

## SUPPLEMENTARY FIGURE LEGENDS

**Supplementary Figure S1. Signaling properties of WT PAPSS1.** Immunoblotting of lysates from 293H cells transfected with empty vector (1µg) or plasmids encoding BRAF V600E-FLAG (80ng) or WT PAPSS1-FLAG (1µg) demonstrates that WT PAPSS1 does not cause MAPK pathway signaling. kDa, kilodalton.

**Supplementary Figure S2. Detection of TRIM24-BRAF.** Four representative spanning sequence reads from genotyping of a second “pan-negative” melanoma with the FoundationOne™ assay shows the alignment of *TRIM24* (purple text) to chromosome 7q32-24 and of *BRAF* (dark blue text) to chromosome 7q34. The reads specifically align to intron 9 of *TRIM24* and to intron 8 of *BRAF*, which indicates a coding sequence break between *TRIM24* exon 9 and *BRAF* exon 9. Below are schematics of wild-type BRAF (blue), wild-type TRIM24 (purple), and the fused TRIM24-BRAF proteins. The RING-finger, B-box types 1 and 2, and coiled-coil domains of TRIM24 and the serine/threonine (S/T) kinase domain of BRAF remain intact in the fused protein. WT, wild-type; ex, exon; RBD, Ras-binding domain; CRD, cysteine-rich domain; RKTR, Arg-Lys-Thr-Arg dimerization domain; RING, Really Interesting New Gene, RING-finger domains play key roles in the ubiquitylation pathway; B-box types 1 & 2 are zinc-finger-like domains; PHD, Plant Homeo Domain, binds tri-methylated lysines on histone H3; BROMO, bromodomain, binds acetylated lysines on histones.

**Supplementary Figure S3. Signaling and inhibition of TRIM24-BRAF.** Immunoblotting of lysates from 293H cells transfected with BRAF V600E-FLAG or TRIM24-BRAF-FLAG show that TRIM24-BRAF activates the MAPK pathway, which can be inhibited by the MEK inhibitor, trametinib (tra), but not the BRAF inhibitor, vemurafenib (vem). Drug dosing: (DMSO vehicle control), 0.1, 0.5, 1 and 5 µmol/L. kDa, kilodalton.

**Supplementary Figure S4. Potential TAX1BP1-BRAF and CDC27-BRAF fusions identified in data from The Cancer Genome Atlas.** Representative RNA sequencing reads from TCGA skin cutaneous melanoma dataset (SKCM) demonstrate the alignment of paired-end reads mapping to **(A)** exon 11 of BRAF (blue text) and exon 8 of TAX1BP1 (green text), both on chromosome 7 or **(B)** to exon 10 of BRAF (blue text) on chromosome 7 and exon 16 of CDC27 (orange text) on chromosome 17. Below each alignment schematic are diagrams of WT BRAF (blue) and the TAX1BP1-BRAF **(A)** or CDC27-BRAF **(B)** fusion proteins. WT, wild-type; ex, exon; RBD, Ras-binding domain; CRD, cysteine-rich domain; RKTR, Arg-Lys-Thr-Arg dimerization domain.

**Supplementary Figure S5. Comparison of reverse phase protein array (RPPA) data in TCGA skin cutaneous melanoma dataset.** Both graphs display box and whisker plots based on RPPA data distributions for **(A)** total or **(B)** phosphorylated MEK1/2 protein levels. Samples are divided based on SNaPshot mutation status into “pan-negative” (NEG), BRAF-mutant (includes the non-V600 BRAF alterations listed in Table S1 as well as BRAF N581T, S467L and G466E), NRAS-mutant, KIT-mutant, or GNAQ/11-status. The last two data points in each graph represent the “pan-negative” samples harboring CDC27-BRAF and TAX1BP1-BRAF. Differences in **(A)** total MEK1/2 protein levels are not statistically significant between any of the sample groups, however, **(B)** the phosphorylated MEK1/2 levels between “pan-negative” and BRAF-mutant or NRAS-mutant samples are statistically significant ( $p < 0.0001$ , Student’s t-test). Although statistical analyses cannot be performed on single samples, the phosphorylated MEK1/2 levels of the “pan-negative” cases harboring CDC27-BRAF and TAX1BP1-BRAF are similar to, or greater than, the levels observed in BRAF- or NRAS-mutant cases.