates induction of angiogenic factors in breast cells. *Neoplasia*, *2*: 18–425, 2000.
26. Prendergast, G. C. Actin'up: RhoB in cancer and apoptosis. *Nat. Rev. Cancer*, *1*: 162–168, 2001.
27. Adnane, J., Muro-Cacho, C., Mathews, L., Sebt, S. M., and Munoz-Antonia, T. Suppression of rho B expression in invasive carcinoma from head and neck cancer patients. *Clin. Cancer Res.*, *8*: 2225–2232, 2002.
28. Ridley, A. J., and Hall, A. The small GTP-binding protein rho regulates the assembly of focal adhesions and actin stress fibers in response to growth factors. *Cell*, *70*: 389–399, 1992.
29. Adamson, P., Marshall, C. J., Hall, A., and Tilbrook, P. A. Post-translational modifications of p21rho proteins. *J. Biol. Chem.*, *267*: 20033–20038, 1992.
30. Jahner, D., and Hunter, T. The ras-related gene rhoB is an immediate-early gene inducible by v-Fps, epidermal growth factor, and platelet-derived growth factor in rat fibroblasts. *Mol. Cell Biol.*, *11*: 3682–3690, 1991.
31. Fritz, G., Kaina, B., and Aktories, K. The ras-related small GTP-binding protein RhoB is immediate-early inducible by DNA damaging treatments. *J. Biol. Chem.*, *270*: 25172–25177, 1995.
32. Lamm, D. L., Riggs, D. R., Traynelis, C. L., and Nseyo, U. O. Apparent failure of current intravesical chemotherapy prophylaxis to influence the long-term course of superficial transitional cell carcinoma of the bladder. *J. Urol.*, *153*: 1444–1450, 1995.
33. Conner, J. P., Rpoportm, F., Olsson, C. A., Sawczuk, I. S., and Benson, M. C. Long-term follow-up patients treated with methotrexate, vinblastine, doxorubicin and cisplatin (M-VAC) for transitional cell carcinoma of urinary bladder: cause for concern. *Urology*, *34*: 353–356, 1990.
34. Uehata, M., Ishizaki, T., Satoh, H., Ono, T., Kawahara, T., Morishita, T., Tamakawa, H., Yamagami, K., Inui, J., Maekata, M., and Narumiya, S. Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature (Lond.)*, *389*: 990–994, 1997.
35. Ishizaki, T., Uehata, M., Tamechika, I., Keel, J., Nonomura, K., Maekawa, M., and Narumiya, S. Pharmacological properties of Y-27632, a specific inhibitor of rho-associated kinases. *Mol. Pharmacol.*, *57*: 976–983, 2000.
36. Moscow, J. A., He, R., Gnarr, J. R., Knutsen, T., Weng, Y., Zhao, W. P., Whang, P. J., Linehan, W. M., and Cowan, K. H. Examination of human tumors for rhoA mutations. *Oncogene*, *9*: 189–194, 1994.
37. Rihet, S., Vielh, P., Camonis, J., Goud, B., Chevillard, S., and de Gunzburg, J. Mutation status of genes encoding RhoA, Rac1, and Cdc42 GTPases in a panel of invasive human colorectal and breast tumors. *J. Cancer Res. Clin. Oncol.*, *127*: 733–738, 2001.
38. Watanabe, N., Madaule, P., Reid, T., Ishizaki, T., Watanabe, G., Kakizuka, A., Saito, Y., Nakao, K., Jockusch, B. M., and Narumiya, S. p140mDia, a mammalian homolog of *Drosophila* diaphanous, is target protein for Rho small GTPase and is a ligand for profilin. *EMBO. J.*, *16*: 3044–3056, 1997.
39. Watanabe, N., Kato, T., Fujita, A., Ishizaki, T., and Narumiya, S. Cooperation between mDia1 and ROCK in Rho-induced actin reorganization. *Nat. Cell Biol.*, *1*: 136–143, 1999.
40. Banyard, J., Anand-Apte, B., Symons, M., and Zetter, B. R. Motility and invasion are differently modulated by Rho family GTPases. *Oncogene*, *19*: 580–591, 2000.

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